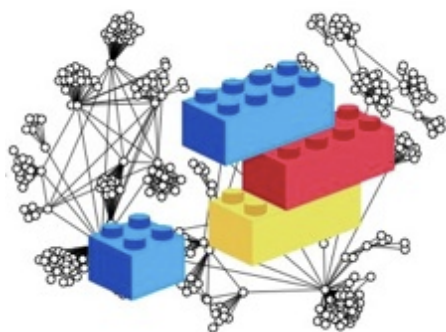


Iterative Network-Guided Connectivity mapping

This website and the linked web pages contain functions, scripts and data objects used in the software enclosed to the paper entitled *A semi-supervised approach for refining transcriptional signatures of drug response and repositioning predictions*, by Francesco Iorio et al, submitted as research paper to PLoS ONE.



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See accompanying file LICENSE.txt or copy at <http://www.gnu.org/licenses/gpl-3.0.html> Paper website: http://www.ebi.ac.uk/~iorio/PLoS_ONE_Submission

Source code and supplementary data

Supplementary Dataset DS1: cMap Drugs prototype ranked lists

Compressed tab delimited txt file containing the 'prototype ranked lists' of genes for all the drug contained in the connectivity map dataset, computed as described in [Iorio et al, PNAS 2010](#).

[SuppDataset_SD1_DRUG_PRLS_txt.zip](#)

How to reproduce results and figures presented in the manuscript?

To start:



Make sure you have R installed. You can download it from <http://cran.ma.imperial.ac.uk/>



We strongly recommend to install and use the RStudio interface to R, downloadable from: <http://www.rstudio.com>

Required libraries:

Make sure you have the following libraries installed (all available on the [CRAN](#) repository):

- [mixtools](#)
- [sROC](#)
- [pheatmap](#)
- [beeswarm](#)

To install them use the following command from the RStudio console:

```
install.packages("[library.name]")
```

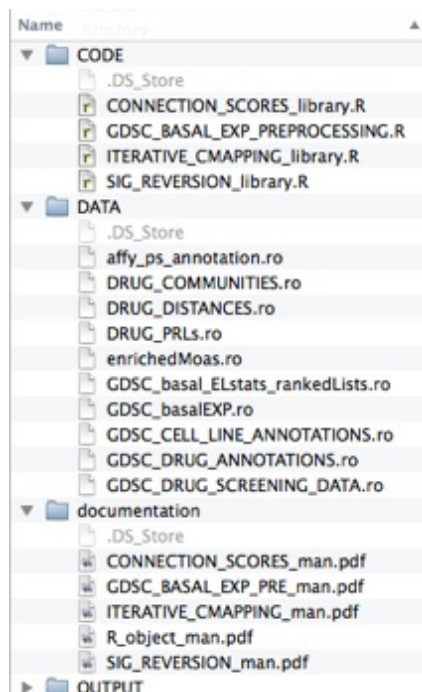
replacing `[library.name]` with each of the library names listed above, in turn.

Working directory creation:



Download and unzip the following compressed folder:
[IorioEtAl_R_code_and_objects.zip](#)

Once uncompressed, the content of this folder and its sub-folders should not be changed. Files in the `OUTPUT` subfolder (initially empty) can be moved and/or deleted.



Working directory setup:

To set the working directory to `IorioEtAl_R_code_and_objects` use the following command from the RStudio console:

```
setwd('[path]/IorioEtAl_R_code_and_objects')
```

replacing `[path]` with the path of the `IorioEtAl_R_code_and_objects` directory.

Ready to go!



To reproduce results and figure presented in our manuscript execute the commands contained in the following pipelines:

Network guided iterative connectivity mapping pipeline

Pipeline for predictive ability validation through the signature reversion paradigm

Iterative network guided connectivity mapping pipeline

A semi-supervised approach for refining transcriptional signatures of drug response and repositioning predictions

(Supplementary Material and Methods: Supplementary Code)

Francesco Iorio - 24 Aprile 2014

Importing libraries of functions needed to compute connectivity scores and to run the iterative network guided connectivity mapping pipeline:

```
options(warn = -1)
source("CODE/CONNECTION_SCORES_library.R")

## Loading required package: boot
## Loading required package: MASS
## Loading required package: segmented
## mixtools package, version 1.0.1, Released January 2014
## This package is based upon work supported by the National Science Foundation under Grant No. SES-0518772.

source("CODE/ITERATIVE_CMAPPING_library.R")
```

Querying the drug network described in Iorio et al (PNAS 2010) using paclitaxel as seed compound:

```
paclitaxelNeighborhood <- DNquery(seed = "paclitaxel", distTh = 0.8065, printToFile = FALSE)
```

Analysing the paclitaxel neighborhood in the drug network (main figure 2 and supplementary table 1):

```
print(paclitaxelNeighborhood[, c("D", "quantile %", "C id", "Adj p-val")])

##           D           quantile % C id           Adj p-val
## demecolcine 0.70572 0.449954794261324 48
## 5252917     0.73307 0.82631885114346 48 0.000236989288084179
## pararosaniline 0.75068 1.26062101237493 62
## MG-132      0.75678 1.46398843106884 40
## parbendazole 0.75768 1.49517688643431 90
## celastrol   0.75776 1.49891482865039 40 0.0182249280466375
## 5224221     0.76625 1.84537534780384 40 0.00169840589958832
## splitomicin 0.76715 1.88836168328883 24
## diltiazem   0.77064 2.05528416537591 73
## cytochalasin_B 0.77165 2.10878346334364 100
## fenbendazole 0.77847 2.5058230131085 69
## gefitinib   0.78277 2.79411180652411 60
## suloctidil  0.78504 2.95157262237672 34
## chlortetracycline 0.78788 3.16907413507521 42
## PHA-00665752 0.7887 3.2295820746981 19
## rotenone    0.78983 3.32606770815082 62 0.0694962846867603
## promethazine 0.7901 3.34697682242205 90 0.331175834577628
## ionomycin   0.79515 3.77742423074317 40 0.00251447929397367
## cyproheptadine 0.79774 4.02120814964852 40 0.000400440318057552
## lynestrenol 0.80095 4.35166560368935 40 9.79332311504551e-05
## perhexiline 0.80485 4.7877199253346 100 0.375201411612571
## terfenadine 0.8058 4.90114310945396 34 0.0559034992586342
```

