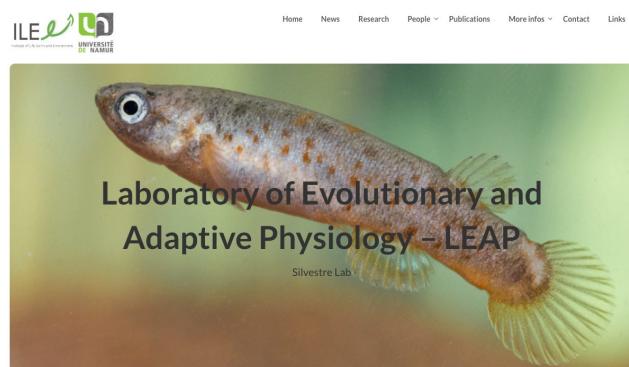


# Bisulfite sequencing analysis Bioinformatic workshop



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[https://github.com/fredsilvestre/bioinfoworkshop\\_epigenetics](https://github.com/fredsilvestre/bioinfoworkshop_epigenetics)

- R Markdown scripts to be fulfilled parts 1 to 4 (copy/paste in Rstudio)
- Dataset for the exercise « \*.csv »
- Context: « README.md »
- studentspack

## Goals of the workshop:

- Learn how to use Bioconductor for bioinformatic analyses
- Learn how to use Rmarkdown
- Learn how to work with DNA sequences in Rstudio and with genomic objects
- Learn how to analyse a methylome from bisulfite sequencing
- Learn how to use bisulfite sequencing to build an epigenetic clock
- Work on real data

### Before the workshop:

- Install R and Rstudio (the most recent versions)
- Run Rstudio and try to get used with the environment ; try to upload some packages on CRAN
- Download the files in the studentspack file on github

### Few online references:

To get familiar with R: <https://www.statmethods.net/index.html>

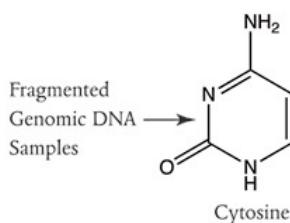
Discussion forum about bioinformatics: <https://stackoverflow.com/>

Reference book: <http://compgenomr.github.io/book/> (mostly Ch10)

## Bisulfite conversion

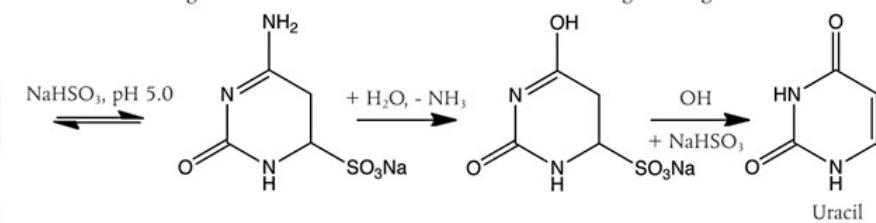
### Step 1

**Denaturation**  
Incubation at 95°C  
fragments genomic DNA



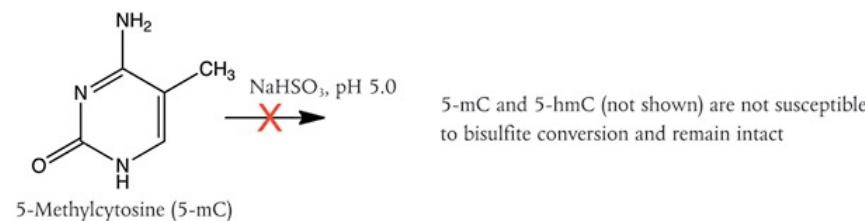
### Step 2

**Conversion**  
Incubation with sodium bisulfite  
at 65°C and low pH (5-6)  
deaminates cytosine residues  
in fragmented DNA



### Step 3

**Desulphonation**  
Incubation at high pH  
at room temperature for 15 min  
removes the sulfite moiety,  
generating uracil



