

Identifying possibly novel sources of antimicrobial resistance in uncultivated bacterial and archeal metagenome-assembled genomes

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Abstract

Antimicrobial resistance (AMR) is a global threat to human health. Efforts to prevent the spread of AMR rely on surveillance of possible AMR determinants. Under-sampled, and uncultured microbial genomes in the form of metagenome data, are a potential source for AMR gene discovery. The Resistance Gene Identifier (RGI) and the Comprehensive Antibiotic Resistance Database (CARD) [1](#) were used to predict resistomes in a data set of 7903 draft quality metagenome assembled genomes [2](#) (MAGs) from various uncultivated bacterial and archeal sources (UBAs). Genes associated with mobile colistin resistance (MCR), and carbapenem resistance, such as Metallo-beta-lactamases (NDM), *Klebsiella pneumoniae* carbapenemases (KPC), and OXA beta-lactamases were found to be represented in the UBAs. [TODO: findings] [TODO: conclusion]

Introduction

Antimicrobials are substances that can halt the growth of and kill microorganisms [3](#). An important part of the microbial adaptive process is the ability to adapt to pressure from antimicrobial agents within the environment. This ability allows microbes to continue to live and grow in the presence of antimicrobial compounds. This is known as antimicrobial resistance (AMR) <https://www.cdc.gov/drugresistance/about.html>. The abuse and misuse of antimicrobials has led to increasing levels of AMR within clinical settings [1](#).

The spread of AMR is a growing global health crisis. AMR within pathogenic microbes is a threat to our treatment of infectious diseases, reducing the efficacy of antibiotics, allowing for prolonged and more severe infections, and an increase in morbidity [4](#). The European Centre for Disease Prevention and Control estimate that 25,000 people die per year due to infections related to AMR [5](#), with reports showing the annual cost to combat AMR infections in just the USA alone to be between \$21 billion and \$34 billion [6](#).

A microbe can acquire AMR through a variety of mechanisms such as intrinsic resistance or lateral gene transfer [7](#). These mechanisms allow for the constant change of the resistome, making determining the scope of AMR within both pathogenic and non-pathogenic bacteria difficult [1](#). To impede the spread of AMR the World Health Organization (WHO) created an action plan which emphasizes the need to strengthen the knowledge and surveillance of AMR. An important aspect of this research is to examine the spread of AMR between environmental and clinical samples [url: https://www.who.int/antimicrobial-resistance/global-action-plan/en/](https://www.who.int/antimicrobial-resistance/global-action-plan/en/). Characterization of many AMR genes is currently being done through curated databases. The Comprehensive Antibiotic Resistance Database (CARD) is an example of one of these curated databases. CARD contains reference data pertaining to the genes, proteins and mutations involved in AMR. CARD allows for the identification of resistance genes within a genome using the Resistance Gene Identifier (RGI). [1](#).

These curated databases contain a large amount of information on the resistomes that are most commonly studied, especially those pertaining to *Staphylococcus aureus*, *Escherichia coli*, *Klebsiella pneumoniae*, *Acinetobacter baumannii*, and *Pseudomonas aeruginosa*. These pathogens are multidrug resistant and are the leading cause of hospital acquired infections [8](#). Most antibiotics have no efficacy on these infections and are therefore treated with carbapenems or polymyxins [5](#).

Carbapenems are broad spectrum beta-lactam antibiotics typically used to treat life threatening, high risk, and multidrug resistant bacterial infections. beta-lactamase producing bacteria have difficulty degrading carbapenems when combined with beta-lactamase inhibitors making them effective treatments against pathogens with multi-drug resistance [5](#). However, there has been a recent emergence of resistance to these antimicrobials, presenting a new threat to public health. One way microbes can develop carbapenem resistance is through the acquisition of carbapenemase genes. [5](#).

Three main carbapenemase genes of concern are *Klebsiella pneumoniae* carbapenemase (KPC), New Delhi metallo-beta-lactamase (NDM) and OXA beta-lactamases [5](#).

