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- 2 Findings from the 2023 Serosurveillance Summit
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Abstract

55 Multiplex-based serological surveillance is a valuable but underutilized tool to 56 understand gaps in population-level exposure, susceptibility, and immunity to 57 infectious diseases. Assays for which blood samples can be tested for antibodies 58 against several pathogens simultaneously, such as multiplex bead immunoassays, 59 can more efficiently integrate public health surveillance in low- and middle-income 60 countries. From March 7-8, 2023, a group of experts representing research institutions, multilateral organizations, private industry, and country partners met 61 62 to discuss experiences, identify challenges and solutions, and create a community 63 of practice for integrated, multi-pathogen serosurveillance using multiplex bead 64 assay technologies. Participants were divided into six working groups: (1) supply 65 chain; (2) laboratory assays; (3) seroepidemiology; (4) data analytics; (5) 66 sustainable implementation; and (6) use case scenarios. These working groups discussed experiences, challenges, solutions, and research needs to facilitate

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integrated, multi-pathogen serosurveillance for public health. Several solutions were proposed to address challenges that cut across working groups.

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Key words: serological surveillance; multiplex bead immunoassay; integrated surveillance; infectious diseases; serology

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Introduction

The World Health Organization (WHO) recently introduced collaborative surveillance as one of five interconnected components of health emergency preparedness, response, and resilience. A core objective of collaborative surveillance is to break down siloed disease surveillance systems and replace them with a collaborative and integrated system across diseases, public and private sectors, and administrative levels.

Serological surveillance, or serosurveillance, complements traditional public health surveillance for infectious diseases through the collection and analysis of specimens (e.g., serum, blood, or oral fluid) to measure antibodies to pathogens and estimate population-level exposure, susceptibility, and immunity to infectious diseases.³ This information can guide public health policies and programs for the control and elimination of several communicable diseases, including vaccine-preventable diseases (VPDs), neglected tropical diseases (NTDs), and emerging infectious diseases (EIDs), among others. While serosurveillance has been used for decades, the COVID-19 pandemic amplified interest in serology.⁴

The development of technologies like multiplex bead immunoassays (MBIAs), which allow for the simultaneous detection of antibodies to more than one pathogen in a single assay, rapidly advanced the ability to efficiently conduct integrated, multipathogen serosurveillance.^{3,5} These technologies enable health systems to monitor exposure, susceptibility, and immunity to multiple pathogens with limited additional resources compared to using single-pathogen assays.⁶

MBIAs have been developed for detecting antibodies against a range of pathogens including VPDs⁷⁻⁹ respiratory pathogens;¹⁰ NTDs;¹¹⁻¹⁵ malaria;^{16,17} sexually transmitted infections;¹⁸ EIDs;¹⁹ arboviruses;²⁰ and SARS-CoV-2.²¹ Integrating

110 serosurveillance across multiple pathogens could efficiently leverage financial 111 resources and personnel as well as metadata obtained from questionnaires. 112 Integration with other surveillance programs (e.g., diagnostic, syndromic, and 113 wastewater surveillance)³ could further improve the efficiency of surveillance 114 systems to control the spread of disease. 115 Despite the MBIA being a powerful tool for public health surveillance, several 116 barriers have prevented the widespread adoption of these technologies, 117 particularly in low- and middle-income countries (LMICs), where they might be 118 especially useful.²² Through partnerships with academic and government research 119 institutions, multi-pathogen serosurveillance has been conducted in sub-Saharan Africa, 23,24 Asia, 3,25,26 and the Americas. 22,27,28 CDC and the Pan American Health 120 121 Organization developed guidance for program managers to design and conduct integrated multi-pathogen serosurveillance.²⁸ 122 123 To discuss the opportunities and complexities with establishing integrated, multi-124 pathogen serosurveillance, experts from the Collaboration on Integrated 125 Biomarkers Surveillance (CIBS) met in 2018 to catalog pathogens for inclusion in 126 multiplex serological assays, lay out objectives for an integrated platform, identify potential use cases, and discuss advocacy. ²⁹ Building on this work, in March 2023, 127 128 the International Vaccine Access Center at the Johns Hopkins Bloomberg School of 129 Public Health, with support from the Bill and Melinda Gates Foundation (BMGF) and 130 collaborators at the Center for Global Health at the University of Colorado Anschutz 131 Medical Campus, convened a serosurveillance summit that further explored topics 132 related to the use of MBIAs for multi-pathogen serosurveillance.

Serosurveillance Summit Meeting

134 Experts across a range of institutions and fields participated, including researchers, 135 multilateral organizations, country partners, private sector companies, and supply 136 chain organizations. Participants were divided into two of six working groups: (1) 137 supply chain, (2) laboratory assays, (3) seroepidemiology, (4) data analytics, (5) 138 sustainable implementation, and (6) use case scenarios. The objectives of the 139 workshop were to share experiences establishing integrated, multi-pathogen 140 serosurveillance with a focus on MBIAs; identify key challenges, potential solutions, 141 and research needs for integrated serosurveillance using MBIAs; and establish a 142 community of practice of technical experts. 143 144 145 Figure 1: A framework depicting key steps for establishing a sustainable 146 integrated serosurveillance system. The steps correspond with the six working 147 groups, except for using data for program action which is part of the sustainable 148 implementation working group. 149 150 **Experiences and challenges** 151 Participants shared their experiences with multi-pathogen serosurveillance (Annex 152 1). Based on these discussions, key issues that countries using multiplex 153 serosurveillance encounter were outlined. 154 155 Public health questions and use cases 156 Because multi-pathogen surveillance involves multiple programs and partners, it 157 can be challenging to generate buy-in from all stakeholders and identify potential 158 programmatic impact early in the planning process. As more pathogens are 159 included, additional groups working on different diseases will need to be engaged.

