



Challenge tests: Validation of a Thermal Treatment

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Objectives

You will learn:

- How to employ the microbial kinetic parameters (obtained from a challenge test or from the literature) to validate a mild thermal treatment
- How to estimate the evolution of microbial concentration under a given temperature profile (i.e., dynamic conditions)

Materials needed:

- This presentation
- Accompanying spreadsheet “Validation – sausage smoking.xls”

Contents

- Aspects of process validation
- Aspects of challenge testing
- Models for thermal treatment validation
 - Primary model: The loglinear microbial death model
 - Secondary model: The Arrhenius model
 - Tertiary model: the small time-interval approach
- Assessing the effectiveness of a mild thermal treatment (smoking)
 - An example for assessing the effectiveness of smoking to inactivate *Salmonella* in raw sausages
 - Scenario 1: Lower initial contamination in sausages
 - Scenario 2: Higher temperature in the smoking chamber
- Conclusions



Documentation and Traceability

Provides a documented history of the microbiological processes, facilitating traceability



Consistency and Reproducibility

Produce repeatably and reliably the desired product within its specified design parameters.



Product Quality Assurance

Allows microbial risks to be controlled throughout the production process in a systematic manner.

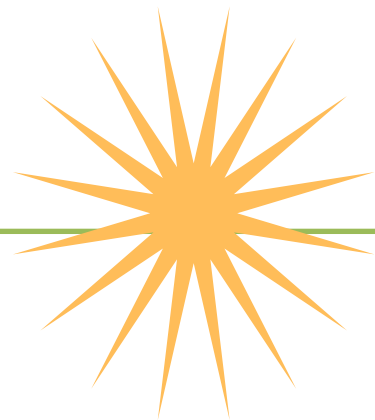
- A microbial challenge test, is used to assess whether a product's preservation or antimicrobial systems are effective
- This test is commonly used to determine a product's ability to resist microbial contamination; and to assess the effectiveness of a thermal or non-thermal treatment to attain certain microbial reduction in the food



Introduction

Aspects of a challenge test

Simulation



Simulates real-life conditions including factors such as temperature, pH, and other environmental conditions that may inhibit microorganisms growth

Methodology



Involves introducing a known quantity of microorganisms into the product and then monitoring the microbial population at regular intervals

Evaluation Criteria



Challenge tests are based on regulatory requirements or industry standards, and compared to predetermined acceptance criteria for microbial load samples in the product under test.

Introduction



The challenge test can be used to evaluate the resistance of a microorganism to a thermal or non-thermal treatment



Results from a challenge test must be first analysed to extract microbial kinetic parameters, which can be later used for prediction and validation of thermal/non-thermal processes

Thermal processes

Raw materials

Preparation

Thermal treatment

In the production of ready-to-eat (RTE) heat-treated food products, the thermal treatment stage is the most common critical control point (CCP) for controlling foodborne pathogens.

Is this step designed to eliminate the hazard or to reduce its occurrence to an acceptable level?

Yes

CCP



Designing a challenge test for thermal treatment

Objective: Validating that the process effectively eliminates target microorganisms at a specified temperature and duration, or at given temperature profile.



Identify target
Microorganism



Establish
Inoculation Level

Designing a challenge test for thermal treatment

Establish Inoculation Levels: Determine the initial microbial levels for the target microorganism. It is essential to maintain a balance between a high enough load to challenge the thermal treatment process but still within a reasonable and allowable range.

Identify Critical Process Parameters: Identify the critical process parameters (CPPs) that significantly influence the effectiveness of the thermal treatment.

Implement Monitoring and Measurement: Incorporate a robust monitoring system to measure critical process parameters during the challenge test.

Calculating death rates based on microbial counts

Fit proper primary and secondary predictive microbiology models: The objective is to characterise how the death rate varies as a function of the temperature (and other relevant intrinsic or extrinsic factors)

Primary model for death:

The loglinear death model is the most common primary model for microbial death used when assessing thermal treatments

Secondary model for death:

The Bigelow model and the Arrhenius model are the most common secondary models used to describe the effect of temperature on microbial death rate

Primary model for death

Loglinear death model: is the time required at certain temperature to reduce the microbial population in one log. It is a measure of thermal resistance.

