

Principles of Phylogenetics

Reading and Inferring Trees

Finlay Maguire

April 1, 2020

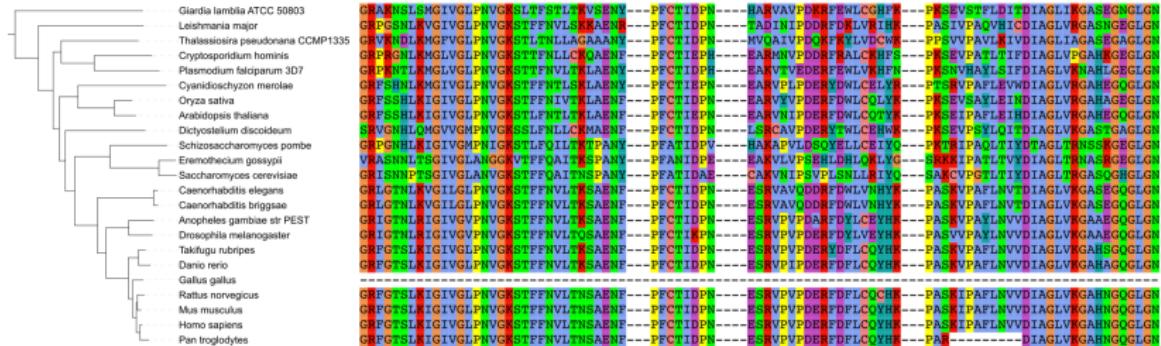
FCS, Dalhousie

Table of contents

1. What are phylogenies?
2. Reading a Tree
3. Making a Tree
4. Tree Inference methods
5. Aside: sources of error
6. Back to inference
7. Conclusion

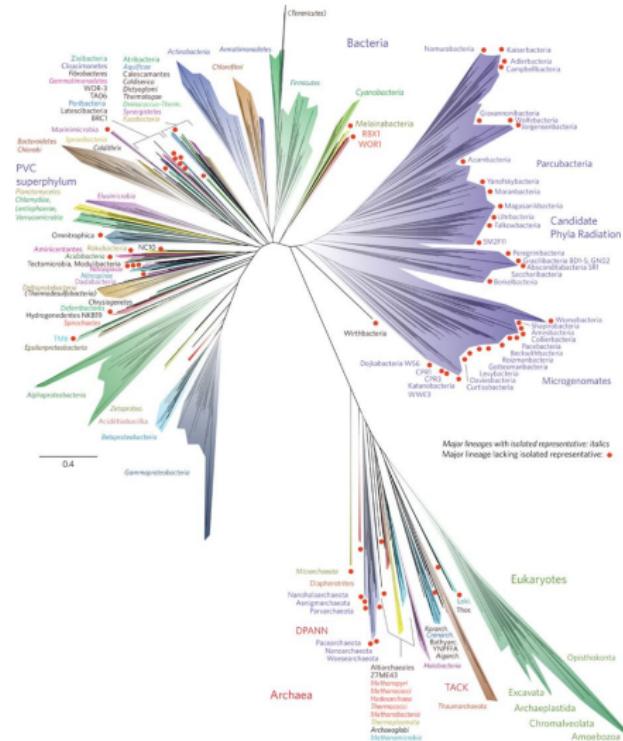
What are phylogenies?

Hypotheses for understanding alignments



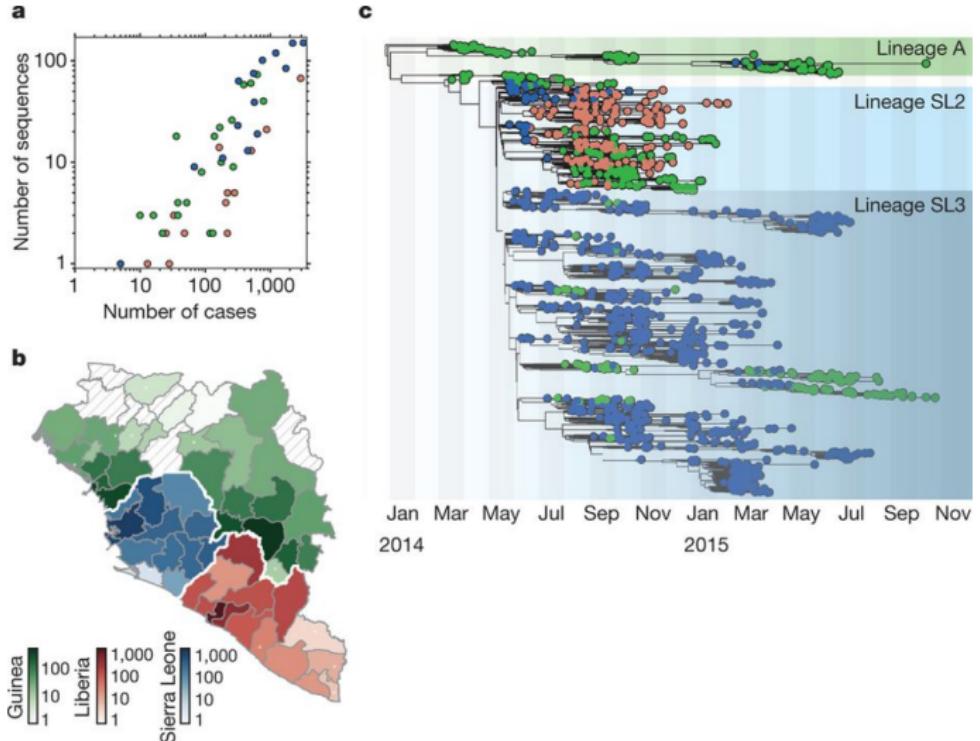
<https://itol.embl.de/help.cgi>

Tree of Life



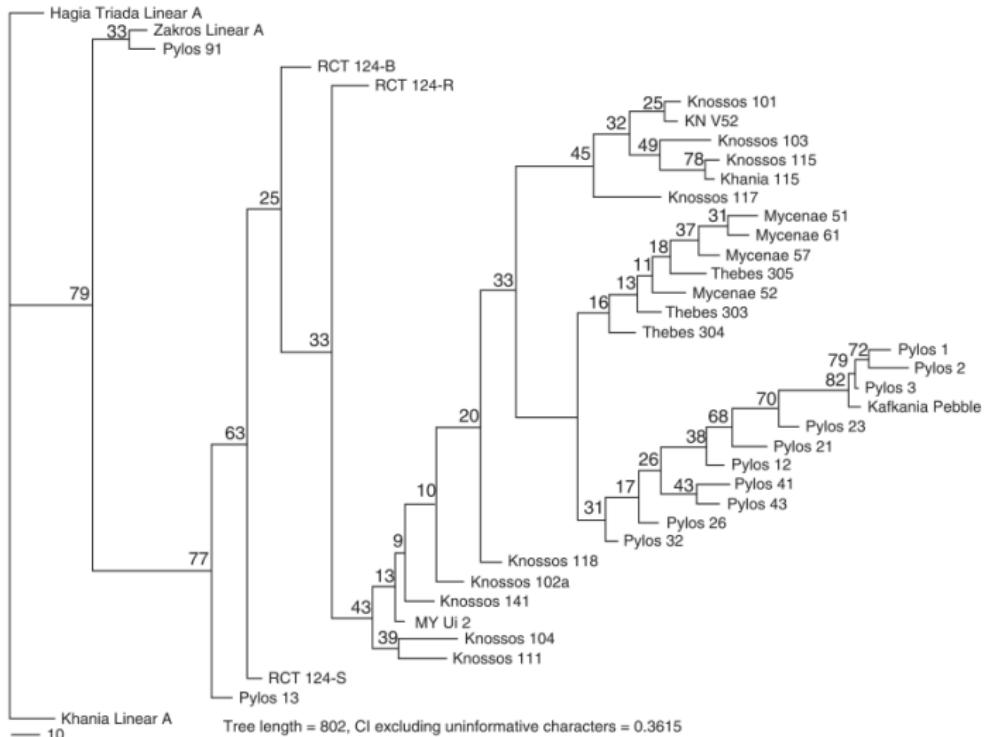
[Hug et al., 2016]

2013- Ebola Outbreak



[Holmes et al., 2016]

Uses outside biology



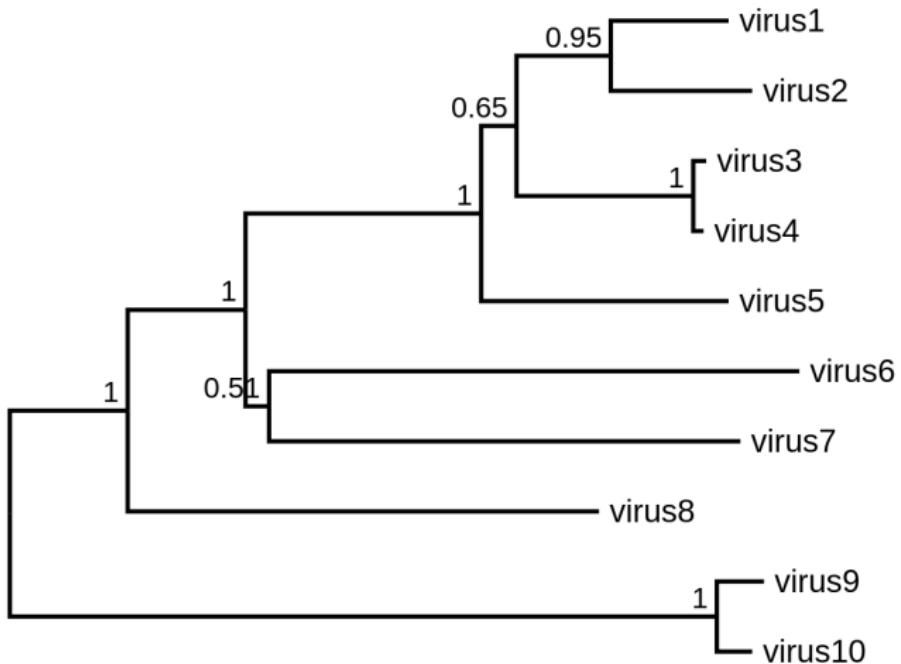
[Skelton, 2008]

Uses outside biology

- Manuscript change [Barbrook et al., 1998]
- Social evolution (many examples, some questionable).
- Plagiarism [Ryu et al., 2008]
- Anything you can measure distances between.