Programmatic or research questions were framed as "use cases" for which

serosurveillance is most likely to add value to existing surveillance systems. Table

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162 1 presents five of the most common use cases identified and linked target
163 pathogen(s) of interest. Several groups of pathogens were considered most
164 relevant, including VPDs, emerging pathogens, NTDs, and other pathogens
165 associated with high disease burden.
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Table 1. Use Cases for Multiplex Serosurveillance

Use Cases and a Representative Example	Example Pathogens
Estimating the burden and distribution of infections to complement or fill gaps in existing surveillance systems ³⁰	NTDs Trypanosoma cruzi (Chagas disease), chikungunya virus, Taenia solium (cysticercosis), Strongyloides spp., Treponema pallidum subspecies pertenue (yaws) Enteric pathogens Campylobacter spp., Vibrio cholerae, Cryptosporidium, Giardia Malaria Plasmodium species HIV Respiratory viruses respiratory syncytial virus (RSV), influenza virus
Identifying emerging and reemerging infections ³¹	Filoviruses Ebolaviruses, Marburg virus Other viruses Lassa virus, mpox virus SARS-CoV-2, Zika virus
Identifying vaccine program reach or gaps and geographic or demographic gaps ³²	Childhood diseases measles virus, polioviruses, rubella virus, diphtheria, tetanus, pertussis Other viruses SARS-CoV-2, yellow fever virus
Assessing changes in pathogen exposure due to behavioral, environmental, or (non-) pharmaceutical interventions or environmental changes ³³	NTDs chikungunya virus, dengue virus, lymphatic filariasis Bacteria Streptococcus pneumoniae spp., Salmonella serotype Typhi (Typhoid) Malaria Plasmodium species
Monitoring peri- and post-elimination	NTDs Chlamydia trachomatis (trachoma), Dracunculus medinensis

settings for diseases with elimination goals³⁴

(Guinea worm), *Leishmania* spp. (visceral leishmaniasis), nematodes (lymphatic filariasis), *Onchocerca volvulus* (onchocerciasis), *Treponema pallidum* subspecies *pertenue* (yaws), *Trypanosoma brucei* (human African trypanosomiasis)

Vaccine-preventable diseases polioviruses

Malaria

Plasmodium spp. (sub-national levels)

Study design

A key issue is identifying the appropriate target population for the question of interest. Because pathogens affect individuals across different demographic characteristics (e.g., age), selecting a target population to cover all pathogens of interest is challenging. Other challenges include defining population sampling strategies and sample sizes to answer multiple questions simultaneously and determining the optimal survey frequency to measure temporal trends across pathogens and address programmatic needs. One viable option includes nesting specimen collection within existing surveys that sample large populations across multiple characteristics (e.g., Demographic and Health Surveys, Multiple Indicator Cluster Surveys). Less resource-intensive sampling strategies like using residual blood specimens (e.g., from health facility laboratories or routine screening of pregnant women) can reduce costs while still capturing a range of populations and time points. Additionally, specimen type (e.g. dried blood spots, venous blood and oral fluid) must balance feasibility of collection and assay validity across all pathogens.

Supplies

One of the biggest challenges raised by multiple groups was related to the supply chain for assay reagents and equipment. Challenges included procuring, maintaining, and repairing platform technology, such as Luminex instruments. Procuring quality-assured beads and assay reagents in a space with few small-scale producers of conjugated beads is also an issue. Other challenges with supply chain sustainability involve understanding and addressing country-specific limitations to importation, maintaining the cold chain, and establishing procurement procedures and processes for reagents and supplies.

Laboratory assays

For multi-pathogen serosurveillance, several issues related to assay development, antigen discovery and validation, identification of proper controls, and assay performance must be addressed. Many research groups have developed standard operating procedures (SOPs) for equipment maintenance, assay techniques, assay development, quality control, and other key multiplex serology operations.

However, resources such as positive and negative controls are not always available and require continuous support to laboratories for appropriate use. Furthermore, quality controls to validate assay runs and track assay performance over time are needed. Standardization holds the promise to make serosurveillance results more comparable between laboratories, but this is currently hampered by the limited availability of reference standards. Available reference reagents are typically calibrated for a specific pathogen, not for a broad range of them. Consequently, to report international units, standards need to be calibrated against each other or an in-house standard must be built and calibrated against multiple standards.

Developing assays requires knowledge of the immunogenicity of antigens, kinetics of antibody responses, and relevance for assessing disease burden or protective immunity (i.e., correlates of immune protection). Because assay development can be technically challenging and requires thorough validation, the number of laboratories that currently undertake assay development is limited. High demand and inadequate capacity limit the availability of antigen-coupled beads, which are currently produced by a small number of research groups, although technology transfer initiatives are ongoing. For countries considering implementing multiplex serosurveillance but unable to develop assays, commercial options are also limited.

