

Felix Boos - 2DTPP

SILAC - three replicates - Version 2

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This analysis pipeline is partly based on an analysis workflow developed by Bernd Klaus.

```
#R setup
```

```
##Defining a working directory
```

We defined the working directory for the analysis. This directory should contain the metadata file and the tab delimited text files containing the data.

```
setwd("~/home/path to the right file location")
```

```
##Loading packages
```

```
library(vsn)
library(limma)
library(MSnbase)
library(gplots)
library(fdrtool)
library(biobroom)
library(tidyverse)
```

```
##Defining some functions and parameters
```

```
customPlot <- list(
  theme_bw(base_size = 12),
  scale_fill_manual(values = c("#e41a1c", "#377eb8", "#4daf4a", "#984ea3", "#ff7f00", "#ffff33", "#a65628"),
  scale_colour_manual(values = c("#e41a1c", "#377eb8", "#4daf4a", "#984ea3", "#ff7f00", "#ffff33", "#a65628")
)
script.version = "V2"
dir_save = "data_analysis_results_V2"
```

```
#Loading data and annotating experimental conditions
```

```
conditions <- read.delim("metadata.csv", sep = ",")
# conditions$ID <- paste0("X", conditions$Temperature, "_", conditions$condition, "_", conditions$rep)
files <- file.path(unique(conditions$file))
files <- files[file.exists(files)]
data <- NULL
for (i in seq_along(files))
{
  print(files[i])
  x <- read_delim(file.path(files[i]), delim = "\t")
  names(x) <- make.names(names(x))
  #keep only proteins with at least two unique peptide matches
  x <- x %>%
```

```

dplyr::filter(qupm >= 2)
#remove keratines
x <- x %>%
  dplyr::filter(!grepl("[K, k][R, r][T, t][0-9]", gene_name))
#remove known contaminants
x <- x %>%
  dplyr::filter(!grepl("#+", protein_id))
x$file <- files[i]
keepnames <- as.character(subset(conditions, file == files[i])$col.name)
x <- x[, c("protein_id", "description", "gene_name", "qupm", "top3", "file", keepnames)]
x <- x %>%
  group_by(protein_id, description, gene_name, qupm, top3, file) %>%
  gather(key = "col.name", value = "value", keepnames)
data <- bind_rows(data, x)
rm(x, keepnames)
}

## [1] "T1_190402_light_only_merged_results_20190403_0919_proteins.txt"
## [1] "T2_190402_light_only_merged_results_20190403_0919_proteins.txt"
## [1] "T3_190502_light_only_merged_results_20190503_0911_proteins.txt"
## [1] "T4_190402_light_only_merged_results_20190402_1343_proteins.txt"
## [1] "T5_190402_light_only_merged_results_20190403_0334_proteins.txt"
## [1] "T6_190402_light_only_merged_results_20190403_0439_proteins.txt"
## [1] "T7_190402_light_only_merged_results_20190403_0730_proteins.txt"
## [1] "T8_190402_light_only_merged_results_20190403_0531_proteins.txt"
## [1] "T9_190405_light_only_merged_results_20190405_2250_proteins.txt"
## [1] "T10_190405_light_only_merged_results_20190405_2326_proteins.txt"
## [1] "T12_190408_light_only_merged_results_20190424_1110_proteins.txt"
## [1] "T1_190403_heavy_only_merged_results_20190403_1440_proteins.txt"
## [1] "T2_190403_heavy_only_merged_results_20190403_1440_proteins.txt"
## [1] "T3_190502_heavy_only_merged_results_20190503_0910_proteins.txt"
## [1] "T4_190403_heavy_only_merged_results_20190403_1623_proteins.txt"
## [1] "T5_190403_heavy_only_merged_results_20190403_1618_proteins.txt"
## [1] "T6_190403_heavy_only_merged_results_20190403_1626_proteins.txt"
## [1] "T7_190403_heavy_only_merged_results_20190403_1628_proteins.txt"
## [1] "T8_190403_heavy_only_merged_results_20190403_1616_proteins.txt"
## [1] "T9_190405_heavy_only_merged_results_20190405_2038_proteins.txt"
## [1] "T10_190405_heavy_only_merged_results_20190405_2122_proteins.txt"
## [1] "T12_190408_heavy_only_merged_results_20190424_1112_proteins.txt"
## [1] "P1111_PH_FB TPP_n2_T1_merged_results_light.txt"
## [1] "P1111_PH_FB TPP_n2_T2_merged_results_light.txt"
## [1] "P1111_PH_FB TPP_n2_T3_merged_results_light.txt"
## [1] "P1111_PH_FB TPP_n2_T4_merged_results_light.txt"
## [1] "P1111_PH_FB TPP_n2_T5_merged_results_light.txt"
## [1] "P1111_PH_FB TPP_n2_T6_merged_results_light.txt"
## [1] "P1111_PH_FB TPP_n2_T7_merged_results_light.txt"
## [1] "P1111_PH_FB TPP_n2_T8_merged_results_light.txt"
## [1] "P1111_PH_FB TPP_n2_T9_merged_results_light.txt"
## [1] "P1111_PH_FB TPP_n2_T10_merged_results_light.txt"
## [1] "P1111_PH_FB TPP_n2_T12_merged_results_light.txt"
## [1] "P1111_PH_FB TPP_n2_T1_merged_results_heavy.txt"
## [1] "P1111_PH_FB TPP_n2_T2_merged_results_heavy.txt"
## [1] "P1111_PH_FB TPP_n2_T3_merged_results_heavy.txt"
## [1] "P1111_PH_FB TPP_n2_T4_merged_results_heavy.txt"

```

```

## [1] "P1111_PH_FB TPP_n2_T5_merged_results_heavy.txt"
## [1] "P1111_PH_FB TPP_n2_T6_merged_results_heavy.txt"
## [1] "P1111_PH_FB TPP_n2_T7_merged_results_heavy.txt"
## [1] "P1111_PH_FB TPP_n2_T8_merged_results_heavy.txt"
## [1] "P1111_PH_FB TPP_n2_T9_merged_results_heavy.txt"
## [1] "P1111_PH_FB TPP_n2_T10_merged_results_heavy.txt"
## [1] "P1111_PH_FB TPP_n2_T12_merged_results_heavy.txt"
## [1] "P1111_PH_FB TPP_n3_T1_merged_results_light.txt"
## [1] "P1111_PH_FB TPP_n3_T2_merged_results_light.txt"
## [1] "P1111_PH_FB TPP_n3_T3_merged_results_light.txt"
## [1] "P1111_PH_FB TPP_n3_T4_merged_results_light.txt"
## [1] "P1111_PH_FB TPP_n3_T5_merged_results_light.txt"
## [1] "P1111_PH_FB TPP_n3_T6_merged_results_light_2ndSP2.txt"
## [1] "P1111_PH_FB TPP_n3_T7_merged_results_light.txt"
## [1] "P1111_PH_FB TPP_n3_T8_merged_results_light.txt"
## [1] "P1111_PH_FB TPP_n3_T9_merged_results_light.txt"
## [1] "P1111_PH_FB TPP_n3_T10_merged_results_light.txt"
## [1] "P1111_PH_FB TPP_n3_T12_merged_results_light.txt"
## [1] "P1111_PH_FB TPP_n3_T1_merged_results_heavy.txt"
## [1] "P1111_PH_FB TPP_n3_T2_merged_results_heavy.txt"
## [1] "P1111_PH_FB TPP_n3_T3_merged_results_heavy.txt"
## [1] "P1111_PH_FB TPP_n3_T4_merged_results_heavy.txt"
## [1] "P1111_PH_FB TPP_n3_T5_merged_results_heavy.txt"
## [1] "P1111_PH_FB TPP_n3_T6_merged_results_heavy_2ndSP2.txt"
## [1] "P1111_PH_FB TPP_n3_T7_merged_results_heavy.txt"
## [1] "P1111_PH_FB TPP_n3_T8_merged_results_heavy.txt"
## [1] "P1111_PH_FB TPP_n3_T9_merged_results_heavy.txt"
## [1] "P1111_PH_FB TPP_n3_T10_merged_results_heavy.txt"
## [1] "P1111_PH_FB TPP_n3_T12_merged_results_heavy.txt"

rm(i, files)
data <- left_join(data, conditions)
# write.csv(data, file.path(dir_save, paste0("unfiltered_raw_data_", script.version, ".csv")))
data <- subset(data, value > 0)

#Protein identification overview

sub <- unique(data[, c("gene_name", "file")])
sub <- merge(sub, unique(conditions[, c("file", "rep", "Temperature", "SILAC")]))
sub$found <- 1
sub$rep <- paste("file", as.numeric(factor(sub$file)))
print(unique(with(sub, paste(rep, "-", file))))

## [1] "file 1 - P1111_PH_FB TPP_n2_T1_merged_results_heavy.txt"
## [2] "file 2 - P1111_PH_FB TPP_n2_T1_merged_results_light.txt"
## [3] "file 3 - P1111_PH_FB TPP_n2_T10_merged_results_heavy.txt"
## [4] "file 4 - P1111_PH_FB TPP_n2_T10_merged_results_light.txt"
## [5] "file 5 - P1111_PH_FB TPP_n2_T12_merged_results_heavy.txt"
## [6] "file 6 - P1111_PH_FB TPP_n2_T12_merged_results_light.txt"
## [7] "file 7 - P1111_PH_FB TPP_n2_T2_merged_results_heavy.txt"
## [8] "file 8 - P1111_PH_FB TPP_n2_T2_merged_results_light.txt"
## [9] "file 9 - P1111_PH_FB TPP_n2_T3_merged_results_heavy.txt"
## [10] "file 10 - P1111_PH_FB TPP_n2_T3_merged_results_light.txt"
## [11] "file 11 - P1111_PH_FB TPP_n2_T4_merged_results_heavy.txt"
## [12] "file 12 - P1111_PH_FB TPP_n2_T4_merged_results_light.txt"

