

AlleleSNP Introduction

1. Background

Allele-specific effects (ASE) are the variations within a single individual, such as differences in chromatin signatures, DNA methylation and gene expression, that are related to the different alleles of a SNP (Birney et al. 2010; Rozowsky et al. 2011). When a heterozygous SNP shows ASE of epigenetic signatures, such as transcription factor (TF) binding or histone modifications, it is a strong indication of its functionality because it shows that within the same cellular environment, the two SNP alleles can behave differently (**Fig 1**). ASE can be identified through examination of the NGS data: for example, we can collect a TF ChIP-seq reads that mapped to a certain SNP, if the number of reads that contain the reference allele and the alternate allele are imbalanced, it might indicate that the SNP might play a critical role in regulating the binding affinity of the TF (**Fig 2**).

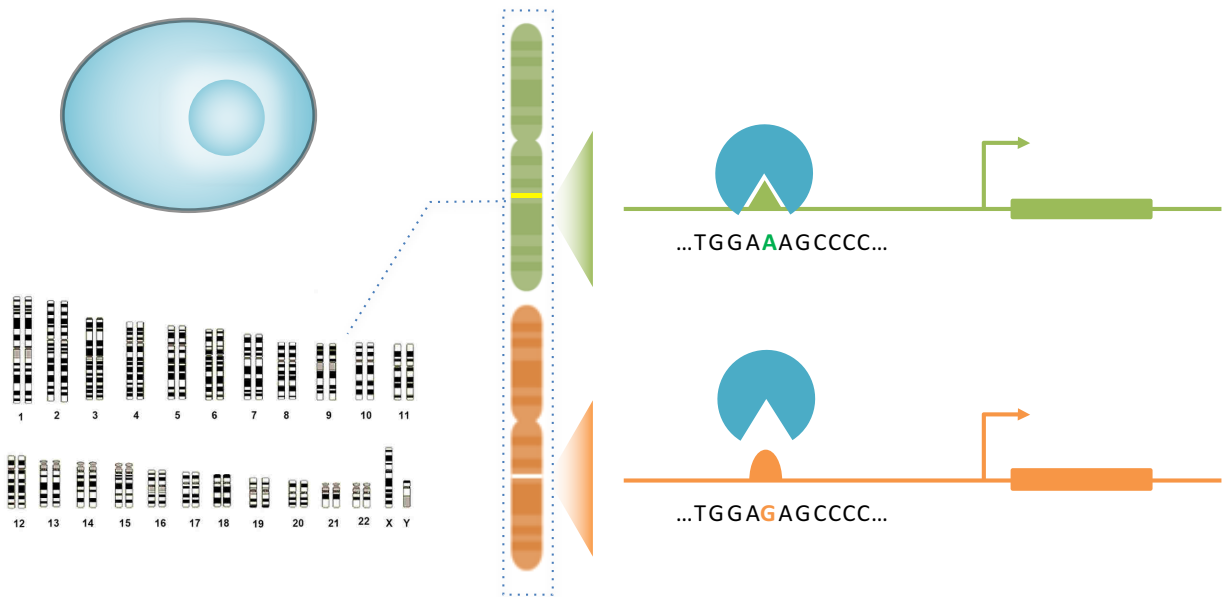


Figure 1. Schematic plot of allele-specific effects

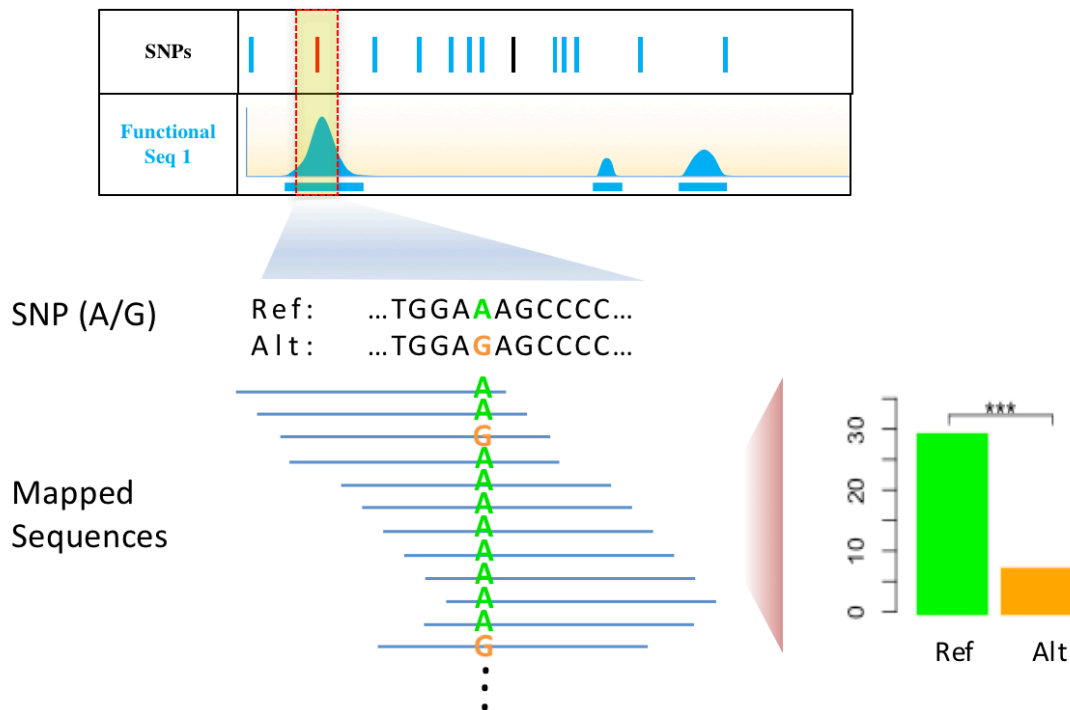


Figure 2. Schematic plot of ASE identification through NGS data

2. Installation

```
if (!require(AlleleSNP)) {
  library(devtools)