Data analysis and interpretation

There are currently no standardized approaches to cleaning raw laboratory data and establishing seropositivity thresholds, as this varies by antigen, the availability of controls, and population. This first stage of analysis includes performing quality control checks, evaluating serial dilution standard curves, and ensuring steps to normalize data are appropriately applied. Translating cleaned data into useful epidemiologic inference requires analytical approaches that model the normalized quantitative values or the establishment of reasonable cut-points for seropositivity that are relevant for the specific use case. Additionally, the interpretation of pathogen-specific age patterns and geographic distributions of seroprevalence requires supplementing laboratory data with demographic characteristics and contextualized epidemiologic understanding of the specific pathogens. The use of serological data for computing age seroprevalence curves and estimating epidemiological parameters, such as forces of infection, is established for well-characterized antigens. A key remaining challenge is selecting and implementing

appropriate analytical approaches (and metadata) to answer questions of interest for less-well-characterized antigens as well as across multiple antigens or pathogens simultaneously. Further, there is a need for data analytic pipelines to facilitate the interpretation of data by balancing detail and complexity and producing user-friendly data visualizations such as spatial maps.

Data for program action

Given the complexities of analyzing multiple pathogens simultaneously, it can be challenging to generate interpretable and easily visualized results for target audiences such as policy makers and program managers. The time from serosurvey data collection to dissemination of results is often long (months), which makes it challenging to use results for decision making. Serosurvey data should be triangulated with other data sources for interpretation of the epidemiological findings for each disease. When serosurveillance for several pathogens are simultaneously analyzed and presented, it can be challenging to contextualize all findings in a succinct, clear manner.

Cross-cutting challenges

Training needs included supply chain logistics and procurement so laboratories can place their supply orders and anticipate shortages. Similarly, training on instrument use and routine maintenance are needed to sustain high instrument performance and minimize downtime and costly repairs. There were also challenges with technology transfer, including training on the MBIA, bead coupling and validation, and quality control procedures. Additionally, there is a strong need for training in data analytics, as multi-pathogen serosurveillance data are complex and most

useful when combined with complementary data such as vaccination coverage or
case-based disease surveillance data.

Table 2 summarizes challenges identified in each working group. Additional details
are available in the full meeting report.

are available in the full meeting report.

Table 2. Key challenges identified by the working groups

Working Group	Key Challenges
Supply Chain	Procuring and maintaining appropriate platform technology, producing and procuring quality-assured beads and assays, commercializing kits, maintaining the cold chain, understanding and addressing country-specific limitations to importation, and limited human and technological capacity to anticipate and avoid supply chain issues
Seroepidemiolog y	Selecting sample populations and sample sizes, establishing the frequency of sampling, identifying and validating less resource-intensive sampling strategies, defining sampling approaches that answer multiple questions, determining core individual- and household-level data to collect, and linking serosurvey antigens to study design for programmatic impact
Laboratory Assays	Supporting technology transfer and training, sharing best practices and protocols, standardizing antigen use across countries, defining quality control standards, developing reference reagents
Data Analytics	Standardizing and cleaning raw laboratory data, translating cleaned data into useful epidemiologic inference by selecting appropriate analytical approaches to answer questions of interest, and developing user friendly analytical and visualization pipelines
Sustainable implementation	Demonstrating added value for initial engagement, generating buy-in across national health systems, ensuring adequate laboratory capacity and procurement, and interpreting data and integrating results for decision making

Proposed Solutions Across Working Groups

275 Creation of an electronic platform to share resources and expertise
 276 There is a strong need for information sharing platforms. New tools and resources
 277 needed to support multiplex serosurveillance include a planning tool that would

allow users to prepare for resources needed and estimate cost; a supply chain "playbook" that details cold chain and labeling requirements and substitutable reagents; protocols that enable adaptive sampling strategies; and ethical considerations for additional testing in serosurveillance studies. Furthermore, case studies demonstrating how countries have used serosurveillance to guide public health actions would help underscore the value of serosurveillance. Additionally, the platform could aid in standardizing pre-processing pipelines between studies and harmonizing data.

A unified platform could host these tools and other resources such as a central repository for antigens and standards, protocols, data packages and scripts, sample size calculation tools, best practices, training resources, and plain-language policy briefs and technical documents. Critically, this platform should also allow users to communicate with one another to troubleshoot problems and share experiences. Creating a single platform to address these needs could help develop and sustain a culture of collaboration while facilitating harmonization efforts where practical.

