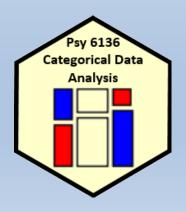


Logistic regression



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Psych 6136

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Model-based methods: Overview

Structure

- Explicitly assume some probability distribution for the data, e.g., binomial, Poisson, ...
- Distinguish between the systematic component— explained by the model— and a random component, which is not
- Allow a compact summary of the data in terms of a (hopefully) small number of parameters

Advantages

- Inferences: hypothesis tests and confidence intervals
- Can test individual model terms (anova ())
- Methods for model selection: adjust balance between goodness-of-fit and parsimony
- Predicted values give model-smoothed summaries for plotting
- Interpret the fitted model graphically

loglm() vs. glm()

With loglm() you can only test overall fit (anova()) or difference between models (Lrstats())

What we can say:

Even the model with all pairwise associations fits poorly

Comparing models with anova () and LRstats ()

```
> anova(berk.mod1, berk.mod2, test="Chisq")
LR tests for hierarchical log-linear models
Model 1:
~Dept * (Gender + Admit)
Model 2:
~ (Admit + Dept + Gender) ^2
         Deviance df Delta(Dev) Delta(df) P(> Delta(Dev)
Model 1 21.74 6
Model 2 20.20 5 1.531
                                              0.21593
Saturated 0.00 0
                       20.204
                                              0.00114
> LRstats(berk.mod1, berk.mod2)
Likelihood summary table:
         AIC BIC LR Chisq Df Pr(>Chisq)
berk.mod1 217 238 21.7 6
                               0.0014 **
berk.mod2 217 240 20.2 5 0.0011 **
Signif. codes: 0 '***' 0.001 '**' 0.01 '*' 0.05 '.' 0.1 '' 1
```

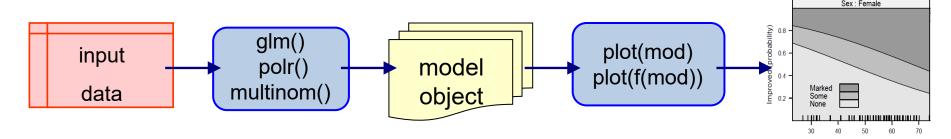
loglm() vs. glm()

With glm() you can test individual terms using anova() or car::Anova()

```
> berkeley <- as.data.frame(UCBAdmissions)</pre>
> berk.glm2 <- glm(Freq ~ (Dept+Gender+Admit)^2, data=berkeley,
                family="poisson")
> anova(berk.glm2, test="Chisq")
Analysis of Deviance Table
Model: poisson, link: log
Response: Freq
Terms added sequentially (first to last)
           Df Deviance Resid. Df Resid. Dev Pr(>Chi)
NULL
                            23
                                    2650
            5 160
                            18
                                    2491 <2e-16 ***
Dept
Gender
            1 163
                            17
                                    2328 <2e-16 ***
                            16 2098 <2e-16 ***
               230
Admit.
            5 1221
                            11 877 <2e-16 ***
Dept:Gender
Dept:Admit 5 855
                                  22 <2e-16 ***
Gender:Admit 1
                                      20
                                            0.22
Signif. codes: 0 '***' 0.001 '**' 0.01 '*' 0.05 '.' 0.1 '' 1
```

Fitting & graphing models: Overview

Object-oriented approach in R:



- Fit model (obj <- glm (...)) → a model object</p>
- \bullet print (obj) and summary (obj) \rightarrow numerical results
- anova (obj) and Anova (obj) → tests for model terms
- update(obj), add1(obj), drop1(obj) for model selection

Plot methods:

- plot (obj) often gives diagnostic plots
- Other plot methods:
 - Mosaic plots: mosaic (obj) for "loglm" and "glm" objects
 - Effect plots: plot (Effect (obj)) for nearly all linear models
 - Influence plots (car): influencePlot (obj) for "glm" objects

Objects & methods

How this works:

• Model objects have a "class" attribute:

```
    loglm(): "loglm"
    glm(): c("glm", "lm") — inherits also from lm()
```

- Class-specific methods have names like method.class, e.g., plot.glm(), mosaic.loglm()
- Generic functions (print (), summary (), plot () ...) call the appropriate method for the class

```
arth.mod <- glm(Better ~ Age + Sex + Treatment, data=Arthritis)
class(arth.mod)
## [1] "glm" "lm"</pre>
```

Objects & methods

Methods for "glm" objects

```
> library(MASS); library(vcdExtra)
> methods(class="qlm")
 [1] add1
                       addterm
                                          anova
                                                            Anova
 [5] asGnm
                                          avPlot
                                                            avPlot3d
                       assoc
                                          brief
                                                            ceresPlot.
 [9] Boot
                       boot.Case
[13] coerce
                       confidenceEllipse confint
                                                            Confint
[17] cooks.distance
                       deviance
                                          drop1
                                                            dropterm
[21] effects
                       extractATC
                                          family
                                                            formula
                                          initialize
[25] gamma.shape
                      influence
                                                            leveragePlot
[29] linearHypothesis
                      logLik
                                         mcPlot.
                                                            mmp
[33] model.frame
                       modFit
                                         mosaic
                                                            ncvTest
[37] nobs
                       predict
                                         print
                                                            profile
                       residualPlot
                                         residualPlots
                                                            residuals
[41] qqPlot
[45] rootogram
                       rstandard
                                        rstudent
[49] show
                                          sigmaHat
                       sieve
                                                            slotsFromS3
[53] summary
                                         weights
                       VCOV
see '?methods' for accessing help and source code
```

There are many, many plot() methods for different types of objects e.g., plot() for a "glm" object $\rightarrow plot.glm()$

```
> methods("plot")
 [1] plot, ANY-method
                        plot, color-method plot.acf*
                        plot.correspondence* plot.data.frame*
[4] plot.ca*
 [7] plot.decomposed.ts* plot.default
                                             plot.dendrogram*
[10] plot.density*
                        plot.ecdf
                                             plot.factor*
[13] plot.formula*
                        plot.function
                                            plot.gnm*
[16] plot.goodfit*
                        plot.hcl palettes* plot.hclust*
[19] plot.histogram*
                        plot.HLtest*
                                             plot.HoltWinters*
[22] plot.isoreq*
                        plot.lda*
                                             plot.lm*
[25] plot.loddsratio*
                     plot.loglm*
                                             plot.mca*
[28] plot.medpolish*
                     plot.mjca*
                                            plot.mlm*
                        plot.prcomp*
[31] plot.ppr*
                                            plot.princomp*
[34] plot.profile*
                        plot.profile.gnm*
                                             plot.profile.nls*
[37] plot.qv*
                        plot.raster*
                                             plot.ridgelm*
[40] plot.rootogram*
                      plot.shingle*
                                             plot.spec*
[43] plot.stepfun
                      plot.stl*
                                             plot.structable*
[46] plot.table*
                    plot.trellis*
                                             plot.ts
[49] plot.tskernel* plot.TukeyHSD* plot.zoo*
see '?methods' for accessing help and source code
```

Modeling approaches: Overview

Association models

- Loglinear models
 (contingency table form)
 [Admit][Gender Dept]
 [Admit Dept][Gender Dept]
 [AdmitDept][AdmitGender][GenderDept]
 - Poisson GLMs
 (Frequency data frame)
 Freq ~ Admit + Gender * Dept
 Freq ~ Admit*Dept + Gender*Dept
 Freq ~ Admit*(Dept + Gender) +
 Gender*Dept
- Ordinal variables
 Freq ~ right + left + Diag(right, left)
 Freq ~ right + left + Symm(right, left)

Response models

- Binary response
- Categorical predictors: logit models logit(Admit) ~ 1 logit(Admit) ~ Dept logit(Admit) ~ Dept + Gender
- Continuous/mixed predictors
- Logistic regression models
 Pr(Admit) ~ Dept + Gender + Age + GRE
- Polytomous response
- Ordinal: proportional odds model
 Improve ~ Age + Sex + Treatment
- General multinomial model
 WomenWork ~ Kids + HusbandIncome