Listing drug communities enriched in the paclitaxel neighborhood (adjusted p-value < 0.05):

```
enriched_cid <- unique(paclitaxelNeighborhood[which(as.numeric(as.character(paclitaxelNeighborhood[,
"Adj p-val"]))) < 0.05], "C id"])
print(enriched_cid)

## [1] 48 40
## Levels: 100 19 24 34 40 42 48 60 62 69 73 90
```

Listing modes-of-action/Drug-features over-represented in the drug communities enriched in the paclitaxel neighborhood:

```
print(as.character(unique(paclitaxelNeighborhood[which(is.element(paclitaxelNeighborhood[,
"C id"], enriched_cid)), "MOAs"])))

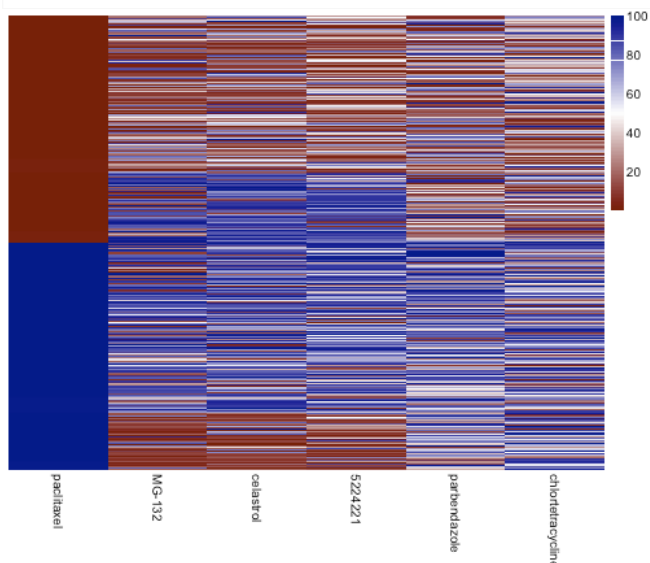
## [1] "plant alkaloids // alkaloid"
## [2] "Proteasome inhibitors and UPS modulators // protein synthesis inhibitors (elongation inhibitors) // calcium signal modulators"
```

Deriving paclitaxel/Proteasome-inhibitors consistent/inconsistent signatures (supplementary table 3)

```
P_PI_consistentSig <- DeriveConsistentSignature(seed = "paclitaxel", otherCompounds = c("MG-132",
"celastrol", "5224221"), PTH = 30, FUZZYNESS = 2, printToFile = FALSE)
P_PI_inconsistentSig <- DeriveInConsistentSignature(seed = "paclitaxel", otherCompounds = c("MG-132",
"celastrol", "5224221"), PTH = 30, FUZZYNESS = 2, printToFile = FALSE)
```

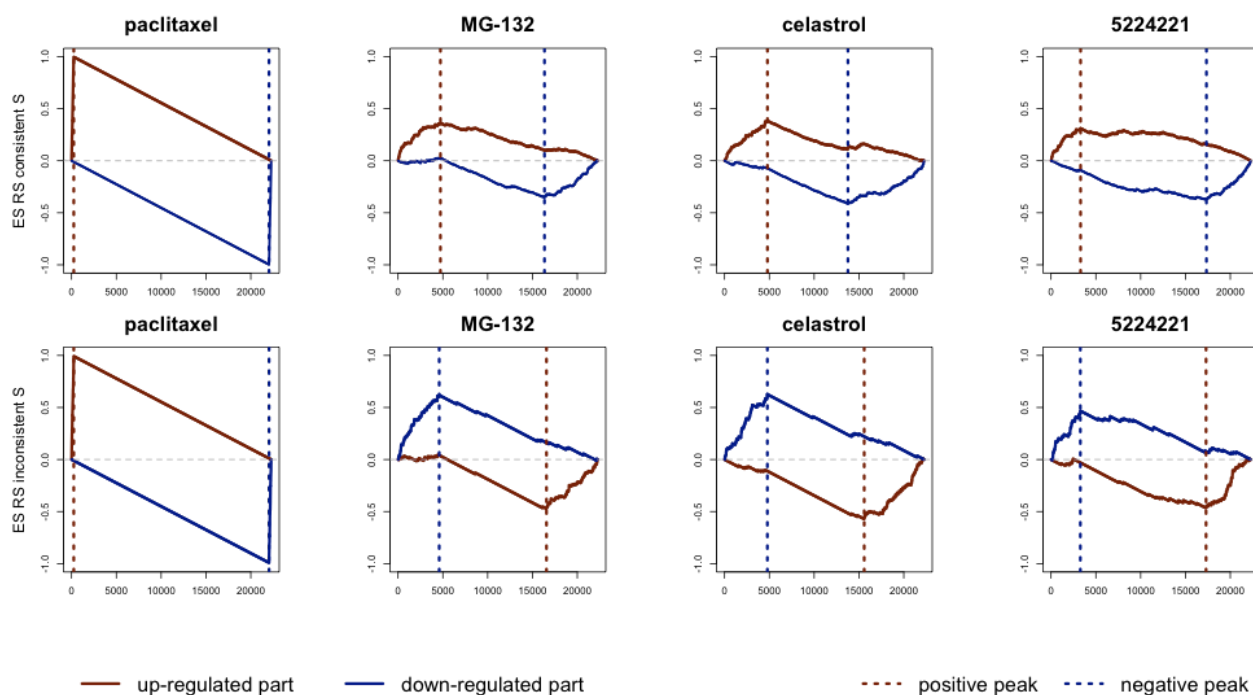
Visualising a heatmap of expression percentiles of the computed signatures along the prototype ranked lists of paclitaxel, the proteasome inhibitors contained in its neighbourhood, and 2 microtubule perturbing drugs included for reference (main figure 3 (A)):

```
percHeatMaps(c(as.character(P_PI_consistentSig$seedUPreg$ProbeSets), as.character(P_PI_inconsistentSig$seedUPreg$ProbeSets),
as.character(P_PI_consistentSig$seedDOWNreg$ProbeSets), as.character(P_PI_inconsistentSig$seedDOWNreg$ProbeSets)),
seed = "paclitaxel", otherCompounds = c("MG-132", "celastrol", "5224221",
"parbendazole", "chlortetracycline"), printToFile = FALSE)
```