Reading a Tree

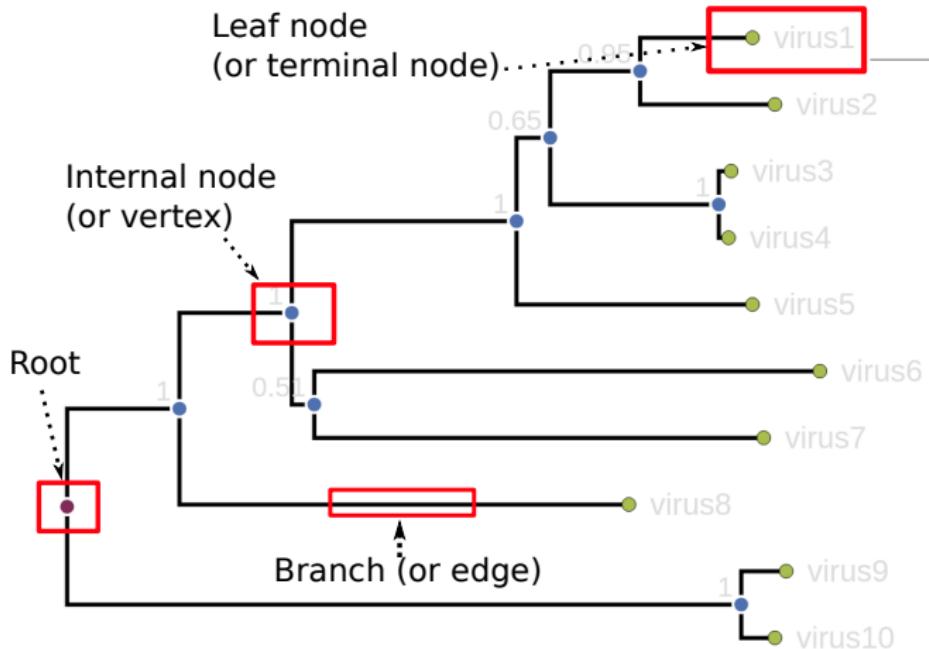
Toy Tree



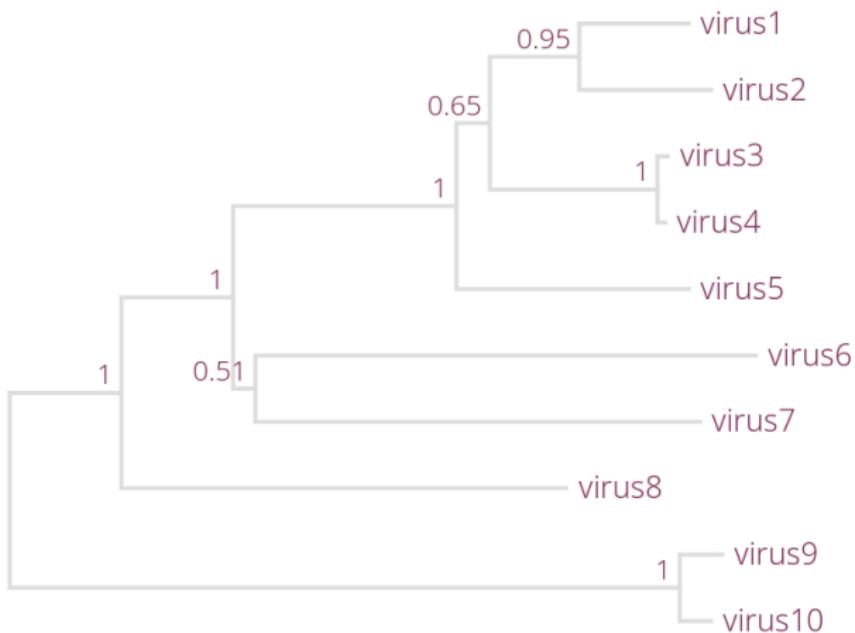
Andrew Rambaut's Tutorial

<http://artic.network/how-to-read-a-tree.html>

Parts of the Tree

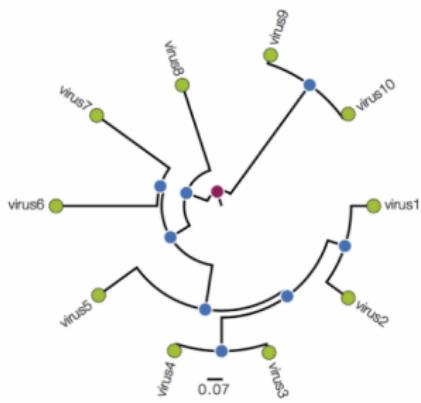


Support Values

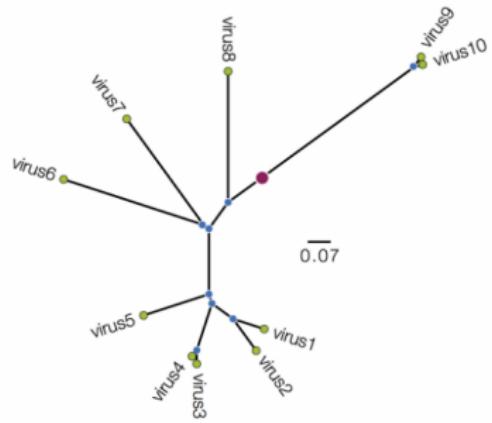


Other formats

A:



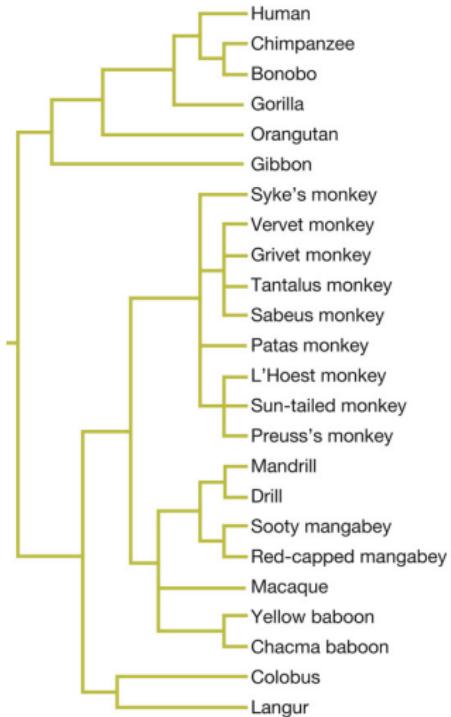
B:



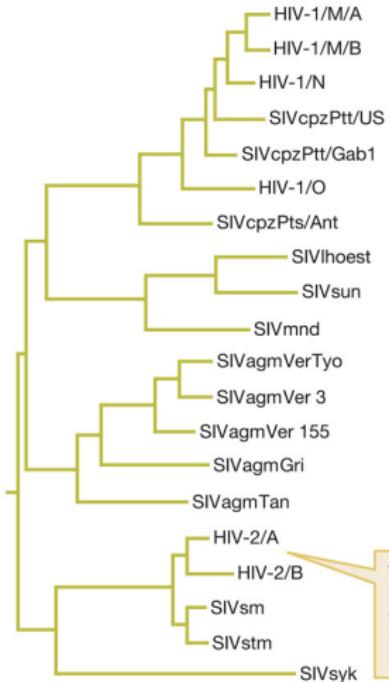
<http://artic.network/how-to-read-a-tree.html>

Meaningful Branch Lengths

A

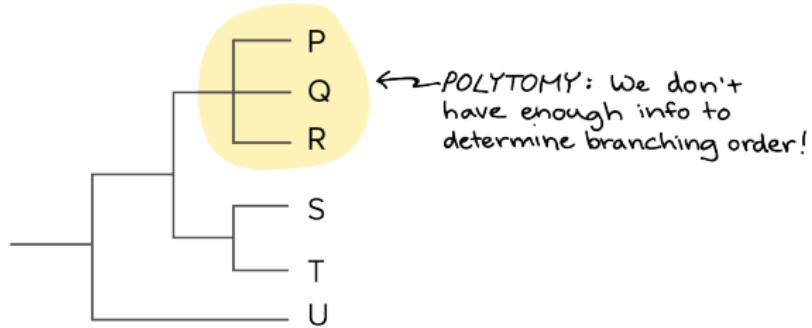


B



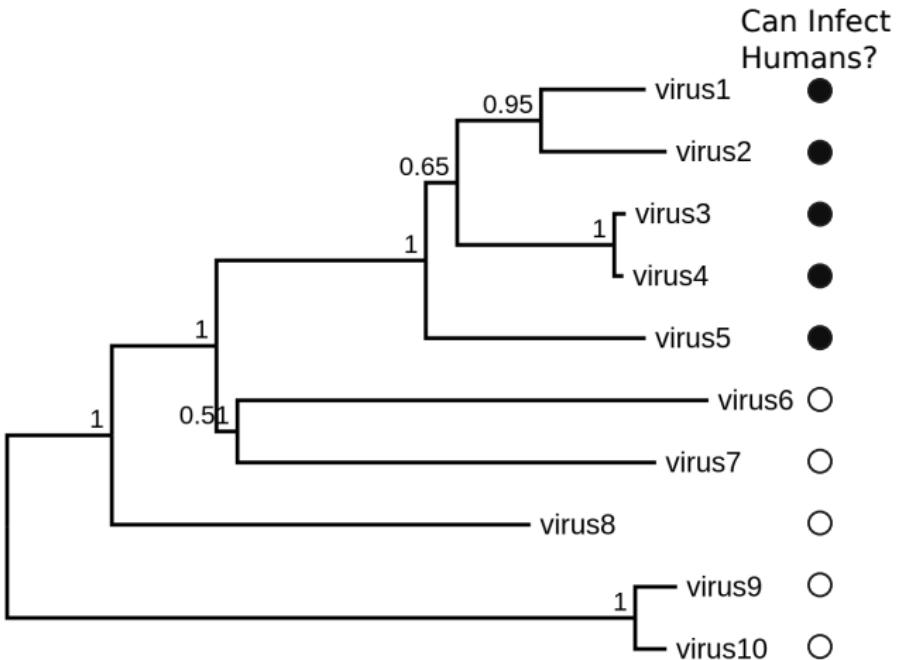
The longer branch leading to HIV-2/B indicates that more change has occurred on that branch than on the branch leading to HIV-2/A