Logistic regression

Response variable

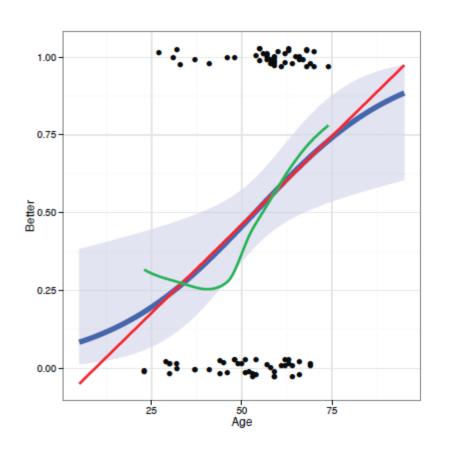
- Binary response: success/failure, vote: yes/no
- Binomial data: x successes in n trials (grouped data)
- Ordinal response: none < some < severe depression
- Polytomous response: vote Liberal, Tory, NDP, Green

Explanatory variables

- Quantitative regressors: age, dose
- Transformed regressors: √age, log(dose)
- Polynomial regressors: age², age³, · · · (or better: splines)
- Categorical predictors: treatment, sex (dummy variables, contrasts)
- Interaction regessors: treatment × age, sex × age

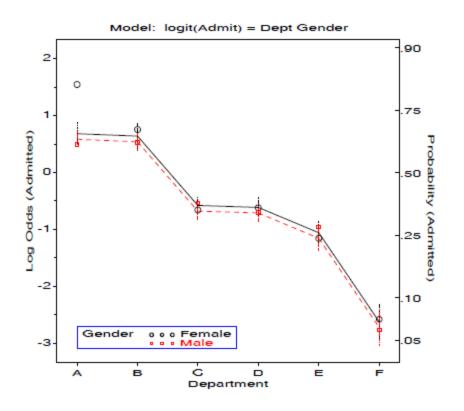
This is exactly the same as in classical ANOVA, regression models

Example: Arthritis treatment



- The response variable, Improved is ordinal: "None" < "Some" < "Marked"</p>
- A binary logistic model can consider just Better = (Improved>"None")
- Other important predictors: Sex, Treatment
- Main Q: how does treatment affect outcome?
- How does this vary with Age and Sex?
- This plot shows the binary observations, with several model-based smoothings

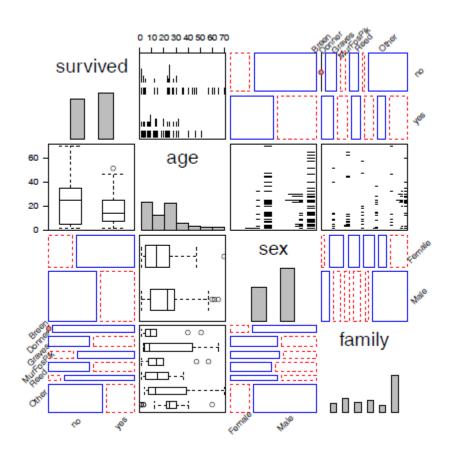
Example: Berkeley admissions



- Admit/Reject can be considered a binomial response for each Dept and Gender
- Logistic regression here is analogous to an ANOVA model, but for log odds(Admit)
- (With categorical predictors, these are often called logit models)
- Every such model has an equivalent loglinear model form.
- This plot shows fitted logits for the main effects model, Dept + Gender

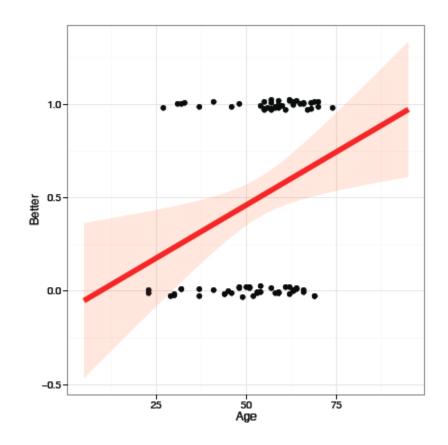
Example: Survival in the Donner party

- Binary response: survived
- Categorical predictors: sex, family
- Quantitative predictor: age
- Q: Is the effect of age linear?
- Q: Are there interactions among predictors?
- This is a generalized pairs plot, with different plots for each pair



Binary response: What's wrong with OLS?

- For a binary response, $Y \in (0, 1)$, want to predict $\pi = Pr(Y = 1 | x)$
- A linear probability model uses classical linear regression (OLS)
- Problems:
 - Gives predicted values and CIs outside $0 < \pi < 1$
 - Homogeneity of variance is violated: $V(\hat{\pi}) = \hat{\pi}(1 \hat{\pi}) \neq$ constant
 - Inferences, hypothesis tests are wrong!



Linear regression vs Logistic regression

OLS regression:

• Assume $y|x \sim N(0, \sigma^2)$

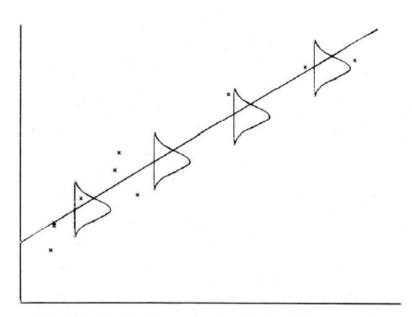


Fig. 2.1. Graphical representation of a simple linear normal regression.

y linear with x constant residual variance

Logistic regression:

Assume Pr(y=1|x) ~ binomial(p)

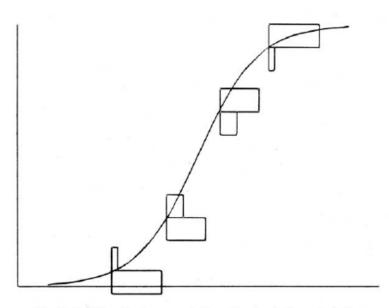
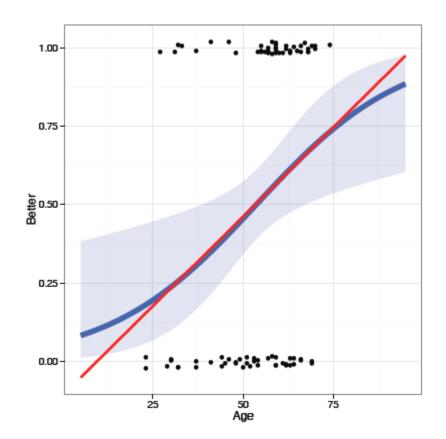


Fig. 2.2. Graphical representation of a simple linear logistic regression.

y ~ logit (x) non-constant residual variance ~ p (1-p)

Logistic regression

- Logistic regression avoids these problems
- Models $logit(\pi_i) \equiv log[\pi/(1-\pi)]$
- logit is interpretable as "log odds" that Y = 1
- A related probit model gives very similar results, but is less interpretable
- For 0.2 ≤ π ≤ 0.8 fitted values are close to those from linear regression.



Logistic regression: One predictor

For a single quantitative predictor, x, the simple linear logistic regression model posits a linear relation between the *log odds* (or *logit*) of Pr(Y = 1) and x,

$$logit[\pi(x)] \equiv log\left(\frac{\pi(x)}{1 - \pi(x)}\right) = \alpha + \beta x$$
.