$$\log_{10} N(t) = \log_{10} N(0) - kt$$

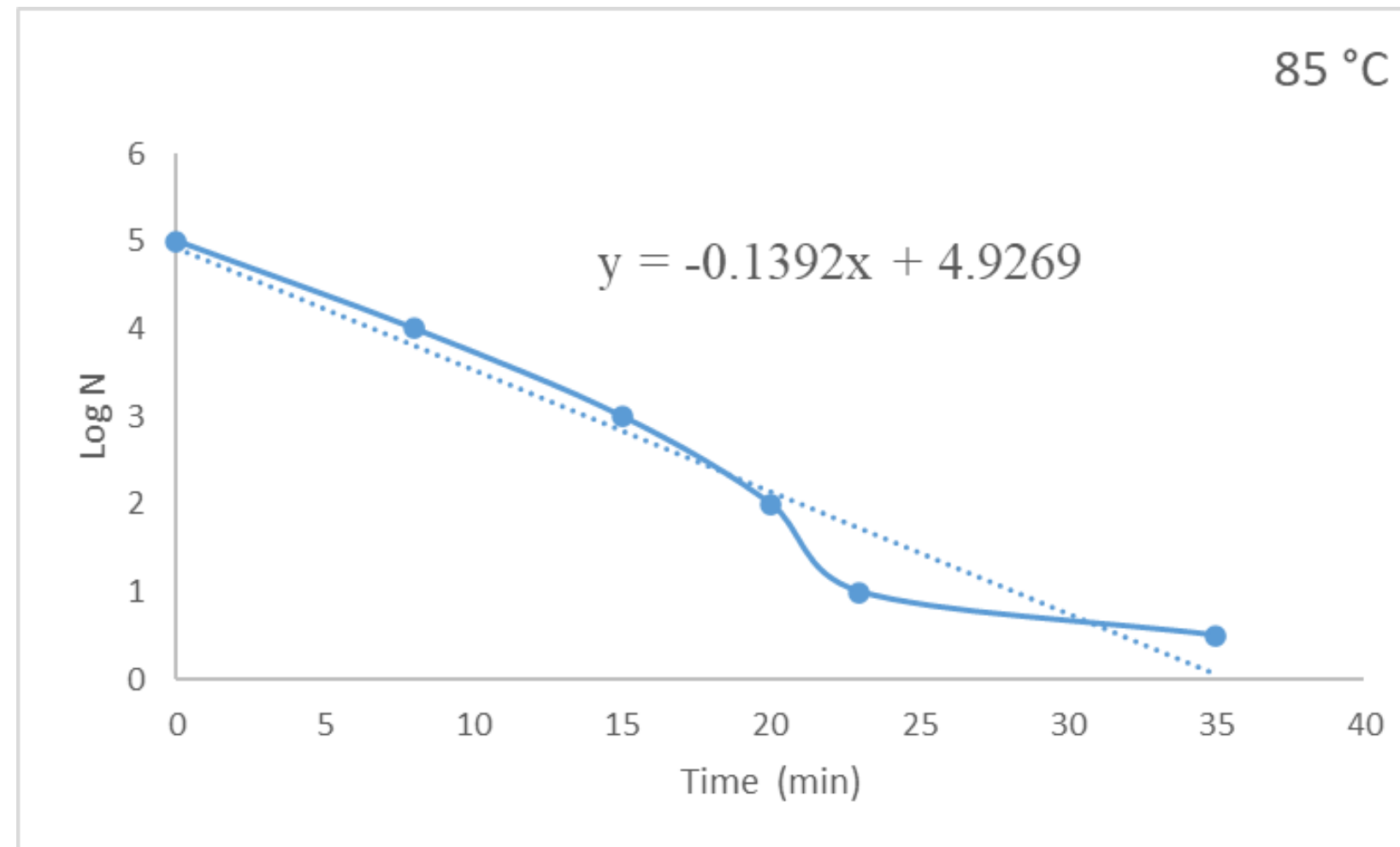
Decimal reduction time (D-Value): is the time required at certain temperature to reduce the microbial population in one log. It is a measure of thermal resistance, and is calculated as the reciprocal of the parameter k

$$D = \frac{1}{K}$$

D values at different storage temperatures are needed to fit the secondary model for death.