Visualising the enrichment score running sums for of the paclitaxel/proteasome-inhibitors consistent/inconsistent signatures along the prototype ranked lists of paclitaxel and the proteasome inhibitors (supplementary figure SF1):

```
plotRunningSums(P_PI_consistentSig, P_PI_inconsistentSig, seed = "paclitaxel",
otherCompounds = c("MG-132", "celastrol", "5224221"), printToFile = FALSE)
```



Computing connectivity scores between the prototype ranked lists of all the cMap drugs and the paclitaxel/proteasome-inhibitors consistent/inconsistent signatures (this may take a while):

```
P_PI_consistent_CS <- CS(P_PI_consistentSig, RANKED_LISTS = DRUG_PRLs, show_progress = FALSE)
```

```
## simulating null model
## number of iterations= 123
## done!
## computing connectivity scores
## Done!
```

```
P_PI_inconsistent_CS <- CS(P_PI_inconsistentSig, RANKED_LISTS = DRUG_PRLs, show_progress = FALSE)
```

```
## simulating null model
## number of iterations= 26
## done!
## computing connectivity scores
## Done!
```

Combining the obtained connectivity scores to refine the paclitaxel neighbourhood:

```
first_nb <- combine_2CS(P_PI_consistent_CS, P_PI_inconsistent_CS, printToFile = FALSE,
  fn = "")
```

Visualising paclitaxel 1st refined neighbourhood (main figure 3 (B) and supplementary table 4):

```
id <- which(first_nb[, "cons S fdr %"] < 5 & first_nb[, "incons S fdr %"] <
  5 & first_nb[, "cons S CS"] > 0 & first_nb[, "incons S CS"] > 0)
print(first_nb[id[2:length(id)], c(4, 8, 9)])
```

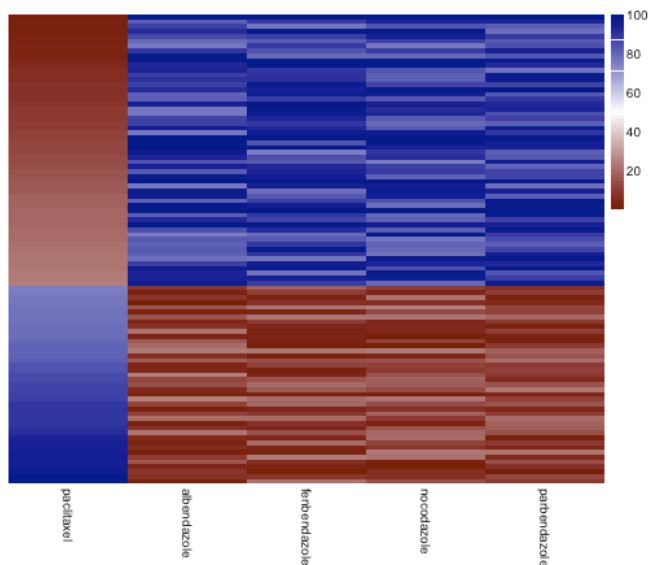
	cons S	NCS	incons S	NCS	avg NCS
## 5252917	4.306	2.719	3.513		
## parbendazole	4.380	1.771	3.076		
## splitomicin	3.657	2.451	3.054		
## fenbendazole	3.916	1.927	2.922		
## gefitinib	3.532	2.228	2.880		
## chlortetracycline	3.429	2.226	2.828		
## rotenone	3.850	1.705	2.778		
## glipizide	3.290	2.185	2.738		
## albendazole	3.240	2.115	2.678		
## bromocriptine	3.167	2.142	2.655		
## diltiazem	3.559	1.739	2.649		
## cyproheptadine	3.620	1.666	2.643		
## moroxydine	2.614	2.471	2.542		
## naloxone	2.936	2.132	2.534		
## perhexiline	3.425	1.596	2.511		
## nilutamide	2.929	1.886	2.407		
## hesperetin	3.169	1.632	2.400		
## nocodazole	2.627	1.980	2.303		
## danazol	2.584	1.788	2.186		
## betulinic acid	2.712	1.616	2.164		
## fluoxetine	2.722	1.603	2.162		
## metolazone	2.159	2.146	2.153		
## hydrastinine	2.517	1.770	2.144		
## practolol	2.385	1.764	2.074		
## genistein	1.984	2.035	2.010		
## monastrol	2.091	1.779	1.935		
## methoxamine	1.958	1.876	1.917		
## primidone	1.874	1.815	1.844		
## 3-hydroxy-DL-kynurenine	1.759	1.921	1.840		
## dehydrocholic acid	1.798	1.854	1.826		
## clozapine	1.933	1.496	1.714		
## thioridazine	1.882	1.533	1.707		
## phenazone	1.739	1.641	1.690		
## epiandrosterone	1.744	1.531	1.638		
## dihydroergotamine	1.681	1.591	1.636		
## chlorphenesin	1.574	1.531	1.552		