- When $\beta > 0$, $\pi(x)$ and the log odds increase as x increases; when $\beta < 0$ they decrease with x.
- This model can also be expressed as a model for the probabilities $\pi(x)$

$$\pi(x) = \text{logit}^{-1}[\pi(x)] = \frac{1}{1 + \exp[-(\alpha + \beta x)]}$$

Thinking logistically:

- Model is for the log odds of the marked response, Y = 1
- Can always back transform with logit⁻¹ to get probability of Y = 1

Logistic regression: One predictor

The coefficients, \mathbb{Z} , β of this model have simple interpretations in terms of odds & log odds

$$\operatorname{logit}[\pi(x)] \equiv \operatorname{log}\left(\frac{\pi(x)}{1 - \pi(x)}\right) = \alpha + \beta x \qquad \operatorname{odds}(Y = 1) \equiv \frac{\pi(x)}{1 - \pi(x)} = \exp(\alpha + \beta x) = e^{\alpha}(e^{\beta})^{x}$$

- β is the change in log odds for a unit increase in x
 - \rightarrow The odds of Y=1 are multiplied by e^{β} for each unit increase in x
- α is the log odds when x=0
 - \rightarrow The odds of Y=1 when x=0 is $e^{\mathbb{Z}}$

In R, use exp (coef (model)) to get these values

Another interpretation: In terms of probability, the slope of the logistic regression curve is $\beta\pi(1-\pi)$

This has the maximum value $\beta/4$ when $\pi = \frac{1}{2}$

Logistic regression: Multiple predictors

- For a binary response, $Y \in (0, 1)$, let \boldsymbol{x} be a vector of p regressors, and π_i be the probability, $\Pr(Y = 1 \mid \boldsymbol{x})$.
- The logistic regression model is a linear model for the log odds, or logit that Y = 1, given the values in x,

$$logit(\pi_i) \equiv log\left(\frac{\pi_i}{1 - \pi_i}\right) = \alpha + \mathbf{x}_i^{\mathsf{T}} \beta$$
$$= \alpha + \beta_1 \mathbf{x}_{i1} + \beta_2 \mathbf{x}_{i2} + \dots + \beta_p \mathbf{x}_{ip}$$

• An equivalent (non-linear) form of the model may be specified for the probability, π_i , itself,

$$\pi_i = \left\{1 + \exp(-[\alpha + \mathbf{x}_i^\mathsf{T} \boldsymbol{\beta}])\right\}^{-1}$$

The logistic model is also a multiplicative model for the odds of "success,"

$$\frac{\pi_i}{1 - \pi_i} = \exp(\alpha + \mathbf{x}_i^\mathsf{T} \beta) = \exp(\alpha) \exp(\mathbf{x}_i^\mathsf{T} \beta)$$

Increasing x_{ij} by 1 increases $logit(\pi_i)$ by β_i , and multiplies the odds by e^{β_i} .

Fitting the logistic regression model

Logistic regression models are the special case of generalized linear models, fit in R using glm(..., family=binomial)

For this example, we define **Better** as any improvement at all

```
> data(Arthritis, package="vcd")
> Arthritis$Better <- as.numeric(Arthritis$Improved > "None")
```

Fit and print:

The summary() method gives details and tests of coefficients

```
> summary(arth.logistic)
Call:
glm(formula = Better ~ Age, family = binomial, data = Arthritis)
Deviance Residuals:
   Min
            10 Median 30 Max
-1.5106 -1.1277 0.0794 1.0677 1.7611
Coefficients:
          Estimate Std. Error z value Pr(>|z|)
(Intercept) -2.6421 1.0732 -2.46 0.014 *
Age 0.0492 0.0194 2.54 0.011 *
Signif. codes: 0 '***' 0.001 '**' 0.01 '*' 0.05 '.' 0.1 '' 1
(Dispersion parameter for binomial family taken to be 1)
   Null deviance: 116.45 on 83 degrees of freedom
Residual deviance: 109.16 on 82 degrees of freedom
```

Interpreting coefficients

```
> coef(arth.logistic)
(Intercept) Age
  -2.64207 0.04925
```

```
> exp(coef(arth.logistic))
(Intercept) Age
    0.07121    1.05048
> exp(10*coef(arth.logistic)[2])
    Age
1.636
```

Interpretations:

- log odds(Better) increase by $\beta = 0.0492$ for each year of age
- odds(Better) multiplied by $e^{\beta} = 1.05$ for each year of age— a 5% increase
- over 10 years, odds(Better) are multiplied by $exp(10 \times 0.0492) = 1.64$, a 64% increase.
- Pr(Better) increases by $\beta/4 = 0.0123$ for each year (near $\pi = \frac{1}{2}$)

Multiple predictors

The main interest here is the effect of Treatment. Sex and Age are control variables. Fit the main effects model (no interactions):

$$logit(\pi_i) = \alpha + \beta_1 x_{i1} + \beta_2 x_{i2} + \beta_2 x_{i2}$$

where x_1 is Age and x_2 and x_3 are the factors representing Sex and Treatment, respectively. R uses dummy (0/1) variables for factors.

$$x_2 = \begin{cases} 0 & \text{if Female} \\ 1 & \text{if Male} \end{cases}$$
 $x_3 = \begin{cases} 0 & \text{if Placebo} \\ 1 & \text{if Treatment} \end{cases}$

- \bullet α doesn't have a sensible interpretation here. Why?
- β_1 : increment in log odds(Better) for each year of age.
- β_2 : difference in log odds for male as compared to female.
- β_3 : difference in log odds for treated vs. the placebo group

Multiple predictors: Fitting

Fit the main effects model. Use I(Age − 50) to center Age, making ② interpretable

lmtest::coeftest() gives just the tests of coefficients provided by summary()

broom::glance() gives model fit statistics

Interpreting coefficients

- $\alpha = -0.578$: At age 50, females given placebo have odds(Better) of $e^{-0.578} = 0.56$.
- $\beta_1 = 0.0487$: Each year of age multiplies odds(Better) by $e^{0.0487} = 1.05$, a 5% increase.
- $\beta_2 = -1.49$: Males $e^{-1.49} = 0.26 \times$ less likely to show improvement as females. (Or, females $e^{1.49} = 4.437 \times$ more likely than males.)
- $\beta_3 = 1.76$: Treated $e^{1.76} = 5.81 \times \text{more likely Better than Placebo}$

Hypothesis testing: Questions

• Overall test: How does my model, $logit(\pi) = \alpha + \mathbf{x}^T \boldsymbol{\beta}$ compare with the null model, $logit(\pi) = \alpha$?

$$H_0: \beta_1 = \beta_2 = \cdots = \beta_p = 0$$

One predictor: Does x_k significantly improve my model? Can it be dropped?

 $H_0: \beta_k = 0$ given other predictors retained

 Lack of fit: How does my model compare with a perfect model (saturated model)?

For ANOVA, regression, these tests are carried out using *F*-tests and *t*-tests. In logistic regression (fit by maximum likelihood) we use

- F-tests \rightarrow likelihood ratio G^2 tests
- t-tests \rightarrow Wald z or χ^2 tests

Maximum likelihood estimation

In classical linear models using lm(), we fit using ordinary least squares. All glm() models use maximum likelihood estimation—better properties

- Likelihood, $\mathcal{L} = \Pr(data \mid model)$, as function of model parameters
- For case i,

$$\mathcal{L}_{i} = \begin{cases} p_{i} & \text{if } Y = 1 \\ 1 - p_{i} & \text{if } Y = 0 \end{cases} = p_{i}^{Y_{i}} (1 - p_{i}^{Y_{i}}) \quad \text{where} \quad p_{i} = 1 / (1 + \exp(\mathbf{x}_{i} \boldsymbol{\beta}))$$

Under independence, joint likelihood is the product over all cases

$$\mathcal{L} = \prod_{i}^{n} p_i^{Y_i} (1 - p_i^{Y_i})$$

 $\bullet \implies$ Find estimates $\widehat{\beta}$ that maximize $\log \mathcal{L}$. Iterative, but this solves the "estimating equations"

$$\boldsymbol{X}^{\mathsf{T}}\boldsymbol{y} = \boldsymbol{X}^{\mathsf{T}}\widehat{\boldsymbol{p}}$$

Overall model tests

Likelihood ratio test (G^2)