Primary model for death

The D-value of bacteria at each temperature can be calculated from the linear regression model for log10 of the surviving bacterial cells and heating time. The D-value is the negative inverse slope of the plot



$$D = \frac{1}{0.1392} = 7.18 \text{ min}$$

Secondary model for death

The Arrhenius equation is a mathematical formula that describes the temperature dependence of reaction rates. It can be applied to biological processes such as how heat affects the inactivation of bacteria. The Arrhenius equation is expressed as follows:

$$D = Ae^{-E_a/RT}$$

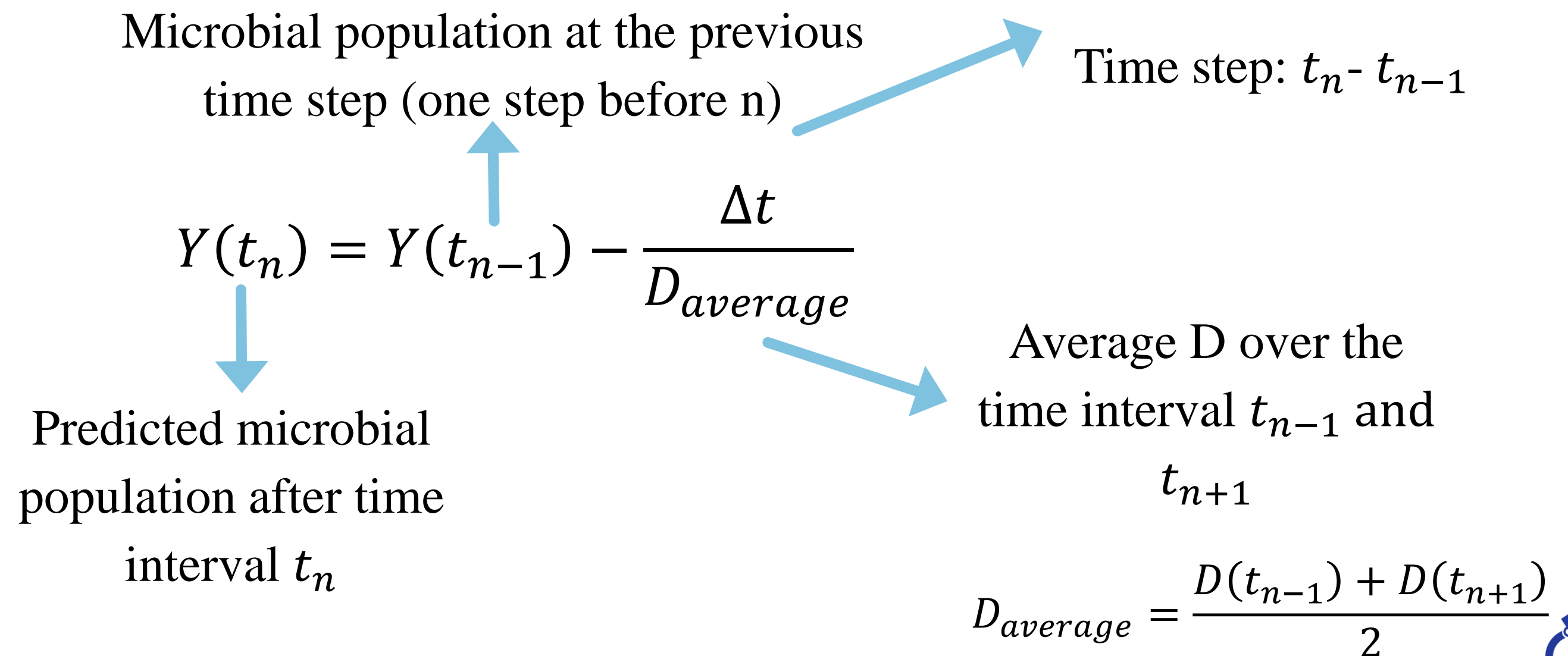
Diagram illustrating the components of the Arrhenius equation:

- D : D value
- A : Rate constant
- E_a : Activation energy (J/mol)
- R : Gas constant (8.314 J/(mol·K))
- T : Absolute temperature in Kelvin

Tertiary model

To predict the change in microbial concentration, as temperature increases (i.e., changes in time), the tertiary model must be solved. This can be done either by a differential equation or by the small time-interval technique

The solution of $Y(t)$ evaluated at a temperature profile, $T(t)$, could be approached as:





Example: Assessing the effectiveness of a mild thermal treatment (smoking) to inactivate *Salmonella* in raw sausages.

Sausages are smoked during 50 minutes in a chamber at air temperature of 65 °C. We wish to know if the time is sufficient to reach a 7.0 log reduction of *Salmonella*.