Deriving a microtubule stabilising signature (supplementary table 5):

```
MS_sig <- DeriveMSTSignature(seed = "paclitaxel", otherCompounds = c("albendazole",
  "fenbendazole", "nocodazole", "parbendazole"), FUZZYNESS = 4, printToFile = FALSE)
```

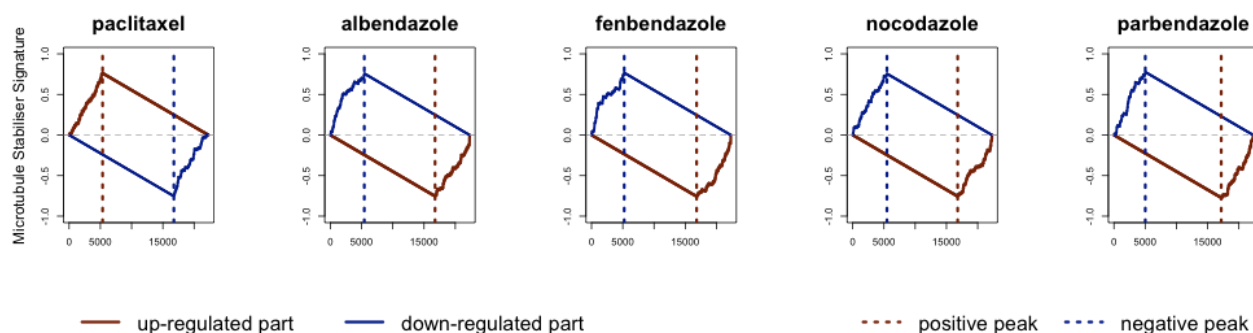
Visualising a heatmap of expression percentiles of the microtubule stabilising signature along the prototype ranked lists of paclitaxel and the three benzimidazoles contained in his first refined neighbourhood (figure 3 (C)):

```
percHeatMaps(c(as.character(MS_sig$seedUPreg$ProbeSets), rev(as.character(MS_sig$seedDOWNreg$ProbeSets))),
  seed = "paclitaxel", otherCompounds = c("albendazole", "fenbendazole", "nocodazole",
  "parbendazole"), printToFile = FALSE)
```



Visualising the enrichment score running sums of the microtubule stabiliser signature along the prototype ranked lists of paclitaxel and the recovered benzimidazoles (supplementary figure SF1):

```
plotRunningSumsMST(MS_sig, seed = "paclitaxel", otherCompounds = c("alendazole",
  "fenbendazole", "nocodazole", "parbendazole"), printToFile = FALSE)
```



Computing connectivity scores between the prototype ranked lists of all the cMap drugs and the microtubule stabilising signature (this may take a while):

```
MST_CS <- CS(MS_sig, RANKED_LISTS = DRUG_PRLs, show_progress = FALSE)
```

```
## simulating null model
## number of iterations= 34
## done!
## computing connectivity scores
## Done!
```

Combining the obtained connectivity scores to finally refine the paclitaxel neighbourhood:

```
previousConnections <- rownames(first_nb)[which(as.numeric(first_nb[, "avg NCS"]) >
  0 & as.numeric(first_nb[, "cons S NCS"]) > 0 & as.numeric(first_nb[, "incons S NCS"]) >
  0 & as.numeric(first_nb[, "cons S fdr %"]) < 5 & as.numeric(first_nb[, "incons S fdr %"]) <
  5)]
final_nb <- combine_3CS(P_PI_consistent_CS, P_PI_inconsistent_CS, MST_CS, previousNeighBr = previousConnections,
  printToFile = TRUE, fn = "final")
```

Visualising the final refined neighbourhood of paclitaxel (main figure 3 (D), table 1 and supplementary table 4):

```
print(final_nb[2:nrow(final_nb), 5])
```

```
##          glipizide          splitomicin          fluoxetine
## "2.19989686915533" "2.04683202514754" "1.96842993269957"
##          metolazone          5252917          diltiazem
## "1.8779827209263" "1.80526353389624" "1.79016488605669"
##          betulinic acid          nilutamide          moroxydine
## "1.77586391018833" "1.75347275374926" "1.61659219321536"
##          perhexiline          gefitinib          bromocriptine
## "1.57596141207419" "1.54301919561025" "1.52192398075313"
##          rotenone          danazol          epiandrosterone
```

##	"1.4943673200471"	"1.47509920854964"	"1.46695129207344"
##	chlortetracycline	thioridazine	cyproheptadine
##	"1.43213750357412"	"1.38190858522147"	"1.36622967073937"
##	hydrastinine	genistein	naloxone
##	"1.35856721388798"	"1.34265532302282"	"1.31001661205284"
##	monastrol	hesperetin	primidone
##	"1.27449217019113"	"1.26817333429531"	"1.26195005404283"
##	phenazone	clozapine	dehydrocholic acid
##	"1.21488168491939"	"1.02664929830309"	"0.982238982351692"
##	methoxamine	3-hydroxy-DL-kynurenine	practolol
##	"0.961273540522456"	"0.914417894562068"	"0.885856416275826"
##	chlorphenesin	dihydroergotamine	parbendazole
##	"0.729772935806824"	"0.550035080493067"	"-0.0926467043760503"
##	fenbendazole	albendazole	nocodazole
##	"-0.159860724573494"	"-0.306185924261493"	"-0.555568320480004"

Pipeline for predictive ability validation through the signature reversion paradigm