```

```

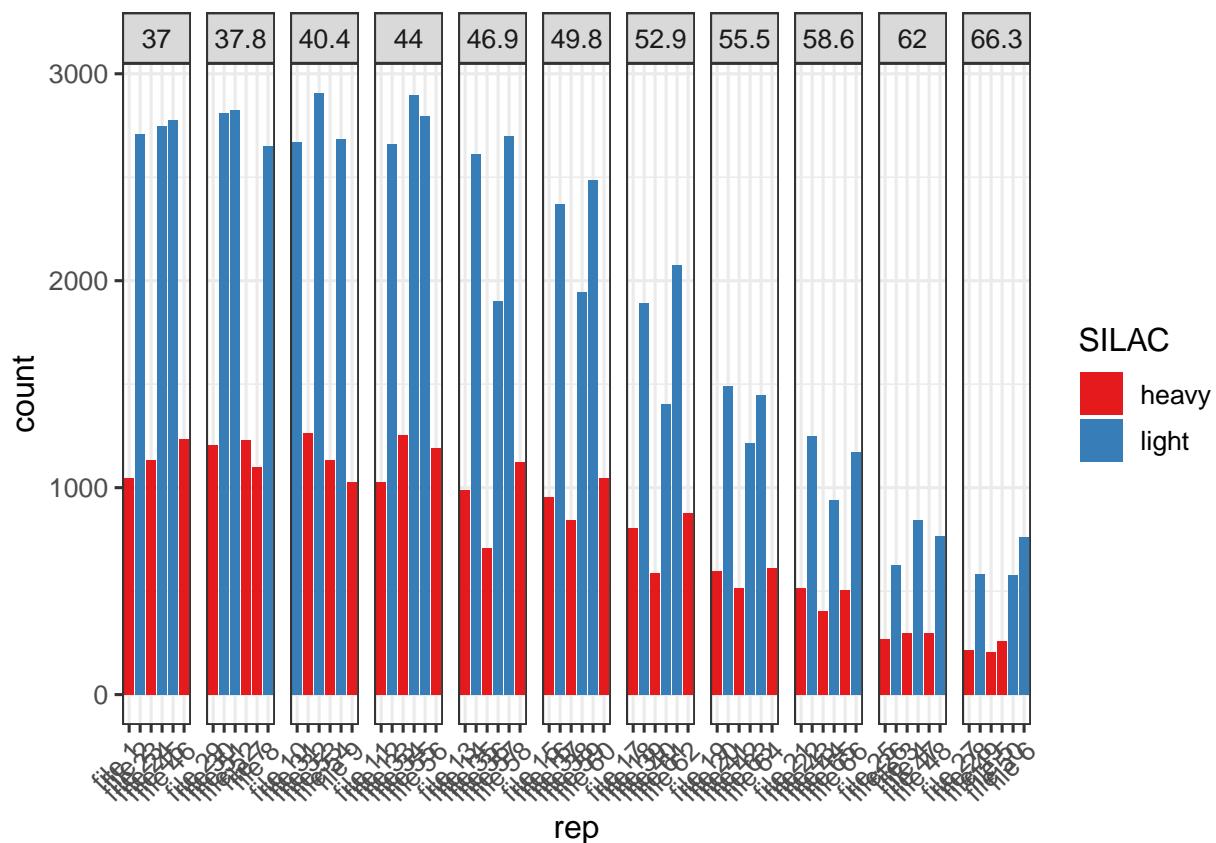
## [13] "file 13 - P1111_PH_FB TPP_n2_T5_merged_results_heavy.txt"
## [14] "file 14 - P1111_PH_FB TPP_n2_T5_merged_results_light.txt"
## [15] "file 15 - P1111_PH_FB TPP_n2_T6_merged_results_heavy.txt"
## [16] "file 16 - P1111_PH_FB TPP_n2_T6_merged_results_light.txt"
## [17] "file 17 - P1111_PH_FB TPP_n2_T7_merged_results_heavy.txt"
## [18] "file 18 - P1111_PH_FB TPP_n2_T7_merged_results_light.txt"
## [19] "file 19 - P1111_PH_FB TPP_n2_T8_merged_results_heavy.txt"
## [20] "file 20 - P1111_PH_FB TPP_n2_T8_merged_results_light.txt"
## [21] "file 21 - P1111_PH_FB TPP_n2_T9_merged_results_heavy.txt"
## [22] "file 22 - P1111_PH_FB TPP_n2_T9_merged_results_light.txt"
## [23] "file 23 - P1111_PH_FB TPP_n3_T1_merged_results_heavy.txt"
## [24] "file 24 - P1111_PH_FB TPP_n3_T1_merged_results_light.txt"
## [25] "file 25 - P1111_PH_FB TPP_n3_T10_merged_results_heavy.txt"
## [26] "file 26 - P1111_PH_FB TPP_n3_T10_merged_results_light.txt"
## [27] "file 27 - P1111_PH_FB TPP_n3_T12_merged_results_heavy.txt"
## [28] "file 28 - P1111_PH_FB TPP_n3_T12_merged_results_light.txt"
## [29] "file 29 - P1111_PH_FB TPP_n3_T2_merged_results_heavy.txt"
## [30] "file 30 - P1111_PH_FB TPP_n3_T2_merged_results_light.txt"
## [31] "file 31 - P1111_PH_FB TPP_n3_T3_merged_results_heavy.txt"
## [32] "file 32 - P1111_PH_FB TPP_n3_T3_merged_results_light.txt"
## [33] "file 33 - P1111_PH_FB TPP_n3_T4_merged_results_heavy.txt"
## [34] "file 34 - P1111_PH_FB TPP_n3_T4_merged_results_light.txt"
## [35] "file 35 - P1111_PH_FB TPP_n3_T5_merged_results_heavy.txt"
## [36] "file 36 - P1111_PH_FB TPP_n3_T5_merged_results_light.txt"
## [37] "file 37 - P1111_PH_FB TPP_n3_T6_merged_results_heavy_2ndSP2.txt"
## [38] "file 38 - P1111_PH_FB TPP_n3_T6_merged_results_light_2ndSP2.txt"
## [39] "file 39 - P1111_PH_FB TPP_n3_T7_merged_results_heavy.txt"
## [40] "file 40 - P1111_PH_FB TPP_n3_T7_merged_results_light.txt"
## [41] "file 41 - P1111_PH_FB TPP_n3_T8_merged_results_heavy.txt"
## [42] "file 42 - P1111_PH_FB TPP_n3_T8_merged_results_light.txt"
## [43] "file 43 - P1111_PH_FB TPP_n3_T9_merged_results_heavy.txt"
## [44] "file 44 - P1111_PH_FB TPP_n3_T9_merged_results_light.txt"
## [45] "file 45 - T1_190402_light_only_merged_results_20190403_0919_proteins.txt"
## [46] "file 46 - T1_190403_heavy_only_merged_results_20190403_1440_proteins.txt"
## [47] "file 47 - T10_190405_heavy_only_merged_results_20190405_2122_proteins.txt"
## [48] "file 48 - T10_190405_light_only_merged_results_20190405_2326_proteins.txt"
## [49] "file 49 - T12_190408_heavy_only_merged_results_20190424_1112_proteins.txt"
## [50] "file 50 - T12_190408_light_only_merged_results_20190424_1110_proteins.txt"
## [51] "file 51 - T2_190402_light_only_merged_results_20190403_0919_proteins.txt"
## [52] "file 52 - T2_190403_heavy_only_merged_results_20190403_1440_proteins.txt"
## [53] "file 53 - T3_190502_heavy_only_merged_results_20190503_0910_proteins.txt"
## [54] "file 54 - T3_190502_light_only_merged_results_20190503_0911_proteins.txt"
## [55] "file 55 - T4_190402_light_only_merged_results_20190402_1343_proteins.txt"
## [56] "file 56 - T4_190403_heavy_only_merged_results_20190403_1623_proteins.txt"
## [57] "file 57 - T5_190402_light_only_merged_results_20190403_0334_proteins.txt"
## [58] "file 58 - T5_190403_heavy_only_merged_results_20190403_1618_proteins.txt"
## [59] "file 59 - T6_190402_light_only_merged_results_20190403_0439_proteins.txt"
## [60] "file 60 - T6_190403_heavy_only_merged_results_20190403_1626_proteins.txt"
## [61] "file 61 - T7_190402_light_only_merged_results_20190403_0730_proteins.txt"
## [62] "file 62 - T7_190403_heavy_only_merged_results_20190403_1628_proteins.txt"
## [63] "file 63 - T8_190402_light_only_merged_results_20190403_0531_proteins.txt"
## [64] "file 64 - T8_190403_heavy_only_merged_results_20190403_1616_proteins.txt"
## [65] "file 65 - T9_190405_heavy_only_merged_results_20190405_2038_proteins.txt"
## [66] "file 66 - T9_190405_light_only_merged_results_20190405_2250_proteins.txt"

```

```

sub <- unique(sub)
ggplot(data = sub, aes(rep, fill = SILAC)) +
  geom_bar() +
  customPlot +
  theme(axis.text.x = element_text(angle = 45, hjust = 1, vjust = 1)) +
  facet_grid(. ~ Temperature, space = "free", scale = "free_x")

```



```

sub$rep <- paste(sub$Temperature, sub$rep)
with(sub, table(rep))

```

```

## rep
##   37 file 1    37 file 2    37 file 23   37 file 24   37 file 45   37 file 46
##     1045        2708       1132        2746       2777       1237
## 37.8 file 29 37.8 file 30 37.8 file 51 37.8 file 52 37.8 file 7 37.8 file 8
##    1208        2811       2826       1230       1101       2650
## 40.4 file 10 40.4 file 31 40.4 file 32 40.4 file 53 40.4 file 54 40.4 file 9
##    2670        1264       2905       1133       2684       1027
## 44 file 11   44 file 12   44 file 33   44 file 34   44 file 55   44 file 56
##    1027        2658       1253       2897       2797       1190
## 46.9 file 13 46.9 file 14 46.9 file 35 46.9 file 36 46.9 file 57 46.9 file 58
##    990         2612       708        1902       2698       1125
## 49.8 file 15 49.8 file 16 49.8 file 37 49.8 file 38 49.8 file 59 49.8 file 60
##    956         2370       845        1947       2486       1047
## 52.9 file 17 52.9 file 18 52.9 file 39 52.9 file 40 52.9 file 61 52.9 file 62
##    803         1893       588        1406       2074       879
## 55.5 file 19 55.5 file 20 55.5 file 41 55.5 file 42 55.5 file 63 55.5 file 64

```

```

##      598       1489       517      1214      1446       609
## 58.6 file 21 58.6 file 22 58.6 file 43 58.6 file 44 58.6 file 65 58.6 file 66
##      517       1250       403       942       504      1170
## 62 file 25   62 file 26   62 file 3    62 file 4   62 file 47   62 file 48
##     267       627        299       842       295      767
## 66.3 file 27 66.3 file 28 66.3 file 49 66.3 file 5 66.3 file 50 66.3 file 6
##     217       583       206       261       579      763

rm(sub.sub)
rm(sub)

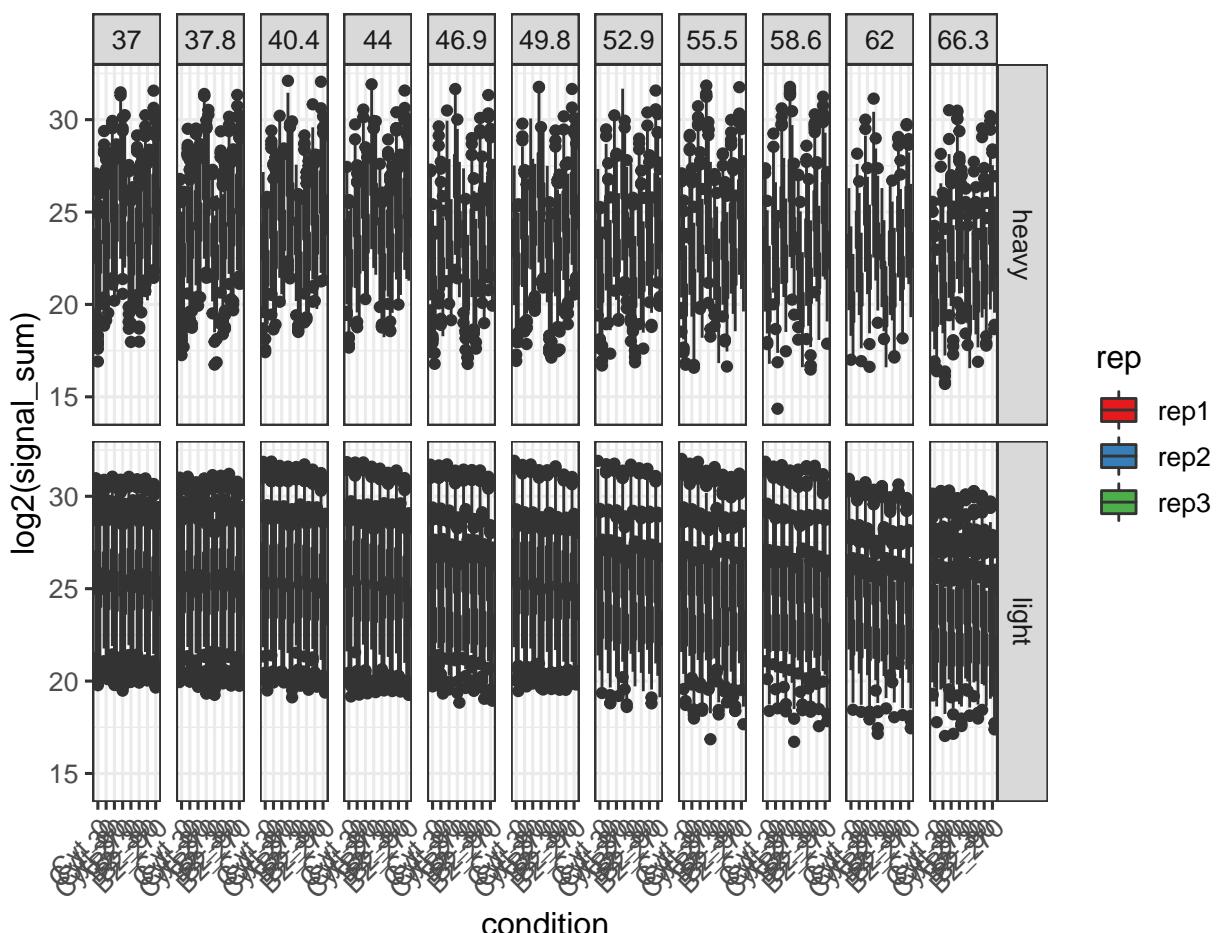
```

#Transforming long data to wide data

```

data$condition <- factor(data$condition, ordered = TRUE, levels =
                           c("Cyt_0", "Cyt_30", "Cyt_90", "Cyt_270",
                             "B2_0", "B2_30", "B2_90", "B2_270"))
ggplot(data = data, aes(condition, log2(value), fill = rep)) +
  geom_boxplot() +
  ylab("log2(signal_sum)") +
  facet_grid(SILAC ~ Temperature, scale = "free_x", space = "free") +
  customPlot +
  theme(axis.text.x = element_text(angle = 45, hjust = 1, vjust = 1))

```



```

cdata <- data %>%
  group_by(gene_name) %>%

```

```

mutate(key = paste("signal_sum", SILAC, Temperature, condition, rep, sep = "_")) %>%
dplyr::select(gene_name, key, value) %>%
group_by(gene_name, key) %>%
summarise(value = sum(value, na.rm = TRUE)) %>%
spread(key = key, value = value)
#found.in.ms.runs
fdata <- data %>%
group_by(gene_name) %>%
dplyr::select(gene_name, file) %>%
unique() %>%
summarise(found.in.ms.runs = n())
fdata_i <- data %>%
group_by(gene_name, sample) %>%
dplyr::select(gene_name, file) %>%
unique() %>%
summarise(found.in.ms.runs = n()) %>%
mutate(sample = paste0("found.in.ms.runs_", sample)) %>%
spread(key = sample, value = found.in.ms.runs)
fdata <- left_join(fdata, fdata_i)
fdata_i <- data %>%
group_by(gene_name, SILAC) %>%
dplyr::select(gene_name, file) %>%
unique() %>%
summarise(found.in.ms.runs = n()) %>%
mutate(SILAC = paste0("found.in.ms.runs_", SILAC)) %>%
spread(key = SILAC, value = found.in.ms.runs)
fdata <- left_join(fdata, fdata_i)
fdata_i <- data %>%
group_by(gene_name, Temperature) %>%
dplyr::select(gene_name, file) %>%
unique() %>%
summarise(found.in.ms.runs = n()) %>%
mutate(Temperature = paste0("found.in.ms.runs_", Temperature)) %>%
spread(key = Temperature, value = found.in.ms.runs)
fdata <- left_join(fdata, fdata_i)
#found.in.reps
fdata_i <- data %>%
group_by(gene_name) %>%
dplyr::select(gene_name, rep) %>%
unique() %>%
summarise(found.in.reps = n())
fdata <- left_join(fdata, fdata_i)
fdata_i <- data %>%
group_by(gene_name, sample) %>%
dplyr::select(gene_name, rep) %>%
unique() %>%
summarise(found.in.reps = n()) %>%
mutate(sample = paste0("found.in.reps_", sample)) %>%
spread(key = sample, value = found.in.reps)
fdata <- left_join(fdata, fdata_i)
#qupm overview
fdata_i <- data %>%
group_by(gene_name) %>%