https://biology-forums.com/gallery/18099_27_04_12_2_16_20.jpeg

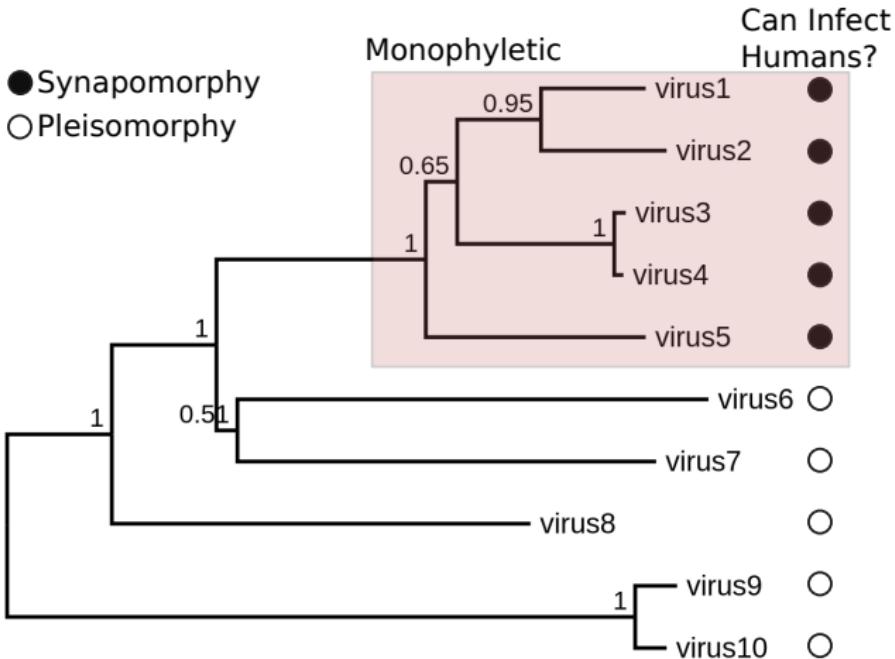


Khan Academy

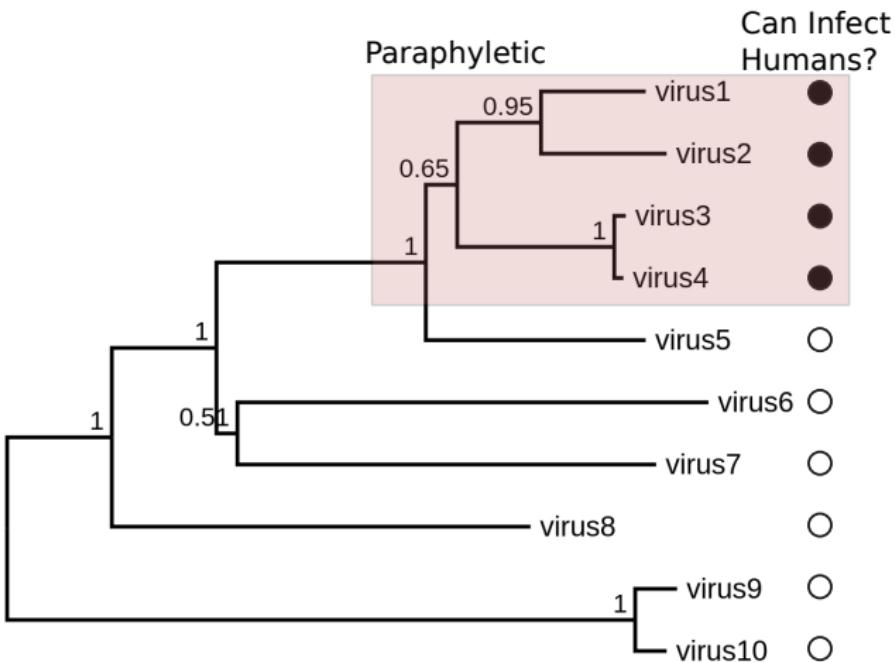
Groupings on the Tree



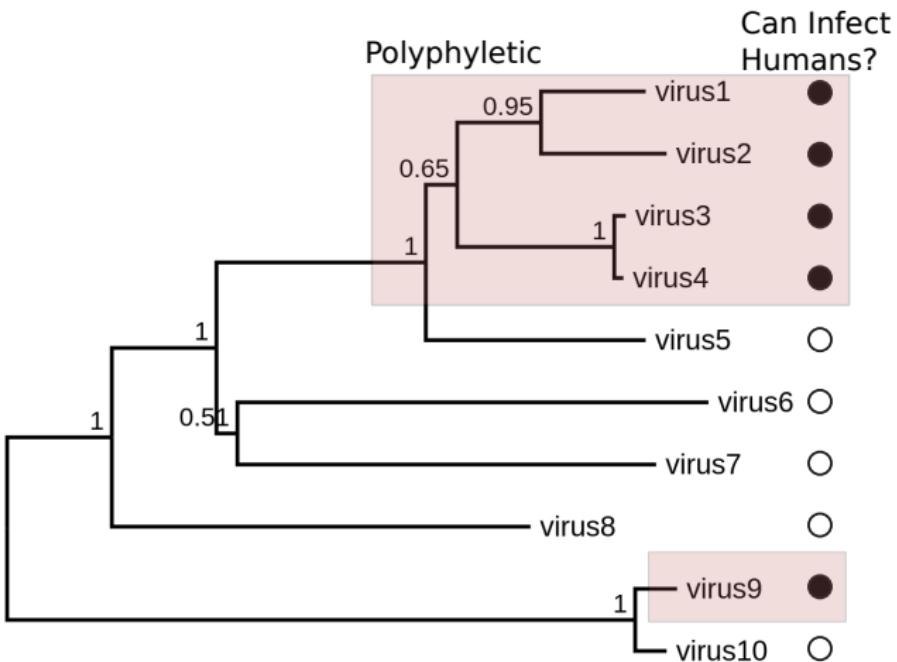
Monophyletic



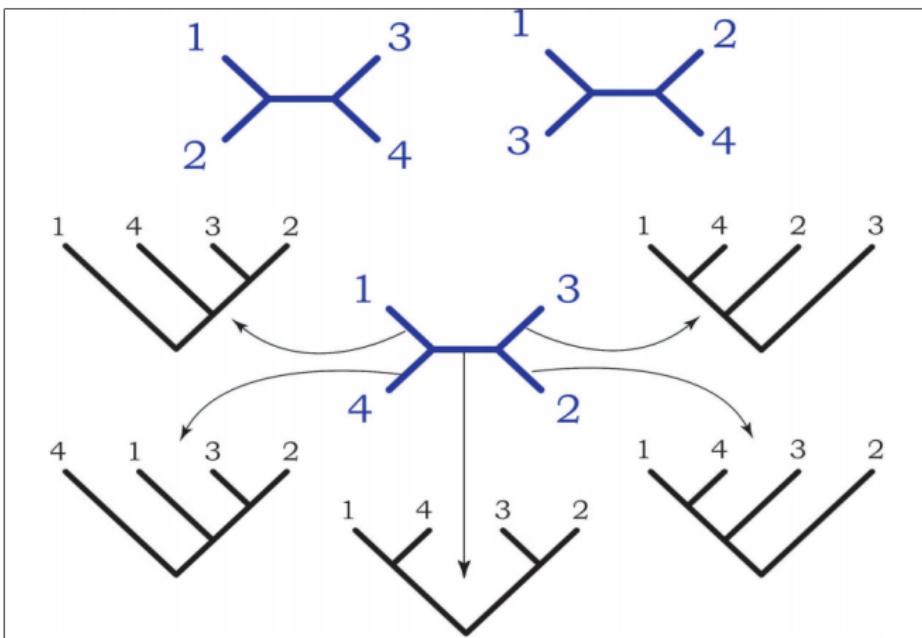
Paraphyletic



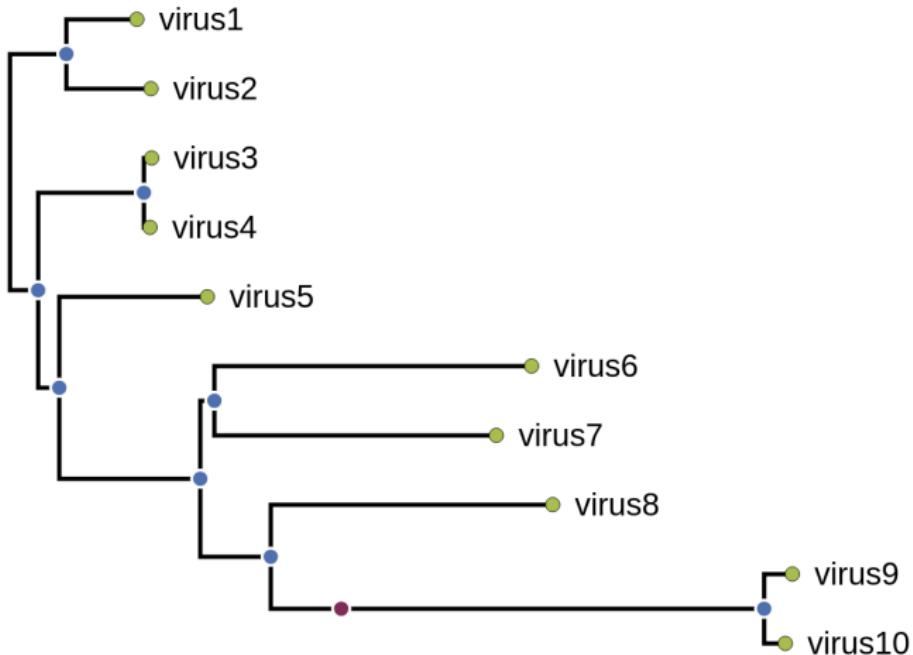
Polyphyletic



Rooting

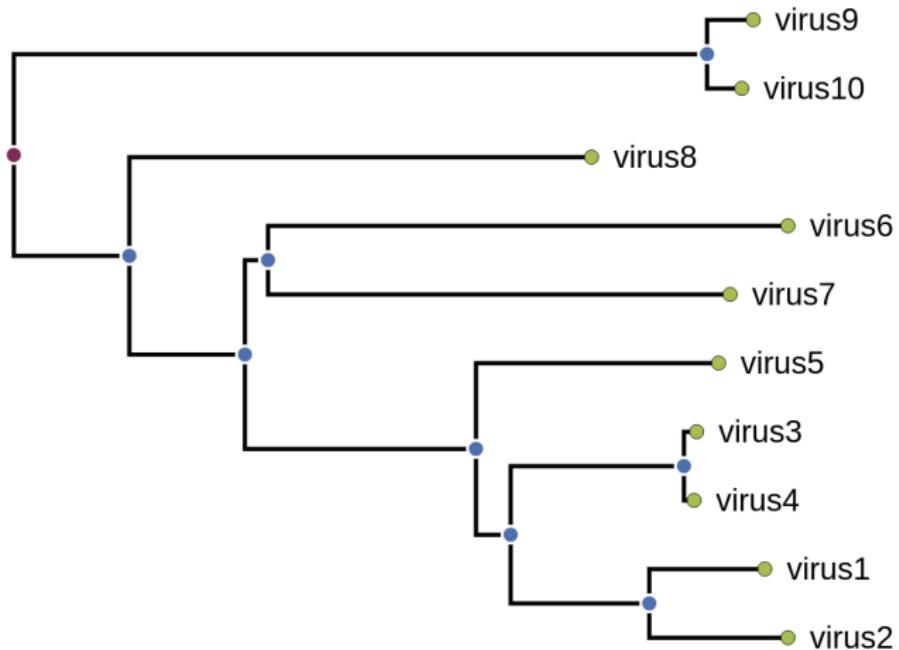


Rooting



<http://artic.network/how-to-read-a-tree.html>

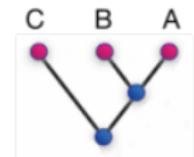
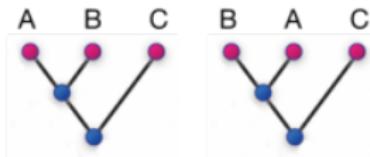
Rooting



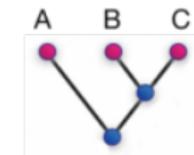
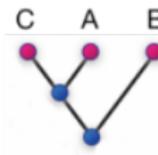
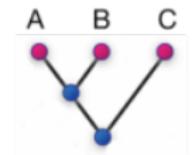
<http://artic.network/how-to-read-a-tree.html>