Building local capacity and training

Expanding the capacity for MBIAs in low-resourced settings will help generate data where it is most needed. Enhanced capacity could also create a more favorable environment for commercialization, enable greater collaboration and country ownership, promote harmonization, and address key bottlenecks. Several areas were identified as priorities for capacity building including the development of regional hubs and use of multiple training approaches such as on-site training,

online training, and train-the-trainer initiatives. These approaches could enable users to perform MBIAs, service and troubleshoot bead-based multiplex platforms, and produce or procure antigen-coupled beads. While discussion of capacity building focused on LMICs, many areas were relevant for users in all countries. Developing quality control and standardized approaches Exploring ways to standardize approaches would allow for comparison of results across countries. However, harmonization can be challenging. Targets for standardization include standardizing approaches to conducting serosurveys; creating or procuring quality assay materials; and best practices for cleaning, analyzing, and presenting data. Developing and validating positive and negative reference controls by antigen (e.g., through the United Kingdom's National Institute for Biological Standards and Control [NIBSC] or using validated recombinant antibodies) would lead to results that are more interpretable across assays, populations, and time-points. Developing a common panel with the most frequently used antigens across regions could also facilitate cross-country comparisons, though customization would still be needed to address country-specific priorities. Establishing a laboratory network and building partnerships A laboratory support network would facilitate knowledge sharing and troubleshooting at country, regional, and global levels, helping to connect laboratory groups. Partnering with private companies would support commercialization of panels and sharing of know-how regarding supply chain constraints. Partnering with supply chain experts would enable procurement and

packaging of common reagents and materials to streamline ordering processes and

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avoid delays caused by stockouts. Regional networks could also allow groups to share limited resources—including access to instruments and materials like antigen-coupled beads—and to pool demand for these resources. Regional hubs could be characterized by function (e.g., coupling antigens to beads and providing quality control) to help meet the needs of different groups, further building a collaborative network.

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Generating political buy-in for multiplex serosurveillance

Participants viewed the establishment of buy-in from governments, funders, and regulatory agencies as essential for the introduction and scale-up of multipathogen serosurveillance. Approaches to achieving support and fostering greater participation from these entities include exploring standardized approval processes for the importation of products necessary for multi-pathogen serosurveillance, developing a taxonomy of pathogen-specific antigens paired to scientific and policy-relevant use cases, involving governmental agencies in training initiatives, and developing analytical and visualization pipelines to aid understanding. Garnering high level regional and international support to develop guidance and recommendations for the implementation and use of integrated serosurveillance was considered a priority. Organizations like PAHO and the US CDC have developed documents that were discussed as starting points.²⁸ This goal could be supported through conversations with decision makers to demonstrate how integrated, multipathogen serosurveillance can complement existing disease surveillance systems and by providing successful case studies. Generating community buy-in through communication of the benefits and limitations of serosurveillance is also critical as exploring the value of integrated serosurveillance hinges on their participation.

Discussion

Serosurveillance provides valuable information to guide public health programs, especially when triangulated with data from other surveillance systems. In isolation, serosurveillance systems are costly to establish and sustain. 28,36 Serosurveillance data is underutilized due to the heterogeneity of assays and the delay in disseminating results to health authorities for meaningful program impact. 37,38 Ideally, integration of serosurveillance with routine public health activities can reduce costs and make it more sustainable but requires sufficient buy-in and funding. 6,14,23,39 The lessons learned from experiences establishing serosurveillance across multiple countries should be shared to promote further investment in this technology.

For serosurveillance to have programmatic impact, data must be available in a timely fashion. Several bottlenecks cause delays: planning epidemiologically relevant serosurveys; procuring materials and equipment; and cleaning, analyzing, and interpreting data.²⁹ Some approaches, such as developing standard operating procedures, addressing supply chain issues, optimizing data analysis pipelines, training local health researchers, and sharing preliminary results with decision makers can shorten the time for data to be used for action.²² Timely serosurveillance data provides insights into disease transmission patterns and population vulnerability to outbreaks to guide control and elimination strategies.

Financial, technical, and political support are also needed to overcome these bottlenecks. For example, the development of a commercial panel for frequently

tested antibodies could address supply chain constraints, but commercialization restricts flexibility to modify the pathogens that can be tested. For commercialization of panels, there will need to be sufficient demand. Without adequate resources, serosurveillance efforts may only be pilots or ad hoc endeavors. Investment in the development of country-led multi-pathogen serosurveillance systems, like PAHO's, 28 can expand the number of countries conducting multi-pathogen serosurveillance.

In addition to the use cases presented, there are additional questions of public health importance that could be explored (e.g., optimizing vaccination schedules). Recently, the most common use case was measuring the spread of SARS-CoV-2; seroprevalence studies were conducted in 149 countries.⁴ This allowed tracking the spread of the virus, identifying transmission dynamics, monitoring population immunity, and evaluating vaccine program performance.^{40,41} Leveraging the capacity building, networking, platforms, and expertise developed during the COVID-19 pandemic, could better prepare us for the next emerging pathogen and support surveillance systems for diseases that are underfunded.

The global response to the COVID-19 pandemic also demonstrated the power of coordination across institutions. Monitoring seroprevalence and population immunity in different settings harnessed learnings across the globe. Though harmonized approaches were feasible for SARS-CoV-2 and allowed for cross-country comparisons, many pathogens need additional research to allow for such comparisons. Some vaccine-preventable diseases like measles and rubella already have standardized international controls, agreed upon correlates of protection,

existing laboratory networks, and clear programmatic actions that can be informed by serological data. As multiplex panels are developed for different pathogens, similar standardization could enable results to be more readily compared across settings. Although VPDs are an area where standardization is within reach, achieving this aim across a diverse array of pathogens–especially considering the unique epidemiological profiles and priorities of different countries–will require more developed serosurveillance systems and international coordination.