## DNA bisulfite conversion

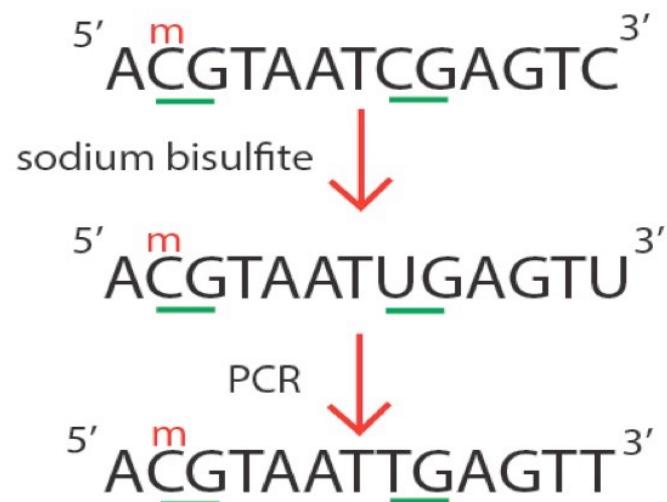
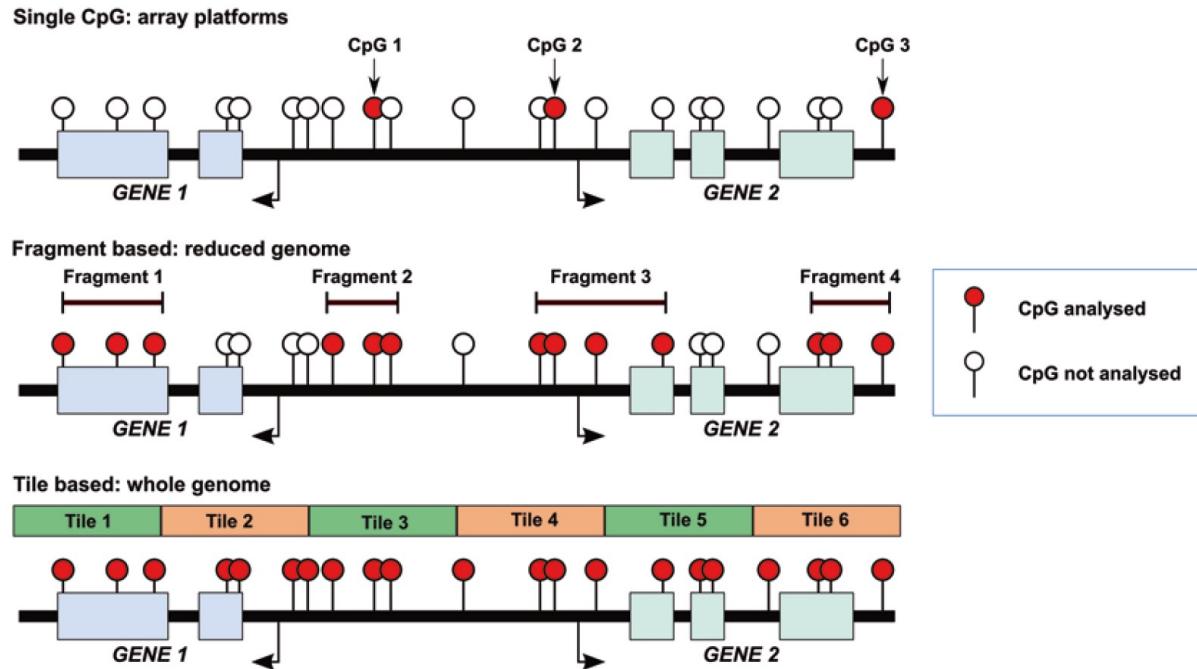