Open the file “Validation – sausage smoking.xls”

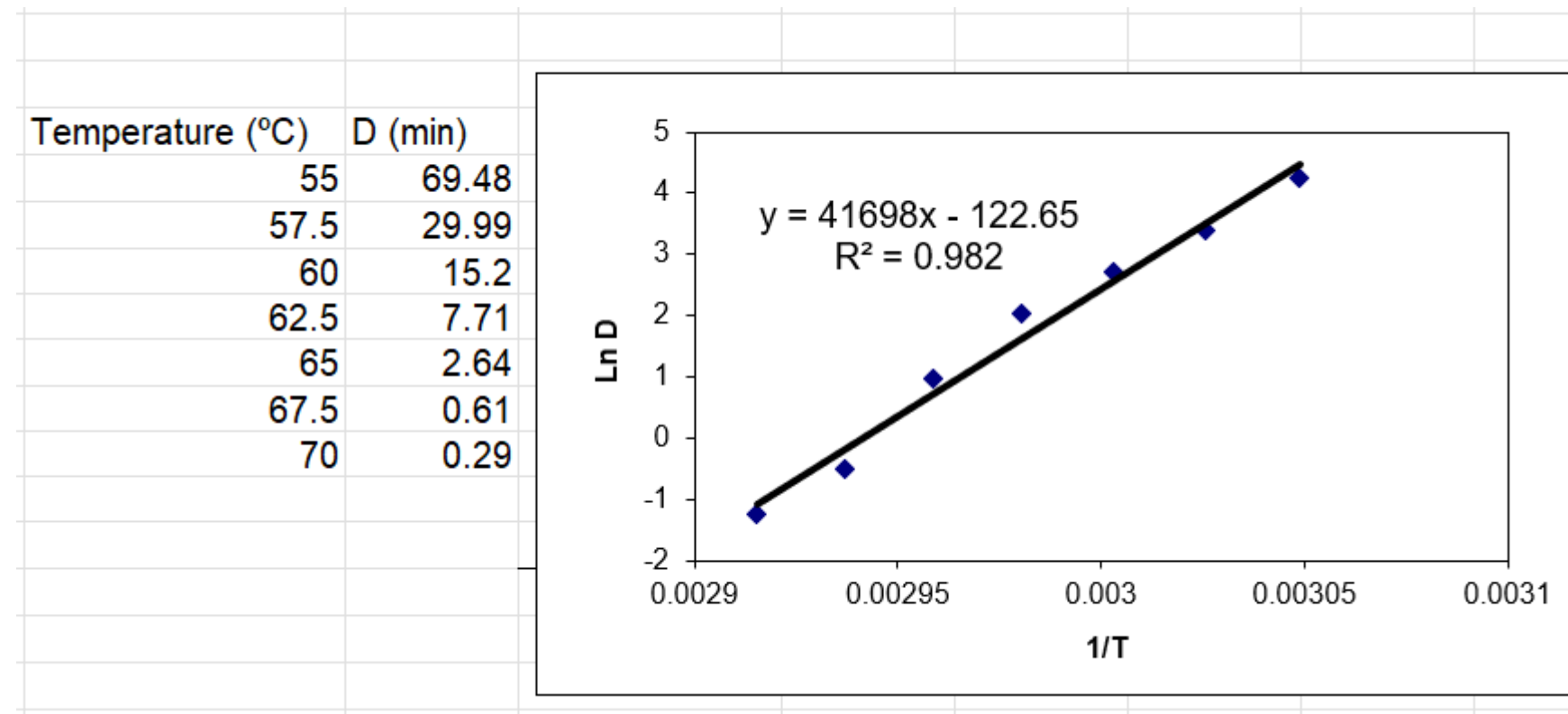
Data on thermal inactivation of *Salmonella* in breaded pork patties was taken from Osaili et al (2007) assuming similar composition to raw sausages

- ❖ Osaili, T. M., Griffis C. L., Martin, E. M., Beard, B. L., Keener, A. E & Marcy, J. A. (2007). Thermal Inactivation of *Escherichia coli* O157:H7, *Salmonella*, and *Listeria monocytogenes* in Breaded Pork Patties. *Journal of Food Science*, 72, 2.