Topology and rotation

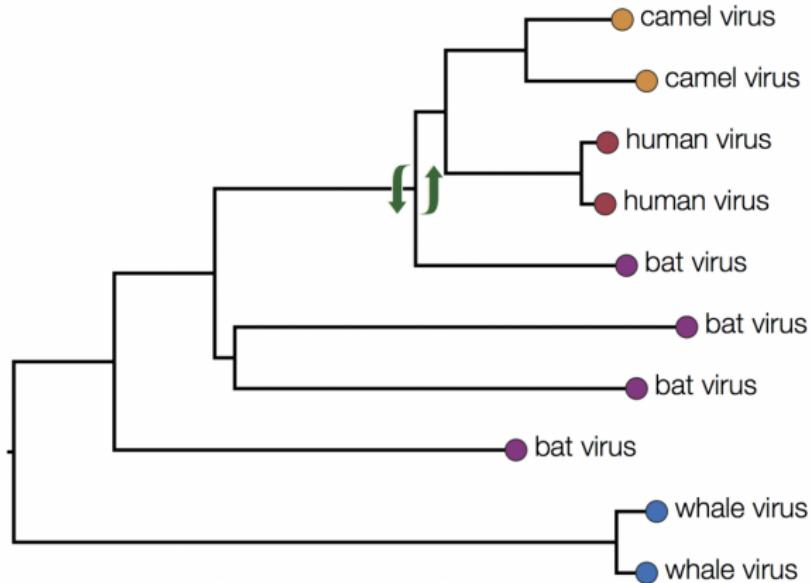
These trees display the same topology



These trees display different topologies

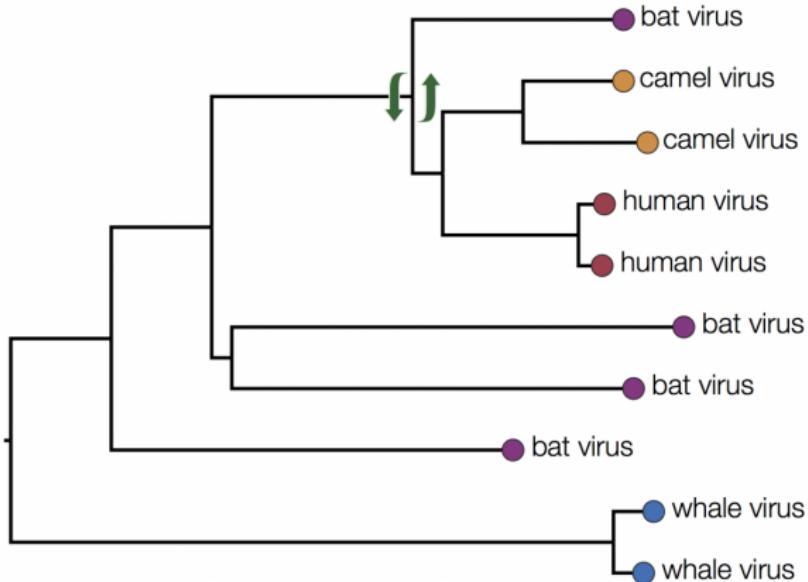


Nodes can rotate

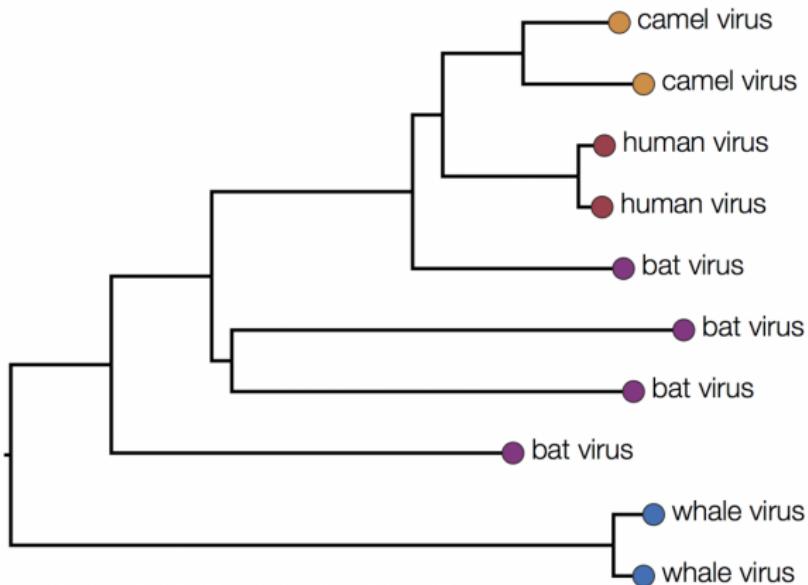


<http://artic.network/how-to-read-a-tree.html>

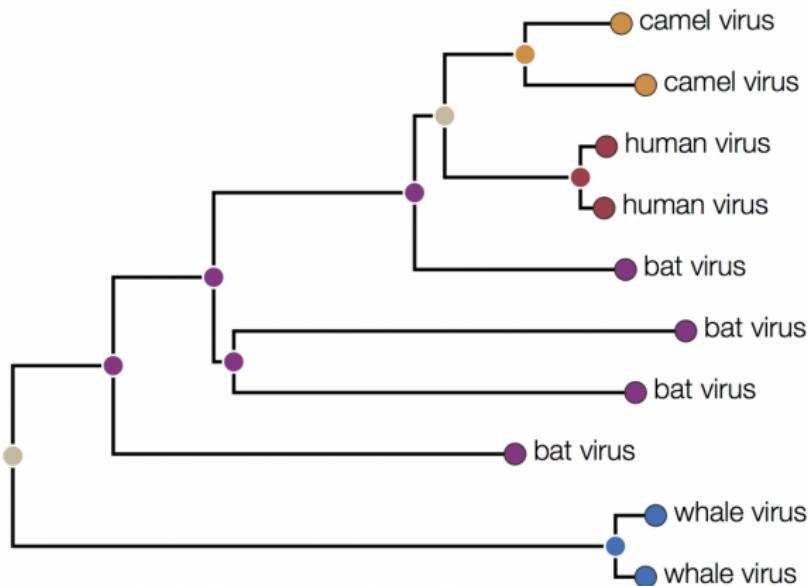
Nodes can rotate



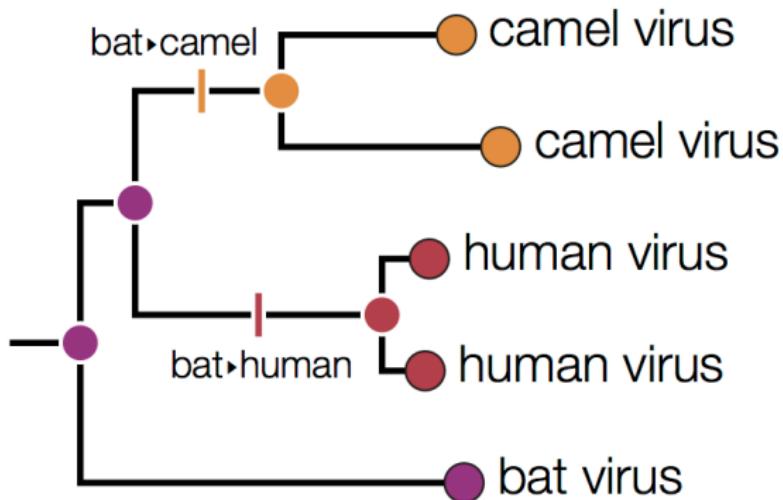
Adding Metadata



Ancestral Node Reconstruction

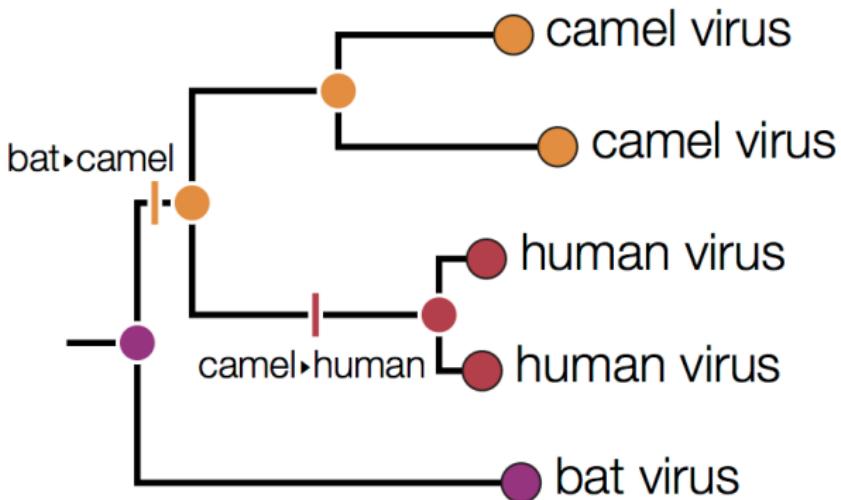


Ancestral Node Reconstruction



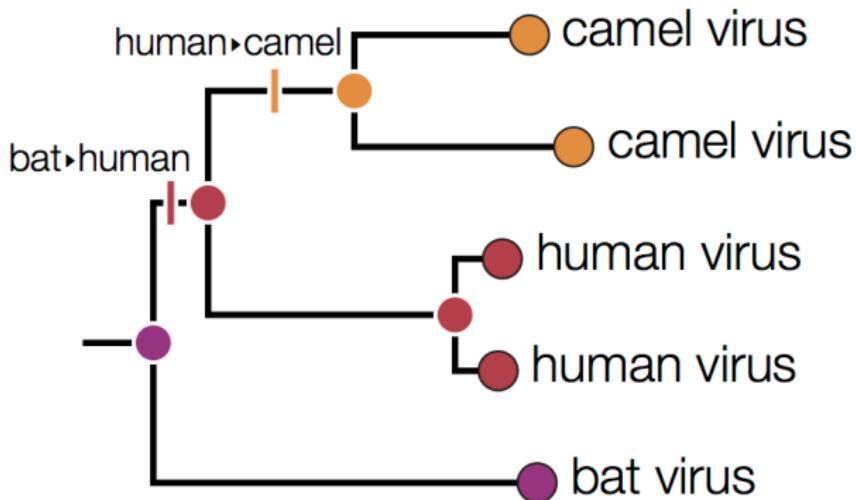
<http://artic.network/how-to-read-a-tree.html>

Ancestral Node Reconstruction



<http://artic.network/how-to-read-a-tree.html>

Ancestral Node Reconstruction



<http://artic.network/how-to-read-a-tree.html>

Making a Tree

Going from data to a tree

- Getting your data
- Aligning your data
- Tree-inference
 - Maximum Parsimony
 - Distance Methods
 - Maximum-Likelihood
 - Bayesian
- Sequence evolution models
- Exploring topology space
- Statistical support

Getting and preparing your data

Finding Similar Sequences

NIH U.S. National Library of Medicine NCBI National Center for Biotechnology Information

BLAST® » blastp suite

Standard Protein BLAST

[blastn](#) [blastp](#) [tblastn](#) [tblastx](#)

Enter Query Sequence

Enter accession number(s), g(i)s, or FASTA sequence(s) [?](#) [Clear](#)

>gb|CAZ39946.1|-NDM-1 [Klebsiella pneumoniae]
MELPNIDM-PVAKLSTALAAALMLSGDMGEIRPITQQQMETGDDRFGLVFRQLAPNWQHTSYLDMPGF
GAVASNLIVRDRGGRLVVDTANTDQQTAQILWNTIKQEENLNVPAVALAVVTHAQDKMGMALHAAGIATY
ANALSNQLARQEGRVAQQISLTFAANGIVEPATAPNFGPLKVFYPRPGHTSDNITYVGIDGTIAFGGCLL
KDSKAKSLSNLGAQDTEHYAASARAFGAFFPKASMIVMSHSAPSRAITHTARMADKL

Query subrange [?](#)

From
To

Or, upload file [Browse...](#) No file selected. [?](#)

Job Title gb|CAZ39946.1|-NDM-1 [Klebsiella pneumoniae]

Enter a descriptive title for your BLAST search [?](#)

BLAST

New Delhi metallo-beta-lactamase 1 [Acinetobacter baumannii]

Sequence ID: [BBA83870.1](#) Length: 261 Number of Matches: 1

Range 1: 3 to 260			GenPept	Graphics	▼ Next Match	▲ Previous Match
Score	Expect	Method	Identities	Positives	Gaps	
498 bits(1282)	2e-177	Compositional matrix adjust.	256/265(97%)	257/265(96%)	7/265(2%)	
Query 4	PNIMHPVAKLSTALAAALMLS GCMPEIRPTIGQOMETGDQRFGDLVFRQLAPNVWQHTS					63
Sbjct 3	PNIMHPVAKLSTALAAALMLS GCMPEIRPTIGQOMETGDQRFGDLVFRQLAPNVWQHTS					62
Query 64	YLDMPGFFGAVASNGLIVRDGGRVLVVDTAWTDDQTAQILNWIKEINLPVALAVVTHAHQ					123
Sbjct 63	YLDMPG GAVASNGLIVRDGGRV+V AWTDQTAQILNWIKEINLPVALAVVTHAHQ					119
Query 124	DKMGGMDALHAAGIATYANALSNQLAPQEGMVAQHSLTFAANGWVEPATAPNFGPLKVF					183
Sbjct 120	DKMGGMDALHAAGIATYANALSNQLAP EGMVAAQHSLTFAANGWVEPATAPNFGPLKVF					178
Query 184	YPGPGBTSDNITVGIDGTDIAFGGCLIKDSKA KSLGNLGDADTEHYAASARAFGAAPKA					243
Sbjct 179	YPGPGBT DNITVGIDGTDIAFGGCLIKDSKA KSLGNLGDADTEHYAASARAFGA -PKA					236
Query 244	SMIVMSHSAPDSRAAITHTARMADK	268				
Sbjct 237	SMIVMSH APDSRAAITHTARMADK	260				

BLAST® » blastp suite » RID-7URVN5C201N

[Edit and Resubmit](#) [Save Search Strategies](#) [Formatting options](#) [Download](#)

Job title: gb|CAZ39946.1|-NDM-1 [Klebsiella pneumoniae]

RID: [7URVN5C201N](#) (Expires on 03-06 00:25 am)

Query ID: lcl|Query_262517

Description: gb|CAZ39946.1|-NDM-1 [Klebsiella pneumoniae]

Molecular weight: amino acid

Query Length: 270

Database Name: nr

Description: All non-redundant GenBank CDS translations+PDB

Program: BLASTP 2.8.1+ [Help](#)

Other reports: [Search Summary](#) [Taxonomy report](#) [Distance tree of results](#) [Related Structures](#) [Multiple alignment](#) [MSA viewed]

Graphic Summary

[Show Conserved Domains](#)

Putative conserved domains have been detected, click on the image below for detailed results.