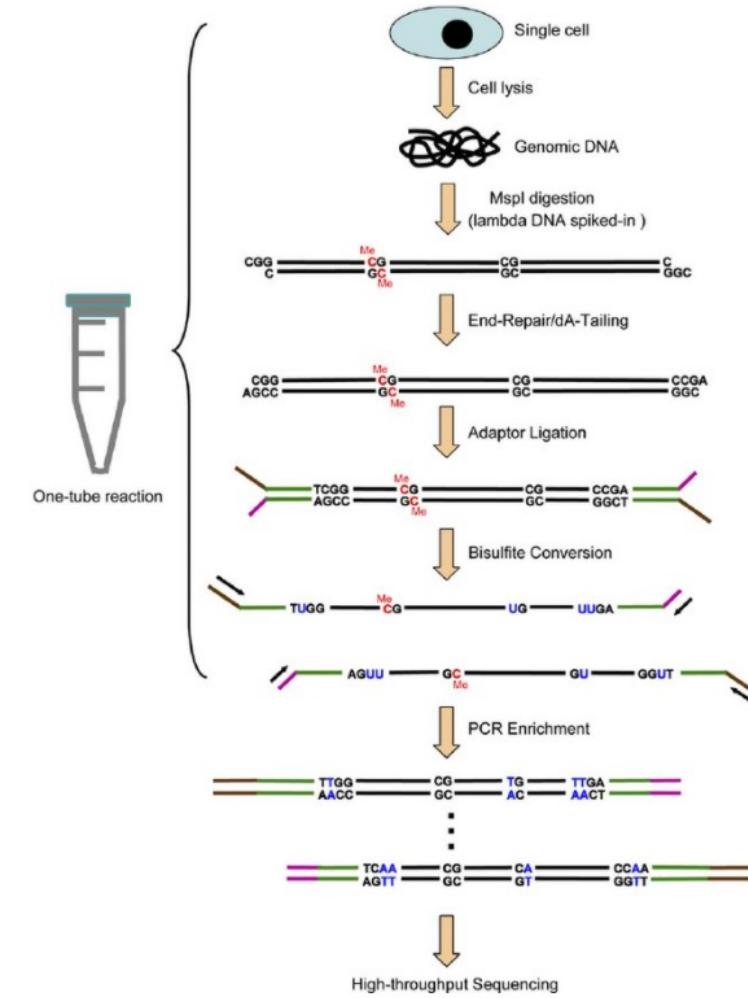
Table 1. Sequences resulting from bisulfite conversion and PCR

	Original sequence	After bisulfite treatment	After PCR amplification
Unmethylated DNA	A-C-G-T-C-G-T-C-A	A-U-G-T-U-G-T-U-A	A-T-G-T-T-G-T-T-A
Methylated DNA	A-C-G-T-C-G-T-C-A	A-C-G-T-C-G-T-U-A	A-C-G-T-C-G-T-T-A