A semi-supervised approach for refining transcriptional signatures of drug response and repositioning predictions

(Supplementary Material and Methods: Supplementary Code)

Francesco Iorio - 24 Aprile 2014

Importing libraries of functions needed to compute connectivity scores and to run the signature reversion pipeline:

```
options(warn = -1)
source("CODE/ITERATIVE_CMAPPING_library.R")
source("CODE/CONNECTION_SCORES_library.R")

## Loading required package: boot
## Loading required package: MASS
## Loading required package: segmented
## mixtools package, version 1.0.1, Released January 2014
## This package is based upon work supported by the National Science Foundation under Grant No. SES-0518772.

source("CODE/SIG_REVERSION_library.R")
```

Loading AffyMetrix probe-set annotations:

```
load("DATA/affy_ps_annotation.ro")
```

Loading the GDSC drug screening data and drug annotations for docetaxel, vinorelbine and paclitaxel:

```
load("DATA/GDSC_DRUG_SCREENING_DATA.ro")
load("DATA/GDSC_DRUG_ANNOTATIONS.ro")
```

Loading the annotation file for the GDSC cell lines:

```
load("DATA/GDSC_CELL_LINE_ANNOTATIONS.ro")
```

Load the basal expression statistic ranked lists:

```
load("DATA/GDSC_basal_ELstats_rankedLists.ro")
```

Or, **alternatively**, recomputing them by executing the following commands:

```
source("DATA/GDSC_basal_ELstats_rankedLists.ro")
ELstats <- EL_statistics(basalEXP)
gdsc_basal_ELstats_rankedLists <- basalRanked_lists(ELstats)
```

Generating the gene signatures to be tested individually and in combinations (note that the functions are called with the default parameters):

1. the paclitaxel optimal signature (supplementary table 2)
2. the paclitaxel/protasome-inh. consistent signature
3. the paclitaxel/protasome-inh. inconsistent signature (supplementary table 3)
4. the microtubule stabilising signature (supplementary table 5)

```
paclitaxel_opt_sig <- DeriveSingleSignature()
paclitaxel_PI_Con_sig <- DeriveConsistentSignature()
paclitaxel_PI_incon_sig <- DeriveInConsistentSignature()
MI_stab_sig <- DeriveMSTSignature()
```

Converting microarray probe-sets to gene symbols:

```
paclitaxel_opt_sig$seedUPreg$ProbeSets <- affy_ps_annotation[as.character(paclitaxel_opt_sig$seedUPreg$ProbeSets),
1]
paclitaxel_opt_sig$seedDOWNreg$ProbeSets <- affy_ps_annotation[as.character(paclitaxel_opt_sig$seedDOWNreg$ProbeSets),
1]
paclitaxel_PI_con_sig$seedUPreg$ProbeSets <- affy_ps_annotation[as.character(paclitaxel_PI_con_sig$seedUPreg$ProbeSets),
1]
paclitaxel_PI_con_sig$seedDOWNreg$ProbeSets <- affy_ps_annotation[as.character(paclitaxel_PI_con_sig$seedDOWNreg$ProbeSets),
1]
paclitaxel_PI_incon_sig$seedUPreg$ProbeSets <- affy_ps_annotation[as.character(paclitaxel_PI_incon_sig$seedUPreg$ProbeSets),
1]
paclitaxel_PI_incon_sig$seedDOWNreg$ProbeSets <- affy_ps_annotation[as.character(paclitaxel_PI_incon_sig$seedDOWNreg$ProbeSets),
1]
MI_stab_sig$seedUPreg$ProbeSets <- affy_ps_annotation[as.character(MI_stab_sig$seedUPreg$ProbeSets),
1]
MI_stab_sig$seedDOWNreg$ProbeSets <- affy_ps_annotation[as.character(MI_stab_sig$seedDOWNreg$ProbeSets),
1]
```


Computing connectivity scores for all the GDSC cell lines and the individual signatures (this may take a while):

```
PACLITAXEL <- CS(paclitaxel_opt_sig, gdsc_basal_ELstats_rankedLists, show_progress = FALSE)

## simulating null model
## number of iterations= 64
## done!
## computing connectivity scores
## Done!

P_PI_CON <- CS(paclitaxel_PI_con_sig, gdsc_basal_ELstats_rankedLists, show_progress = FALSE)

## simulating null model
## number of iterations= 686
## done!
## computing connectivity scores
## Done!

P_PI_INCON <- CS(paclitaxel_PI_incon_sig, gdsc_basal_ELstats_rankedLists, show_progress = FALSE)

## simulating null model
## number of iterations= 80
## done!
## computing connectivity scores
## Done!

MI <- CS(MI_stab_sig, gdsc_basal_ELstats_rankedLists, show_progress = FALSE)

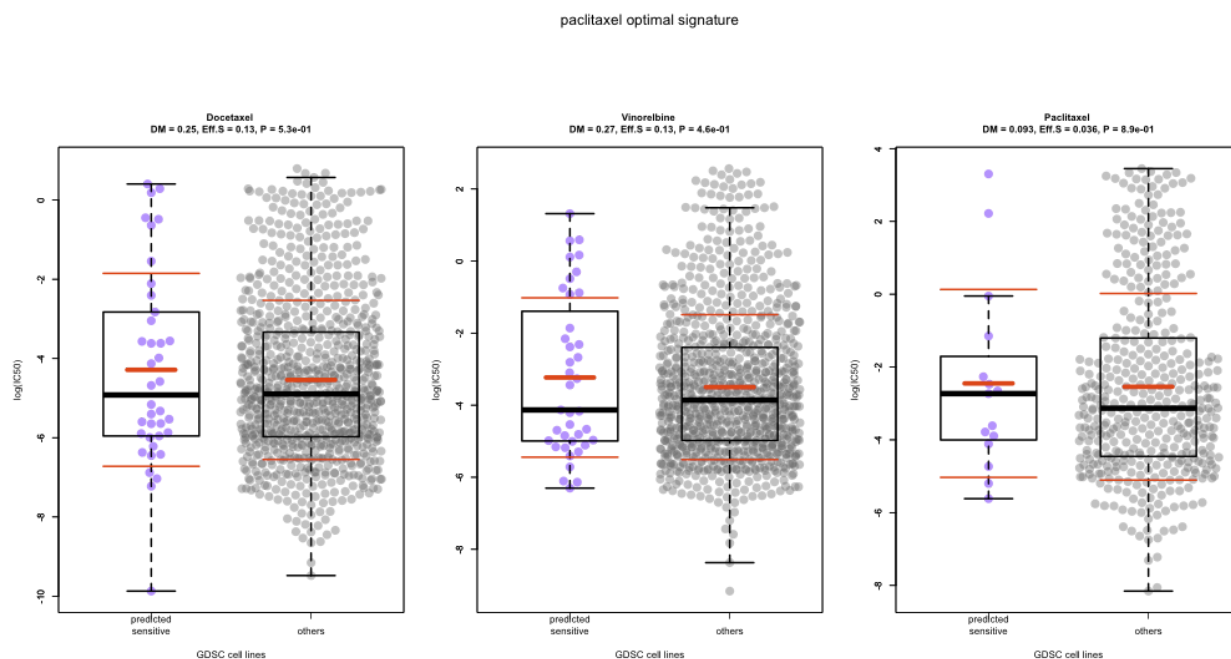
## simulating null model
## number of iterations= 33
## done!
## computing connectivity scores
## Done!
```