- Compare nested models, similar to F tests in OLS
- Let L_1 = maximized value for our model $logit(\pi_i) = \beta_0 + \mathbf{x}^T_i \boldsymbol{\beta}$ w/ k predictors
- Let L_0 = maximized likelihood for the null model $logit(\pi_i) = \beta_0$ under H_0 : $\beta_1 = \beta_2 = ... = \beta_k$
- Likelihood ratio test statistic:

$$G^{2} = -2\log\left(\frac{L_{0}}{L_{1}}\right) = 2(\log L_{1} - \log L_{0}) \sim \chi_{k}^{2}$$

Wald tests & confidence intervals

- Analogous to t-tests in OLS
- Test H_0 : $\beta_i = 0$

$$z = \frac{b_i}{s(b_i)} \sim \square (0,1)$$
 or $z^2 \sim \chi_1^2$

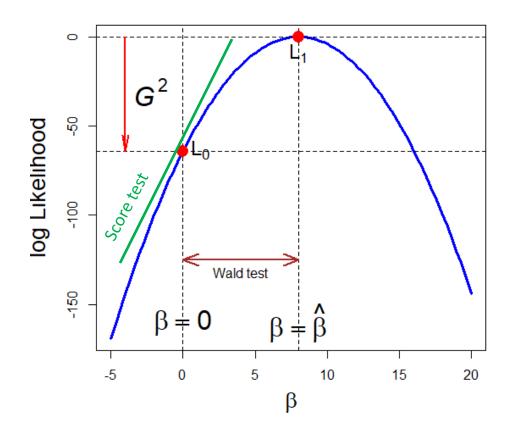
Confidence interval

$$b_i \pm z_{1-\alpha/2} s(b_i)$$

LR, Wald & Score tests

	Testing	Global	Null	Hypothesis:	BETA=0
Test		Chi-Squ	uare	DF	Pr > ChiSq
Likelihood 1	Ratio	24.3	3859	3	<.0001
Score		22.0	0051	3	<.0001
Wald		17.5	5147	3	0.0006

H0:
$$\beta_1 = \beta_2 = \beta_3 = 0$$



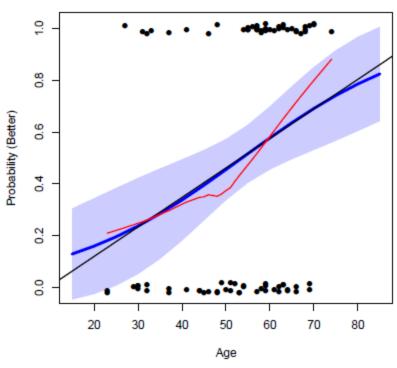
Different ways to measure departure from H_0 : $\beta = 0$

- LR test: diff^{ce} in log L
- Wald test: $(\beta \beta_0)^2$
- Score test: slope at $\beta = 0$

Plotting logistic regression data

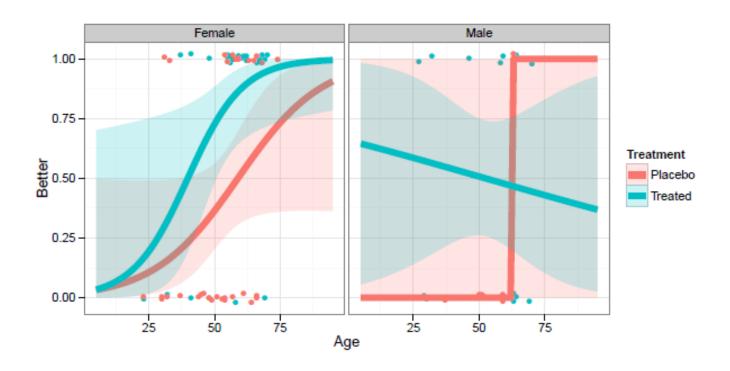
Plotting a binary response together with a fitted logistic model can be difficult because the 0/1 response leads to much overplottling.

- Need to jitter the points
- Useful to show the fitted logistic curve
- Confidence band gives a sense of uncertainty
- Adding a non-parametric (loess) smooth shows possible nonlinearity
- NB: Can plot either on the response scale (probability) or the link scale (logit) where effects are linear



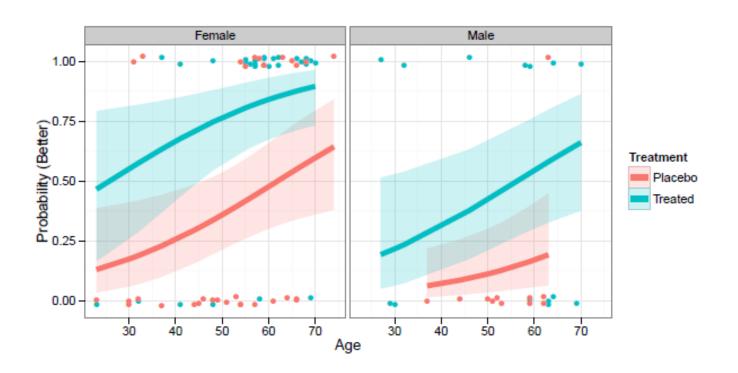
Types of plots

 Conditional plots: Stratified plot of Y or logit(Y) vs. one X, conditioned by other predictors--- only that subset is plotted for each panel



Types of plots

 Full-model plots: Plot of fitted response surface, showing all effects; usually shown in several panels



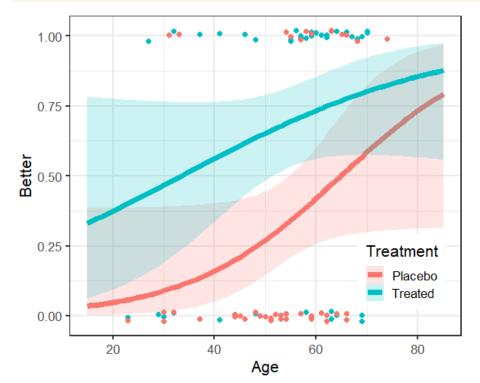
Types of plots

 Effect plots: plots of predicted effects for terms in the model, averaged over predictors not shown in a given plot



Conditional plots with ggplot2

Plot Arthritis data by Treatment, ignoring Sex; overlay fitted logistic reg. lines

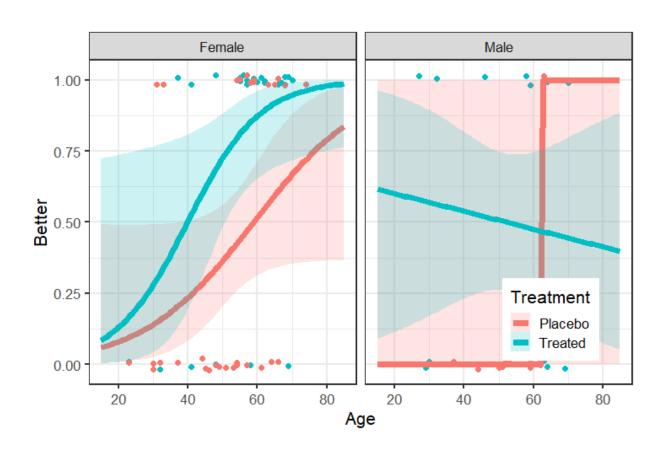


geom_jitter() shows the observations
more distinctly

Fitted lines use method="glm", family=binomial

Conditional plots with ggplot2

Can show the conditional plots for M & F, simply by faceting by Sex



Only the data for each Sex is used in each plot

Plotting the data points shows that the data for males is too thin to give good estimates of separate regression

Full-model plots

Full-model plots show the fitted values on the logit scale or on the response scale (probability), usually with confidence bands. This often requires a bit of custom programming.