While multi-pathogen serosurveillance has traditionally been used in high-income countries (HICs),⁴³ it has also been used in LMICs, often with a high degree of technical support from organizations based in HICs. Some studies include samples from LMICs that were tested entirely in an HIC,⁴⁴ in both LMICs and HICs,^{21,45,45} and entirely in an LMIC.^{23,46-49} To ensure the promotion of country ownership, initiatives are needed to build local capacity to couple beads, perform MBIAs, and analyze data that is coordinated with national governments and aligned with their priorities. More recently, efforts to transfer technology and build capacity in countries in the Americas^{22,28} and Africa²³ have paved the way for future endeavors to scale-up multiplex serosurveillance. To move towards routine serosurveillance globally, additional funding is needed to fill research gaps and advance implementation in additional settings, including bolstering capacity in laboratories that do not yet have the technologies used in multi-pathogen serosurveillance.

Building on the momentum from previous efforts, the 2023 Serosurveillance Summit provides further impetus to advance collaboration among countries to conduct multi-pathogen serosurveillance. Participants will continue serving on working groups to put into practice the proposed solutions outlined above. This community of practice brings together a network of scientists and practitioners to facilitate knowledge sharing and develop a platform for multi-pathogen, multi-country serosurveillance. These established networks and relationships could facilitate rapid response efforts for future emerging pathogens. As the world moves to reclaim the progress against infectious diseases that was disrupted by the COVID-19 pandemic-and to enhance preparedness to prevent or mitigate the next pandemic-the appetite for establishing multi-pathogen serosurveillance systems has never been greater.

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470 References

- 471 1. World Health Organization., 2023. Defining collaborative surveillance: a core
- concept for strengthening the global architecture for health emergency
- preparedness, response, and resilience (HEPR). World Health Organization
- 474 2. Archer BN, Abdelmalik P, Cognat S, Grand PE, Mott JA, Pavlin BI, Barakat A,
- Dowell SF, Elmahal O, Golding JP, Gongal G, Hamblion E, Hersey S, Kato M, Koua
- 476 EL, et al., 2023. Defining collaborative surveillance to improve decision making
- for public health emergencies and beyond. The Lancet 401: 1831–1834
- 478 3. Arnold BF, Scobie HM, Priest JW, Lammie PJ., 2018. Integrated Serologic
- Surveillance of Population Immunity and Disease Transmission. Emerg Infect Dis 24: 1188–1194
- 481 4. Arora RK, Joseph A, Wyk JV, Rocco S, Atmaja A, May E, Yan T, Bobrovitz N,
- Chevrier J, Cheng MP, Williamson T, Buckeridge DL., 2021. SeroTracker: a global
- 483 SARS-CoV-2 seroprevalence dashboard. Lancet Infect Dis 21: e75-e76
- 484 5. Elshal MF, McCoy JP., 2006. Multiplex bead array assays: performance evaluation
- and comparison of sensitivity to ELISA. Methods San Diego Calif 38: 317–323
- 486 6. Solomon AW, Engels D, Bailey RL, Blake IM, Brooker S, Chen J-X, Chen J-H,
- Churcher TS, Drakeley CJ, Edwards T, Fenwick A, French M, Gabrielli AF, Grassly
- 488 NC, Harding-Esch EM, et al., 2012. A Diagnostics Platform for the Integrated
- Mapping, Monitoring, and Surveillance of Neglected Tropical Diseases: Rationale
- and Target Product Profiles. PLoS Negl Trop Dis 6: e1746
- 491 7. Dhiman N, Jespersen DJ, Rollins LO, Harring JA, Beito EM, Binnicker MJ., 2010.
- Detection of IgG-class antibodies to measles, mumps, rubella, and varicella-
- zoster virus using a multiplex bead immunoassay. Diagn Microbiol Infect Dis 67: 346–349
- 495 8. Hayford K, Mutembo S, Carcelen A, Matakala HK, Munachoonga P, Winter A,
- Wanyiri JW, Searle K, Mwansa FD, Mwiche A, Phiri C, Book C, Thuma PE, Moss
- 497 WJ., 2019. Measles and rubella serosurvey identifies rubella immunity gap in
- 498 young adults of childbearing age in Zambia: The added value of nesting a
- serological survey within a post-campaign coverage evaluation survey. Vaccine 37: 2387–2393
- 501 9. de Voer RM, Schepp RM, Versteegh FGA, van der Klis FRM, Berbers GAM., 2009.
- 502 Simultaneous Detection of Haemophilus influenzae Type b Polysaccharide-
- 503 Specific Antibodies and Neisseria meningitidis Serogroup A, C, Y, and W-135
- 504 Polysaccharide-Specific Antibodies in a Fluorescent-Bead-Based Multiplex
- 505 Immunoassay. Clin Vaccine Immunol CVI 16: 433-436
- 506 10. Mahony J, Chong S, Merante F, Yaghoubian S, Sinha T, Lisle C, Janeczko R., 2007.
- 507 Development of a respiratory virus panel test for detection of twenty human
- respiratory viruses by use of multiplex PCR and a fluid microbead-based assay. J
- 509 Clin Microbiol 45: 2965-2970
- 510 11. Kim JS, Oldenburg CE, Cooley G, Amza A, Kadri B, Nassirou B, Cotter SY, Stoller
- NE, West SK, Bailey RL, Keenan JD, Gaynor BD, Porco TC, Lietman TM, Martin DL.,
- 512 2019. Community-level chlamydial serology for assessing trachoma elimination
- in trachoma-endemic Niger. PLoS Negl Trop Dis 13: e0007127
- 514 12. Martin DL, Bid R, Sandi F, Goodhew EB, Massae PA, Lasway A, Philippin H,
- Makupa W, Molina S, Holland MJ, Mabey DCW, Drakeley C, Lammie PJ, Solomon
- 516 AW., 2015. Serology for trachoma surveillance after cessation of mass drug
- administration. PLoS Negl Trop Dis 9: e0003555
- 518 13. Goodhew EB, Morgan SMG, Switzer AJ, Munoz B, Dize L, Gaydos C, Mkocha H,
- West SK, Wiegand RE, Lammie PJ, Martin DL., 2014. Longitudinal analysis of