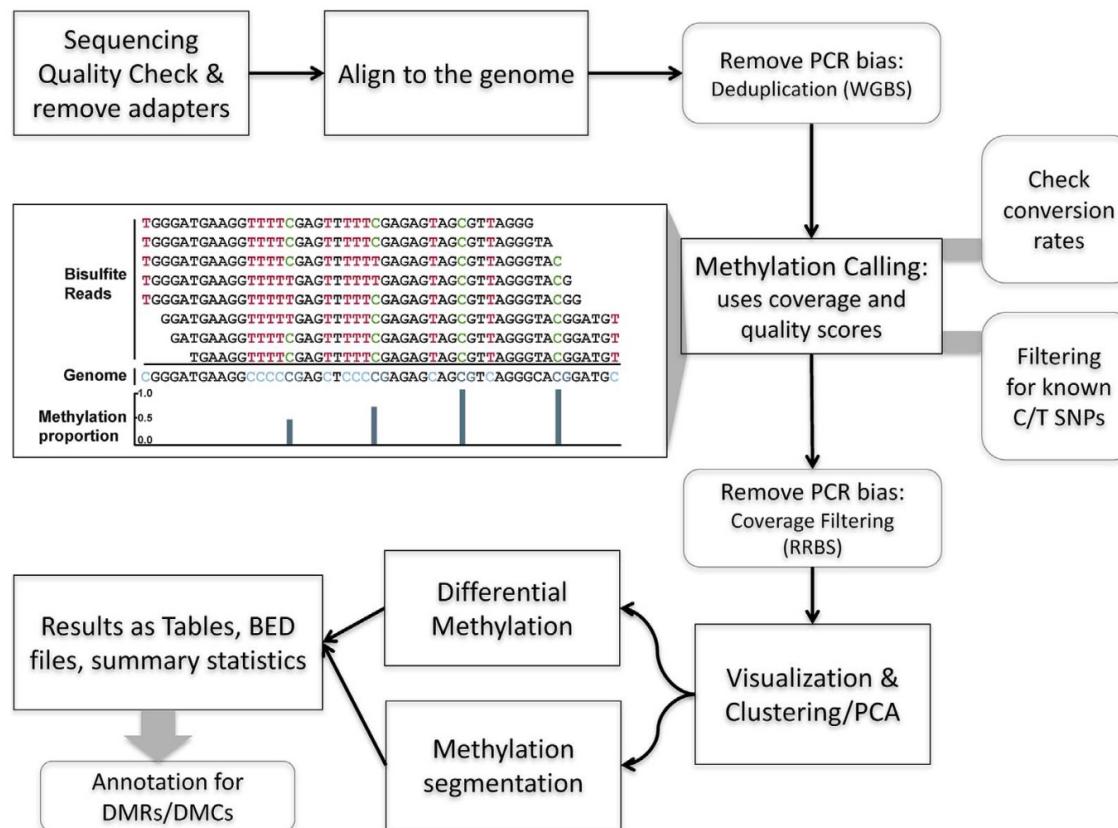


**Fig. 1** Major analysis approaches for genome-wide DNA methylation analysis. There are several approaches for analyzing differential methylation between different groups and conditions. These approaches differ based on the unit of analysis: (1) the single CpG site approach independently analyzes each CpG site in investigated samples; (2) for RRBS, Mspl-digested fragments can be used as the unit of analysis (implemented in DMAP [32] package); (3) a common approach for large bisulfite sequencing data is to investigate regions with fixed size genomic windows. It is possible to use sliding windows based on user-specified criteria

## RRBS workflow



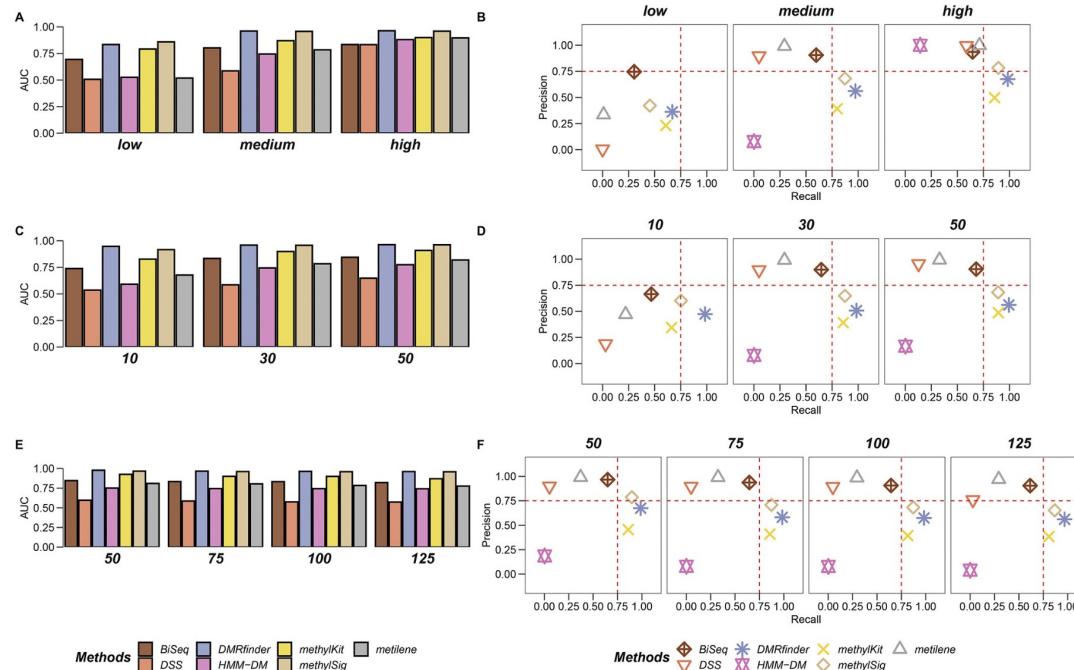
## The workflow of bioinformatic analysis:



**Fig. 1.** Workflow for analysis of DNA methylation using data from bisulfite sequencing experiments.

Wreczycka et al 2017

Recall (=sensitivity) = ratio of true DMRs detected by a tool to the total true DMRs >  $TP / (TP + FN)$   
 Precision = ratio of true DMRs detected by a tool to all DMRs detected by itself >  $TP / (TP + FP)$