[CD search result summary](#)

Distribution of the top 100 Blast Hits on 100 subject sequences [?](#)

Mouse over to see the title, click to show alignments

Color key for alignment scores:

- <40
- 40-50
- 50-80
- 80-200
- >=200

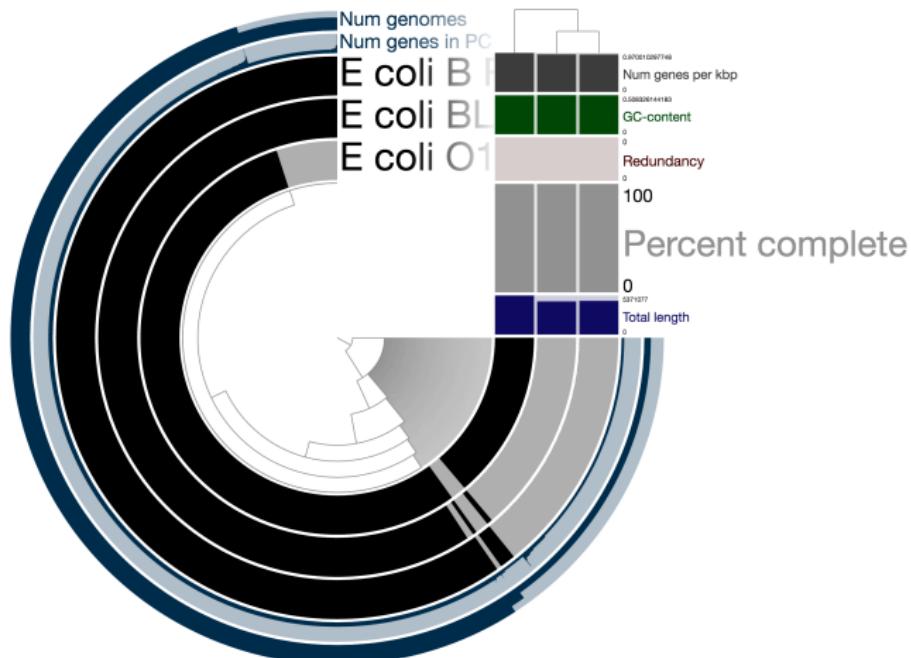
Query

1 50 100 150 200 250

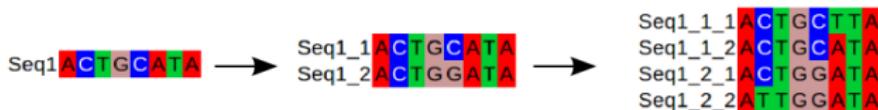
Core Genome Inference

Pangenome of three *E. coli*'s

Tree order: Tree (D: Unknown; L: Unknown) | Current view: single | Sample order: protein_clusters



Multiple Sequence Alignment



<https://bioinf.comav.upv.es/courses/biotech3/theory/multiple.html>

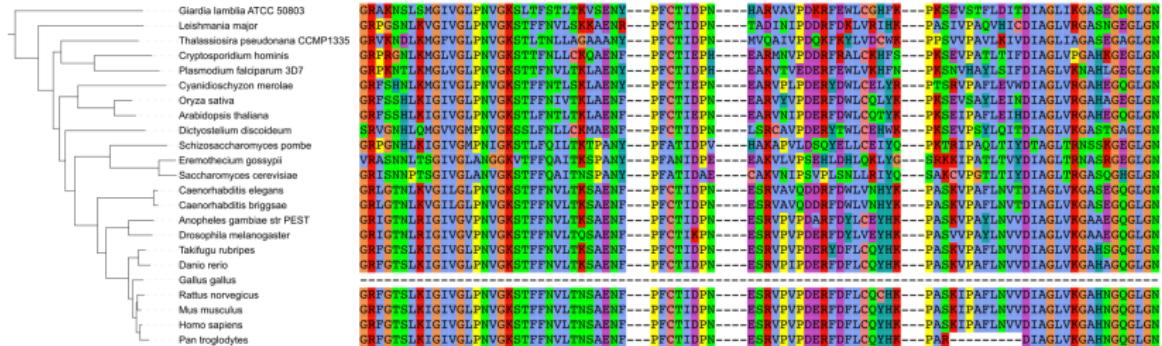
Multiple Sequence Alignment

A multiple sequence alignment of 12 protein sequences. The sequences are color-coded by residue type: A (green), C (blue), D (red), E (purple), F (yellow), G (light green), H (orange), I (pink), K (brown), M (grey), P (cyan), R (dark blue), S (teal), T (light blue), V (light pink), W (light orange), and Y (light purple). The alignment shows a highly conserved motif across all sequences, indicated by identical colors at each position. The motif consists of: Position 1: Y (light purple), Position 2: K (brown), Position 3: V (light pink), Position 4: V (light pink), Position 5: P (cyan), Position 6: D (red), Position 7: D (red), Position 8: V (light pink), Position 9: V (light pink), and Position 10: G (light green).

SYKVKLITPDGPIEFDCPDDVYILDQAEAAAGHDLPYSCRAGSCSSCAGKIAAGGAVDQTDGNFLLDD	SYKVKLITPDGPIEFDCPDDVYILDQAEAAAGHDLPYSCRAGSCSSCAGKIAAGGAVDQTDGNFLLDD	SYKVKLITPDGPIEFDCPDDVYILDQAEAAEGHDLPYSCRAGSCSSCAGKVTAGSVVDOSDGNFLDDE	SYKVKLITPDGPIEFDCPDDVYILDQAEAAEGHDLPYSCRAGSCSSCAGKVTAGTVDQSDGKFLLDD	SYKVKLVTDPGTOEFCPSDVYILDHAEEEVGIDL PYSCRAGSCSSCAGKVVGGEVDOSDGSFLDD	TYKVKLITPEGPQEFDCCPDDVYILDAAEEAGIDL PYSCRAGSCSSCAGKVNNGNVNQEDGSFLDD	AYKVTLVTPEGKQELCPCDDVYILDAAEEAGIDL PYSCRAGSCSSCAGKVTSGSVNODDGSFLDD	AYKVTLVTPGTGNVEFOCPDDVYILDAAEEEGIDL PYSCRAGSCSSCAGKLKTGSLNQDDOSFLDD	TYKVKFITPEGEQEVECDDDVYVLDAAEEAGIDL PYSCRAGSCSSCAGKVVSGSVDOSDOSFLDD	TYKVKFITPEGELEVECDDDVYVLDAAEEAGIDL PYSCRAGSCSSCAGKVVSGSVDQSDOSFLDD	TYKVKFITPEGEQEVECDDDVYVLDAAEEAGIDL PYSCRAGSCSSCAGKVVSFGFVDQSDOSFLDD	TYKVKFITPEGEQEVECEEDDVYVLDAAEEAGLDL PYSCRAGSCSSCAGKVVSGSIDOSDOSFLDD	TYKVKFITPEGEQEVECEEDDVYVLDAAEEAGIDL PYSCRAGSCSSCAGKVVSGSIDOSDOSFLDD	TYNVKLITPEGEVELQVPPDVYILDQAEEDGIDL PYSCRAGSCSSCAGKVVSGSVVDQSDOSYLLDD
--	--	--	---	--	---	--	---	--	--	---	---	---	--

<https://bioinf.comav.upv.es/courses/biotech3/theory/multiple.html>

Alignment Trimming



<https://itol.embl.de/help.cgi>

Trimmed Alignment

SYKVKLITPDGPIEFDCPDDVYILDQAEAAAGHDLPYSCRAGSCSSCAGKIAAGGAVDQTDGNFLLDD
SYKVKLITPDGPIEFDCPDDVYILDQAEAAAGHDLPYSCRAGSCSSCAGKIAAGGAVDQTDGNFLLDD
SYKVKLITPEGPIEFECPPDDVYILDQAEAAEGHDLPYSCRAGSCSSCAGKVTAGSVVDOSDGNFLDDE
SYKVKLITPDGPIEFECPPDDVYILDQAEAAEGHDLPYSCRAGSCSSCAGKVTAGTVDQSDGKFLLDD
SYKVKLVTPDGTOEFCPSDVYILDHAEEEVGIDL PYSCRAGSCSSCAGKVVVGGEVDDOSDGSFLDDE
TYKVKLITPEGPOEFDCPDDVYILDHAEEVGIDL PYSCRAGSCSSCAGKVNNGNVNQEDGSFLDDE
AYKVTLVTPEGKQELCPCDDVYILDAAEEAGIDL PYSCRAGSCSSCAGKVTSGSVN0DDGSFLDDE
AYKVTLVTPGTGNVEFOCPDDVYILDAAEEEGIDL PYSCRAGSCSSCAGKLKTGSLNQDDOSFLDDE
TYKVKFITPEGEQEVECDDDVYVLDAAEEAGIDL PYSCRAGSCSSCAGKVVSGSVDOSDOSFLDDE
TYKVKFITPEGELEVECDDDVYVLDAAEEAGIDL PYSCRAGSCSSCAGKVVSGSVDQSDOSFLDDE
TYKVKFITPEGEQEVECEEDDVYVLDAAEEAGIDL PYSCRAGSCSSCAGKVVSGFVDQSDOSFLDDE
TYKVKFITPEGEQEVECEEDDVYVLDAAEEAGIDL PYSCRAGSCSSCAGKVVSGSIDOSDOSFLDDE
TYNVKLITPEGEVELQVPPDVYILDQAEEDGIDL PYSCRAGSCSSCAGKVVSGSVDQSDOSYLLDD

<https://bioinf.comav.upv.es/courses/biotech3/theory/multiple.html>

Tree Inference methods

Difficulties

- Huge number of possible trees
- unrooted = $\frac{2n-5!}{2^{n-3}(n-3)!}$
- rooted = $\frac{2n-3!}{2^{n-2}(n-2)!}$
- 10 taxa = 2,027,025 unrooted and 34,459,425 rooted topologies.
- 50 taxa = 2.84e74 unrooted and 2.75e76 rooted topologies.
- Topology space geometry is large and awkward to traverse.
- Large number of parameters to optimise
- How do you choose which tree is optimal? Which criterion?

Parsimony

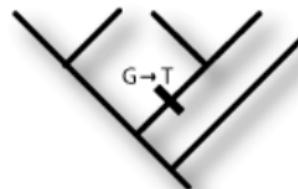
Using Maximum Parsimony to Choose Between Two Possible Trees

Sample:

1 2 3 4 5

Observation:

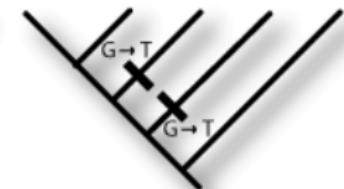
G G T T G



1 change required
→ better tree

1 2 3 4 5

G G T T G



2 changes required
→ poorer tree

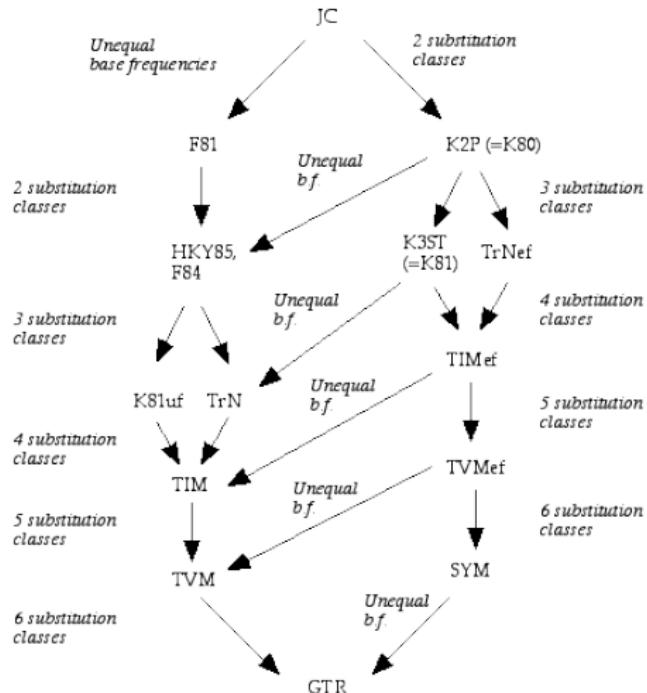
Intuitive: minimise the number of changes needed.

Parsimony Pros/Cons

- Advantages:
 - Very simple
 - Works on any type of data (no explicit model).
- Disadvantages:
 - Very simple
 - Requires informative sites with consistent signal.
 - Poor handling of multiple substitutions.
 - Can't incorporate extra information.
 - Not consistent for certain tree shapes (misleading support values).