Carbapenemases are classified as one of four classes (A-D) based on their molecular structure [9](#). Class A, C, and D both use a serine residue to hydrolyze beta-lactam antibiotics [10](#). KPC genes are a part of class A. Many variants of KPC are spreading and being discovered, however carbapenemases within this class are more rare than the other three classes [5](#). Class D contains the OXA family of beta-lactamases. Unlike KPC and NDM, this family is characterized by phenotype rather than genotype. This family is characterized by specific hydrolases that can hydrolyze oxacillin and cloxacillin, with the enzyme being poorly inhibited by clavulanic acid <https://card.mcmaster.ca/ontology/36026>. This results in a low amount of sequence homology within the family. A subset of this family, OXA-48 possesses carbapenem hydrolyzing activity [11](#). The OXA-48 subfamily consists of five variants: OXA-48, OXA-162, OXA-163, OXA-181, OXA-204, and OXA-232 [12](#). Class B beta-lactamases are metalloenzymes that use a zinc active site to hydrolyze beta-lactam antibiotics. The NDM gene is classified as a class B beta-lactamase [5](#).

When pathogens become resistant to carbapenems, the last line of defense is colistin. Colistin is a polymyxin antibiotic. Polymyxins act via disrupting membrane permeability [13](#). Colistin is limited to a last line defense against carbapenem resistant infections due to its neurotoxic and nephrotoxic effects [13](#). Despite its already limited use, mobilized colistin resistance (MCR) have been discovered [14](#). Due to the emergence of resistance to many last-line antibiotics, it is important to characterize the total diversity and spread of these resistance determinants.

The UBA dataset contains the data needed to expand AMR surveillance to a wide range of environmental samples. Coupling RGI predictions of the UBA metagenomes with phylogenetic analysis of the predicted AMR genes enables the possibility to discover previously unseen AMR diversity in genomes not yet analyzed for AMR, and provides insight into the diversity of AMR across non-clinical samples.

Methods

AMR Prediction

A dataset of 7903 draft quality MAGs which were recovered from the Sequence Read Archive by Parks [2](#) were used for this analysis. RGI version 5.0 with CARD database version 3.02 [1](#) was run on the contigs of the 7903 MAGs with the inclusion of loose, perfect, and strict hits.

Phylogenetic Analysis

MCR

Obtaining candidate and reference MCR genes

24 canonical sequence variants as indicated by CARD for the MCR phosphoethanolamine transferase [AMR Gene Family], MCR 3.11, 6.1, 2.2, 3.6, 3.10, 3.5, 1.10, 1.9, 3.9, 3.8, 3.7, 3.2, 3.1, 1.6, 1, 2, 3, 1.2, 1.3, 1.4, 8, 4, 5, 7.1 were obtained from the CARD database.

Output containing only AMR gene prediction data for the MCR phosphoethanolamine transferase gene family was produced by filtering RGI output based on its association with 'MCR phosphoethanolamine'. The filtered data were translated to a blast database. The 24 CARD canonical sequences were used to perform a multiple query BLASTP against the UBA blast database with a e-value threshold of 1e-170 and a query coverage of 99%. Redundant results were filtered from these BLASTP results by retrieving only the longest sequence for each uniquely labeled result.

Additional putative MCR phosphoethanolamine transferase gene family sequences were accumulated by querying the CARD prevalence data version 3.0.4 ("based on sequence data acquired from NCBI on 28-Feb-2019, analyzed using RGI 4.2.2 (DIAMOND homolog detection) and CARD 3.0.1"). The prevalence data was translated to a BLAST database. The 24 CARD canonical sequences were used to perform a multiple query BLASTP against the prevalence blast database with a e-value threshold of 1e-180 and a query coverage of 99%. Redundant results were filtered from these BLAST results by retrieving only the longest sequence for each uniquely labeled result.