```

```

dplyr::select(gene_name, qupm) %>%
unique() %>%
summarise(max.qupm = max(qupm, na.rm = TRUE))
fdata <- left_join(fdata, fdata_i)
fdata_i <- data %>%
group_by(gene_name, sample) %>%
dplyr::select(gene_name, sample, qupm) %>%
unique() %>%
summarise(max.qupm = max(qupm, na.rm = TRUE)) %>%
mutate(sample = paste0("max.qupm_", sample)) %>%
spread(key = sample, value = max.qupm)
fdata <- left_join(fdata, fdata_i)
#abundance overview
fdata_i <- data %>%
group_by(gene_name) %>%
dplyr::select(gene_name, top3) %>%
unique() %>%
summarise(average.top3 = mean(top3, na.rm = TRUE))
fdata <- left_join(fdata, fdata_i)
fdata_i <- data %>%
group_by(gene_name, sample) %>%
dplyr::select(gene_name, sample, top3) %>%
unique() %>%
summarise(average.top3 = mean(top3, na.rm = TRUE)) %>%
mutate(sample = paste0("average.top3_", sample)) %>%
spread(key = sample, value = average.top3)
id_data <- data %>%
ungroup() %>%
dplyr::select(gene_name, protein_id, description) %>%
unique()
dups <- id_data %>%
group_by(gene_name) %>%
summarise(n = n()) %>%
dplyr::filter(n > 1) %>%
dplyr::select(gene_name)
if (nrow(dups) > 0)
{
  dups <- dups$gene_name
  dup.data <- subset(id_data, gene_name %in% dups)
  id_data <- subset(id_data, !gene_name %in% dups)
  combine_ids <- function(ids)
  {
    ids <- as.character(ids)
    return(paste(
      sort(unique(unlist(strsplit(ids, split = "[|]")))), collapse = "|"))
  }
  dup.data <- dup.data %>%
    group_by(gene_name) %>%
    summarise(protein_id = combine_ids(protein_id),
              description = combine_ids(description))
  id_data <- bind_rows(id_data, dup.data)
  rm(dup.data, combine_ids)
}

```

```

rm(dups)
fdata <- left_join(id_data, fdata)
rm(id_data)
fdata <- left_join(fdata, fdata_i)
cdata <- left_join(fdata, cdata)
rm(fdata_i, fdata)

#Filter data

Only proteins that were quantified with two unique peptide matches are kept for the analysis. Proteins were filtered according to these condition already when loading the data. Moreover, only proteins were kept if they were quantified in at least 2/3 of the replicates.

dim(cdata)

## [1] 3775 636
min.found.in.ms.runs <- 2
min.qupm <- 2
for (s in unique(conditions$sample))
{
  cname <- paste0("keep_", s)
  cdata[, cname] <- ifelse(
    cdata[, paste0("found.in.reps_", s)] >=
      min.found.in.ms.runs & cdata[, paste0("max.qupm_", s)] >= min.qupm, TRUE, FALSE)
  cdata[is.na(cdata[, cname]), cname] <- FALSE
  rm(cname)
}
rm(s)
cdata$keep <- FALSE
for (i in seq_along(rownames(cdata)))
{
  cdata$keep[i] <- any(as.logical(cdata[i, grep("keep_", names(cdata), value = TRUE)]))
}
rm(i)
table(cdata$keep)

##
## FALSE TRUE
## 545 3230
cdata <- subset(cdata, keep)
dim(cdata)

## [1] 3230 659
rm(min.found.in.ms.runs, min.qupm)
cdata <- subset(cdata, (found.in.ms.runs_heavy >= 8 | found.in.ms.runs_light >= 8) &
  (found.in.ms.runs_37 >= 2 | found.in.ms.runs_37.8 >= 2))
dim(cdata)

## [1] 2804 659

#Building an expression set object
#Constructing assay data
raw_data <- cdata %>%
  dplyr::select(starts_with("signal_sum")) %>%
  as.data.frame()

```

```

rownames(raw_data) <- cdata$gene_name
names(raw_data) <- gsub("signal_sum_", "", names(raw_data))

#Constructing metadata
conditions_i <- data.frame(ID = names(raw_data))
# conditions_i$ID <- paste0("X", conditions_i$ID)
conditions_i <- left_join(conditions_i, conditions)

#Constructing fdata
fdata <- cdata %>%
  dplyr::select(-starts_with("signal_sum")) %>%
  as.data.frame()

#Defining ID columns
rownames(fdata) <- fdata$gene_name
rownames(conditions_i) <- conditions_i$ID
colnames(raw_data) <- conditions_i$ID

#Log2-transformation of raw_ signal_sums
raw_data_m <- log2(as.matrix(raw_data))
raw_data_m[is.infinite((raw_data_m))] <- NA
raw_data_m[is.na((raw_data_m))] <- NA

#Creating an expression set
raw_dataE <- ExpressionSet(assayData = raw_data_m,
                           phenoData = AnnotatedDataFrame(conditions_i),
                           featureData = AnnotatedDataFrame(fdata))
validObject(raw_dataE)

## [1] TRUE
rm(raw_data, raw_data_m, conditions_i, fdata)

#Batch effect removal
batchcl_raw_dataE <- raw_dataE
for (s in unique(conditions$sample))
{
  print(s)
  batchcl_raw_dataE_i <-
    batchcl_raw_dataE[fData(batchcl_raw_dataE)[, paste0("keep_", s)], ,
                      batchcl_raw_dataE$sample == s]

  exprs(batchcl_raw_dataE_i) <-
    removeBatchEffect(exprs(batchcl_raw_dataE_i),
                      batch = as.character(pData(batchcl_raw_dataE_i)$rep),
                      design = model.matrix(~0 + as.character(pData(batchcl_raw_dataE_i)$condition)))

  for (cn in colnames(batchcl_raw_dataE_i))
  {
    exprs(batchcl_raw_dataE)[fData(batchcl_raw_dataE)[, paste0("keep_", s)], cn] <-
      exprs(batchcl_raw_dataE_i)[, cn]
  }
  rm(cn)
}

```

```

## [1] "light_37"
## [1] "light_37.8"
## [1] "light_40.4"
## [1] "light_44"
## [1] "light_46.9"
## [1] "light_49.8"
## [1] "light_52.9"
## [1] "light_55.5"
## [1] "light_58.6"
## [1] "light_62"
## [1] "light_66.3"
## [1] "heavy_37"
## [1] "heavy_37.8"
## [1] "heavy_40.4"
## [1] "heavy_44"
## [1] "heavy_46.9"
## [1] "heavy_49.8"
## [1] "heavy_52.9"
## [1] "heavy_55.5"
## [1] "heavy_58.6"
## [1] "heavy_62"
## [1] "heavy_66.3"

rm(s)

```

#Data normalization

The vsn package from Wolfgang Huber is used to apply a variance stabilization normalization method on the log2 raw data.

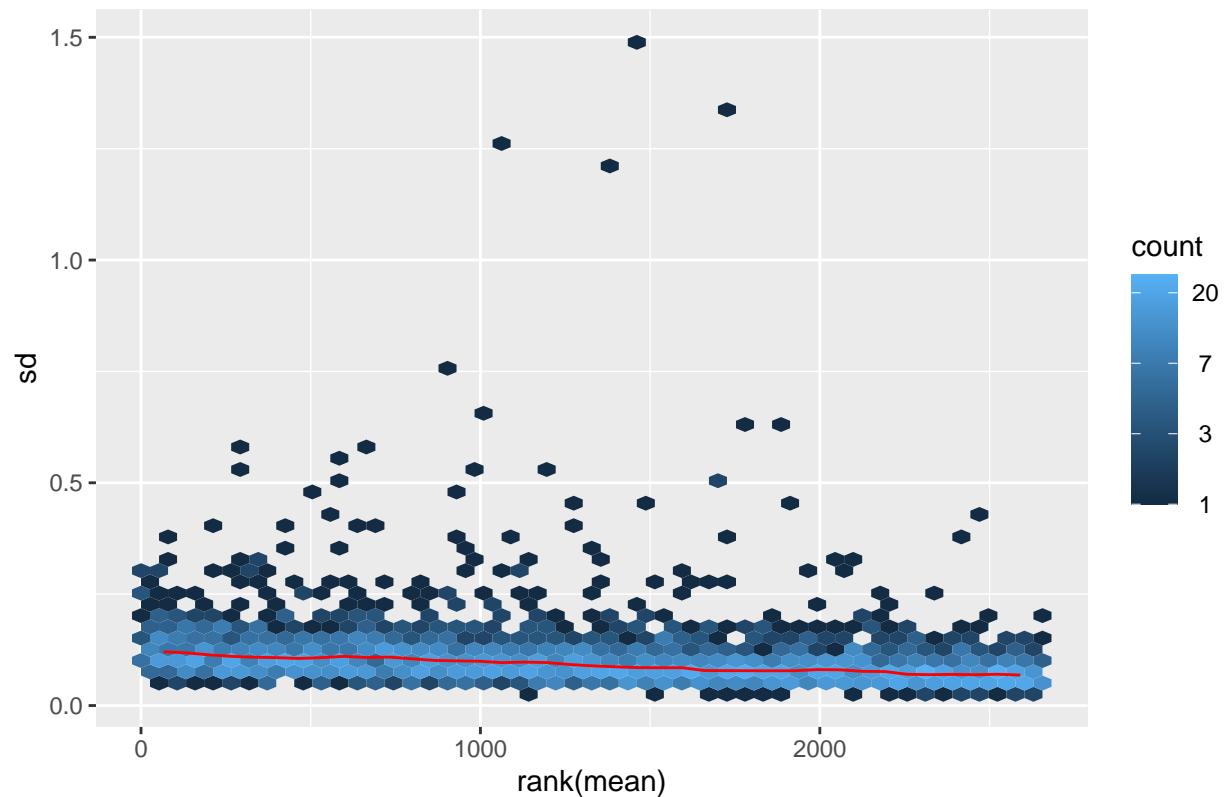
```

norm_batchcl_raw_dataE <- batchcl_raw_dataE
for (s in unique(conditions$sample))
{
  print(paste0(s))
  vsn.fit <- vsn2(2^exprs(norm_batchcl_raw_dataE[
    fData(norm_batchcl_raw_dataE)[, paste0("keep_", s)],
    norm_batchcl_raw_dataE$sample == s]))
  norm_batchcl_raw_dataE_i <- predict(vsn.fit, 2^exprs(norm_batchcl_raw_dataE[
    fData(norm_batchcl_raw_dataE)[, paste0("keep_", s)],
    norm_batchcl_raw_dataE$sample == s]))
  for (cn in colnames(norm_batchcl_raw_dataE_i))
  {
    exprs(norm_batchcl_raw_dataE)[fData(norm_batchcl_raw_dataE)[, paste0("keep_", s)], cn] <-
      norm_batchcl_raw_dataE_i[, cn]
  }
  rm(cn, norm_batchcl_raw_dataE_i)
  sdplot.object <- meanSdPlot(vsn.fit, plot = FALSE)
  print(sdplot.object$gg + ggtitle(s))
  rm(vsn.fit, sdplot.object)
}

## [1] "light_37"

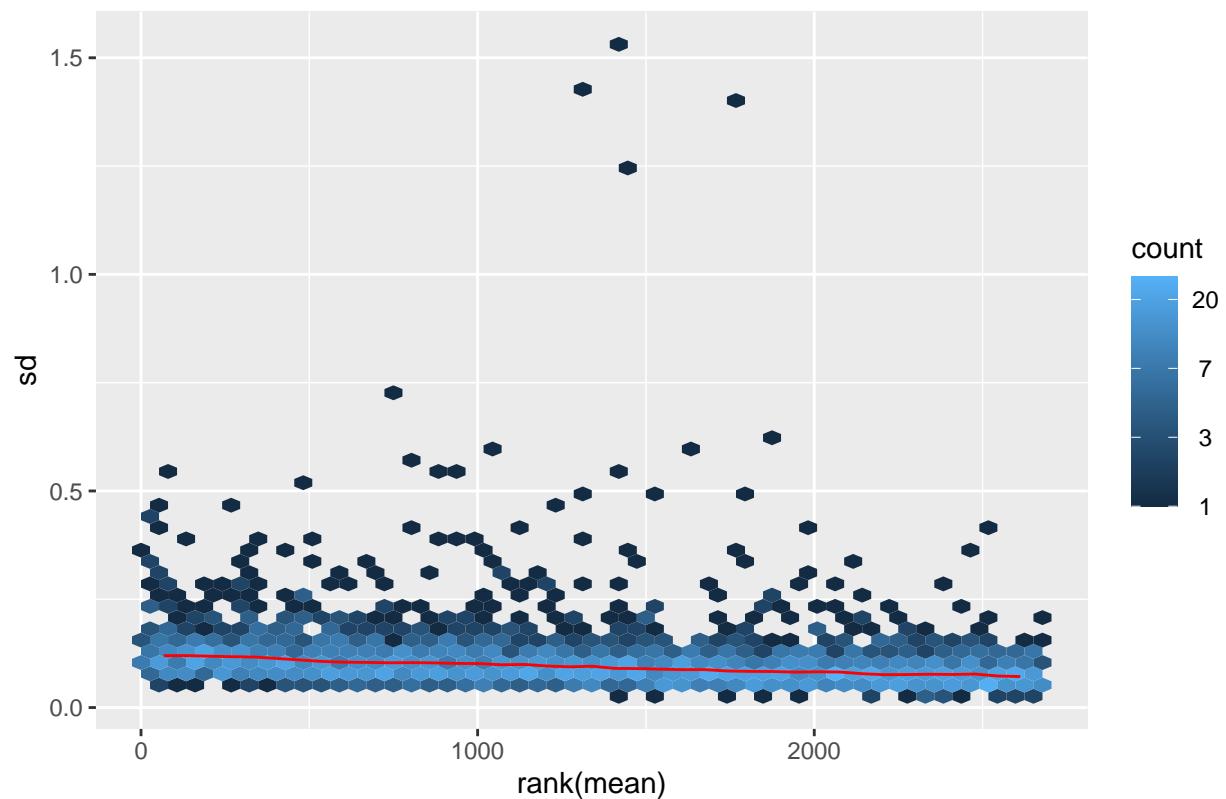
```