Evolutionary Models

Sequence Evolution Models



Sequence Evolution Models

	HIV-W _m	HIV-W _m +F	HIV-B _m	HIV-B _m +F	REV-1 step	JTT+F	JTT	WAG+F	MtMAM+F	rtREV	mtREV 24+F	WAG	Dayhoff+F	rtREV+F	Dayhoff	Equal Input	mtREV 24	mtMAM	REV
HIV-W _m	0	45	44	46	47	46	47	47	47	46	47	47	47	47	47	47	47	47	47
HIV-W _m +F	1	0	45	46	46	46	46	47	47	47	47	47	47	47	47	47	47	47	47
HIV-B _m	0	1	0	15	43	30	39	43	46	46	46	46	46	47	47	47	47	47	47
HIV-B _m +F	0	0	15	0	43	37	40	44	47	46	47	46	47	47	47	47	47	47	47
REV-1 step	0	1	4	4	0	6	6	11	31	32	22	14	17	24	28	35	41	43	47
JTT+F	0	0	8	5	40	0	28	47	46	46	47	47	47	47	47	47	47	47	47
JTT	0	0	3	3	38	4	0	35	44	46	45	47	47	46	47	47	47	47	47
WAG+F	0	0	3	1	34	0	5	0	43	44	43	39	42	46	47	47	47	47	47
MtMAM+F	0	0	0	0	16	0	0	2	0	14	2	6	4	7	12	31	47	47	46
rtREV	0	0	0	0	12	0	1	2	29	0	8	1	3	3	4	39	47	47	47
MtREV 24+F	0	0	0	0	18	0	1	1	41	37	0	7	7	22	25	47	47	47	47
WAG	0	0	0	1	29	0	0	2	40	45	35	0	30	39	43	46	47	47	47
Dayhoff+F	0	0	0	0	26	0	0	0	39	43	29	8	0	36	43	46	47	47	47
rtREV+F	0	0	0	0	19	0	0	0	35	41	20	2	1	0	20	46	47	47	47
Dayhoff	0	0	0	0	18	0	0	0	32	39	17	0	1	17	0	44	47	47	47
Equal Input	0	0	0	0	11	0	0	0	14	2	0	1	0	1	2	0	41	46	47
mtREV 24	0	0	0	0	4	0	0	0	0	0	0	0	0	0	0	4	0	43	45
mtMAM	0	0	0	0	4	0	0	0	0	0	0	0	0	0	0	0	1	0	44
REV	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	2	3	0

[†]Models are arranged by decreasing rank performance (see Table 2)

doi:10.1371/journal.pone.0000503.t003

[Nickle et al., 2007]

How do we select a model?

- Which model is most likely given the data?
- Information Criterion (regularisation to penalise overly complex models)
- Decision Theory: risk minimisation.

What happens theoretically if the wrong model is specified?

- Increased Inaccuracy (wrong tree more often)
- Inconsistency (adding more data converges to wrong tree)
- Wrong branch lengths (important for certain analyses)
- Wrong tree support values

What actually happens

[Abadi et al., 2019]

- Almost always use the most flexible model (GTR+I+G/LG)
- Criteria are inconsistent (BIC/AIC disagree in 62% of cases)
- Different models change the distance matrix trivially.
- ALL models lead to very similar topologies.
- Model only really important if branch length matters to you.

Distance Matrix

The diagram illustrates the construction of a distance matrix from a sequence alignment matrix. On the left, a yellow-highlighted block shows a sequence alignment matrix with four rows labeled 1, 2, 3, and 4, and three columns of sequence data. An arrow points from this matrix to a distance matrix on the right.

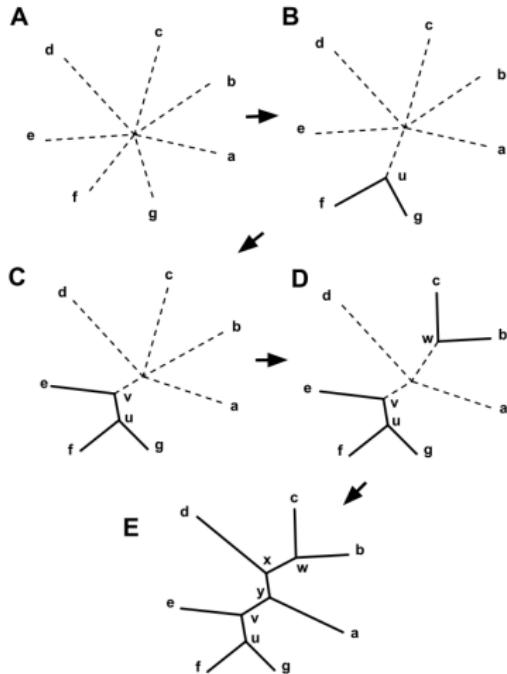
On the right, the distance matrix is represented as:

$$\begin{matrix} & \begin{matrix} 1 & 2 & \dots & n \end{matrix} \\ \begin{matrix} 1 \\ 2 \\ \vdots \\ n \end{matrix} & \begin{pmatrix} 0 & 0.3 & \dots & 0.5 \\ 0.3 & 0 & \dots & 0.3 \\ \vdots & \vdots & \ddots & \vdots \\ 0.5 & 0.4 & \dots & 0 \end{pmatrix} \end{matrix}$$

<https://slideplayer.com/slide/4422868/>

Neighbour-Joining

Iteratively pair off branches that minimise the total sum of branch lengths



Distance Approaches Pros/Cons

- Advantages:
 - Very fast (often used as starting point)
 - Works well for clock-like and closely related sequences
- Disadvantages:
 - Requires a sequence evolution model
 - Pairwise distance isn't always error-free estimate of evolutionary distance (bigger problem with divergent sequences).
 - Doesn't use all available information
 - Cannot reconstruct character histories

Aside: sources of error

Sources of Error

- Bad data

Sources of Error

- Bad data
- Sampling error

Sources of Error

- Bad data
- Sampling error
- Misleading evolutionary events

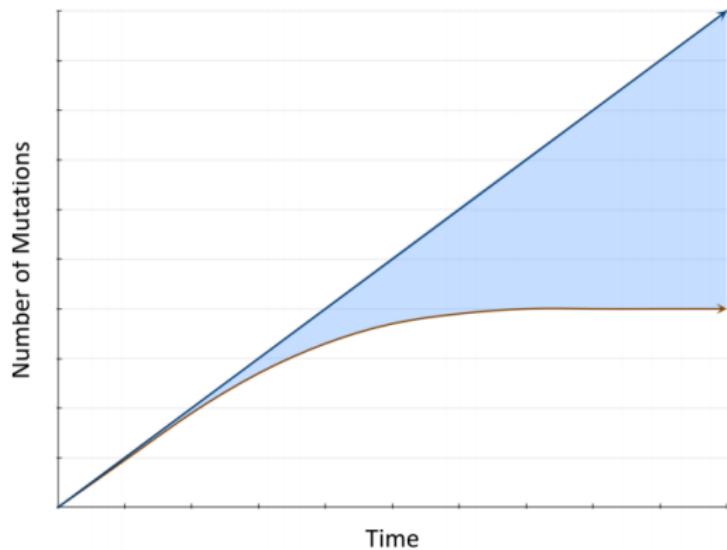
Sources of Error

- Bad data
- Sampling error
- Misleading evolutionary events
- Misspecified models

Sources of Error

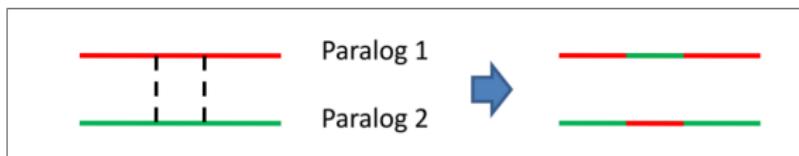
- Bad data
- Sampling error
- Misleading evolutionary events
- Misspecified models
- Inappropriate inference

Saturation

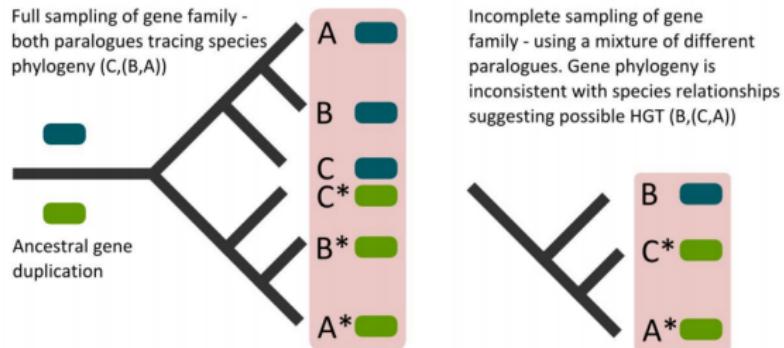


[Leonard, 2010]

Misleading Signal: Recombination

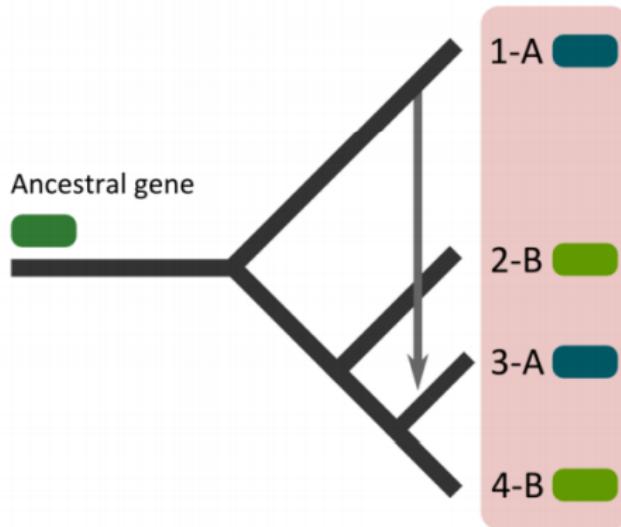


Misleading Signal: Hidden Paralogy/Incomplete Sampling



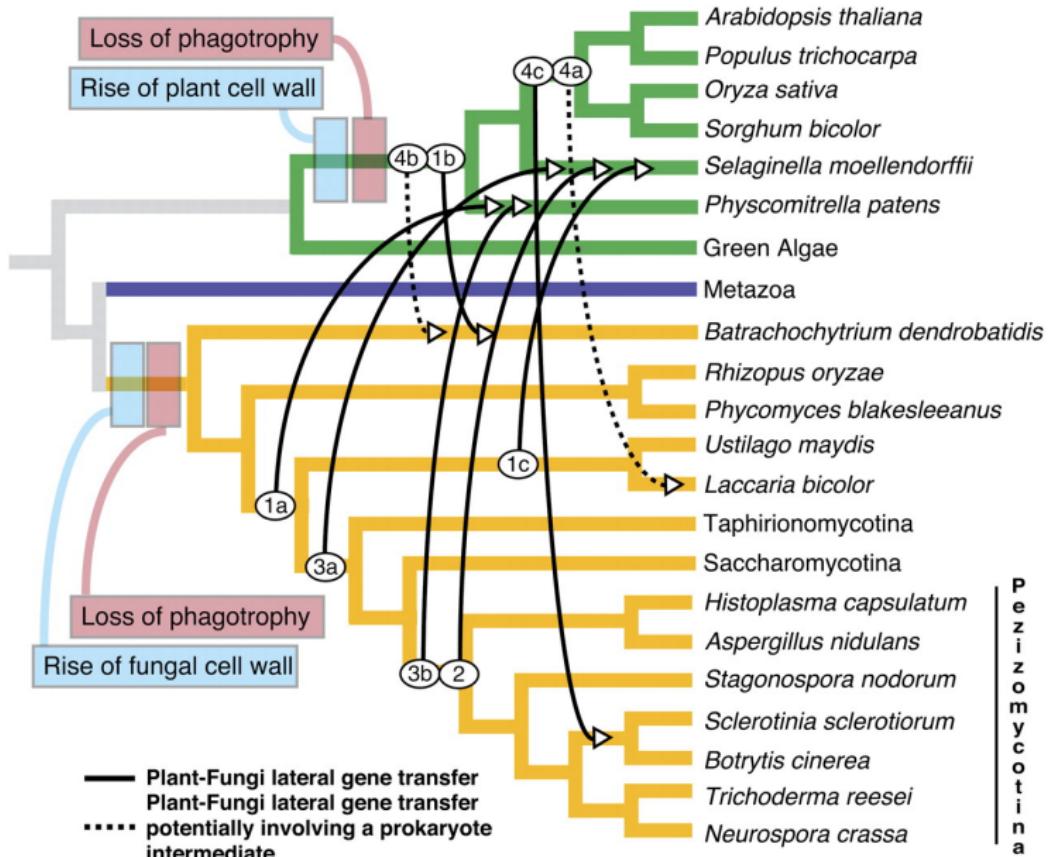
[Leonard, 2010]

Misleading Signal: Horizontal Gene Transfer



[Leonard, 2010]

Misleading Signal: Horizontal Gene Transfer

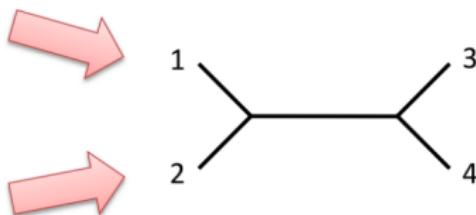


Tree not always correct paradigm

Ask for a tree get a tree.

