**Freeman Chris Lewis Jr.**

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# Education

University of California Davis Bachelor of Science in Physiological Psychology with an area of emphasis in Neurology, Physiology & Behavior. (2008-2013)

Tulane University School of Public Health & Tropical Medicine Master of Public Health in Global Community Health and Behavioral Sciences

(2020-present [Expected graduation date: May 2022])

# NOTABLE COURSE WORK AND LAB EXPERIENCE

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| --- | --- | --- |
| Organic Chemistry (Lab) | Molecular Biology (Lab) | General Biology and Chemistry (Lab) |
| Physiological Psychology (Lab) | Biochemistry (Lab) | Neurobiology, Physiology, and Behavior (Lab) |
| Hormones and Behavior (Lab) | Cognitive Neuroscience | Biological Psychology |
| Developmental Disorders | Human Development | Comparative Neurobiology (Lab) |

**NOTABLE WORKSHOPS AND SELF-PACED LEARNING**

Stanford’s Institute for Computation and Mathematical Engineering (ICME)

* Linear Algebra
* Introduction to Mathematical Optimization
* Introduction to Machine Learning
* Introduction to Deep Learning
* Deep Learning for Natural Language Processing
* Data Visualization in Tableau

# SUMMARY OF QUALIFICATION AND SKILLS

* Advance proficiency with research techniques including those involved in behavioral neuroscience, physiology, microscopy, genomics and cell biology.
* Expertise in designing and executing aseptic techniques and stereotaxic administration of neuroanatomical tracers into localized brain regions.
* Excellent ability to conduct research experiments while accurately maintaining and meticulously recording procedures and results.
* Capable of recreating animal models of disease to test proprietary compounds.
* Above average communication skills with the aptitude to effectively relay information to an audience.
* PC Skills: Microsoft Office (Word, Excel, PowerPoint, OneNote, Outlook, Publisher, Access), Statistica, Sigmaplot. and Ethovision (automated tracking software), Tableau.
* Programing Languages – SAS, STATA, R Programming.

# PROFESSIONAL EXPERIENCE

*Senior Research Associate In Vivo Physiology/Pre-Clinical Translational Pharmacology, Calico Life Science (Alphabet Company)*

(Dr. Ganesh Kolumam & Dr. Nick Van Bruggen, 2015-Present)

* Independently created experimental designs, prepared samples, constructed libraries and generated data of gene expression patterns in animal models of disease using next generation Illumina sequencing.
* Familiarity with droplet-based and plate-based single cell assays and platforms (10x Genomics), spatial transcriptomics, laboratory robotics, large-scale experiments and process development.
* Purified DNA/RNA for -omic’s analysis (both single cell and bulk -omic’s sequencing).
* Experience with cell culture and tissue processing including isolation of Mouse Embryonic Fibroblasts (MEF).
* Independently performed animal dosing (routes: Intravenous, Intraperitoneal, Subcutaneous, Intramuscular, & Oral) and animal blood collections (Retro Orbital, Cardiac Puncture, Tail Nick, Submandibular) in mice to perform Pharmacodynamic, Pharmacokinetic, and Therapeutic Efficacy studies.
* Harvested tissues during necropsy’s for further downstream analysis.
* Autonomously performed Intracerebrovrentricular (ICV) Stereotaxic injections and subsequent tissue processing (brain sectioning, immunohistochemical staining)
* Performed microdissections of tissues/organs of the nervous, digestive, cardiovascular and musculoskeletal systems.
* Developed in vivo assays to test drug target engagement to be subsequently used for human clinical studies.
* Independently completed in vitro assays to test for target engagement through secondary biomarker measurements.
* Familiarity with Prometheion Metabolic Cages and Vium Video Monitoring Systems for large scale analyses of physiological phenotypes.

*Research Scientist Intern, Genentech (Roche Group)*

(Dr. Kimberly Scearce-Levie, 2015)

* Autonomously constructed equipment, refined training protocols, trained subjects, analyzed data, and communicated results of cognitive
* Validated behavioral phenotypes in a rodent model of Alzheimer’s disease using novel touchscreen operant conditioning technology.
* Developed skills in molecular biology – specifically immunohistochemistry techniques (perfusion, dissection, and sectioning).
* Mastered pharmaceutical compound administration through various dosing routes (intraperitoneal, subcutaneous, and oral gavage).