- antibody responses to trachoma antigens before and after mass drug administration. BMC Infect Dis 14: 216
- 522 14. Priest JW, Jenks MH, Moss DM, Mao B, Buth S, Wannemuehler K, Soeung SC,
- Lucchi NW, Udhayakumar V, Gregory CJ, Huy R, Muth S, Lammie PJ., 2016.
- 524 Integration of Multiplex Bead Assays for Parasitic Diseases into a National,
- Population-Based Serosurvey of Women 15-39 Years of Age in Cambodia. PLoS Negl Trop Dis 10: e0004699
- 527 15. Lima MM, Sarquis O, de Oliveira TG, Gomes TF, Coutinho C, Daflon-Teixeira NF,
- Toma HK, Britto C, Teixeira BR, D'Andrea PS, Jansen AM, Bóia MN, Carvalho-Costa FA., 2012. Investigation of Chagas disease in four periurban areas in
- northeastern Brazil: epidemiologic survey in man, vectors, non-human hosts and reservoirs. Trans R Soc Trop Med Hyg 106: 143–149
- 532 16. Rogier E, van den Hoogen L, Herman C, Gurrala K, Joseph V, Stresman G,
- Presume J, Romilus I, Mondelus G, Elisme T, Ashton R, Chang M, Lemoine JF,
- Druetz T, Eisele TP, et al., 2019. High-throughput malaria serosurveillance using a one-step multiplex bead assay. Malar J 18: 402
- 536 17. Rogier E, Moss DM, Chard AN, Trinies V, Doumbia S, Freeman MC, Lammie PJ.,
- 537 2017. Evaluation of Immunoglobulin G Responses to Plasmodium falciparum and
- Plasmodium vivax in Malian School Children Using Multiplex Bead Assay. Am J Trop Med Hvg 96: 312–318
- 540 18. Yufenyuy EL, Vedapuri S, Zheng A, Cooley G, Danavall D, Mayur S, Kodani M, Chen C, Tun Y, Fakile YF, Martin D, Kamili S, Karem K, Parekh BS., 2022.
- Development of a Bead-Based Multiplex Assay for Use in Multianalyte Screening
- and Surveillance of HIV, Viral Hepatitis, Syphilis, and Herpes. J Clin Microbiol 60: 60234821
- 545 19. Rogier E, Plucinski M, Candrinho B, Moss DM, Gibbons A, Colborn J, Higgins J,
- Chambe G, Muchanga J, Muguande O, Matsinhe G, Mathe G, Doyle T, Zulliger R,
- Saifodine A, et al., 2022. Adaptation to a Multiplex Bead Assay and
- Seroprevalence to Rift Valley Fever N Protein: Nampula Province, Mozambique, 2013-2014. J Virol 96: e0067222
- 550 20. Bailly S, Rousset D, Fritzell C, Hozé N, Ben Achour S, Berthelot L, Enfissi A,
- Vanhomwegen J, Salje H, Fernandes-Pellerin S, Saout M, Lavergne A, Manuguerra
- J-C, Carod J-F, Djossou F, et al., 2021. Spatial Distribution and Burden of Emerging Arboviruses in French Guiana. Viruses 13: 1299
- 554 21. Gwyn S, Abubakar A, Akinmulero O, Bergeron E, Blessing UN, Chaitram J,
- Coughlin MM, Dawurung AB, Dickson FN, Esiekpe M, Evbuomwan E, Greby SM,
- 556 Iriemenam NC, Kainulainen MH, Naanpoen TA, et al., 2022. Performance of
- 557 SARS-CoV-2 Antigens in a Multiplex Bead Assay for Integrated Serological
- Surveillance of Neglected Tropical and Other Diseases. Am J Trop Med Hyg 107: 260–267
- 560 22. Saboyá-Díaz M-I, Castellanos LG, Morice A, Ade MP, Rey-Benito G, Cooley GM,
- Scobie HM, Wiegand RE, Coughlin MM, Martin DL., 2023. Lessons learned from
- the implementation of integrated serosurveillance of communicable diseases in the Americas. Rev Panam Salud Pública 47: e53
- 564 23. Njenga SM, Kanyi HM, Arnold BF, Matendechero SH, Onsongo JK, Won KY, Priest
- JW., 2020. Integrated Cross-Sectional Multiplex Serosurveillance of IgG Antibody
- Responses to Parasitic Diseases and Vaccines in Coastal Kenya. Am J Trop Med Hyg 102: 164–176
- 568 24. Arzika AM, Maliki R, Goodhew EB, Rogier E, Priest JW, Lebas E, O'Brien KS, Le V, Oldenburg CE, Doan T, Porco TC, Keenan JD, Lietman TM, Martin DL, Arnold BF.,