Selecting an fdr threshold and the drugs to be tested:

```
th <- 0.3
DRUGS <- rownames(DRUG_PROPS)
```

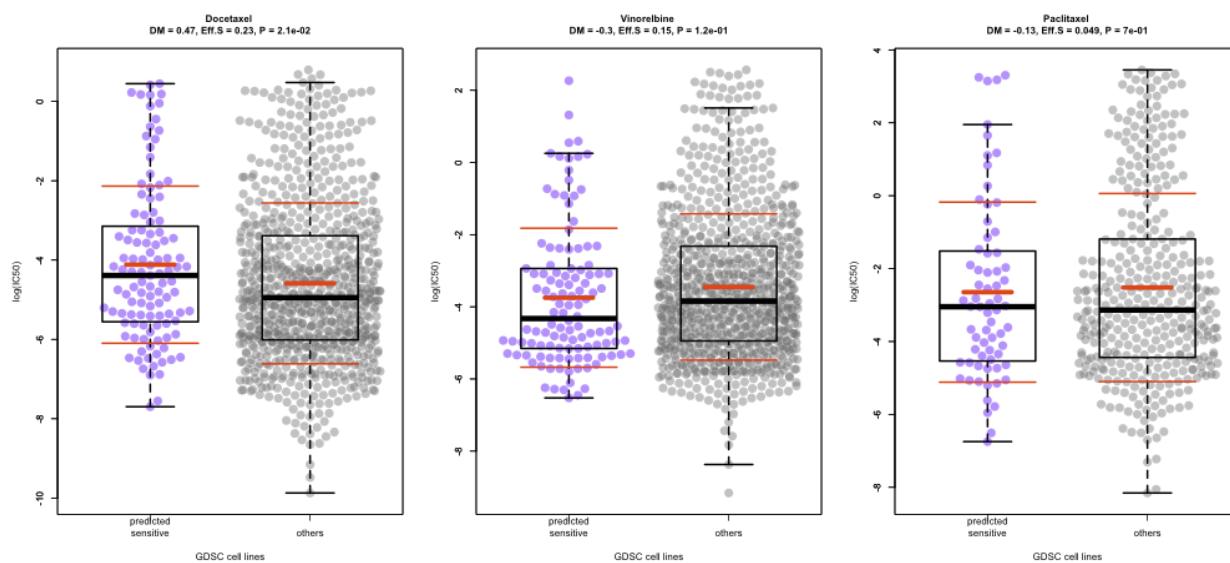
Testing the predictive ability of the individual signatures and storing performance scores (supplementary figure SF6) :

```
totRES <- test_pred_ability(list(PACLITAXEL), DRUGS = DRUGS, mainTitle = "paclitaxel optimal signature")
```



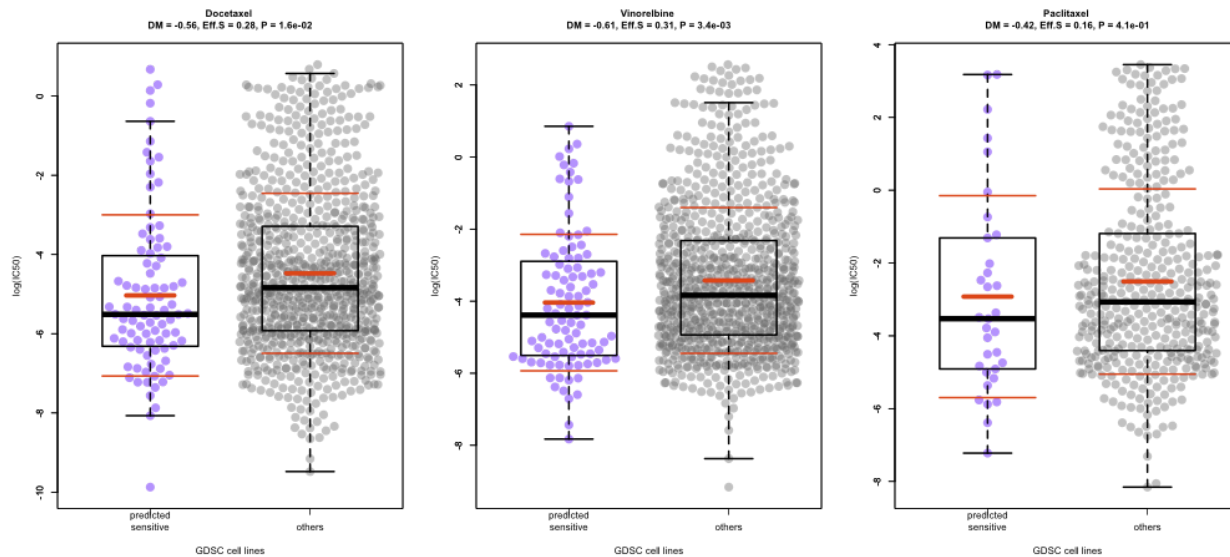
```
totRES <- rbind(totRES, test_pred_ability(list(P_PI_CON), DRUGS = DRUGS, mainTitle = "paclitaxel/proteasome-inh. consistent signature"))
```