light_37



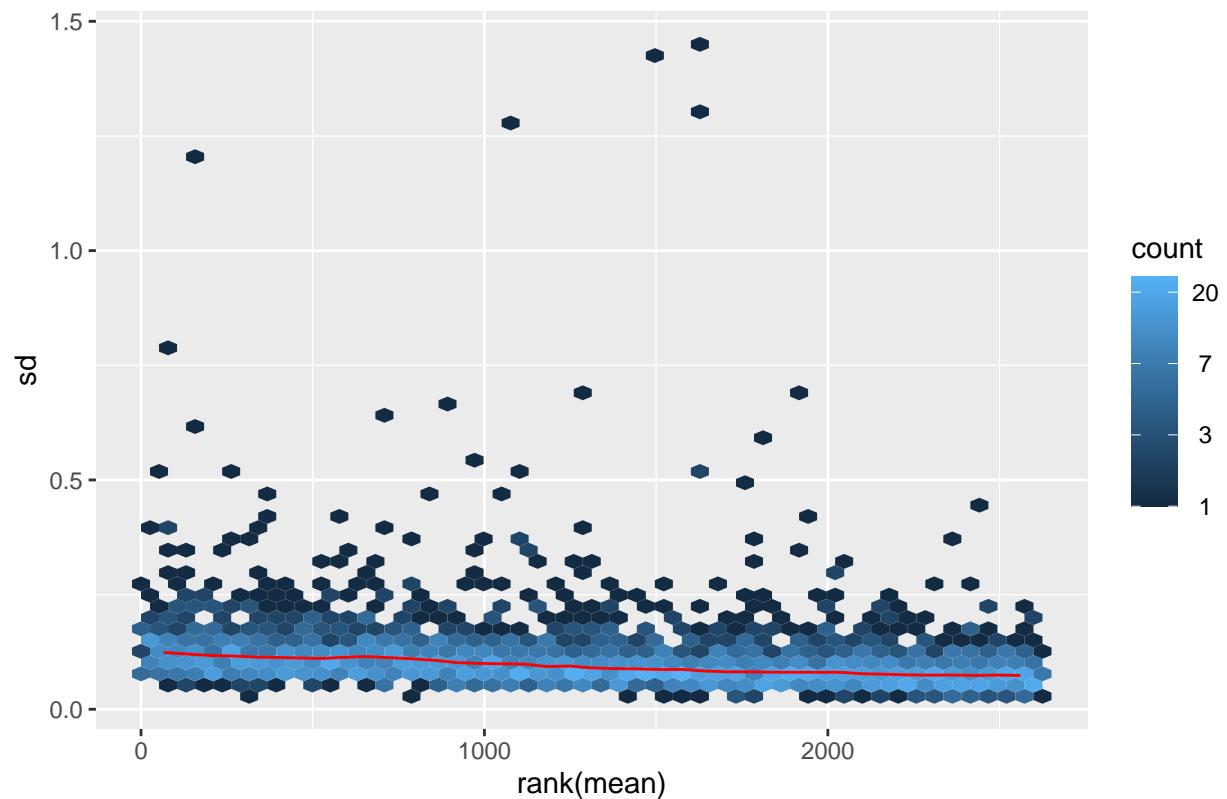
```
## [1] "light_37.8"
```

light_37.8



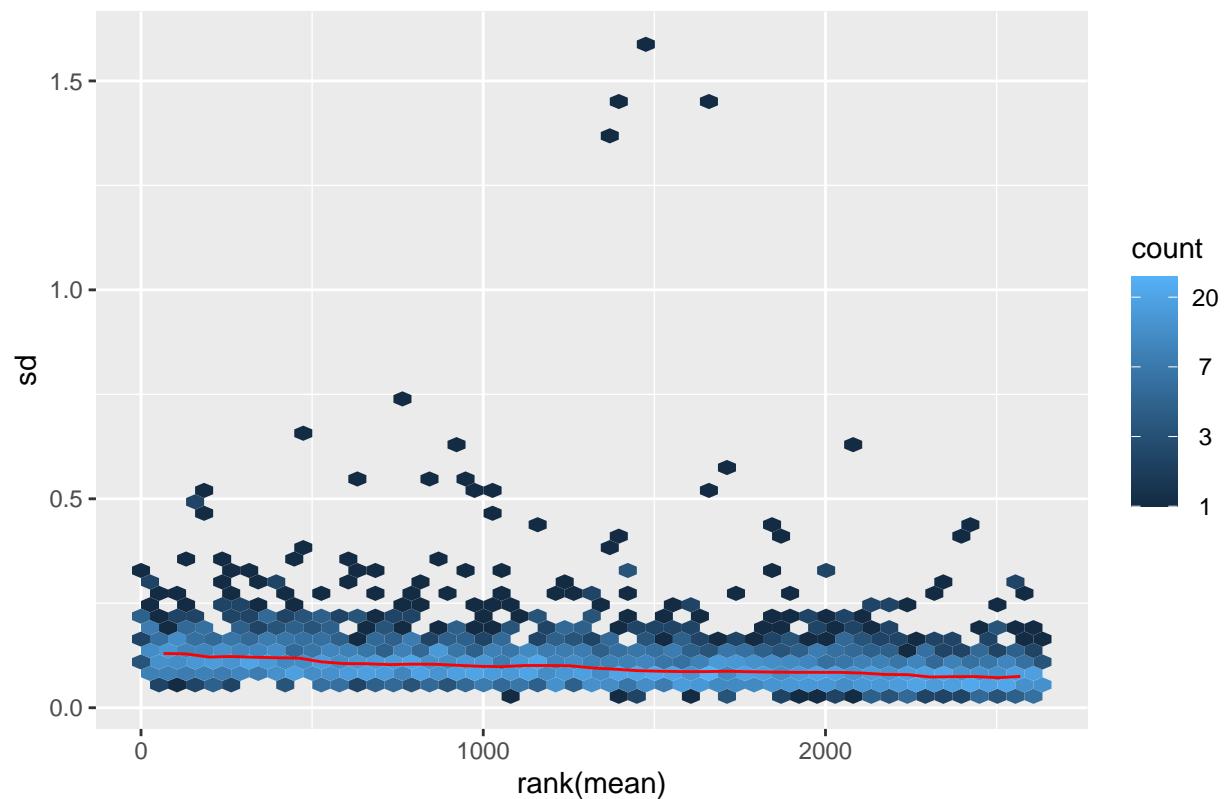
```
## [1] "light_40.4"
```

light_40.4



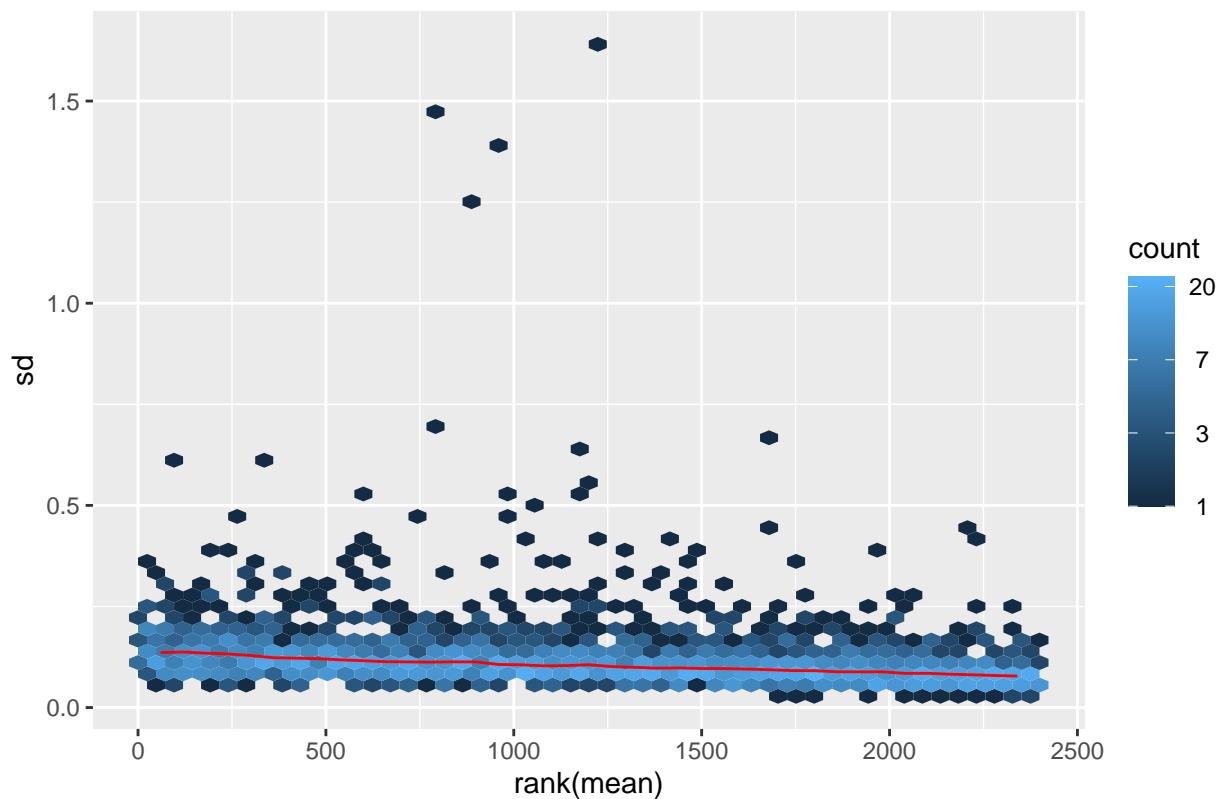
```
## [1] "light_44"
```

light_44



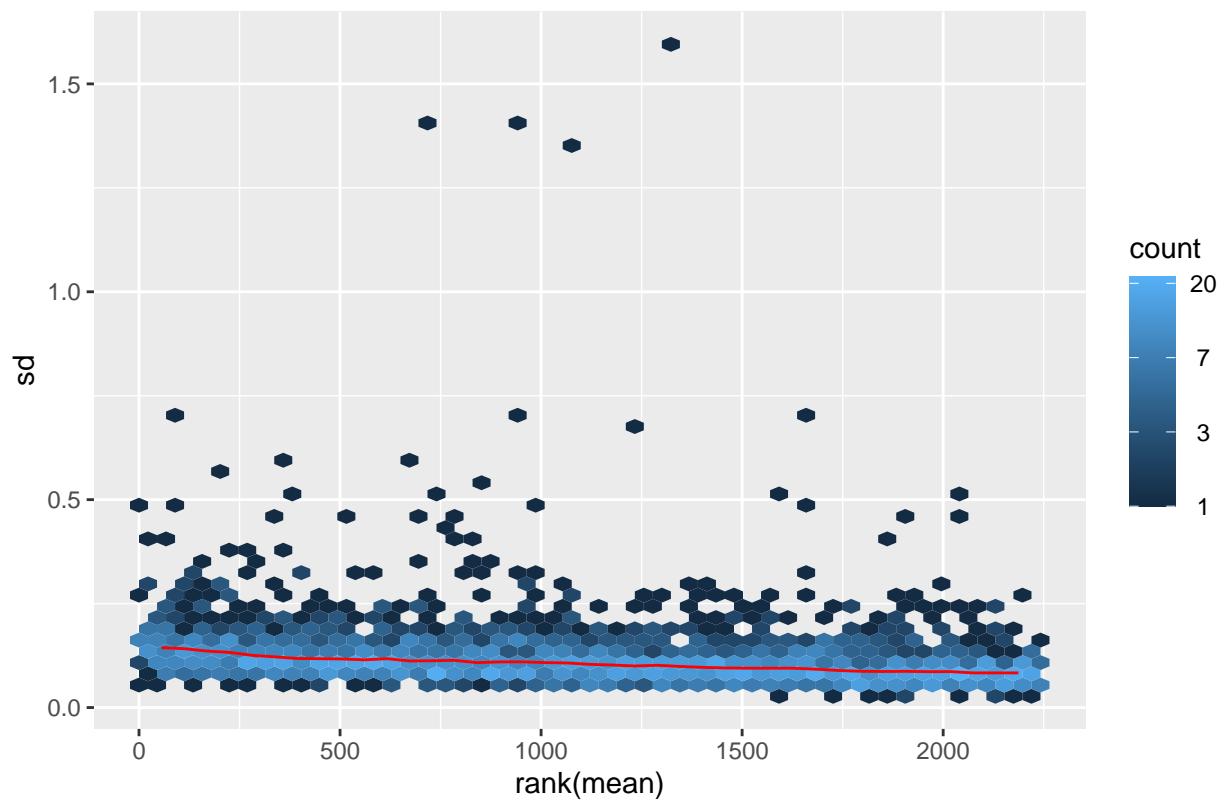
```
## [1] "light_46.9"
```

light_46.9



