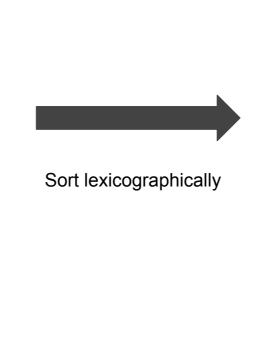
#### **Compression:** The Burrows Wheeler Transform

A classical structure: the **Burrows Wheeler transform** (BWT)

GATTAGATACAT\$ ATTAGATACAT\$G TTAGATACAT\$GA TAGATACAT\$GAT AGATACAT\$GATT GATACAT\$GATTA ATACAT\$GATTAG TACAT\$GATTAGA ACAT\$GATTAGAT CAT\$GATTAGATA AT\$GATTAGATAC T\$GATTAGATACA **\$GATTAGATACAT** 





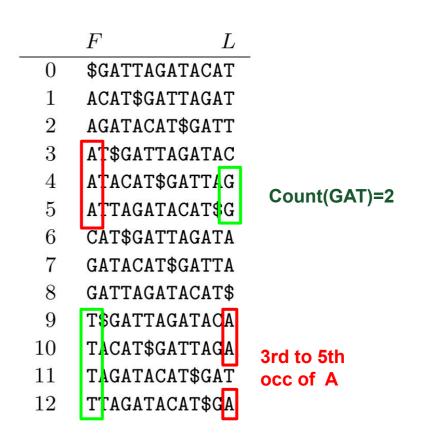
# **Self Indexing:** The FM-index

#### **Searching for GAT in GATTAGATACAT:**



- Count(P) in O(|P|)
- Locate(P) in O(|P| + occ)

Applied in short read aligners such as **Bowtie** and **BWA**!



# **Self Indexing:** Runs

The BWT can be quite repetitive:

BWT(GATTAGATACAT\$) = TTTCGGAA\$AATA

A **run** is a **substring of identical letters**, for example AAABBBA has 3 runs.

BWT(GATTAGATACAT\$) = TTTCGGAA\$AATA => 8 runs

Method	Space	Count(P)	Locate(P)
The Run-Length FM index [Mäkinen, Navarro, Siren, and Välimäki.]	O(r)	O( P log T )	/ (not in O(r))
The r-index [Gagie, Navarro, and Prezza.]	O(r)	O( P )	O( P +occ)

#### Indexing of a collection: read sets

TTAGA
TAGATA
GATTAGATACAT
GATTA ATACAT
GATAC

- Could we have a similar structure with space depending on the number of runs for an aligned readset?
- Can we find a link between the number of runs of that similar structure for the readsets and the runs in the BWT of the genome it is aligned to?

#### Indexing of a collection: Naive approach

Concatenate the reads with a separator "\$" and build the FM-index of the entire string.

#### **Example:**

S = GATTA\$TTAGA\$TAGATA\$GATAC\$ATACAT\$
BWT(S) = CATAATGTTTTTCGG\$GAAAA\$\$AATAAT\$A\$ => 20 runs.

#### Issues:

- Computing the BWT for such a long string is challenging (and the context before the dollars is not relevant).
- The \$ break some runs as in CGG\$G in the example.

# Indexing of a collection: The EBWT

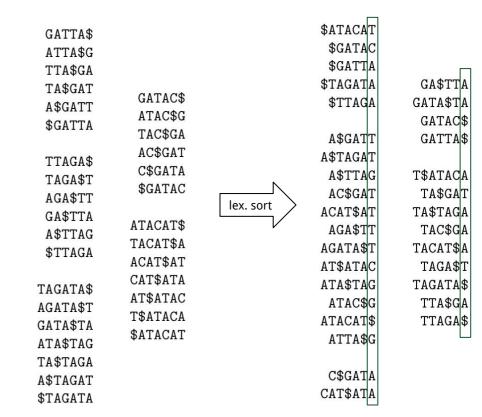
**Extended BWT:** lexicographically sort all rotations of each string.

[Mantaci, Restivo, Rosone, and Sciortino, An extension of the Burrows–Wheeler Transform, 2007]

#### **Example:**

EBWT(S) = TCAAATTGTTTTCGG\$GAAAA\$\$ATAAAT\$A\$ => 19 runs.