Initial Data:

Linearising Arrhenius equation: $\ln D = \ln A - \frac{E_a}{R} \left(\frac{1}{T} \right)$



$\ln A$ and E_a/R are parameters extracted from the fitted line, now to be used in prediction mode for solving the tertiary model

Step 1

In the excel sheet, the initial parameters **Y0** and **Air temperature** must be set. In addition, set the death kinetic parameters **Ln A** and **Ea/R**, previously estimated

Concentration pre-smoking Y0 (log CFU/g)	Air temperature in smoking chamber (°C)	Ln A (1/min)	Ea/R
7	65 °C	-122.6	41698.09

The table below in the excel sheet is made to provide the predictions of Y(t) for the temperature profile (first and second column)


Microbial death to be solved in small time intervals				
Time (min)	Temperature (°C)	D (min)	Y(t) (log10 CFU/g)	C(t) (log10 CFU/g)
0.000	25			
0.017	25.02			
0.100	25.16			
0.183	25.32			

Step 2

Implement the two equations (1) and (2) in Excel to calculate death rate and microbial population

$$D = \exp\left(\ln A - \frac{Ea}{R} \left(\frac{1}{T}\right)\right) \quad (1)$$

$$Y(t_n) = Y(t_{n-1}) - \frac{\Delta t}{D_{average}} \quad (2)$$



Implement the equation
(1) if $T > 55 \text{ }^{\circ}\text{C}$

We can assume death rate=0
when $T < 55 \text{ }^{\circ}\text{C}$ (assuming
growth below this temperature
is negligible)

Step 2

$$D = \exp\left(\ln A - \frac{Ea}{R} \left(\frac{1}{T}\right)\right) \quad Y(t_n) = Y(t_{n-1}) - \frac{\Delta t}{D_{average}}$$

Microbial death to be solved in small time intervals

Time (min)	Temperature (°C)	D (min)	Y(t) (log10 CFU/g)	C(t) (log10 CFU/g)
7.6	62.96859442	8.273486556	6.868969668	7395536.22
7.6833333	63.09213091	7.904530738	6.858450083	7218551.902
7.7666667	63.20932289	7.56997756	6.847463967	7038238.292
7.85	63.32041199	7.266131311	6.83601687	6855148.546
7.9333333	63.42563644	6.9897553	6.824115627	6669843.238
8.0166667	63.52523077	6.738007818	6.811768249	6482883.981
8.1	63.61942538	6.508387856	6.798983825	6294827.377
8.1833333	63.70844627	6.298688982	6.785772404	6106219.395

Drag theses formula down for the subsequent rows

Step 3

Results

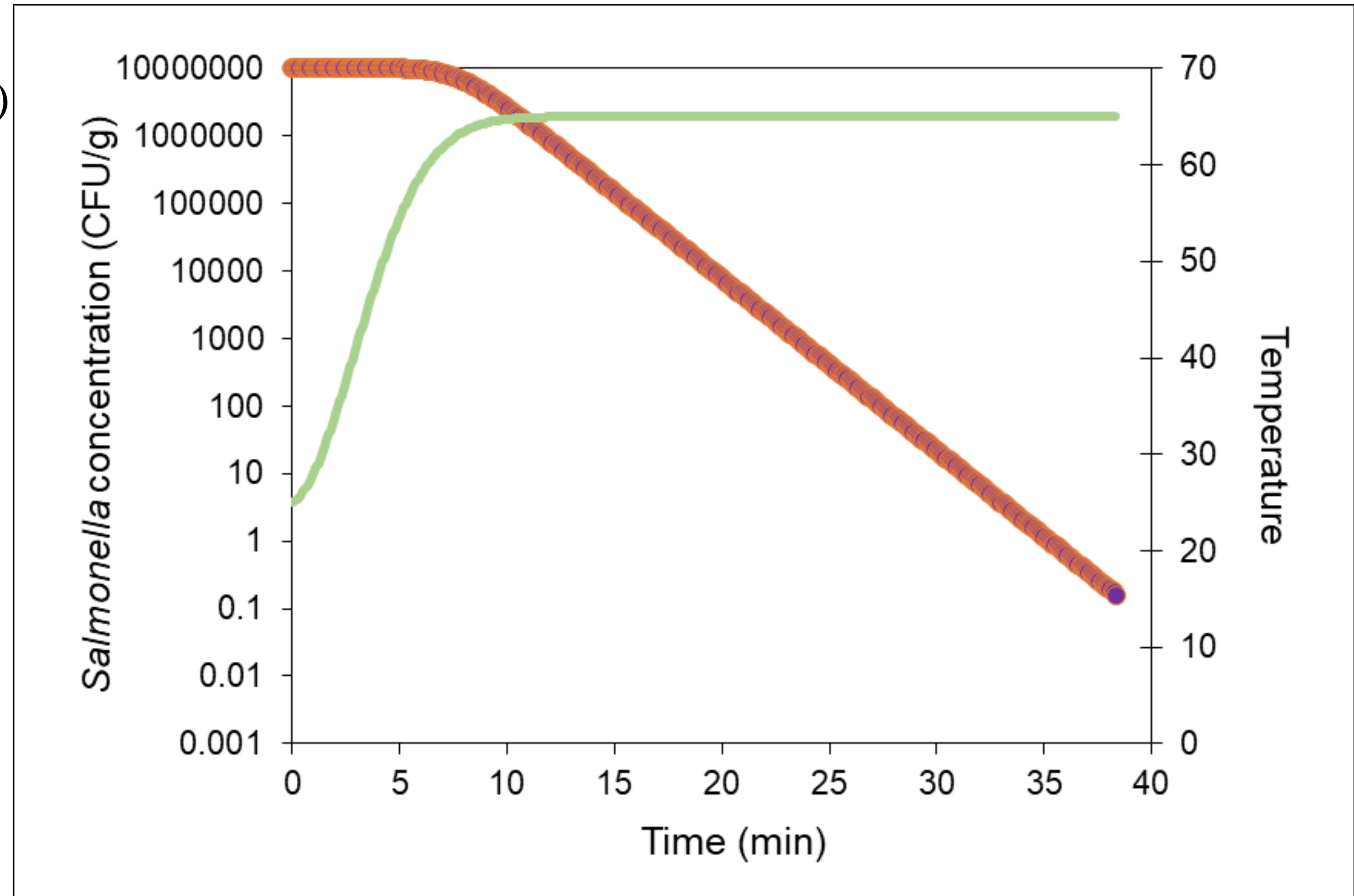
In this scenario, *Salmonella* concentration pre-smoking Y0 was assumed high (7.0 log CFU/g)