The 24 CARD canonical sequences were used to perform a multiple query BLASTP against the NCBI non-redundant database blast database with a e-value threshold of 1e-180 and a query coverage of 90%. Redundant results were filtered from these BLAST results by retrieving only the longest sequence for each uniquely labeled result.

The filtered sequences from NCBI non-redundant data, CARD prevalence data, UBA data were all concatenated to one multi-FASTA format file with the canonical sequences. These the concatenated amino acid sequences were aligned with MAFFT FFT-NS-2. The aligned sequences were trimmed by trimal using the automated1 option. IQ-TREE was then used to build a bootstrapped tree with -bb 1000 with the LG+R10 model of substitution. all leaves of the trees were labeled with as much taxonomic information as possible for each rank, based on information from metadata.

The result was carried to subsequent analysis.

Results

Phylogenetic analysis of putative MCR sequences

The phylogenetic relationships of the CARD canonical sequences, and the CARD prevalence sequences involving the MCR family were investigated. A total of 30 genes, an out-group, the 23 canonical sequences, and the 6 prevalence sequences, were selected for analyses. The tree in Figure 1 shows several distinct clades. Each MCR variant forms its own clade. MCR-1, MCR-2, and MCR-6 form a clade, appearing to more closely related to one another than with the other MCR family members.

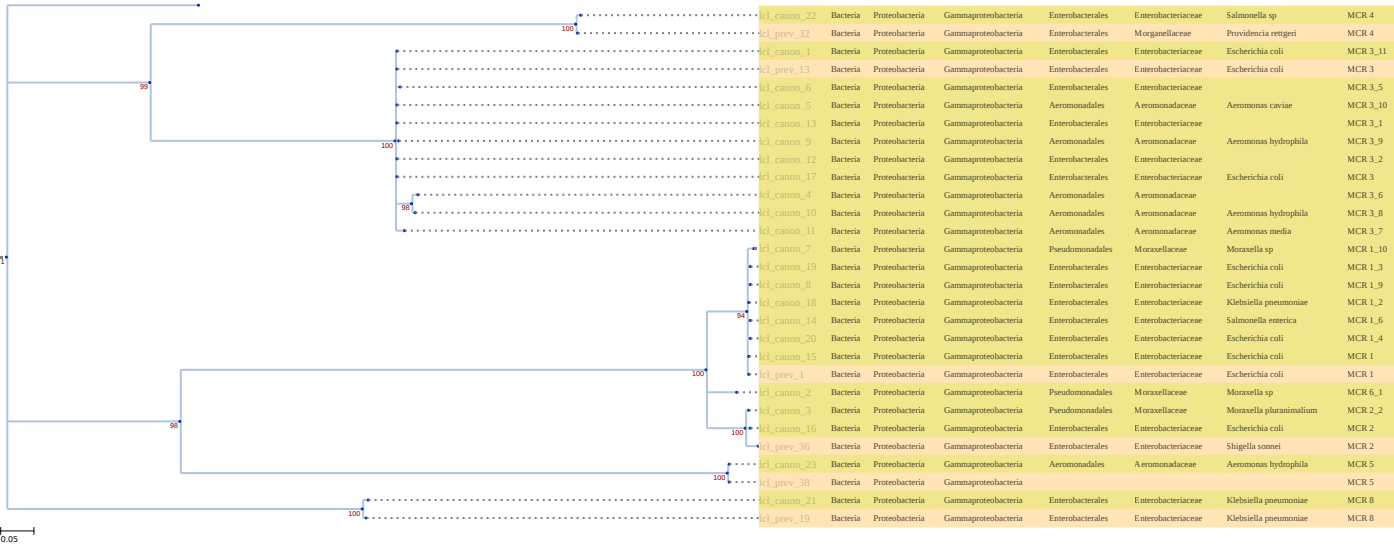


Figure 1: Phylogenetic relationship for 23 canonical (lcl_canon) MCR family sequences along with 6 prevalence (lcl_prev) sequences.