  install_github("foreverycc/AlleleSNP_Package")
}
library(AlleleSNP)
```

3. Identify ASE by bam files

The first way to identify ASE is through examination of bam file.

For each bam file (ChIP-seq, DNase-seq, ATAC-seq, FAIRE-seq, etc.), AlleleSNP will search for heterozygous given SNPs, then extract the number of reads that contain either the reference or the alternate allele, and finally perform statistical tests (**Fig 2**).

3.1 Input SNP format

```
assnp_dir = .libPaths()
index_snp_file = paste0(assnp_dir, "/AlleleSNP/extdata/input_snps/input_snp_example2.csv")
read.csv(index_snp_file, header = F)
```

```
##          V1  V2
## 1   rs1051730 EUR
## 2   rs10937405 ASN
## 3   rs8034191 EUR
## 4   rs8042374 EUR
## 5   rs9387478 ASN
## 6   rs402710 EUR
## 7   rs139852726 EUR
## 8   rs7741164 ASN
```

3.2 Input bam files

```
bam_dir = paste0(assnp_dir, "/AlleleSNP/extdata/sample/A549/bam_files")
list.files(bam_dir)
```

```
## [1] "A549_H3K27ac.bam"      "A549_H3K27ac.bam.bai" "A549_H3K4me1.bam"
## [4] "A549_H3K4me1.bam.bai" "A549_H3K4me3.bam"     "A549_H3K4me3.bam.bai"
## [7] "A549_H3K9ac.bam"      "A549_H3K9ac.bam.bai"  "A549_PolIII.bam"
## [10] "A549_PolIII.bam.bai"
```