It produces a slower decline in population, assuming other factors remain constant.

Notice that a 7-log reduction is attained in ~35 min, therefore lower than 50 min.

The smoking process has been validated



Other scenarios

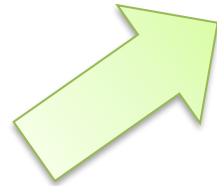
- Let us now compare different scenarios by adjusting parameters such as the microbial concentration pre-smoking and the temperature of the smoking chamber

1) Using the same spreadsheet, simulate *Salmonella* inactivation over time until sausages reach a temperature of **65 °C**, now with an initial concentration of **2 log CFU/g**.

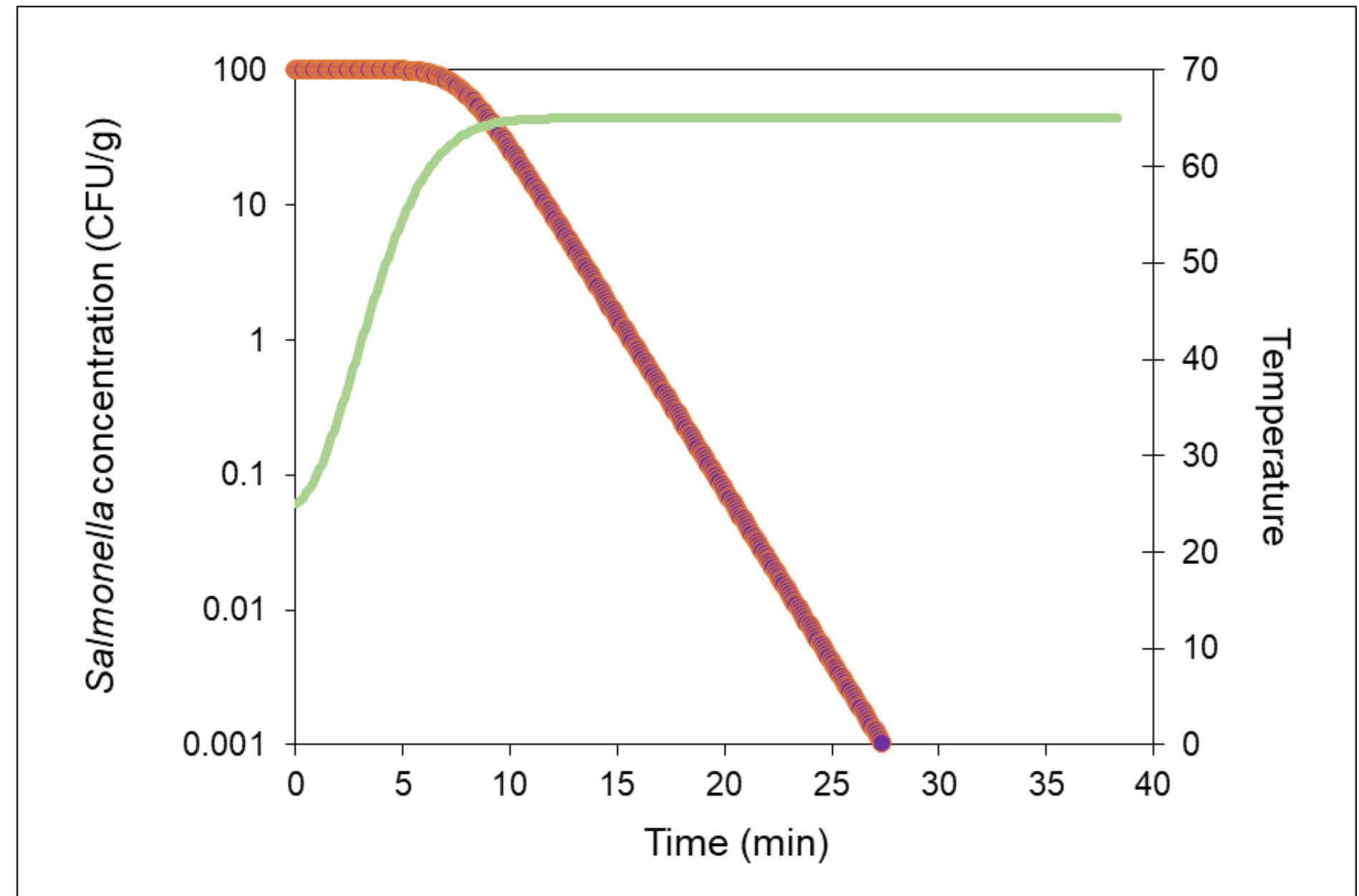
Other scenarios

Parameters used to solve the tertiary model for smoking

Y0 (log CFU/g)	LnA (1/min)	Ea (J/mol)
2	-122	41698.08898



Results

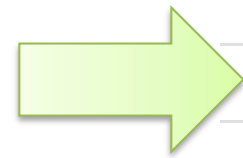


The initial inoculum represents the starting population of microorganisms before smoking. Now *Salmonella* population will die-off well before the end of smoking

Other scenarios

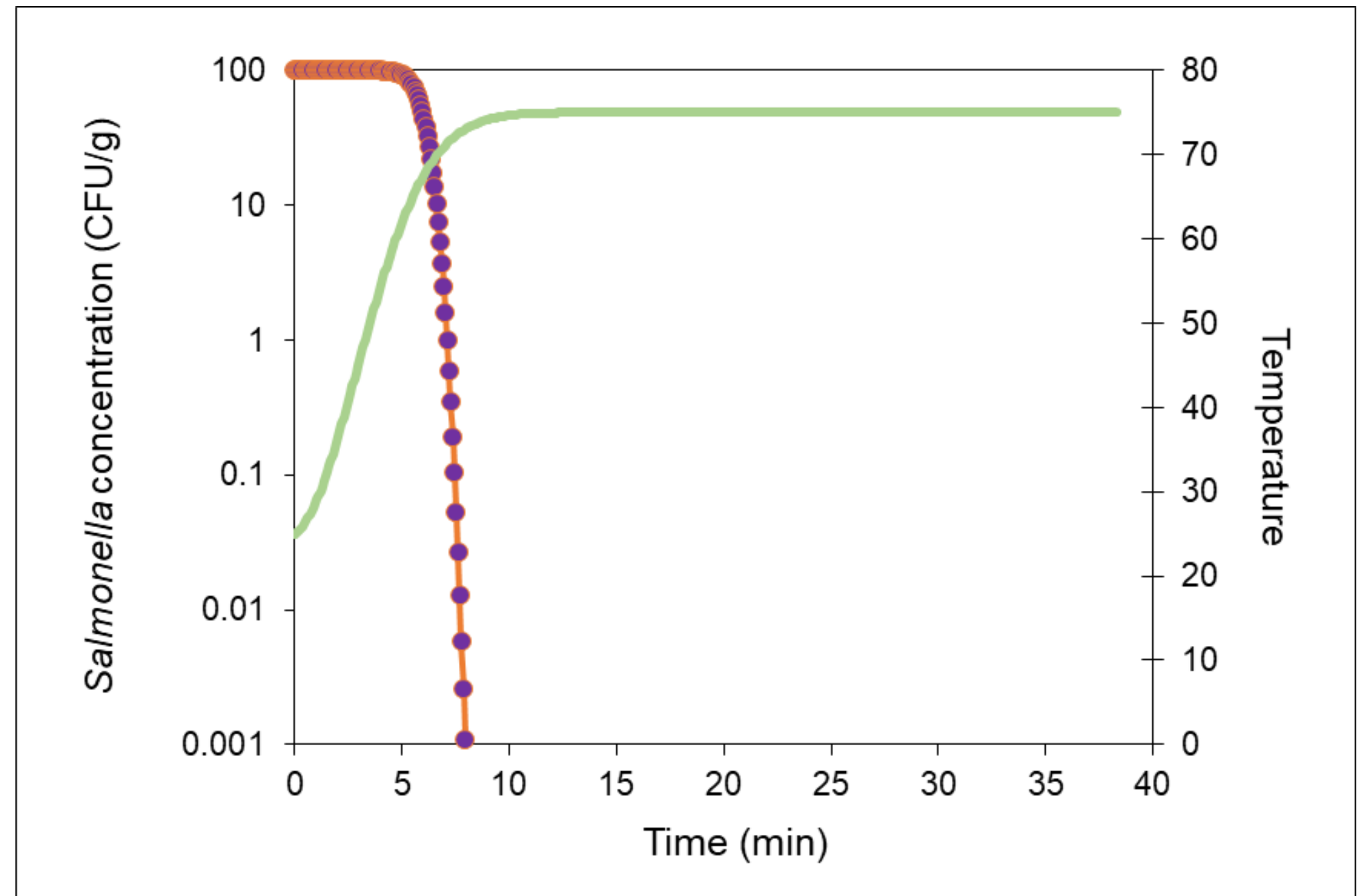
2) Now imagine the smoking chamber temperature has changed to 75 °C.
Simulate *Salmonella* inactivation in smoking sausages over time with an initial concentration of **2 log CFU/g**