- Easier to build and update than the naive approach.
- Still has more than double the number of run than BWT(GATTAGATACAT).



#### Indexing of a collection: the heuristics on order

**RLO**: Reorganize the reads in the reversed lexicographic order (co-lexicographic order)

#### **Example:**

RLO(S) = {GATTA\$, TTAGA\$, TAGATA\$, GATAC\$, ATACAT\$} EBWT(RLO(S)) = AAACTGTTTTTTCGG\$GAAAA\$\$AATAAT\$A\$ => 19 runs

#### SPRING:

[ SPRING: a next-generation compressor for FASTQ data, Chandak, Tatwawadi, Ochoa, Hernaez, and Weissman, 2018]

Attempts to reorder reads according to their position in the genome.

EBWT(SPRING(S))= ACATATTGTTTTCGG\$GAAAA\$\$ATAAAT\$A\$ => 22 runs

#### **Indexing of a tree:** The XBWT

**eXtended BWT:** Generalization of the BWT for labeled trees where nodes are sorted by their label from the node to the root and we output the outgoing edges.

$$XBWT(T) = ab abc $c $ $ aacaa$$$$

(Can be seen as a sub-case of a Wheeler graph.)

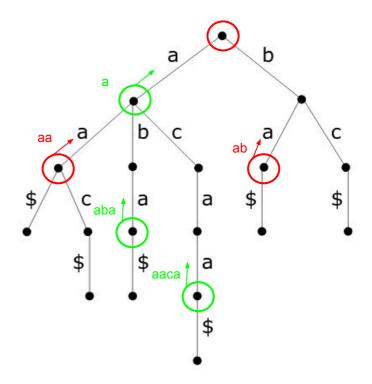


Figure: XBWT Tricks, Giovanni Manzini

### Our approach: using alignment and XBWT

If the reads correspond to a known assembled genome, use the genome as additional context for better compression.

**Example:** 

TTAGA
TAGATA
GATTAGATACAT
GATTA ATACAT
GATAC

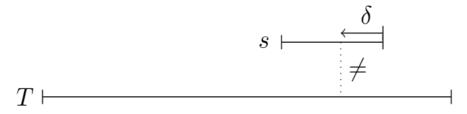
T =

XBWT(T)=GGTTTTTTTTCCCGGGGAAAAAAAAATTTTAAAAAAAA => 7 runs.

#### Our approach: Theoretical guaranties

If we create such a tree, where the **reads are errorlessly sampled** and aligned to the **reference**, then the XBWT of the tree has the **same number of runs** as the BWT of the reverse of the reference.

But in reality the reads are not perfectly matching the assembled genome ..?



If the reads differ from the reference string and that the average **distance from first difference** (insertion, deletion, or substitution) **to the end** of the read is  $\delta$ , then, the XBWT of the tree will have at most  $2\delta$  additional runs per reads.

### Our approach: Theoretical guaranties

```
If we create a labeled tree T as explained, let:
r be the number of runs in the XBWT,
t be the number of reads,
Then in O(r+t) words of space,
We can:
Count(P) in O(|P|log log(|T|))
Locate(P) in O((|P|+occ)log log(|T|))
```

What are the runs in the XBWT in practice? And how do we build the XBWT?

#### For scalability: prefix-free parsing construction

The reference and genomes are typically tens of Gb of data...

Consequently, algorithms designed to work in RAM may not be practical.

We chose to adapt a previous technique **Prefix free parsing**.

**Prefix free construction** takes advantage of the **highly repetitive** nature of genomic databases using **context-triggered piecewise hashing**.

[Boucher, Gagie, Kuhnle, Langmead, Manzini and Mun, Prefix-free parsing for building big BWTs, WABI 2017]

[PFP Data Structure, Boucher, Cvacho, Gagie, Holub, Manzini, Navarro, Rossi, 2020]

**54 GB peak memory** for 1000 variants of human chromosome 19, initially occupying roughly **56 GB**.

To aim for a scalable structure we adapted Prefix free parsing to build the XBWT.

# **Experiments:** comparison to the state of the art

A preliminary comparison only on the number of runs for now.

We compared to:

- **EBWT** (using the ropebwt2 implementation), with and without \$.
- **SPRING + EBWT**, with and without \$.
- RLO + EBWT, with and without \$.