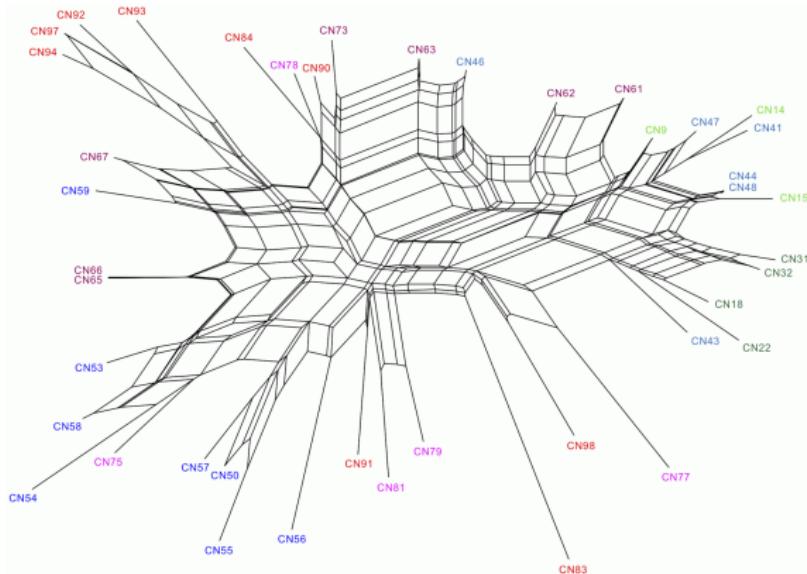
1 ACCGAGCAA
2 ACCGAGCAA
3 ACCGAGCAA
4 ACCGAGCAA

1 ACCGAATGA
2 ACCGAGCAG
3 GTTAGGCAG
4 GTTAGATGA



Tree not always correct paradigm

Ask for a tree get a tree.



Reanalysis of [Marwick, 2012] from
<http://phylonetworks.blogspot.ca/2013/02/>

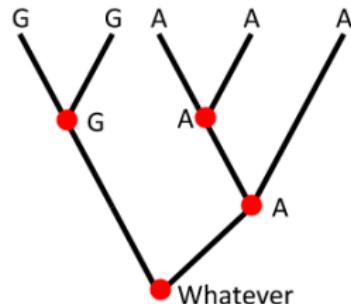
Back to inference

Maximum-Likelihood

- Likelihood = $p(\text{data} \mid \text{topology, branch, evolutionary model}) = p(D|\tau, \theta)$
- Maximum likelihood is the topology, branch lengths and model parameters with the highest likelihood.
- Performed site by site, search topology space then finding optimal tree parameters.
- Too expensive to exhaustively search likelihood surface so heuristics.
- Most methods start with distance-based starting tree and greedily traverse model space.

Maximum-Likelihood

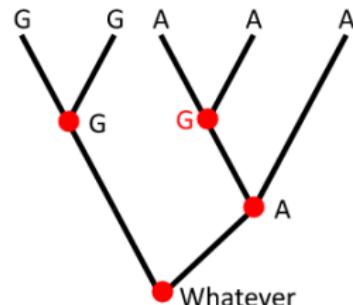
Seq1	A
Seq2	A
Seq3	A
Seq4	G
Seq5	G



Parsimony's answer for internal states

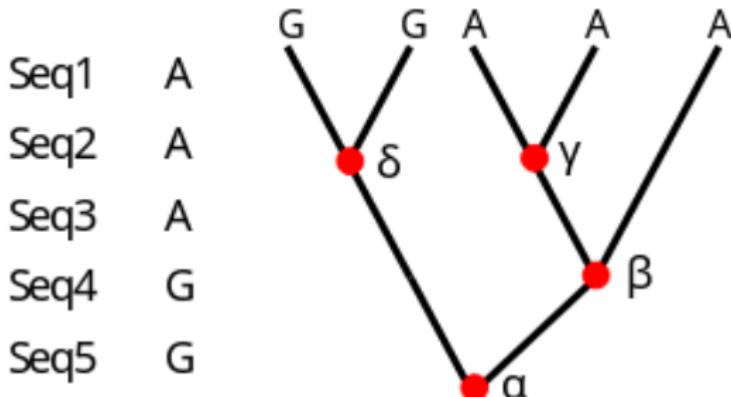
Maximum-Likelihood

Seq1	A
Seq2	A
Seq3	A
Seq4	G
Seq5	G



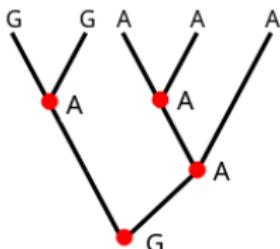
But likelihood will consider this too
(and all other possibilities)

The “intuition” version:



$$\cdot p(D|\tau, \theta) = \sum_{\alpha} \sum_{\beta} \sum_{\gamma} \sum_{\delta} = p(A, A, A, G, G, \alpha, \beta, \gamma, \delta | \tau, \theta)$$

Maximum-Likelihood



Likelihood of a given tree
(branching order AND branch lengths)

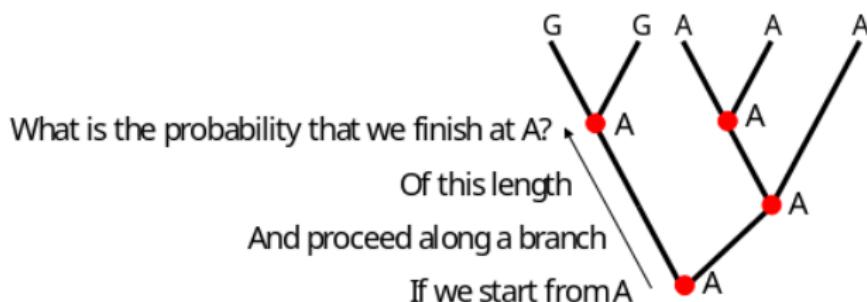
Probability of the states, given the tree

$$P(Data | T) = \sum_{\alpha} \sum_{\beta} \sum_{\gamma} \sum_{\delta} P(A, A, A, G, G, \alpha, \beta, \gamma, \delta | T)$$

Sum over ALL possible internal states

Maximum-Likelihood

$$P(A, A, A, G, G, \alpha, \beta, \gamma, \delta | T)$$



Depends on SUBSTITUTION MATRIX
and BRANCH LENGTH

Maximum-Likelihood Pros/Cons

- Advantages:
 - Maximum use of information in data
 - Explicit Model
 - Can handle complex models
 - Robust and consistent (for correct model)
 - Allows comparison of trees (which is ‘best’ and by how much)
- Disadvantages:
 - Default treatment sites as independent.
 - Very slow for exhaustive search
 - Model misspecification issues
 - Difficult to extend.
 - Question formulation can be unintuitive

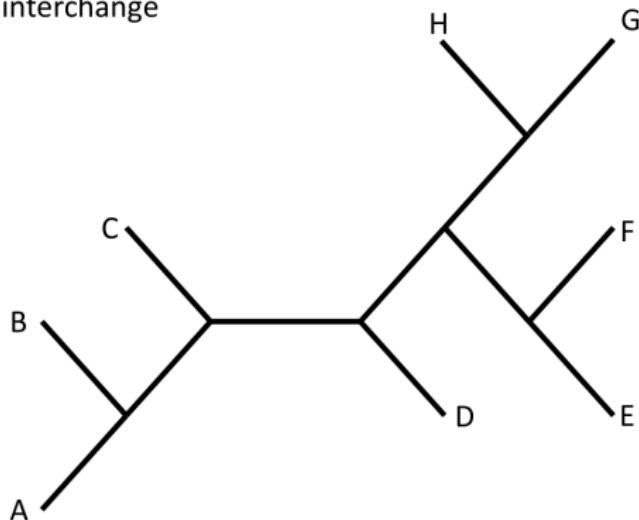
- Bayes Rule: $p(\theta|X) = \frac{p(D|\theta)p(\theta)}{\int p(D|\theta)p(\theta)d\theta}$
- For trees: $p(\theta, \tau|D) = \frac{p(D|\theta, \tau)p(\theta)p(\tau)}{\int^\theta \int^\tau p(D|\theta, \tau)p(\theta)p(\tau)d\tau d\theta}$
- Approximate marginal probability using Markov-Chain Monte-Carlo
- Run multiple chains to estimate convergence

Bayesian Pros/Cons

- Advantages:
 - Fast (relatively)
 - Can infer many different parameters
 - More flexible framework
 - More intuitive formulation
- Disadvantages:
 - Choice of priors
 - Difficulty determining convergence
 - Model misspecification issues.