Other scenarios



Parameters fixed	
a=	-2.55E-04
b=	-5.66E-06
Air temperature (°C)	75
Initial temperature (°C)	25

Results



Increasing the smoking temperature accelerates the death of *Salmonella* population. From an initial contamination of 100 CFU/g, the concentration dropped to 0.001 CFU/g (1 cell in 1000 kg sausages) in about 8 minutes

Conclusion

- ✓ To validate a thermal (or non-thermal) treatment, microbial kinetic parameters must be obtained from challenge test studies; yet they can be available from the literature or from databases.
- ✓ Many product-related factors (proximate composition, physicochemical properties, packaging) affect the death kinetics of the target microorganism in the food. The microbial kinetic parameters chosen must describe the target microorganism in that particular food.
- ✓ The thermal treatment regime to validate implies that the heating temperature and holding time must be known. Such temperature profile will drive the solution of the microbial concentration in time.
- ✓ The estimation of the microbial concentration in time at a given temperature profile requires the solution of a tertiary model (primary plus secondary), which can be approached by solving the equation for small time intervals.

Further reading

- ❖ Murphy, R.Y., Beard, B.L., Martin, E.M., Duncan, L.K. and Marcy, J.A. (2004). Comparative Study of Thermal Inactivation of *Escherichia coli* O157:H7, *Salmonella*, and *Listeria monocytogenes* in Ground Pork. *Journal of Food Science*.
<https://doi.org/10.1111/j.1365-2621.2004.tb06351.x>.
- ❖ Osaili, T.M., Hasan, F., Dhanasekaran, D.K., Obaid, R.S., Al-Nabulsi, A.A., Rao, S., Fatima, H., Ayyash, M., Savvaidis, I. and Holley, R., (2020). Thermal inactivation of *Escherichia coli* O157:H7 strains and *Salmonella* spp. in camel meat burgers. *LWT*.
<https://doi.org/10.1016/j.lwt.2019.108914>.
- ❖ Pérez-Rodríguez, F., Valero, A. (2013). Predictive Microbiology in Foods. In: Predictive Microbiology in Foods. Springer Briefs in Food, Health, and Nutrition, vol 5. Springer, New York, NY. https://doi.org/10.1007/978-1-4614-5520-2_1.
- ❖ Liu, S., Wei, X., Tang, J., Qin, W. and Wu, Q. (2021). Recent developments in low-moisture foods: microbial validation studies of thermal pasteurization processes, *Critical Reviews in Food Science and Nutrition*.
<https://doi.org/10.1080/10408398.2021.2016601>
- ❖ Wason, S., Verma, T. and Subbiah, J. (2021). Validation of process technologies for enhancing the safety of low-moisture foods. A review. *Compr Rev Food Sci Food Saf*. [Validation of process technologies for enhancing the safety of low-moisture foods](#).



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