*Junior Specialist, University of California, Davis Department of Psychiatry and Behavioral Neuroscience*

*(MIND Institute)*

(Dr. Jacqueline Crawley, 2012 - 2015)

* Independently conducted assessments of animal health and diagnosed health concerns when applicable.
* Conducted a multitude of behavioral assays including but not limited to the measurement of ultrasonic vocalizations in transgenic models of Autism.
* Followed proper IACUC protocol involving the testing and handling of animal subjects.
* Catalogued and classified experimental data using various software packages.
* Prepared cohesive analyses of tabulated experimental data for article publication.

*Undergraduate Lab Research Assistant, University of California Davis Psychology Department of Comparative Neurobiology*

(Dr. Karen Bales, 2012-2013)

* Handled Prairie Vole subjects while administering Oxytocin intranasal injections.
* Collected blood samples from Prairie Vole subjects and performed immunohistochemistry on the samples.
* Conducted behavioral assays and scored behavioral traits of Prairie Vole subjects.
* Quantified neurostaining within Prairie Vole brains through the use of neuroimaging techniques.
* Surgically removed Prairie Vole brain for experimentation.

*Emergency Room Intern, University of California Davis Medical Center*

(Dr Mary L. Bing, 2012)

* Developed professional relationships with and conveyed feelings of sensitivity towards patients and their comfortability by performing basic duties (maintained the cleanliness of patients’ rooms, motivated rehabilitation and recovery through conversation, and perceived non- verbal social cues for assistance).
* Initiated new ideas as well as coordinated tasks in order to provide support to medical staff by respectfully delegating specific duties and spatial locations to fellow undergraduate peers within the Emergency Room.
* Provided alternative solutions to patients in the waiting area by facilitating discussion and listening attentively to the feelings and issues they described.

## Publications:

Yang M, Mahrt, E J, **Lewis, FC**, Foley, G, Portmann, T, Dolmetsch, RE, Portfors, CV, Crawley, JN (2015). 16p11.2 Deletion Syndrome Mice Display Sensory Deficits and Reduced Ultrasonic Vocalizations during Social Interactions. *Autism Research*.

Yang M, **Lewis F**, Foley G, Crawley JN (2015). In Tribute to Bob Blanchard: Divergent Behavioral Phenotypes of 16p11.2 Deletion Mice Reared in Same-Genotype Versus Mixed-Genotype Cages. *Physiology and Behavior*.

Yang M, **Lewis F**, Sarvi M, Foley G, Crawley J. 16p11.2 Deletion Mice Display Cognitive Deficits in Touchscreen Learning and Novelty Recognition Tasks. *Learning and Memory*.

Deurloo MHS, Chen W, Yang M, Lin YW, Tam E, Huang YC, Wu M, **Lewis, FC**, Foley GM, Crawley JN, Sun HS, Osborne LR, Feng ZP. General Transcription Factor 2I (Gtf2i) copy number regulates neuronal maturation and cognition through TRPC3 channel and muscarinic

receptor- dependent mechanisms. *Brain Research*.

## Conference Presentation:

Yang M, Mahrt J, **Lewis F**, Foley G, Portmann T, Dolmetsch R, Portfors C, Crawley J. 16p11.2 deletion syndrome mice display ultrasonic vocalization deficits during social interactions. *Society for Neuroscience (SfN) Annual Meeting*. Washington,D.C.

Yang M, **Lewis F**, Foley G, Portmann T, Dolmetsch R, Crawley J. 16p11.2 Deletion Mice Display Cognitive Deficits in Novelty DiscriminationTasks. *The International Meeting for Autism Research (IMFAR)*. Salt Lake City, Utah. 2015

Yang M, **Lewis F**, Foley G, Crawley J. 16p11.2 Deletion Mice Display Cognitive Deficits in Novelty Discrimination Tasks.

*International Behavioral Neuroscience Society (IBNS) Annual Meeting*. British Columbia, Canada, 2015

**Abstract Submissions:**

Ambade A, Purified DNA/RNA for -omic’s analysis (both single cell and bulk -omic’s sequencing). A, Aditya, **Lewis F**, Kolumam G, Morrison J, Cornicelli J. Defining the Onset of Fibrosis in a CDAA Diet Induced Mouse Model of Non-Alcoholic Steatohepatitis using RNA-seq / pathway profiling. *American Association for the Study of Lier Diseases (AALSD)*. Digital Experience, 2020.

**References available on request**