Steps:

- Obtain fitted values with predict (model, se.fit=TRUE) type="link" (logit) is the default
- Can use type="response" for probability scale
- Join this to your data (cbind())
- Plot as you like: plot (), ggplot (), ···

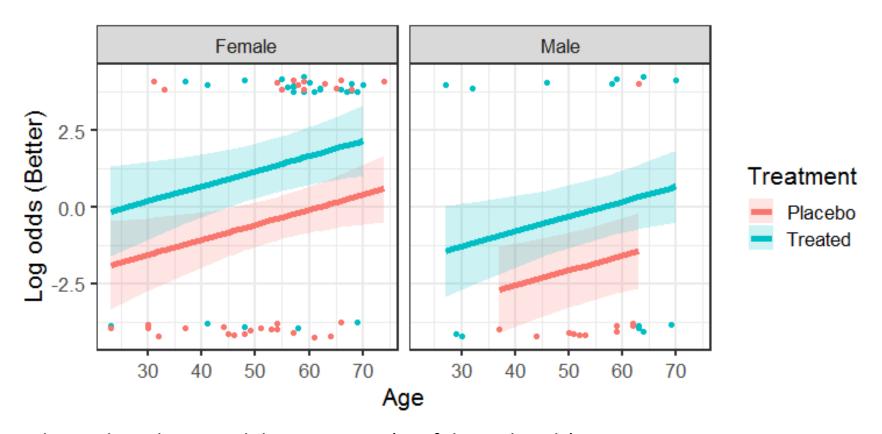
Plotting with ggplot2

Plot the fitted log odds, confidence band and observations

Using color=Treatment gives separate points and lines for the two groups

Full-model plot

Plotting on the logit scale shows the additive effects of age, treatment and sex NB: easier to compare the treatment groups within the same panel

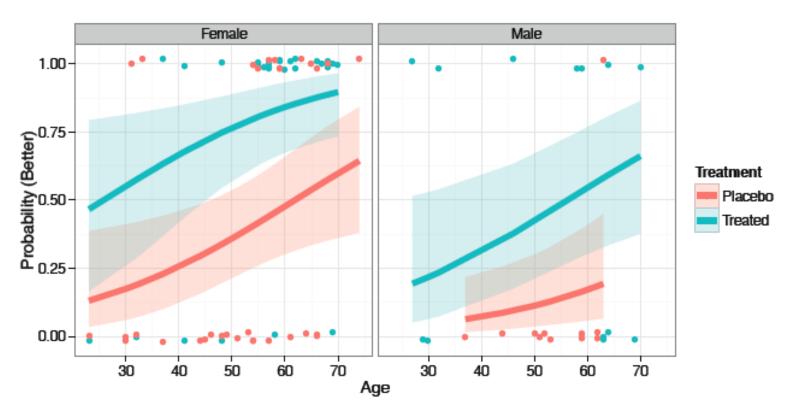


These plots show model uncertainty (confidence bands) Jittered points show the data

Full-model plot

Plotting on the probability scale may be simpler to interpret

Use predict (... type = "response") to get fitted probabilities



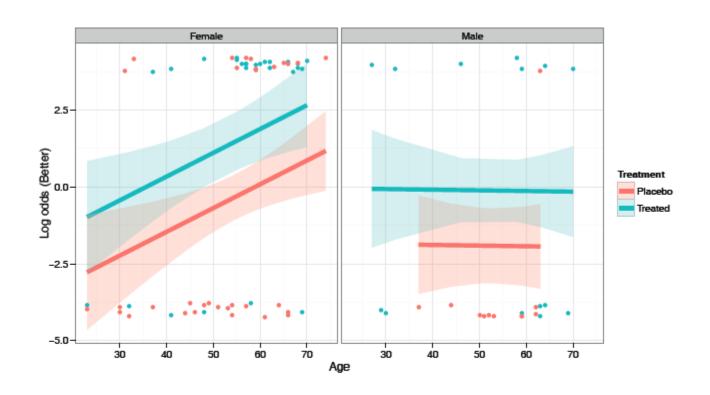
Models with interactions

Is the linear effect of age the same for females, males?

- We can test this by adding an interaction of Sex × Age
- update() makes it easy to add/subtract terms from a model
- car::Anova() gives partial tests of each term after all others

The interaction term Age: Sex is not quite significant, but plot the fitted model anyway

Models with interactions

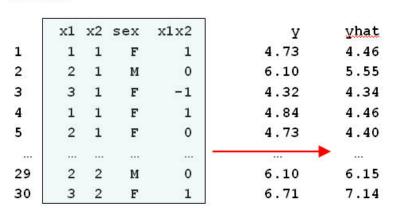


- Only the model changes
- predict () automatically incorporates the revised model terms
- Plotting steps remain the same
- This interpretation is quite different!

Effect plots: Basic ideas

Show a given marginal effect, controlling / adjusting for other model effects

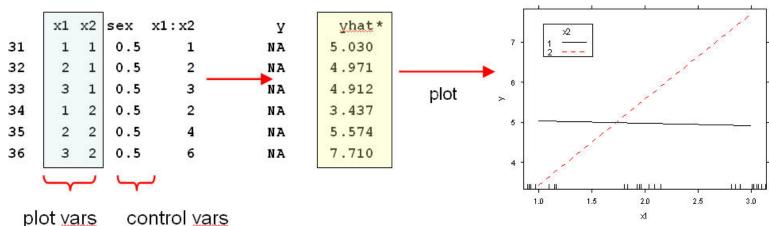
Data



- Xβ̂⇒ŷ Fit data:
- •Score data $\mathbf{X} * \hat{\mathbf{\beta}} \Rightarrow \hat{\mathbf{y}} *$
 - plot vars: vary over range
 - control yars: fix at means

x1*x2 effect plot

Score data



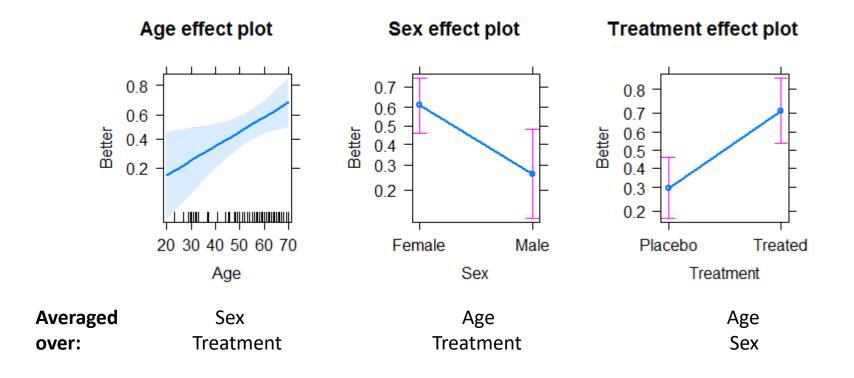
Effect plots: Details

- For simple models, full model plots show the complete relation between response and all predictors.
- Fox(1987)— For complex models, often wish to plot a specific main effect or interaction (including lower-order relatives)— controlling for other effects
 - Fit full model to data with linear predictor (e.g., logit) $\eta = \mathbf{X}\beta$ and link function $g(\mu) = \eta \rightarrow \text{estimate } \mathbf{b}$ of β and covariance matrix $\widehat{V(\mathbf{b})}$ of \mathbf{b} .
 - Construct "score data"
 - Vary each predictor in the term over its' range
 - Fix other predictors at "typical" values (mean, median, proportion in the data)
 - → "effect model matrix," X*
 - Use predict () on X*
 - Calculate fitted effect values, η̂* = X* b.
 - Standard errors are square roots of diag $X^* \widehat{V(b)} X^{*T}$
 - Plot $\hat{\eta}^*$, or values transformed back to scale of response, $g^{-1}(\hat{\eta}^*)$.
- Note: This provides a general means to visualize interactions in all linear and generalized linear models.