paclitaxel/proteasome-inh. consistent signature



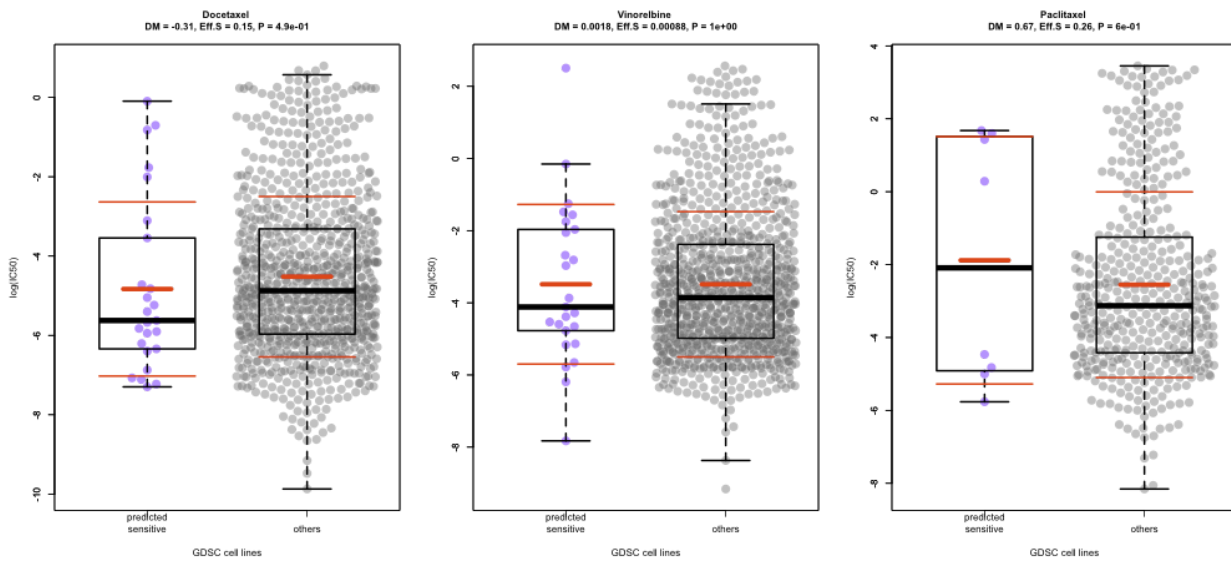
```
totRES <- rbind(totRES, test_pred_ability(list(P_PI_INCON), DRUGS = DRUGS, mainTitle = "paclitaxel/proteasome-inh. inconsistent signature"))
```

paclitaxel/proteasome-inh. inconsistent signature



```
totRES <- rbind(totRES, test_pred_ability(list(MI), DRUGS = DRUGS, mainTitle = "Microtubule stabilising signature"))
```

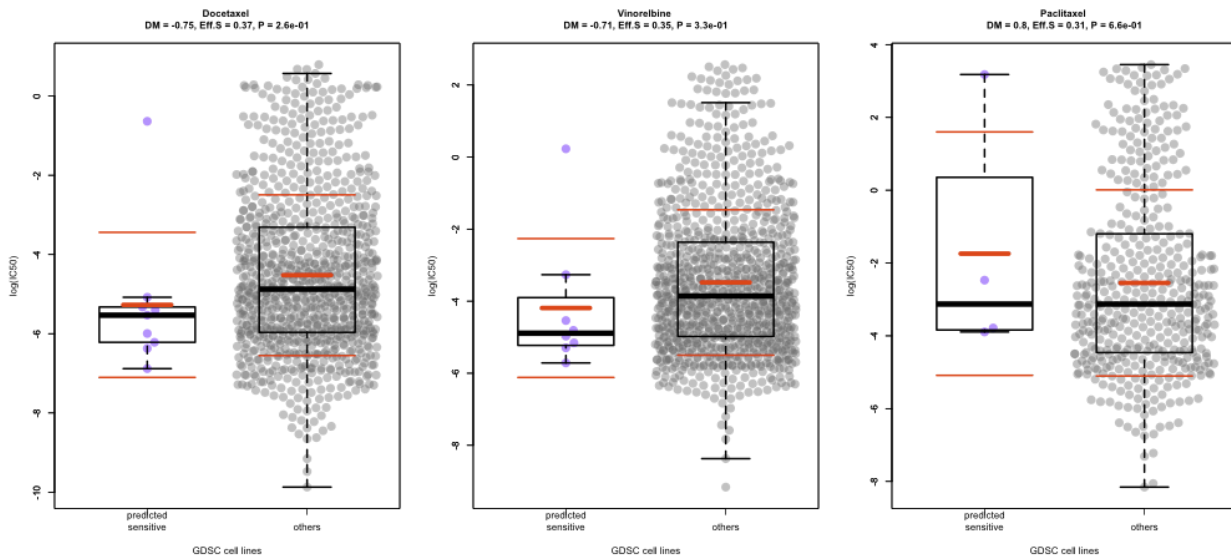
Microtubule stabilising signature



Testing the predictive ability of the combined signatures and storing performance scores (main figure 5 (B) and supplementary figure SF6):

```
totRES <- rbind(totRES, test_pred_ability(list(P_PI_CON, P_PI_INCON), DRUGS = DRUGS,
  mainTitle = "paclitaxel/proteasome-inh. consistent + inconsistent signatures"))
```