- 570 2022. Effect of biannual azithromycin distribution on antibody responses to 571 malaria, bacterial, and protozoan pathogens in Niger. Nat Commun 13: 976
- 572 25. Feldstein LR, Bennett SD, Estivariz CF, Cooley GM, Weil L, Billah MM, Uzzaman 573 MS, Bohara R, Vandenent M, Adhikari JM, Leidman E, Hasan M, Akhtar S, Hasman
- 574 A, Conklin L, et al., 2020. Vaccination coverage survey and seroprevalence
- 575 among forcibly displaced Rohingya children, Cox's Bazar, Bangladesh, 2018: A 576 cross-sectional study. PLoS Med 17: e1003071
- 577 26. Cooley GM, Feldstein LR, Bennett SD, Estivariz CF, Weil L, Bohara R, Vandenent 578 M, Mainul Hasan A, Akhtar MS, Uzzaman MS, Billah MM, Conklin L, Ehlman DC,
- 579 Asiedu K, Solomon AW, et al., 2021. No Serological Evidence of Trachoma or
- 580 Yaws Among Residents of Registered Camps and Makeshift Settlements in Cox's 581 Bazar, Bangladesh. Am J Trop Med Hyg 104: 2031-2037
- Pan American Health Organization., 2020. Multiplex Bead Assay for Integrated 582 27. Serological Surveillance of Communicable Diseases in the Region of the 583
- 584 Americas. Report of the third regional meeting (Cuernavaca, 4-5 March 2020) -585 PAHO/WHO | Pan American Health Organization
- 586 28. Anon. Toolkit for Integrated Serosurveillance of Communicable Diseases in the 587 Americas - PAHO/WHO | Pan American Health Organization. Available at:
- 588 https://www.paho.org/en/documents/toolkit-integrated-serosurveillance-
- 589 communicable-diseases-americas. Accessed
- 590 29. Wiens KE, Jauregui B, Arnold BF, Banke K, Wade D, Hayford K, Denis AC-S, Hall RH. Salie H. Rodriguez-Barraguer I, Azman AS, Vernet G, Leung DT, Surveillance 591 592 on behalf of the C on IB., 2022. Building an integrated serosurveillance platform
- 593 to inform public health interventions: Insights from an experts' meeting on
- 594 serum biomarkers. PLoS Negl Trop Dis 16: e0010657
- 595 30. Salie H, Paul KK, Paul R, Rodriguez-Barraguer I, Rahman Z, Alam MS, Rahman M, 596 Al-Amin HM, Heffelfinger J, Gurley E., 2019. Nationally-representative serostudy 597 of dengue in Bangladesh allows generalizable disease burden estimates. eLife 8: 598 e42869
- 599 31. Basto-Abreu A, Carnalla M, Torres-Ibarra L, Romero-Martínez M, Martínez-
- 600 Barnetche J, López-Martínez I, Aparicio-Antonio R, Shamah-Levy T, Alpuche-Aranda C, Rivera JA, Barrientos-Gutierrez T., 2022. Nationally representative 601
- 602 SARS-CoV-2 antibody prevalence estimates after the first epidemic wave in
- 603 Mexico. Nat Commun 13: 589
- 604 32. Murhekar MV, Gupta N, Hasan AZ, Kumar MS, Kumar VS, Prosperi C, Sapkal GN,
- 605 Thangaraj JWV, Kaduskar O, Bhatt V, Deshpande GR, Thankappan UP, Bansal AK,
- Chauhan SL, Grover GS, et al., 2022. Evaluating the effect of measles and rubella 606 607 mass vaccination campaigns on seroprevalence in India: a before-and-after
- 608 cross-sectional household serosurvey in four districts, 2018-2020. Lancet Glob 609 Health 10: e1655-e1664
- 610 33. Plucinski MM, Candrinho B, Chambe G, Muchanga J, Muguande O, Matsinhe G,
- 611 Mathe G, Rogier E, Doyle T, Zulliger R, Colborn J, Saifodine A, Lammie P, Priest
- 612 JW., 2018. Multiplex serology for impact evaluation of bed net distribution on
- burden of lymphatic filariasis and four species of human malaria in northern 613
- 614 Mozambique. PLoS Negl Trop Dis 12: e0006278
- 615 34. Oguttu D, Byamukama E, Katholi CR, Habomugisha P, Nahabwe C, Ngabirano M,
- Hassan HK, Lakwo T, Katabarwa M, Richards FO, Unnasch TR., 2014. 616
- 617 Serosurveillance to Monitor Onchocerciasis Elimination: The Ugandan
- Experience. Am J Trop Med Hyg 90: 339-345 618