Plotting main effects

allEffects() calculates effects for all high-order terms in the model The response is plotted on the logit scale, but labeled with probabilities

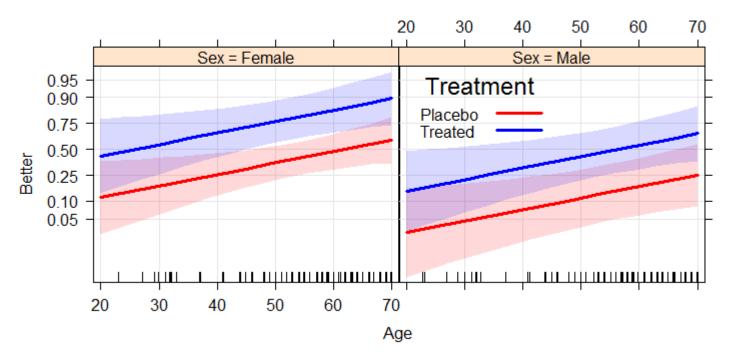
```
library(effects)
arth.eff2 <- allEffects(arth.logistic2)
plot(arth.eff2, rows=1, cols=3, lwd=2)</pre>
```



Full-model plot

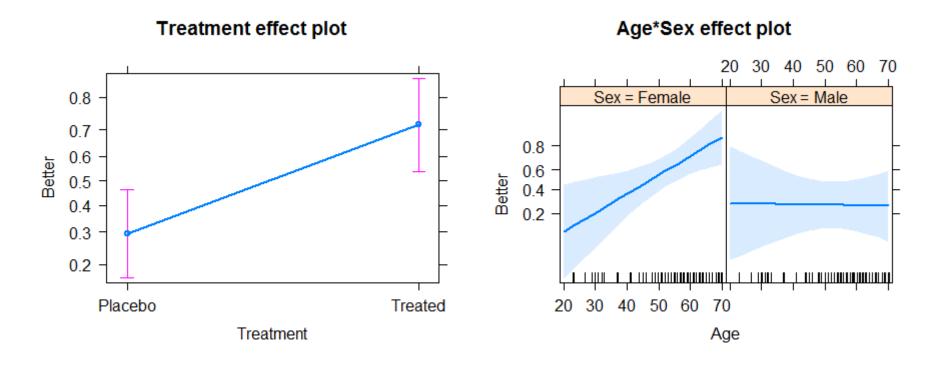
The full-model plot is simply the **Effect()** of the highest-order interaction of factors

Age*Treatment*Sex effect plot



Model with interaction of Age × Sex

```
arth.eff4 <- allEffects(arth.logistic4)
plot(arth.eff4, lwd=2)</pre>
```



Only the high-order terms: Treatment & Age * Sex are shown & need to be interpreted Q: How would you describe this?

Race & Crime

Toronto Star investigation of racial disparities in treatment by Toronto Police Services

FOI request \rightarrow > ½ M arrests, 1997-2002

Evidence for racial profiling?

Only look at discretionary charges:

Simple marijuana possession Non-moving auto infractions

THE SATURDAY STAR



Periods of rain; windy. High 14 C

- Waterfront: Dreams of what could be, B1, B4-5
- Hydro woes: Insulating against price spikes, E1
- Wheels: The Bug goes roofless, G1 Paul Martin: The man who would be king, H1
- Carol Shields: Her last novel? Unless . . . J1

thestar .com ONTARIO EDITION

Chrétien

expected

to keep

cabinet

minister

Ethics report has 'wiggle room' to

save MacAulay BY TIM HARPER

AND LES WHITTINGTON OTTAWA - Jean Chrétien receives a report from his ethics counsellor today that is expected to give him enough "wiggle room" to keep his solicitor-general, Lawrence MacAulay, in the federal cabinet. Ethics counsellor Howard

Wilson completed his report and delivered it to the Prime Minister's Office last night where it was received by Chrétien's chief of staff, Percy It was then to be relayed to Chrétien by secure fax to Bei-rut, where the Prime Minister

is attending a summit of French-speaking nations. It was 1:30 a.m. in Beirut when

the fax arrived so Chrétien

Senior sources said last night that unless there is a surprise in Wilson's report, the Prince Edward Island minister will remain, Chrétien will return to Ottawa and weather the inevi-

table storm of opposition and media protest and forge ahead

with an ethics package by mid-

ing whether MacAulay broke ethics guidelines for cabinet

ministers in the awarding of a

\$100,000 to Everett Roche, a

Charlottetown political friend of the solicitor-general's.

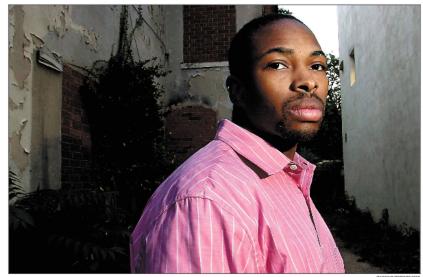
Chrétien will not fire MacAu-

lay unless he is given incontro-

vertible evidence of wrongdo-

ing for two key reasons, sourc

AN INVESTIGATION INTO RACE AND CRIME



SUING POLICE: Jason Burke, falsely accused of dealing drugs during Caribana two years ago, says he was a victim of racial profiling.

Singled out Star analysis of police crime data shows

Telling numbers

Police records show that a black person in Toronto arrested on a single drug possession charge was **less likely** to be released at the scene...



a white person on

to be held for a ball hearing, compared to

Blacks arrested by Toronto po-lice are treated more harshly than crime data shows.

Black people, charged with sim-ple drug possession, are taken to police stations more often than

bail hearing, at twice the rate of

ing a traffic stop. This difference, cess to these numbers was grantsay civil libertarians, community ed to anyone outside the police

■ Managing Editor's notebook, A2

justice is different for blacks and whites

leaders and criminologists, suggests police use racial profiling in deciding whom to pull over.

The evidence is contained in a whites facing the same charge. massive police database recording Once at the station, accused more than 480,000 incidents in blacks are held overnight, for a which an individual was arrested, police. or ticketed, for an offence dating back to 1996. It included almost The Toronto crime data also 800,000 criminal and other charg- person's skin has nothing to do shows a disproportionate number es. The Star obtained that data with h of black motorists are ticketed for through a freedom of information ficers. violations that only surface follow- request, marking the first time ac-

Police are forbidden, by their overning board, from analyzing this data in terms of race, but The Star has no such restriction. The findings provide hard evidence of what blacks have long suspected race matters in Canadian society especially when dealing with

Chief Julian Fantino disputed the findings, saying the colour of a "We don't treat people different-

Please see Toronto, A12



F Please see MacAulay, A8

Case study: Arrests for marijuana

- In Dec. 2002, the *Toronto Star* examined the issue of racial profiling, by analyzing a data base of 600,000+ arrest records from 1997-2002.
- They focused on a subset of arrests for which police action was discretionary, e.g., simple possession of small quantities of marijuana, where the police could:
 - Release the arrestee with a summons like a parking ticket
 - Bring to police station, hold for bail, ... -- harsher treatment
- Response variable: released: "Yes", "No"
 - Main predictor of interest: skin-colour of arrestee (black, white)
 - Other predictors: year, age, sex, ...

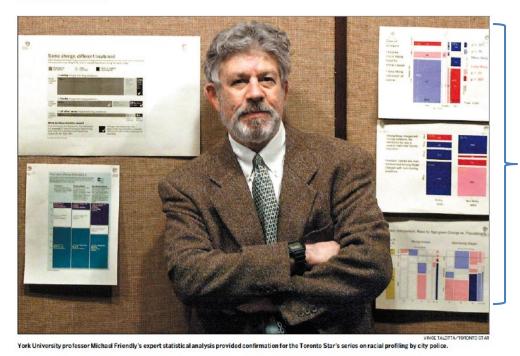
Toronto Star meets mosaic displays

How to communicate these results most effectively?

• What is the message? What features are directly comprehensible to the audience?