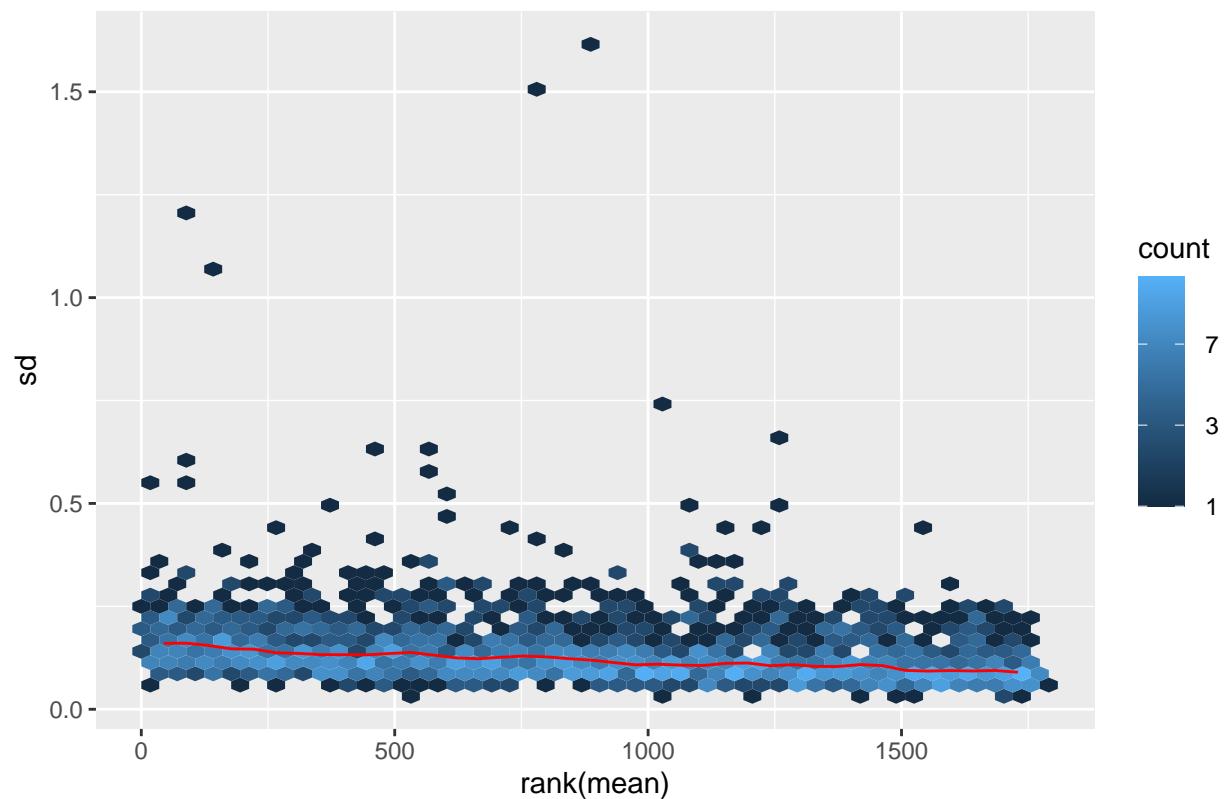
```
## [1] "light_49.8"
```

light_49.8



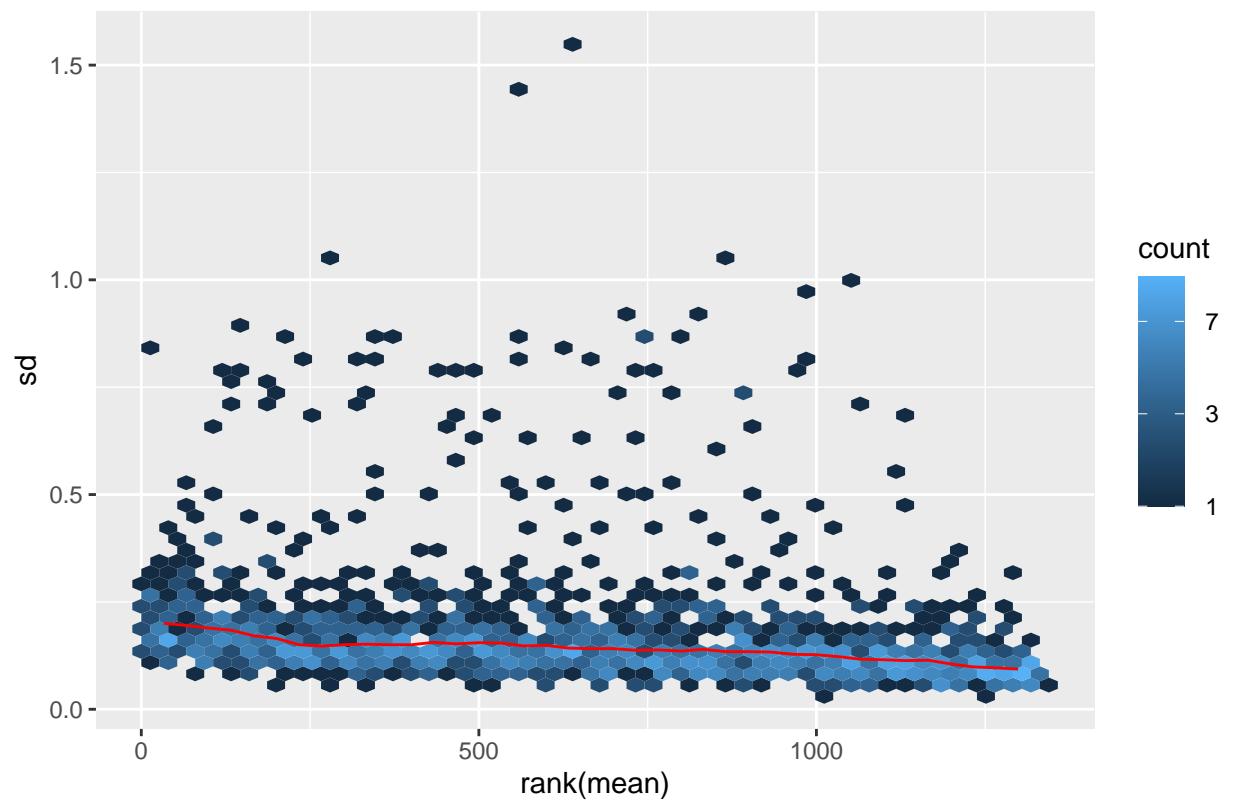
```
## [1] "light_52.9"
```

light_52.9



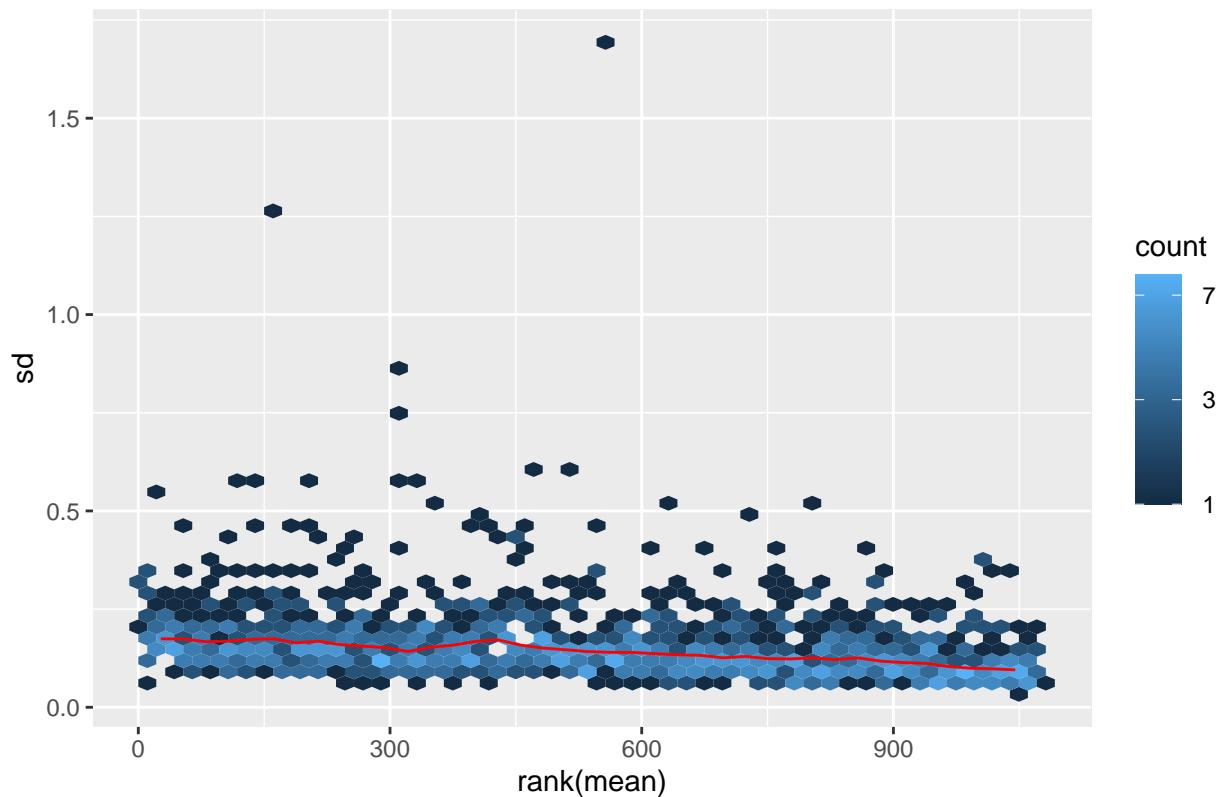
```
## [1] "light_55.5"
```

light_55.5



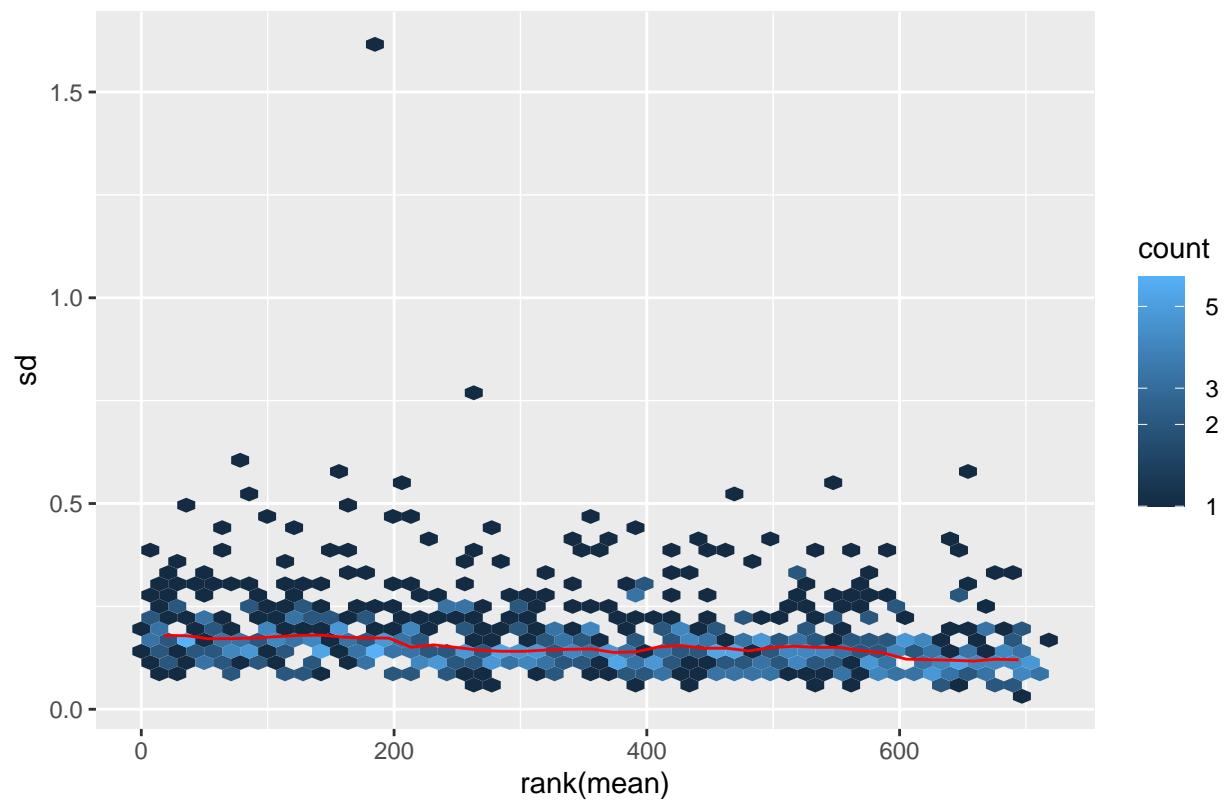
```
## [1] "light_58.6"
```

light_58.6