- 619 35. Carcelen, Andrea, Kong, Alex, Hegde, Sonia, Takahashi, Saki, Moss, William.,
- 620 2023. Serosurveillance Summit Meeting Report. Baltimore, Maryland, USA: Johns 621
- Hopkins Bloomberg School of Public Health International Vaccine Access Center 622 36. Turner HC, Wills BA, Rahman M, Quoc Cuong H, Thwaites GE, Boni MF, Clapham
- HE., 2018. Projected costs associated with school-based screening to inform 623
- 624 deployment of Dengyaxia: Vietnam as a case study. Trans R Soc Trop Med Hyg 112: 369-377 625
- 626 37. Donnici C, Ilincic N, Cao C, Zhang C, Deveaux G, Clifton D, Buckeridge D, 627 Bobrovitz N, Arora RK., 2022. Timeliness of reporting of SARS-CoV-2
- 628 seroprevalence results and their utility for infectious disease surveillance.
- 629 Epidemics 41: 100645
- 630 38. Fink RV, Fisher L, Sulaeman H, Dave H, Levy ME, McCann L, Di Germanio C,
- 631 Notari IV EP, Green V, Cyrus S, Williamson P, Saa P, Haynes JM, Groves J, Mathew
- 632 S, et al., 2022. How do we...form and coordinate a national serosurvey of SARS-
- 633 CoV-2 within the blood collection industry? Transfusion (Paris) 62: 1321-1333
- 634 39. Lammie PJ, Moss DM, Brook Goodhew E, Hamlin K, Krolewiecki A, West SK, Priest
- 635 IW., 2012. Development of a new platform for neglected tropical disease 636 surveillance. Int J Parasitol 42: 797-800
- 637 40. Murhekar MV, Kumar MS, Kamaraj P, Khan SA, Allam RR, Barde P, Dwibedi B,
- Kanungo S, Mohan U, Mohanty SS, Roy S, Sagar V, Savargaonkar D, Tandale BV, 638
- 639 Topno RK, et al., 2020. Hepatitis-B virus infection in India: Findings from a
- 640 nationally representative serosurvey, 2017-18. Int | Infect Dis 100: 455-460
- 641 41. Knezevic I, Mattiuzzo G, Page M, Minor P, Griffiths E, Nuebling M, Moorthy V.,
- 642 2022. WHO International Standard for evaluation of the antibody response to
- 643 COVID-19 vaccines: call for urgent action by the scientific community. Lancet 644 Microbe 3: e235-e240
- 645 42. den Hartog G, van Binnendijk R, Buisman A-M, Berbers GAM, van der Klis FRM.,
- 646 2020. Immune surveillance for vaccine-preventable diseases. Expert Rev
- 647 Vaccines 19: 327-339
- 648 43. Hoes J, Knol MJ, Mollema L, Buisman A, de Melker HE, van der Klis FRM., 2019.
- 649 Comparison of antibody response between boys and girls after infant and
- 650 childhood vaccinations in the Netherlands. Vaccine 37: 4504-4510
- 651 44. Swarthout TD, Henrion MYR, Thindwa D, Meiring JE, Mbewe M, Kalizang'Oma A,
- 652 Brown C, Msefula J, Moyo B, Mataya AA, Barnaba S, Pearce E, Gordon M,
- 653 Goldblatt D, French N, et al., 2022. Waning of antibody levels induced by a 13-
- 654 valent pneumococcal conjugate vaccine, using a 3 + 0 schedule, within the first
- 655 year of life among children younger than 5 years in Blantyre, Malawi: an
- 656 observational, population-level, serosurveillance study. Lancet Infect Dis 22:
- 657 1737-1747
- 658 45. Scobie HM, Patel M, Martin D, Mkocha H, Njenga SM, Odiere MR, Pelletreau S,
- 659 Priest IW, Thompson R, Won KY, Lammie Pl., 2017. Tetanus Immunity Gaps in
- Children 5-14 Years and Men ≥ 15 Years of Age Revealed by Integrated Disease 660
- 661 Serosurveillance in Kenya, Tanzania, and Mozambique. Am J Trop Med Hyg 96:
- 662 415-420
- 663 46. Arnold BF, Kanyi H, Njenga SM, Rawago FO, Priest JW, Secor WE, Lammie PJ, Won
- 664 KY, Odiere MR., 2020. Fine-scale heterogeneity in Schistosoma mansoni force of
- 665 infection measured through antibody response. Proc Natl Acad Sci U S A 117:
- 23174-23181 666
- 667 47. Iriemenam NC, Ige FA, Greby SM, Okunoye OO, Uwandu M, Aniedobe M, Nwaiwu 668 SO, Mba N, Okoli M, William NE, Ehoche A, Mpamugo A, Mitchell A, Stafford KA,

Thomas AN, et al., 2023. Comparison of one single-antigen assay and three multi-antigen SARS-CoV-2 IgG assays in Nigeria. J Clin Virol Plus 3: 100139

671 48. Steinhardt LC, Ige F, Iriemenam NC, Greby SM, Hamada Y, Uwandu M, Aniedobe M, Stafford KA, Abimiku A, Mba N, Agala N, Okunoye O, Mpamugo A,

673 Swaminathan M, Onokevbagbe E, et al. Cross-Reactivity of Two SARS-CoV-2

Serological Assays in a Setting Where Malaria Is Endemic. J Clin Microbiol 59: e00514-21

676 49. Herman C, Leonard CM, Uhomoibhi P, Maire M, Moss D, Inyang U, Abubakar A, Ogunniyi A, Mba N, Greby SM, Okoye MI, Iriemenam NC, Maikore I, Steinhardt L,

Rogier E., 2023. Non-falciparum malaria infection and IgG seroprevalence among

children under 15 years in Nigeria, 2018. Nat Commun 14: 1360