3.3 Get allele-specific binding events by examination of the bam files

```
get_assnp_byBam(index_snp_file = index_snp_file, bam_dir = bam_dir, sample_name = "A549_singleBam")
```

```
##          rsID    biofeature ref alt ref_rmdup alt_rmdup genotype_singleBam
## 1   rs3813570   A549_PolII  49  74         21         26             TRUE
## 2   rs31490    A549_H3K4me3  25  16         14         11             TRUE
## 3   rs27996    A549_H3K4me3  30  21         15         13             TRUE
## 4   rs57064725 A549_H3K27ac   9  15          7          9             TRUE
## 5   rs57064725 A549_H3K4me3  23  30         17         15             TRUE
## 6   rs684513   A549_H3K4me3  46  38         18         15             TRUE
## 7   rs3813570   A549_H3K4me3  54  51         20         22             TRUE
## 8   rs503464    A549_PolIII  33  31         21         16             TRUE
## 9   rs59133824 A549_H3K4me3  78  78         29         28             TRUE
## 10  rs59683676 A549_H3K4me3  77  77         29         28             TRUE
##      genotype_sample genotype_vcf genotype_final biofeature_overlap_names
## 1                NA              NA              TRUE                  NA
## 2                NA              NA              TRUE                  NA
## 3                NA              NA              TRUE                  NA
## 4                NA              NA              TRUE                  NA
## 5                NA              NA              TRUE                  NA
## 6                NA              NA              TRUE                  NA
## 7                NA              NA              TRUE                  NA
## 8                NA              NA              TRUE                  NA
## 9                NA              NA              TRUE                  NA
## 10               NA              NA              TRUE                  NA
##      biofeature_overlap_num biofeature_overlap ref_count alt_count ref_cnv
## 1                NA              NA              NA              NA              1
## 2                NA              NA              NA              NA              1
## 3                NA              NA              NA              NA              1
## 4                NA              NA              NA              NA              1
## 5                NA              NA              NA              NA              1
## 6                NA              NA              NA              NA              1
## 7                NA              NA              NA              NA              1
## 8                NA              NA              NA              NA              1
## 9                NA              NA              NA              NA              1
```

			NA		NA		NA	NA	1
##	10								
##		alt_cnv	p.val.raw	p.val.cnv	p.val.cnv.bh	p.val.cnv.bonf			
##	1	1	0.03004691	0.03004691	0.3004691	0.3004691			
##	2	1	0.21102360	0.21102360	0.7420078	1.0000000			
##	3	1	0.26243754	0.26243754	0.7420078	1.0000000			
##	4	1	0.30745625	0.30745625	0.7420078	1.0000000			
##	5	1	0.41010272	0.41010272	0.7420078	1.0000000			
##	6	1	0.44520467	0.44520467	0.7420078	1.0000000			
##	7	1	0.84537032	0.84537032	1.0000000	1.0000000			
##	8	1	0.90065325	0.90065325	1.0000000	1.0000000			
##	9	1	1.00000000	1.00000000	1.0000000	1.0000000			
##	10	1	1.00000000	1.00000000	1.0000000	1.0000000			

4. Identify ASE by sample

If you have more information of a sample, you could also identify ASE is through integrating multiple data types of a sample.

Here AlleleSNP provided a way to integrate bam files, peak files, and vcf files together to identify ASE. This mode may identify heterozygous SNPs that are not called using single bam file. But it also requires more prerequisite work, such as peak calling, vcf calling.

4.1 In the sample mode, we can incorporate more types of data, including:

- bam files
- peak files (.bed)
- vcf files

```
sample_dir = paste0(assnp_dir, "/AlleleSNP/extdata/sample/A549")
for (dir in list.files(sample_dir, full.names = T)) {
  cat (dir, "\n")
  for (file in list.files(dir)) {
    cat (file, "\n")
  }
}
```

```
##
/Library/Frameworks/R.framework/Versions/3.3/Resources/library/AlleleSNP/extdata/sample/A549/bam_files
## A549_H3K27ac.bam
## A549_H3K27ac.bam.bai
## A549_H3K4me1.bam
## A549_H3K4me1.bam.bai
## A549_H3K4me3.bam
## A549_H3K4me3.bam.bai
## A549_H3K9ac.bam
## A549_H3K9ac.bam.bai
## A549_PolIII.bam
## A549_PolIII.bam.bai
##
/Library/Frameworks/R.framework/Versions/3.3/Resources/library/AlleleSNP/extdata/sample/A549/peak_files
## A549_H3K27ac_peaks.bed
```

```
## A549_H3K4me1_peaks.bed
## A549_H3K4me3_peaks.bed
## A549_H3K9ac_peaks.bed
## A549_PolIII_peaks.bed
##
/Library/Frameworks/R.framework/Versions/3.3/Resources/library/AlleleSNP/extdata/sample/A549/vcf_files
## A549_ChIPseq_GATK.vcf
## A549_ChIPseq_Samtools.vcf
## A549_WGS_GATK.vcf
```