Race and Crime

Graphic designer's early attempts

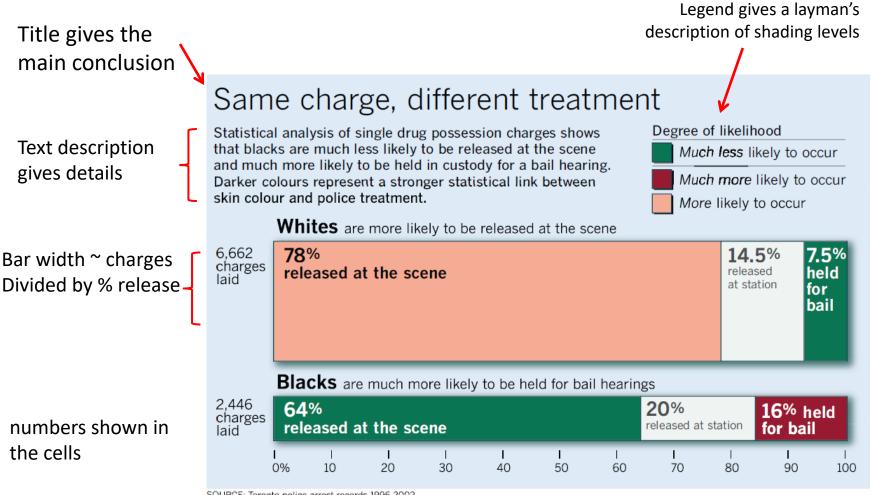


My early attempts

Man behind the numbers

Racial profiling: Presentation graphic

Together, we created this (nearly) self-explaining infographic



Arrests for marijuana: Data

Response variable: released

Control variables:

- year, age, sex
- employed, citizen: Yes, No
- checks: # of police databases (previous arrests, convictions, parole status) where the arrestee's name was found

```
> library(car)
                    # for Anova()
> data(Arrests, package = "carData")
> some(Arrests)
    released colour year age sex employed citizen checks
              White 2000 24 Male
218
         Yes
                                      Yes
                                              Yes
                                                       0
1301
          No Black 1999 17 Male
                                      Yes
                                               No
1495
         Yes White 1998 23 Male
                                      Yes
                                              Yes
1732
         Yes Black 2000 18 Male
                                      Yes
                                              Yes
1838
         Yes Black 1997 27 Male
                                       No
                                              Yes
2257
          No White 2001 19 Male
                                       No
                                              Yes
3100
          No Black 2000 19 Male
                                       No
                                              Yes
3843
         Yes White 1999
                          20 Male
                                      Yes
                                              Yes
         Yes Black 1999 26 Male
4580
                                      Yes
                                              Yes
4833
         Yes Black 1998 38 Male
                                      Yes
                                              Yes
                                                       0
```

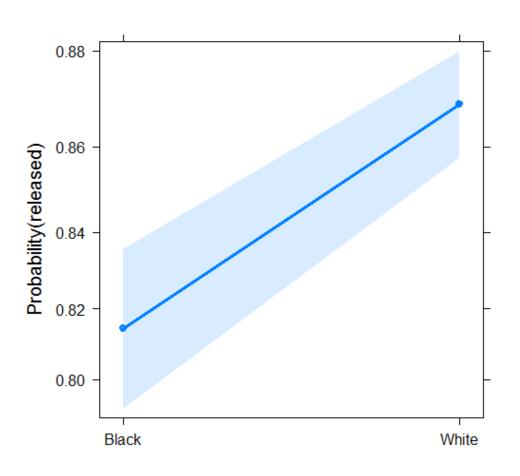
Arrests for marijuana: Model

year is numerical. But may be non-linear. Convert to a factor
Fit model with all main effects, but allow interactions of colour:year and colour:age

```
> Arrests$year <- as.factor(Arrests$year)</pre>
> arrests.mod <- glm(released ~ employed + citizen + checks +
                       colour*year + colour*age,
                       family=binomial, data=Arrests)
> Anova(arrests.mod)
Analysis of Deviance Table (Type II tests)
Response: released
           LR Chisq Df Pr(>Chisq)
              72.7 1 < 2e-16 ***
employed
             25.8 1 3.8e-07 ***
citizen
         205.2 1 < 2e-16 ***
19.6 1 9.7e-06 ***
checks
colour
              6.1 5 0.29785
year
age 0.5 1 0.49827 colour:year 21.7 5 0.00059 ***
colour:age 13.9 1 0.00019 ***
Signif. codes: 0 '***' 0.001 '**' 0.01 '*' 0.05 '.' 0.1 '' 1
```

Effect plot: Skin colour

plot(Effect("colour", arrests.mod), lwd=3, ci.style="bands", ...)



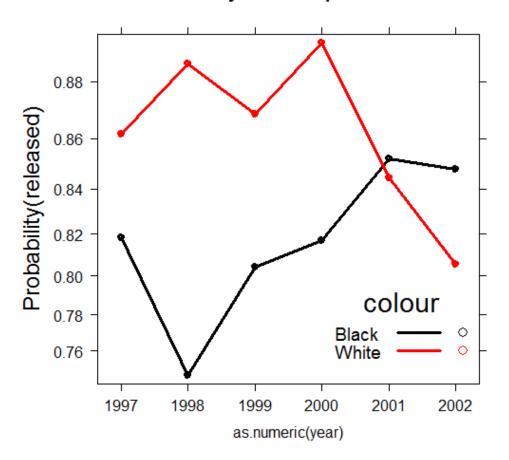
- Effect plot for colour shows average effect controlling (adjusting) for all other factors simultaneously
- (The *Star* analysis controlled for these one at a time.)
- → Evidence for different treatment of blacks & whites
- Even Francis Nunziata could understand this.
- However, effect smaller than reported by the Star

Effect plots: Interactions

The story turned out to be more nuanced than reported by the *Toronto Star*

```
plot(Effect(c("colour", "year"), arrests.mod), multiline=TRUE, ...)
```

colour*year effect plot



Up to 2000, strong evidence for differential treatment of blacks & whites

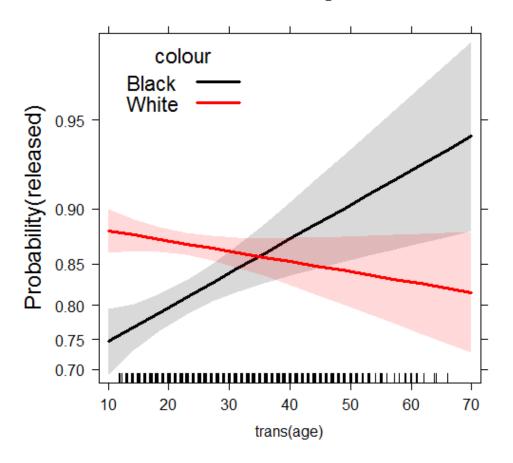
Also: evidence to support Police claim of effect of training to reduce racial effects in treatment

Effect plots: Interactions

A more surprising finding ...

```
plot(Effect(c("colour", "year"), arrests.mod), multiline=TRUE, ...)
```

Effects of skin colour and age on release



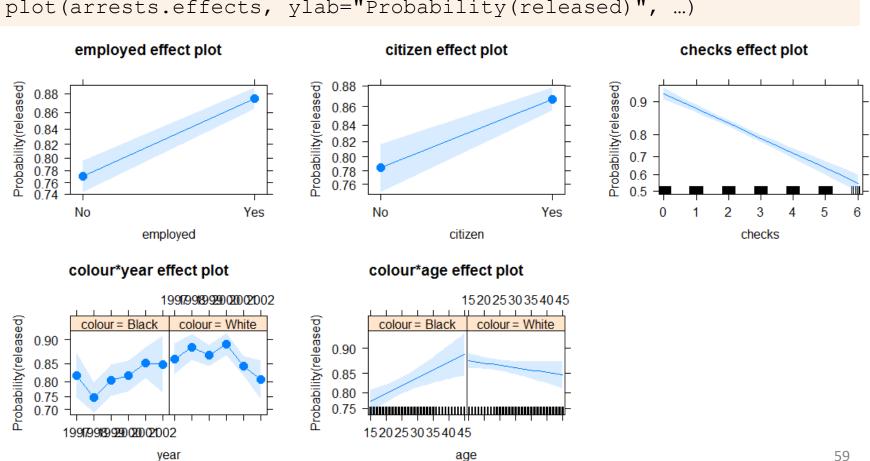
Opposite age effects for blacks & whites:

- Young blacks treated more harshly than young whites
- Older blacks treated less harshly than older whites

Effect plots: allEffects

All high-order terms can be viewed together using plot(allEffects(mod))

```
arrests.effects <- allEffects(arrests.mod,
xlevels=list(age=seq(15,45,5)))
plot(arrests.effects, ylab="Probability(released)", ...)</pre>
```



Model diagnostics

As in regression and ANOVA, the validity of a logistic regression model is threatened when:

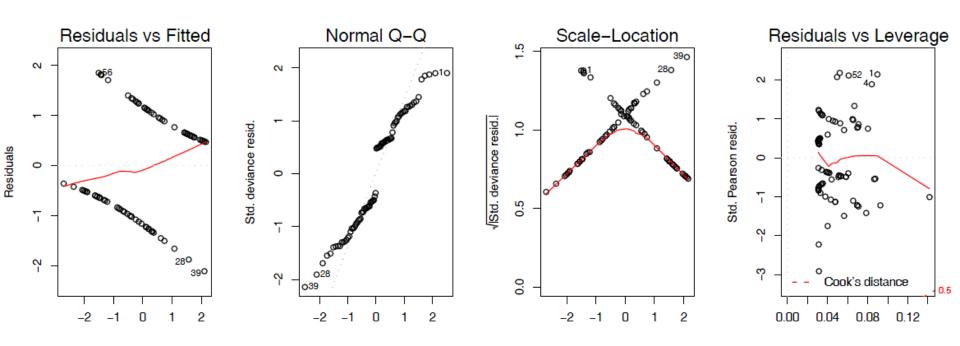
- Important predictors have been omitted from the model
- Predictors assumed to be linear have non-linear effects on Pr(Y = 1)
- Important interactions have been omitted
- A few "wild" observations have a large impact on the fitted model or coefficients

Model specification: Tools and techniques

- Use non-parametric smoothed curves to detect non-linearity
- Consider using polynomial terms (X², X³,...) or regression splines (e.g., ns (X, 3))
- Use update (model, ...) to test for interactions—formula: ...

Diagnostic plots in R

In R, plotting a glm object gives the "regression quartet" – 4 basic diagnostic plots

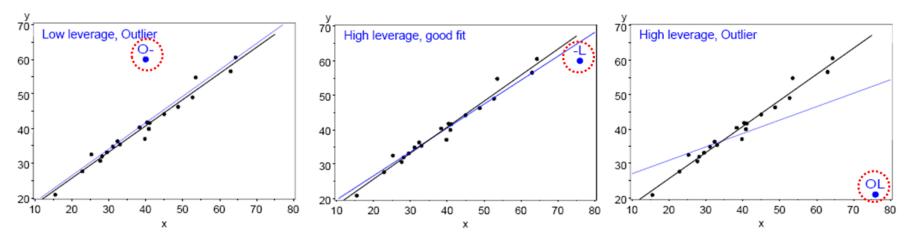


These plots often look peculiar for logistic regression models Better versions are available in the car package

Unusual data: Leverage & Influence

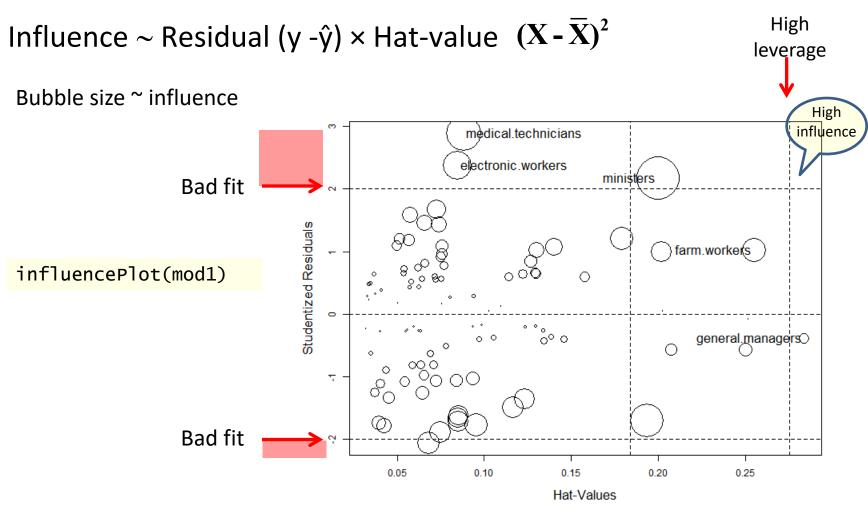
- "Unusual" observations can have dramatic effects on least-squares estimates in linear models
- Three archetypal cases:
 - Typical X (low leverage), bad fit -- Not much harm
 - Unusual X (high leverage), good fit -- Not much harm
 - Unusual X (high leverage), bad fit -- BAD, BAD, BAD
- Influential observations: unusual in both X & Y
- Heuristic formula:

Influence = X leverage × Y residual



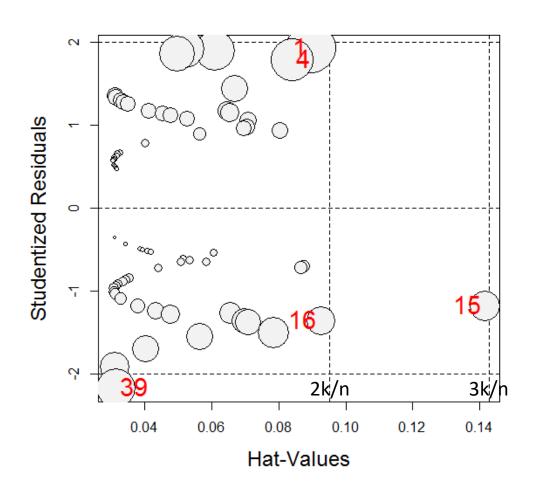
Influence plots

Influence (Cook's D) measures impact of individual obs. on coefficients, fitted values



Influence plots in R

library(car)
influencePlot(arth.logistic2, ...)



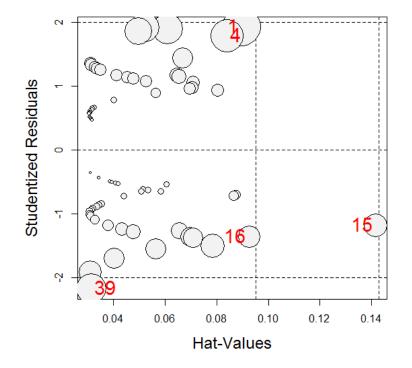
X axis: Leverage ("hat values") notable values: > 2k/n, 3k/n

Y axis: Studentized residuals

Bubble size ~ Cook's D (influence on coefficients)

Which cases are influential?

	Treatment	Sex	Age	Better	StudRes	Hat	CookD
1	Treated	Male	27	1	1.92	0.0897	0.1128
4	Treated	Male	32	1	1.79	0.0840	0.0818
15	Treated	Female	23	0	-1.18	0.1416	0.0420
16	Treated	Female	32	0	-1.36	0.0926	0.0381
39	Treated	Female	69	0	-2.17	0.0314	0.0690



case 1: younger male: moderate Hat, better than predicted → large Cook D

case 39: older female: small Hat, but did not improve with treatment

Looking ahead

- Logistic regression models need not always have linear effects— models nonlinear in Xs sometimes useful
- Polytomous outcomes can be handled as well
 - e.g., Improved = {"None", "Some", "Marked"}
- If ordinal,
 - the proportional odds model is a simple extension
 - nested dichotomies provides an alternative approach
- Otherwise, multinomial logistic regression is the way

Summary

- loglm() provides only overall tests of model fit
- Model-based methods, glm(), provide hypothesis tests, Cls & tests for individual terms
- Logistic regression: A glm() for a binary response
 - linear model for the log odds Pr(Y=1)
 - All similar to classical ANOVA, regression models
- Plotting
 - Conditional, full-model plots show data and fits
 - Effect plots show predicted effects averaged over others
- Model diagnostics
 - Influence plots are often informative