Phylogenetic analysis of NCBI data

Sequences from the NCBI non-redundant data were added to the analysis to be compared with the canonical and prevalence sequences. This resulted in a total of 409 sequences for subsequent analysis, all labeled as phosphoethanolamine lipid A transferase genes, where 104 hits were labeled as MCR family genes.

One of the BLAST results was a variant which was not included in the CARD canonical or prevalence sequences. The variant, MCR-9, was reported in the literature to be a novel MCR homologue with similarity to MCR-3 [15](#). The gene was shown to confer resistance to colistin when expressed in *Escherichia coli*.

The phylogeny in Figure [2](#) shows that the, the MCR 3 clade branches with MCR-9. This MCR-3 and MCR-9 clade is separated by a large amount diversity from the MCR 1,2,4,5, and 8 clades. This diversity is represented by many bacteria containing phosphoethanolamine lipid A transferase with none reported to be in the MCR family.

RGI produced 1457246 results AMR determinants under the loose cutoff from the UBA data, 7171 for the strict cutoff, and 310 for the loose cutoff. It was hoped that phylogenetic analysis could find AMR

determinants would be found in under the loose criterion that may have been missed by RGI-CARD. The UBA BLAST results were included in the phylogeny in Figure 2 for the analysis. The diversity between clades, swathes of which are not reported to be in the MCR family, contain multiple sources from UBAs. It is unclear if these relationships could be involved with the MCR family, but there are more enticing relationships present in the tree. The remainder of the analysis deals with relationships which are deemed to be interesting based on the locations of the UBAs between MCR family clades.

MCR-5, MCR-1,2,6 clades

The MCR-5, MCR1,2,6 clades shown in figure ?? share, with 100 bootstrap support, a common ancestor. Branching between these two clades, as a descendant to this ancestor, is a clade of *Psychrobacter* species bacteria. This clade includes two hits from the non-redundant database, and two hits from the UBA data. According to the Parks 2 data, the identity of these samples are, UBA3068, A *Psychrobacter* sp., sampled from Oil polluted marine microbial communities from Coal and Oil in Point Santa Barbara, California, USA and, UBA4193, a *Psychrobacter* sp, sampled from the New York City MTA subway samples. The quality information for these sequences, shown in Table ??, shows that UBA3068 is near complete, while UBA4193 is only partial. It is encouraging to see that even with the quality difference, these two sequences branch in the expected clade. *Psychrobacter* 16 is a Genus is widespread and includes many cold adapted bacteria, it is an opportunistic pathogen, and has been found sometimes be a cause of infections in humans, animals, and fish. Many new species of *Psychrobacter* have been discovered in cold climates 17. Some of the species have been shown to be resistant to colistin, like *Psychrobacter vallis* ps. nov. and *Psychrobacter aquaticus* ps. nov. 18.

MCR-3 clade

A single result, UBA705, which the Parks data 2 reports as a Comparative metagenome analyses of anode-associated microbial communities developed in rice paddy field-soil microbial fuel cells, is reported to be *Aeromonas hydrophilia*. This present within the clade alone with several other canonical and non-redundant *Aeromonadaceae*. *Aeromonas hydrophilia* is a species which has been found to have an MCR-3 gene. Again, even though qualities vary (Table ??), the sequences branch in the expected location.

MCR 3,9 clade

This clade contains 8 UBAs from the loose CARD hits under the given constraints, and several NCBI non-redundant hits not reported as MCR family genes. 7 Loose hits for MCR in a */Leclercia/* *adecarboxylata* branch within this clade, branching closely with the NCBI non-redundant MCR9. These */Leclercia/* branch below the common ancestor of MCR-9 and MCR-3 which are well supported. The */Leclercia/* UBAs were all sampled from New York City MTA subway samples Metagenome. *Acinetobacter* is another opportunistic pathogen 19 which is becoming resistant to many antimicrobials.

Discussion

Conclusion

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