4.2 Get allele-specific binding events by integrating all the data of the sample

```
get_assnp_bySample(index_snp_file = index_snp_file, sample_name = "A549", sample_dir = sample_dir)
```

```
##      rsID      biofeature ref alt ref_rmdup alt_rmdup genotype_singleBam
## 1  rs3813570  A549_PolIII  49  74         21         26              TRUE
## 2  rs57064725 A549_H3K27ac  9  15          7          9              TRUE
## 3  rs684513   A549_H3K4me3 46  38         18         15              TRUE
## 4  rs27996    A549_H3K4me3 30  21         15         13              TRUE
## 5  rs57064725 A549_H3K4me3 23  30         17         15              TRUE
## 6  rs31490    A549_H3K4me3 25  16         14         11              TRUE
## 7  rs3813570  A549_H3K4me3 54  51         20         22              TRUE
## 8  rs503464   A549_PolIII  33  31         21         16              TRUE
## 9  rs59133824 A549_H3K4me3  78  78         29         28              TRUE
## 10 rs59683676 A549_H3K4me3  77  77         29         28              TRUE
##      genotype_sample genotype_vcf genotype_final biofeature_overlap
## 1              TRUE              TRUE              TRUE              TRUE
## 2              TRUE              TRUE              TRUE              TRUE
## 3              TRUE              TRUE              TRUE              TRUE
## 4              TRUE              TRUE              TRUE              TRUE
## 5              TRUE              TRUE              TRUE              TRUE
## 6              TRUE              TRUE              TRUE              TRUE
## 7              TRUE              TRUE              TRUE              TRUE
## 8              TRUE              TRUE              TRUE              TRUE
## 9              TRUE              TRUE              TRUE              TRUE
## 10             TRUE              TRUE              TRUE              TRUE
##      biofeature_overlap_num
## 1              4
## 2              4
## 3              4
## 4              3
## 5              4
## 6              4
## 7              4
## 8              4
## 9              4
## 10             4
##
##      biofeature_overlap_names
## 1  A549_H3K27ac_peaks,A549_H3K4me3_peaks,A549_H3K9ac_peaks,A549_PolIII_peaks
## 2  A549_H3K27ac_peaks,A549_H3K4me3_peaks,A549_H3K9ac_peaks,A549_PolIII_peaks
## 3  A549_H3K27ac_peaks,A549_H3K4me3_peaks,A549_H3K9ac_peaks,A549_PolIII_peaks
## 4  A549_H3K27ac_peaks,A549_H3K4me3_peaks,A549_H3K9ac_peaks
## 5  A549_H3K27ac_peaks,A549_H3K4me3_peaks,A549_H3K9ac_peaks,A549_PolIII_peaks
## 6  A549_H3K27ac_peaks,A549_H3K4me1_peaks,A549_H3K4me3_peaks,A549_H3K9ac_peaks
```

```

## 7      A549_H3K27ac_peaks,A549_H3K4me3_peaks,A549_H3K9ac_peaks,A549_PolIII_peaks
## 8      A549_H3K27ac_peaks,A549_H3K4me3_peaks,A549_H3K9ac_peaks,A549_PolIII_peaks
## 9      A549_H3K27ac_peaks,A549_H3K4me3_peaks,A549_H3K9ac_peaks,A549_PolIII_peaks
## 10     A549_H3K27ac_peaks,A549_H3K4me3_peaks,A549_H3K9ac_peaks,A549_PolIII_peaks
##      ref_count alt_count ref_cnv alt_cnv  p.val.raw  p.val.cnv p.val.cnv.bh
## 1         48         36    1501    1558 0.03004691 0.05533592    0.5533592
## 2         29         39    1501    1558 0.30745625 0.35532555    0.8862402
## 3         45         46    1501    1558 0.44520467 0.35803028    0.8862402
## 4         27         15    1754      929 0.26243754 0.40944408    0.8862402
## 5         29         39    1501    1558 0.41010272 0.49624781    0.8862402
## 6         30          8    1754      929 0.21102360 0.67303236    0.8862402
## 7         48         36    1501    1558 0.84537032 0.70660674    0.8862402
## 8         24         38    1501    1558 0.90065325 0.78816140    0.8862402
## 9         42         55    1501    1558 1.00000000 0.88474339    0.8862402
## 10        40         53    1501    1558 1.00000000 0.88624024    0.8862402
##      p.val.cnv.bonf
## 1         0.5533592
## 2         1.0000000
## 3         1.0000000
## 4         1.0000000
## 5         1.0000000
## 6         1.0000000
## 7         1.0000000
## 8         1.0000000
## 9         1.0000000
## 10        1.0000000

```