Removing the \$ reduced the number of runs between 2.7% and 29.2%.

#### **Experiments:** datasets and protocol

Only reads matched to the genome, not to the reverse complement.

Reads and corresponding genome:

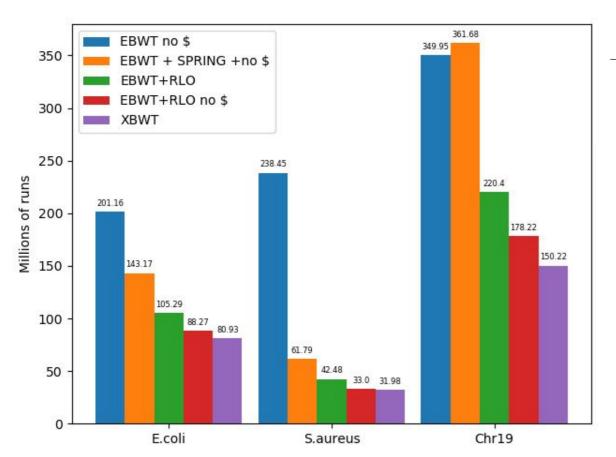
- **E.coli**: from the single cell dataset
- **S.aureus**: from the single cell dataset
- R.sphaeroides: from the Gage-b dataset

Reads aligned to a reference genome:

• Chr19: a reference genome and the reads of a HiSeq 2000 readsets that aligned

**bwa-mem** used to align the reads to the genomes.

#### **Experiments:** results



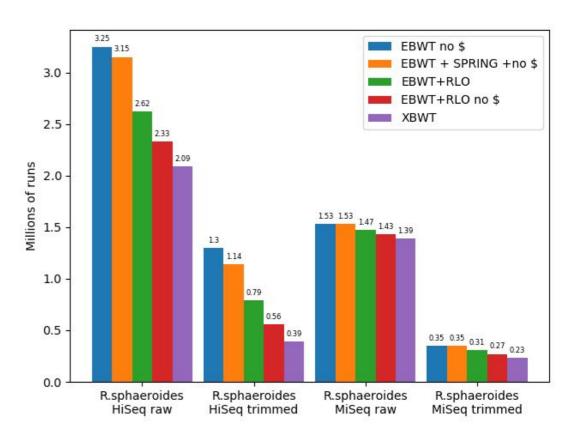
Dataset	Nb. reads	Coverage	$\delta$	errorless reads
E.coli	14M	$304 \times$	13	57%
S.aureus	26M	$927 \times$	7	88%
Chr19	34M	$57 \times$	15	71%

- RLO+EBWT has much less runs than the plain EBWT.
- XBWT performs best (but RLO is close)
- On S.aureus, RLO, RLO no \$
   and XBWT are very close.

# **Experiments:** Gage-b R.sphaeroides

Dataset	Number of reads	Read length	Coverage
HiSeq			
raw	166 820	101	$46 \times$
$\operatorname{trimmed}$	$134\ 207$	up to 101	$37 \times$
MiSeq			
raw	23 102	251	$24 \times$
trimmed	20 046	up to $251$	$20 \times$

Dataset	$\delta$	Errorless reads	Error rate
HiSeq			
raw	27	31.34%	0.04%
$\operatorname{trimmed}$	6	83.26%	0.01%
MiSeq			
raw	122	0.25%	0.15%
$\operatorname{trimmed}$	29	63.55%	0.03%



#### **Summary of our contributions:**

- Looking at the genome for additional context for better compression is worth investigating!
  - We provide theoretical time and space guaranties depending on the number of reads and the number of runs.
  - We show an upper bound on the number of runs depending on the errors in the reads compared to the genome.
  - The experimental number of runs is comparatively small.
- Prefix-free construction of the BWT can be adapted for the XBWT.
- A similar approach could be used to improve the space usage of the hybrid index. (Not explored in this talk)

#### //**TODO**:

- Larger scale analysis (on human genome, on long reads)
- FM-index, implementation and time analysis
- Time comparison of PFP construction of the XBWT compared to other construction
   [BWT-tunneling by Uwe Baier, Wheeler sort by Jarno Alanko]