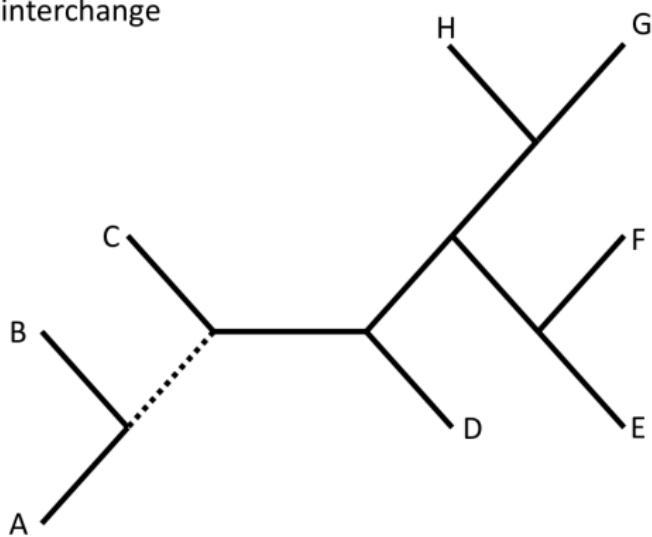
Searching Tree-Space: NNI

Nearest-neighbor interchange



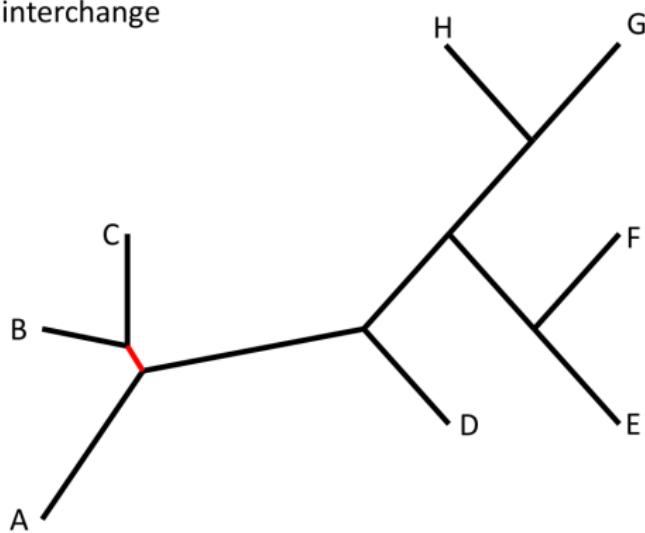
Searching Tree-Space

Nearest-neighbor interchange



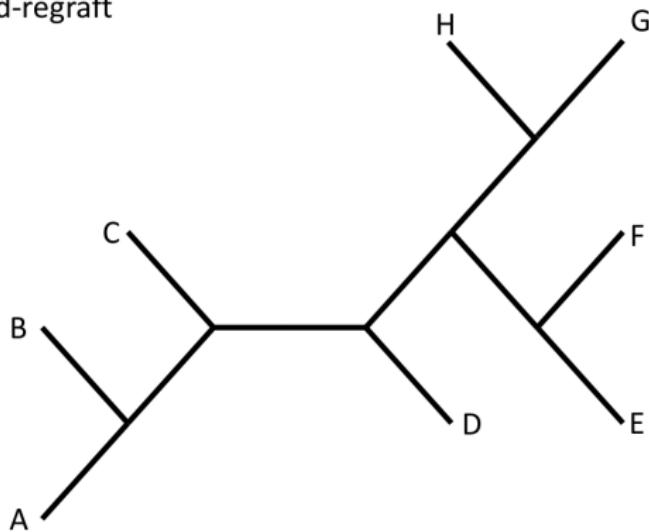
Searching Tree-Space

Nearest-neighbor interchange



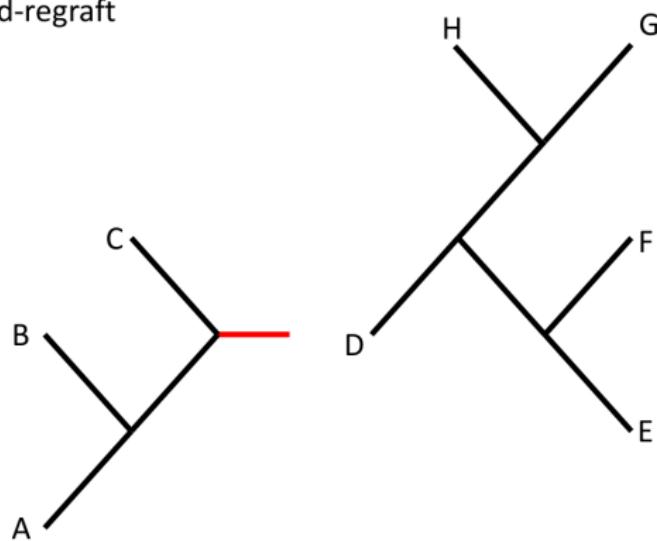
Searching Tree-Space

Subtree prune-and-regraft



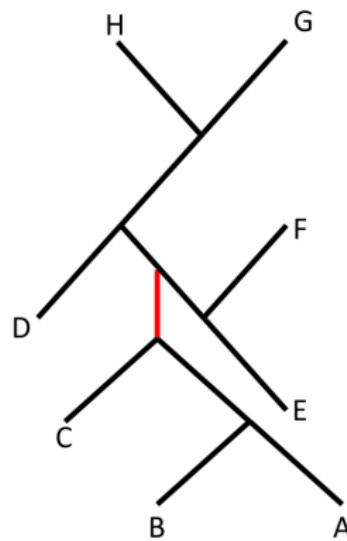
Searching Tree-Space

Subtree prune-and-regraft



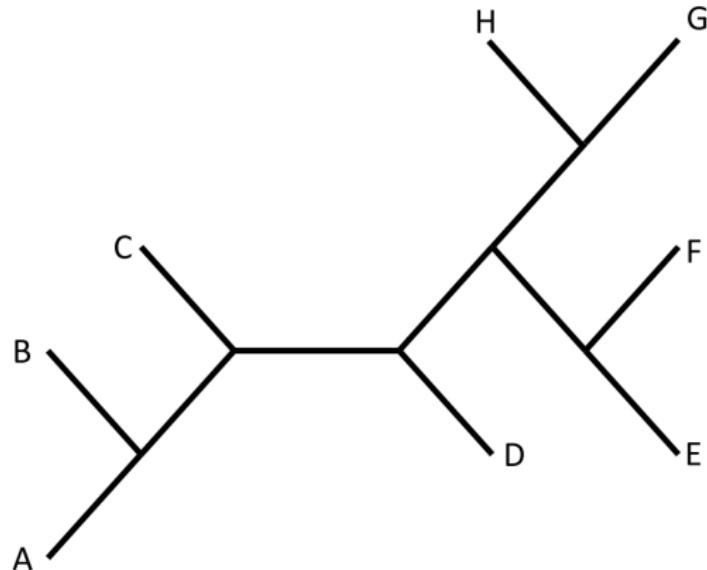
Searching Tree-Space

Subtree prune-and-regraft



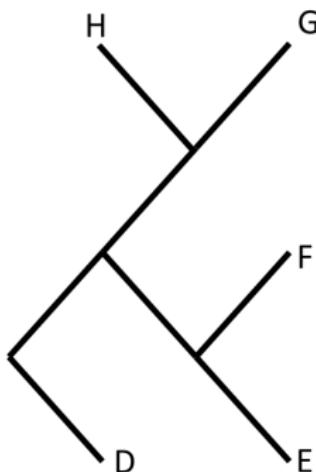
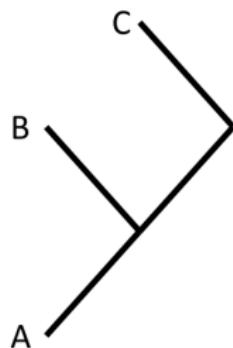
Searching Tree-Space

Tree bisection and reconnection



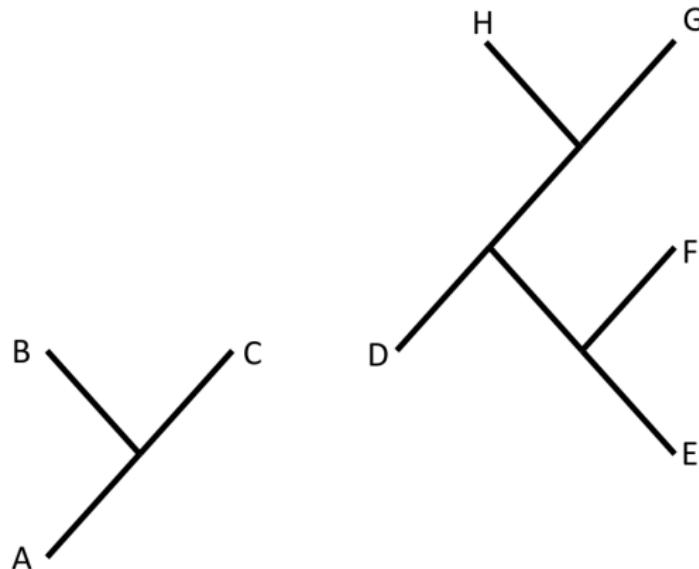
Searching Tree-Space

Tree bisection and reconnection



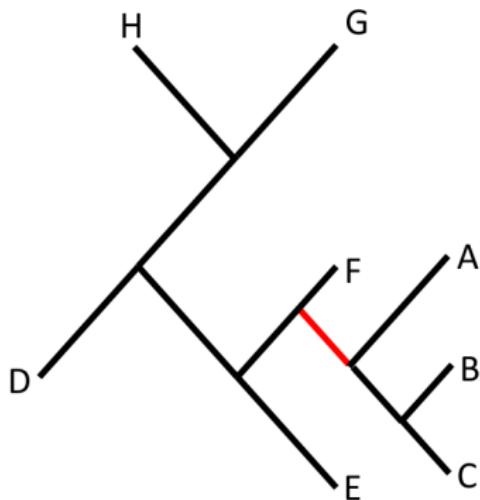
Searching Tree-Space

Tree bisection and reconnection



Searching Tree-Space

Tree bisection and reconnection



Conclusion

Summary

- Phylogenetics are a useful tool to investigate the relations between sequences

Summary

- Phylogenetics are a useful tool to investigate the relations between sequences
- There are some tricks to interpretation of trees.

Summary

- Phylogenetics are a useful tool to investigate the relations between sequences
- There are some tricks to interpretation of trees.
- Inferring a phylogeny requires: data, alignment, trimming, method selection.

Summary

- Phylogenetics are a useful tool to investigate the relations between sequences
- There are some tricks to interpretation of trees.
- Inferring a phylogeny requires: data, alignment, trimming, method selection.
- Parsimony is simplest but easily misled.

Summary

- Phylogenetics are a useful tool to investigate the relations between sequences
- There are some tricks to interpretation of trees.
- Inferring a phylogeny requires: data, alignment, trimming, method selection.
- Parsimony is simplest but easily misled.
- Distance, ML, and Bayesian need an evolutionary model.

Summary

- Phylogenetics are a useful tool to investigate the relations between sequences
- There are some tricks to interpretation of trees.
- Inferring a phylogeny requires: data, alignment, trimming, method selection.
- Parsimony is simplest but easily misled.
- Distance, ML, and Bayesian need an evolutionary model.
- Distance methods are fast but naive.

Summary

- Phylogenetics are a useful tool to investigate the relations between sequences
- There are some tricks to interpretation of trees.
- Inferring a phylogeny requires: data, alignment, trimming, method selection.
- Parsimony is simplest but easily misled.
- Distance, ML, and Bayesian need an evolutionary model.
- Distance methods are fast but naive.
- ML and Bayesian methods treat phylogenetics as a statistics problem.

Summary

- Phylogenetics are a useful tool to investigate the relations between sequences
- There are some tricks to interpretation of trees.
- Inferring a phylogeny requires: data, alignment, trimming, method selection.
- Parsimony is simplest but easily misled.
- Distance, ML, and Bayesian need an evolutionary model.
- Distance methods are fast but naive.
- ML and Bayesian methods treat phylogenetics as a statistics problem.
- Allow probabilistic reconstruction of ancestral states and population parameters.

Summary

- Phylogenetics are a useful tool to investigate the relations between sequences
- There are some tricks to interpretation of trees.
- Inferring a phylogeny requires: data, alignment, trimming, method selection.
- Parsimony is simplest but easily misled.
- Distance, ML, and Bayesian need an evolutionary model.
- Distance methods are fast but naive.
- ML and Bayesian methods treat phylogenetics as a statistics problem.
- Allow probabilistic reconstruction of ancestral states and population parameters.
- Tree topology space is non-trivial to search.

Questions?

References i

-  Abadi, S., Azouri, D., Pupko, T., and Mayrose, I. (2019).
Model selection may not be a mandatory step for phylogeny reconstruction.
Nature Communications, 10(1):934.
-  Barbrook, A. C., Howe, C. J., Blake, N., and Robinson, P. (1998).
The phylogeny of the canterbury tales.
Nature, 394(6696):839.
-  Holmes, E. C., Dudas, G., Rambaut, A., and Andersen, K. G. (2016).
The evolution of ebola virus: Insights from the 2013–2016 epidemic.
Nature, 538(7624):193.

References ii

-  Hug, L. A., Baker, B. J., Anantharaman, K., Brown, C. T., Probst, A. J., Castelle, C. J., Butterfield, C. N., Hernsdorf, A. W., Amano, Y., Ise, K., et al. (2016).
A new view of the tree of life.
Nature microbiology, 1(5):16048.
-  Leonard, G. (2010).
Development of fusion and duplication finder blast (fdfblast): a systematic tool to detect differentially distributed gene fusions and resolve trifurcations in the tree of life.
-  Marwick, B. (2012).
A cladistic evaluation of ancient thai bronze buddha images: six tests for a phylogenetic signal in the griswold collection.
Connecting empires, pages 159–176.

References iii

-  Nickle, D. C., Heath, L., Jensen, M. A., Gilbert, P. B., Mullins, J. I., and Pond, S. L. K. (2007).
Hiv-specific probabilistic models of protein evolution.
PLoS One, 2(6):e503.
-  Richards, T. A., Soanes, D. M., Foster, P. G., Leonard, G., Thornton, C. R., and Talbot, N. J. (2009).
Phylogenomic analysis demonstrates a pattern of rare and ancient horizontal gene transfer between plants and fungi.
The Plant Cell, 21(7):1897–1911.
-  Ryu, C.-K., Kim, H.-J., Ji, S.-H., Woo, G., and Cho, H.-G. (2008).
Detecting and tracing plagiarized documents by reconstruction plagiarism-evolution tree.
In *2008 8th IEEE International Conference on Computer and Information Technology*, pages 119–124. IEEE.

-  Skelton, C. (2008).
Methods of using phylogenetic systematics to reconstruct the history of the linear b script.
Archaeometry, 50(1):158–176.