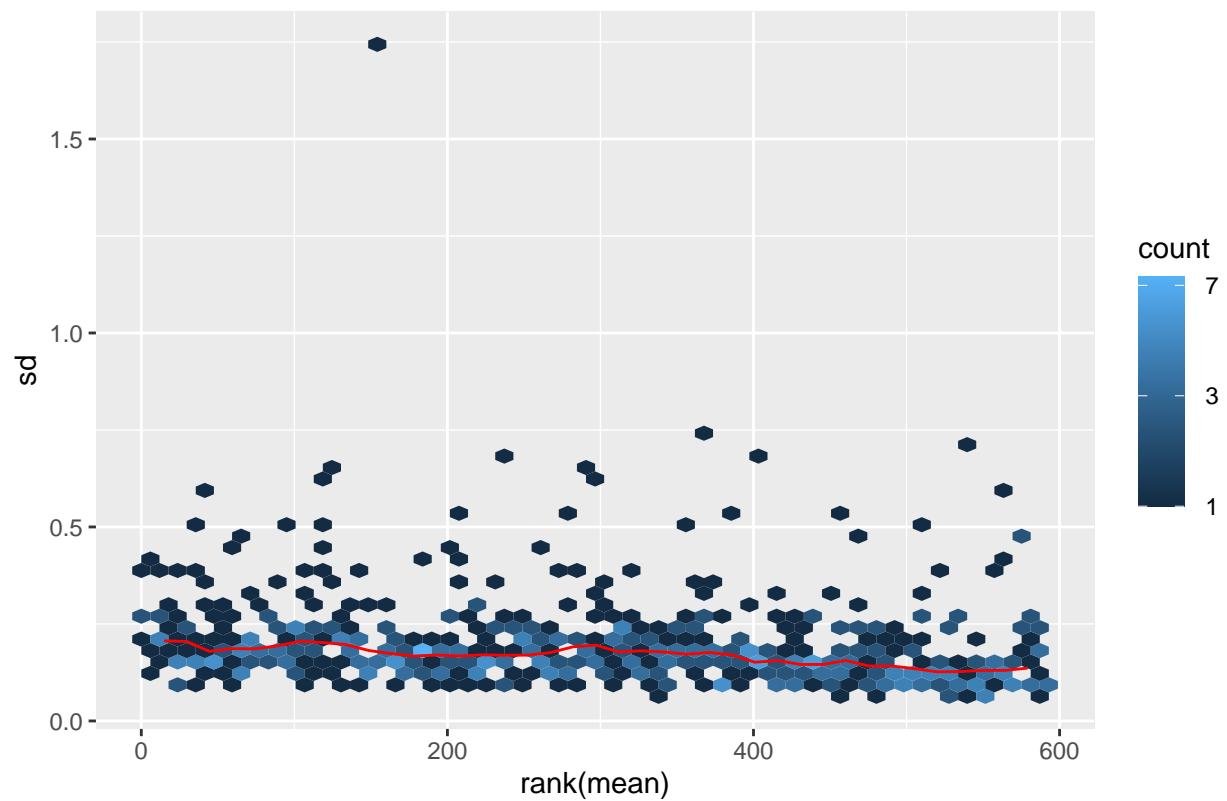
```
## [1] "light_62"
```

light_62



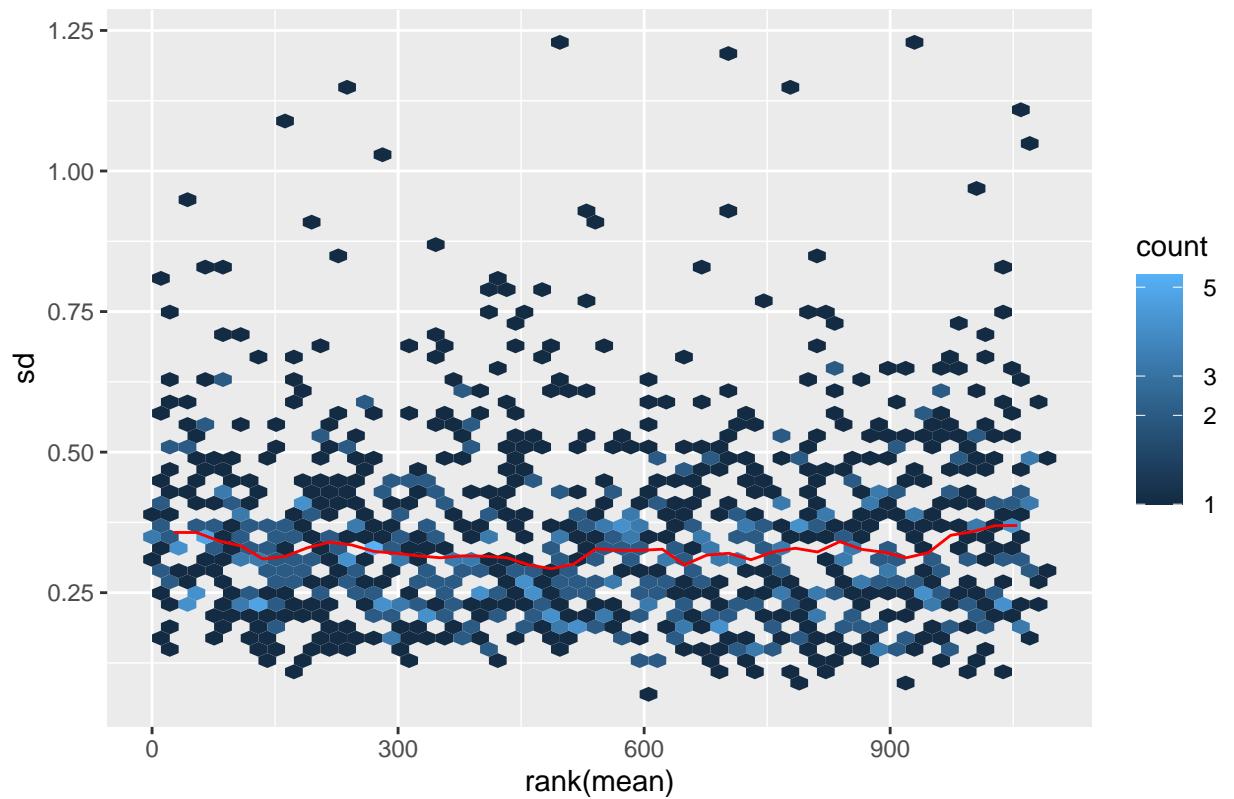
```
## [1] "light_66.3"
```

light_66.3



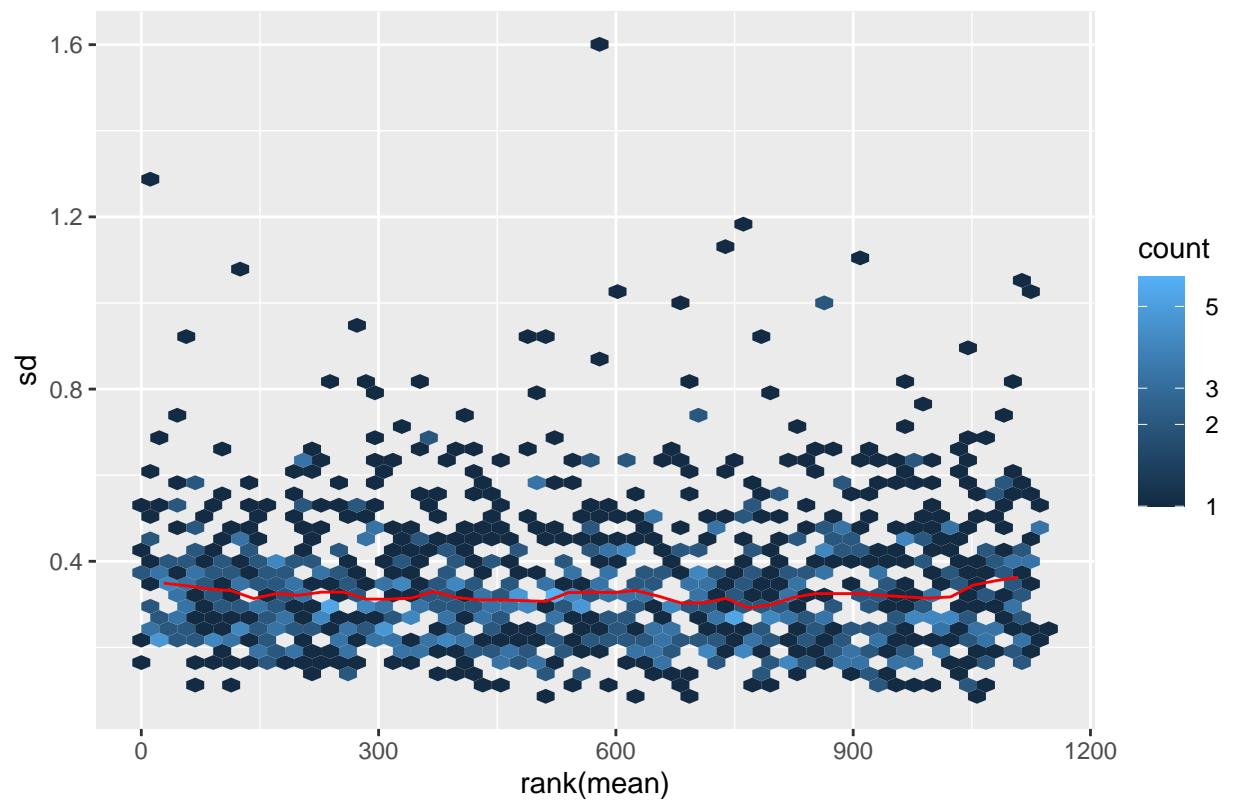
```
## [1] "heavy_37"
```

heavy_37



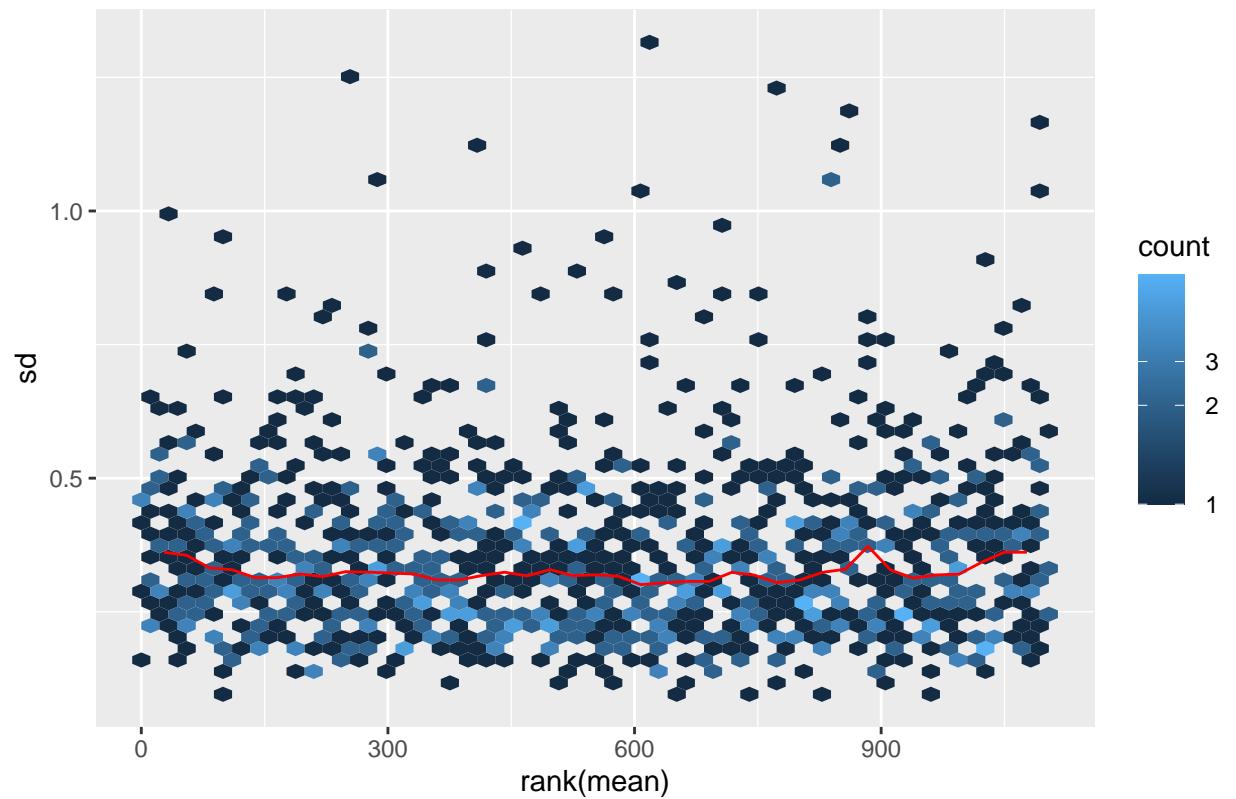
```
## [1] "heavy_37.8"
```

heavy_37.8



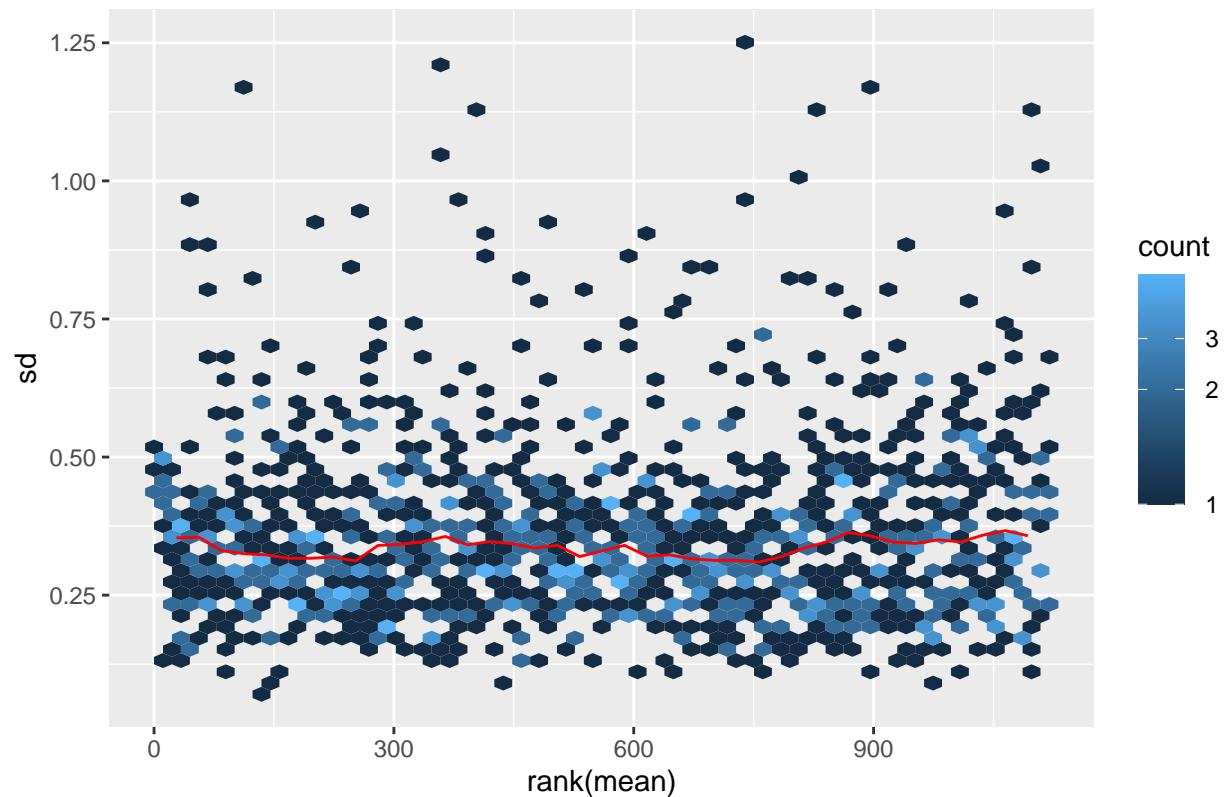