paclitaxel/proteasome-inh. consistent + inconsistent signatures

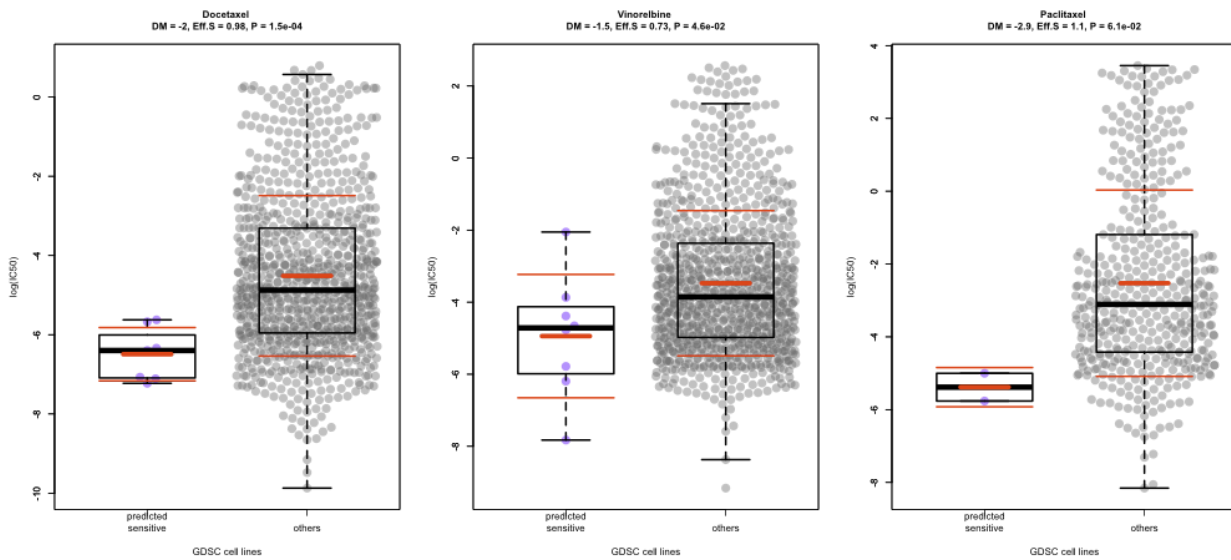


```
totRES <- rbind(totRES, test_pred_ability(list(P_PI_CON, MI), DRUGS = DRUGS,
  mainTitle = "paclitaxel/proteasome-inh. consistent + microtubule stabilising signature"))
```

paclitaxel/proteasome-inh. consistent + microtubule stabilising signature

```
totRES <- rbind(totRES, test_pred_ability(list(P_PI_INCON, MI), DRUGS = DRUGS,
  mainTitle = "paclitaxel/proteasome-inh. inconsistent + microtubule stabilising signature"))
```

paclitaxel/proteasome-inh. inconsistent + microtubule stabilising signature



```
totRES <- rbind(totRES, test_pred_ability(list(P_PI_CON, P_PI_INCON, MI), DRUGS = DRUGS,
  mainTitle = "paclitaxel/proteasome-inh. consistent + inconsistent + microtubule stabilising signature"))
```

paclitaxel/proteasome-inh. consistent + inconsistent + microtubule stabilising signature

Summarising the predictive performances for all the tested signatures and signature combinations:

```
performanceMatrix <- matrix(NA, nrow = length(unique(totRES$used signature(s))),
  ncol = 3, dimnames = list(unique(totRES$used signature(s)), unique(totRES$drug)))
signatures <- unique(totRES$used signature(s))

for (i in 1:length(signatures)) {
  idxs <- which(totRES$used signature(s) == signatures[i])
  performanceMatrix[i, ] <- log10(as.numeric(as.character(totRES[idxs, "p-val"]))) *
    sign(as.numeric(as.character(totRES[idxs, "deltaMean"])))
}
```

Plotting the performance summary (supplementary figure SF7):

```
par(las = 2)
barplot(t(performanceMatrix), ylab = "association signed p-value", names.arg = c("a",
  "b", "c", "d", "b + c", "b + d", "c + d", "b + c + d"), beside = TRUE, col = c("#000000",
  "#8B8378", "#C1CDCD"), ylim = c(-max(abs(c(performanceMatrix))), na.rm = TRUE),
  max(abs(c(performanceMatrix))), na.rm = TRUE), border = NA)
abline(h = -log10(0.05), lty = 2, col = "darkgray")
abline(h = log10(0.05), lty = 2, col = "darkgray")
abline(h = 0, lty = 1, col = "black")
legend("topleft", c("docetaxel", "vinorelbine", "paclitaxel"), fill = c("#000000",
  "#8B8378", "#C1CDCD"), border = NA, title = "Drugs")
legend("bottomright", c("a = paclitaxel optimal", "b = paclitaxel/proteasome-inh. consistent",
  "c = paclitaxel/proteasome-inh. inconsistent", "d = microtubule stabilising"),
  title = "Signatures")
```