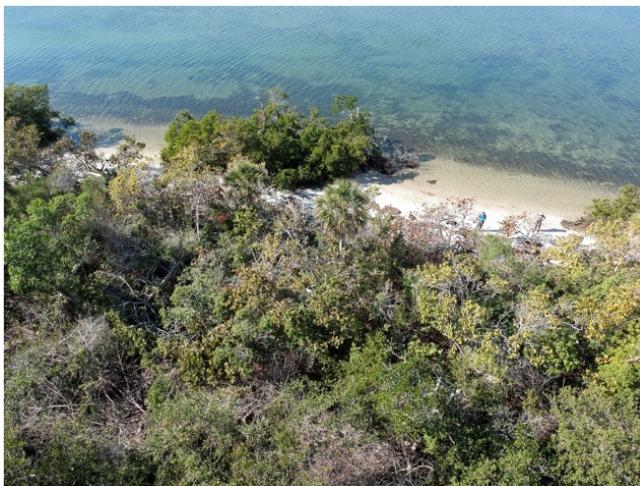
**Fig. 4.** Comprehensive evaluations of tools for detecting DMRs. (A) AUC of various tools for detecting DMR with different methylation difference between two tested groups. The methylation difference in DMR between two groups was set to 3 levels: low (methylation difference between two groups < 0.2), medium (methylation difference between two groups was 0.2–0.4), and high (methylation difference between two groups > 0.4). (B) Recall and precision of various tools for detecting DMRs with different methylation difference between two tested groups. (C) AUC of various tools for detecting DMRs under a different depth of sequencing coverage. Three types of depth of sequencing coverage were compared, including 10 $\times$ , 30 $\times$ , and 50 $\times$ . (D) Recall and precision of various tools for detecting DMRs under a different depth of sequencing coverage. (E) AUC of various tools for detecting DMRs using different sequencing read lengths. (F) Recall and precision of various tools for detecting DMRs using different sequencing read lengths.

Liu et al 2020

## *The mangrove rivulus: surviving in the mangroves*



*Kryptolebias marmoratus*



## *The mangrove rivulus: surviving in the mangroves*



Rivulus microhabitat and sampling



## *The mangrove rivulus: surviving in the mangroves*



Simultaneous  
Hermaphrodites

Males (1-25%)  
(Primary and secondary males)

***K. marmoratus* = Only known  
self-fertilizing vertebrate**

= androdioecy  
(≠ parthenogenesis)

## *The mangrove rivulus: surviving in the mangroves*



Simultaneous  
Hermaphrodites



Self-fertilization

↔  
Outcrossing

Males (1-25%)  
(Primary and secondary males)

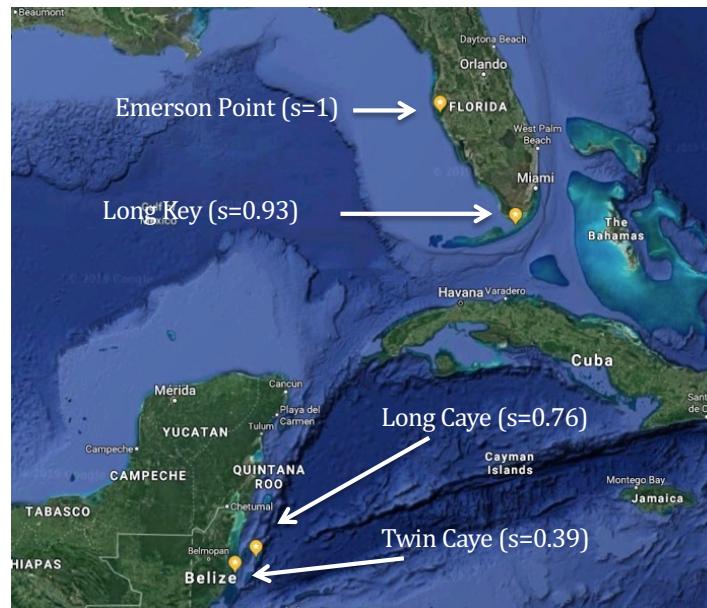
Variable OC (low OC in Florida ; higher in Belize)

# ***The mangrove rivulus: surviving in the mangroves***



Scientific field missions 2019: Belize and Florida

Valentine Chapelle thesis (FRIA since 10/2018): “*The epigenetic origin of behavioral traits variability in a self-fertilizing fish : the mangrove rivulus*”



## *The mangrove rivulus: surviving in the mangroves*



## *The mangrove rivulus: surviving in the mangroves*



Population	N total fish	N males	N herma	N juveniles	% of male
Twin Caye (PG)	177	74	103	0	<b>41.8</b>
Long Caye (LC)	31	3	26	2	10.3
Emerson Point (EPP)	540	2	538	0	0.4
Long Key (LK)	44	1	35	8	2.8