```
## [1] "heavy_40.4"
```

heavy_40.4



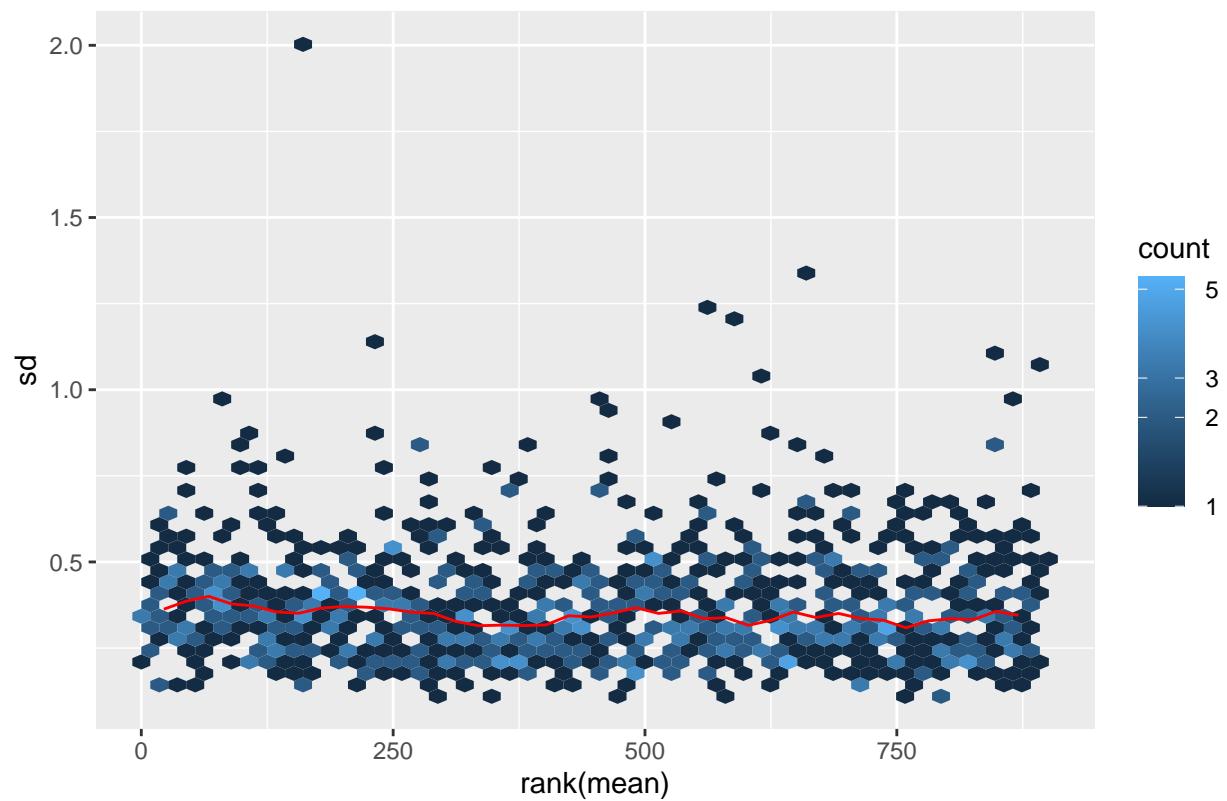
```
## [1] "heavy_44"
```

heavy_44



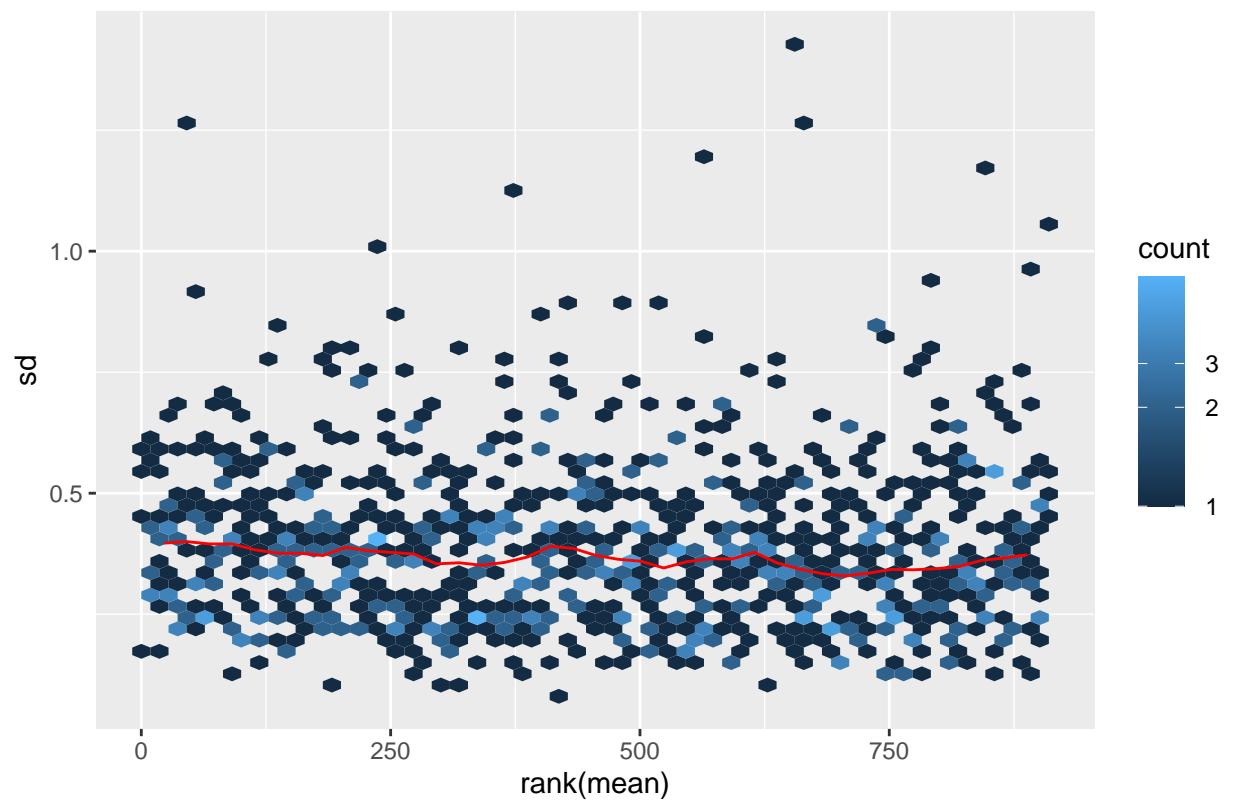
```
## [1] "heavy_46.9"
```

heavy_46.9



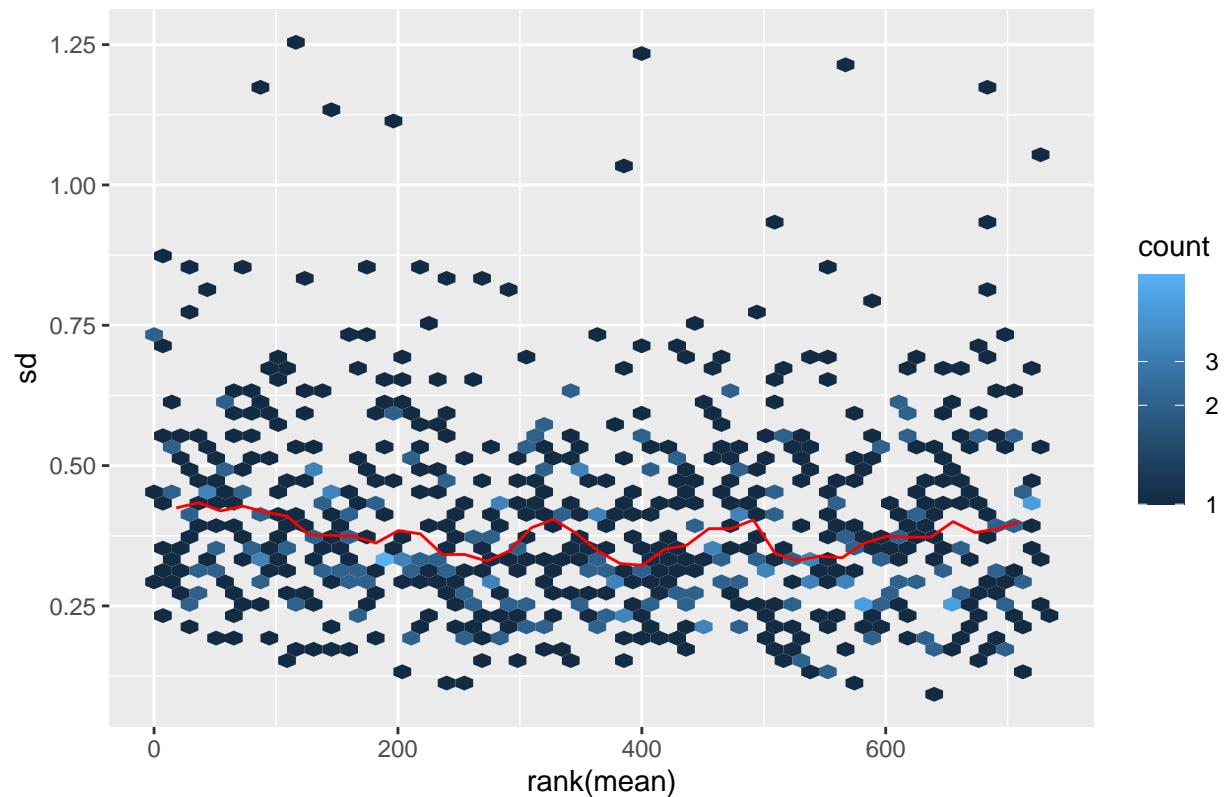
```
## [1] "heavy_49.8"
```

heavy_49.8



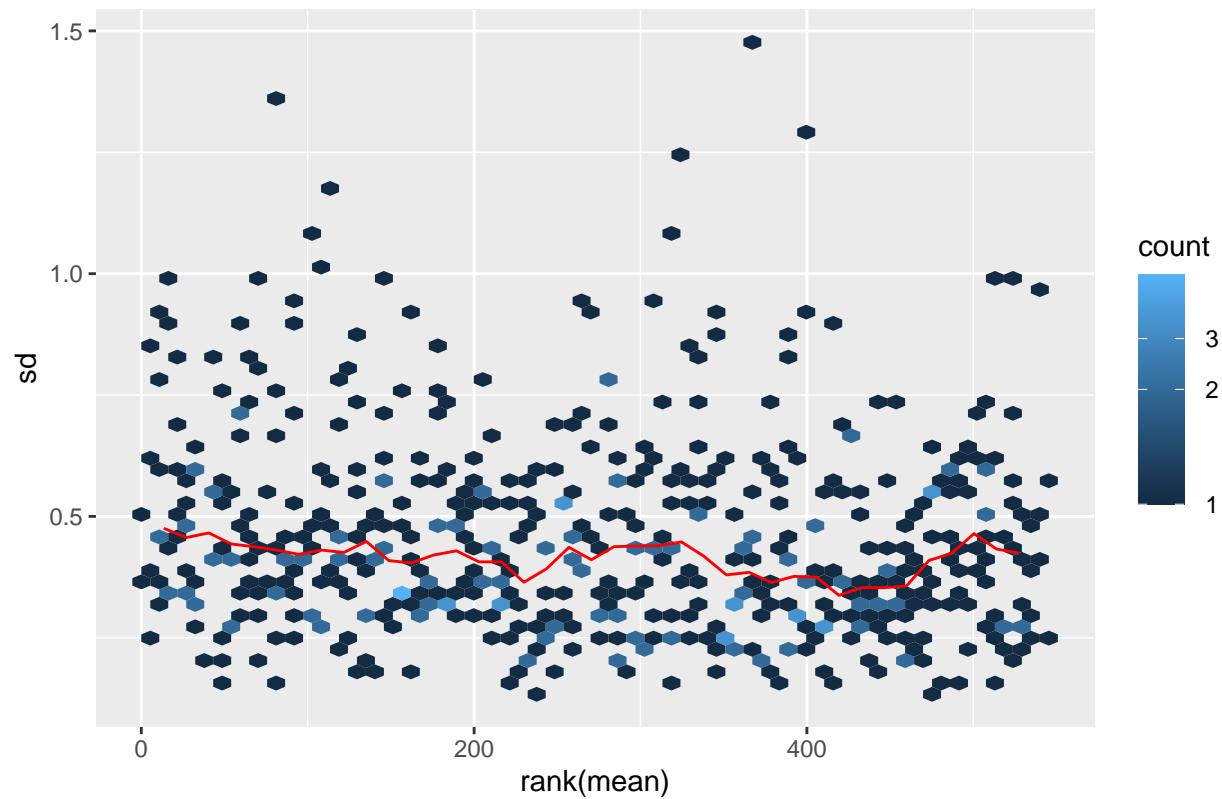
```
## [1] "heavy_52.9"
```

heavy_52.9



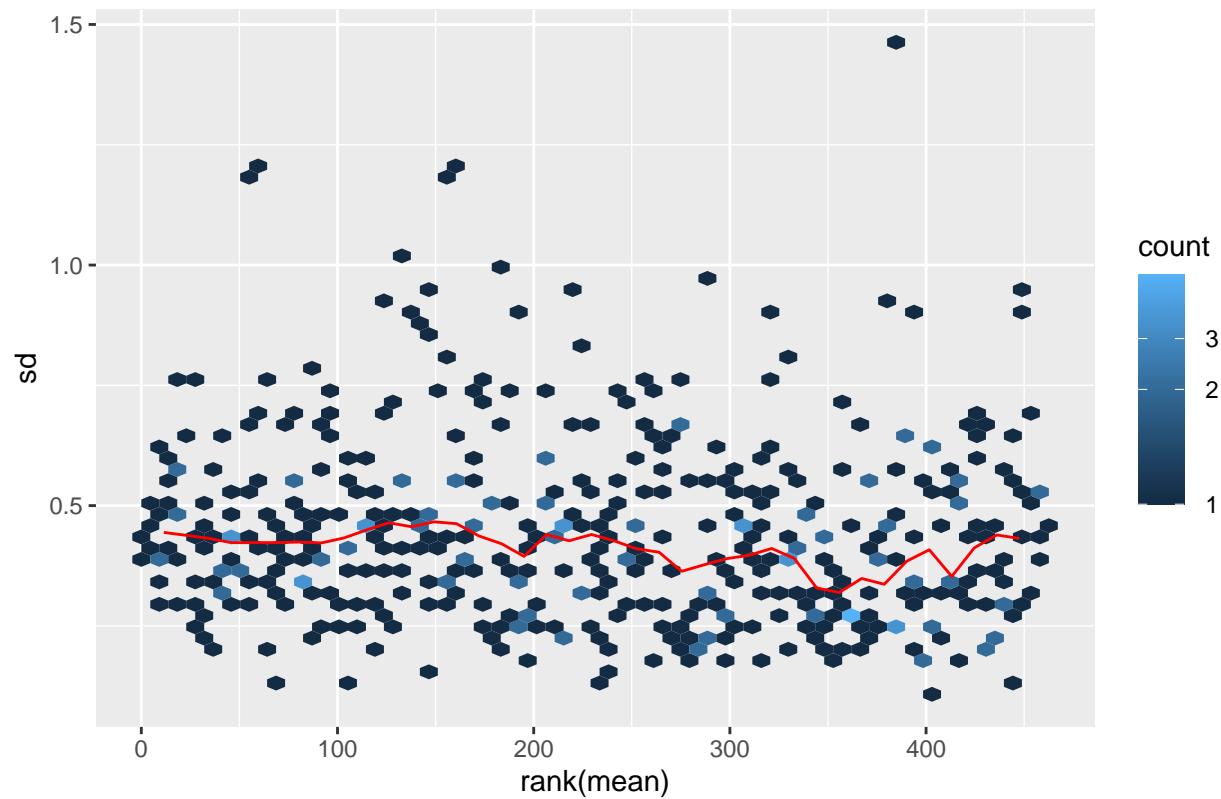
```
## [1] "heavy_55.5"
```

heavy_55.5



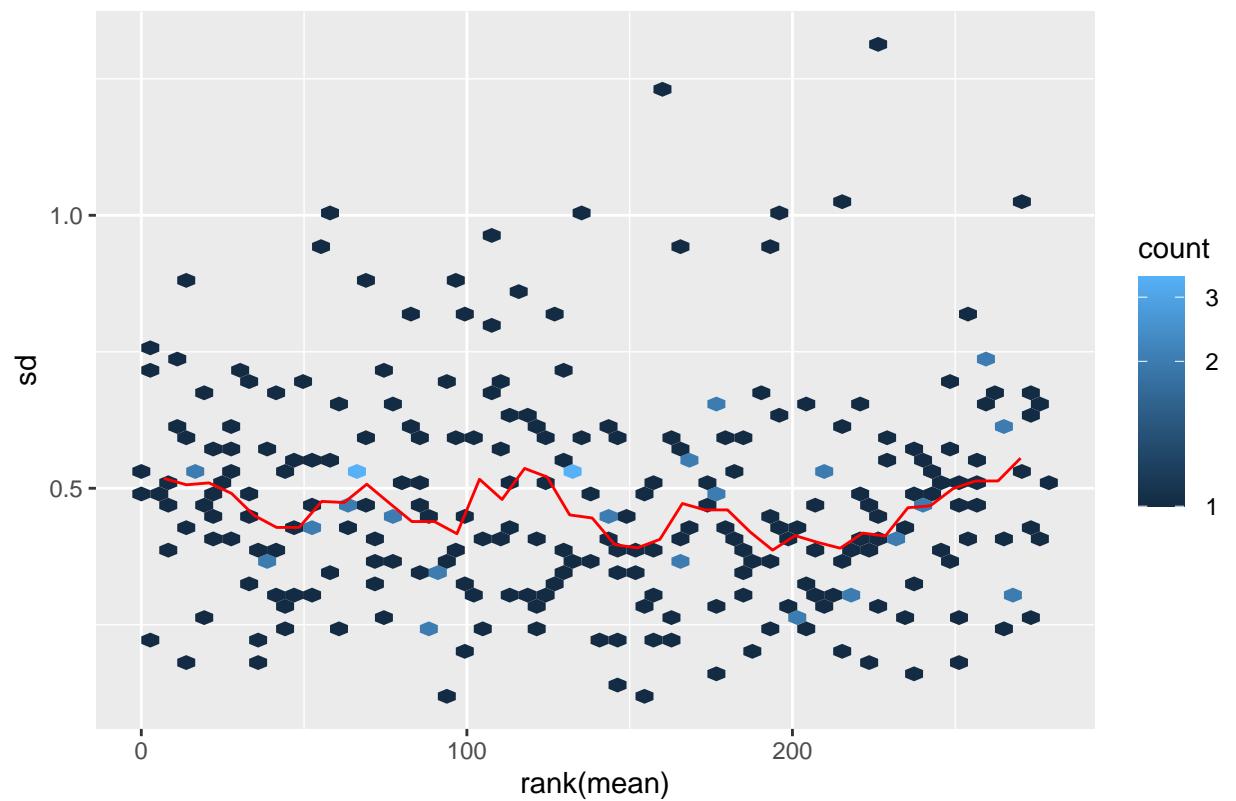
```
## [1] "heavy_58.6"
```

heavy_58.6



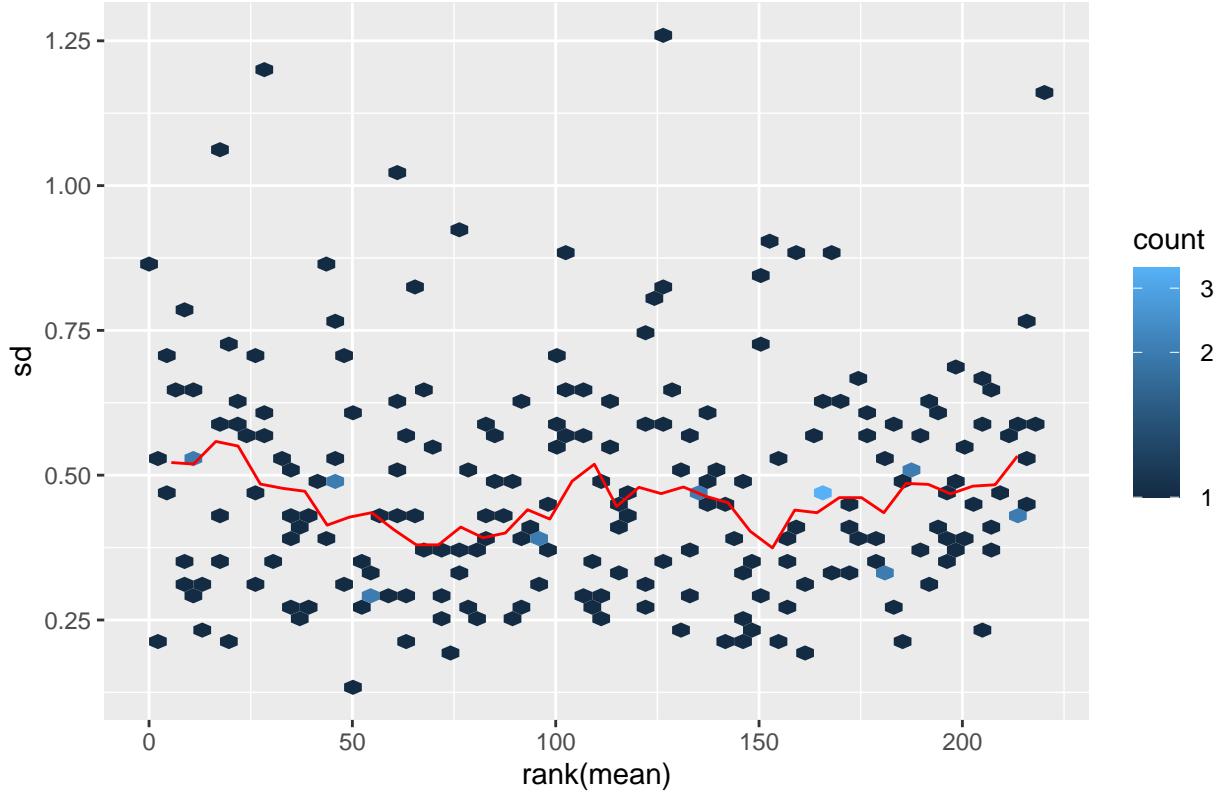
```
## [1] "heavy_62"
```

heavy_62



```
## [1] "heavy_66.3"
```

heavy_66.3



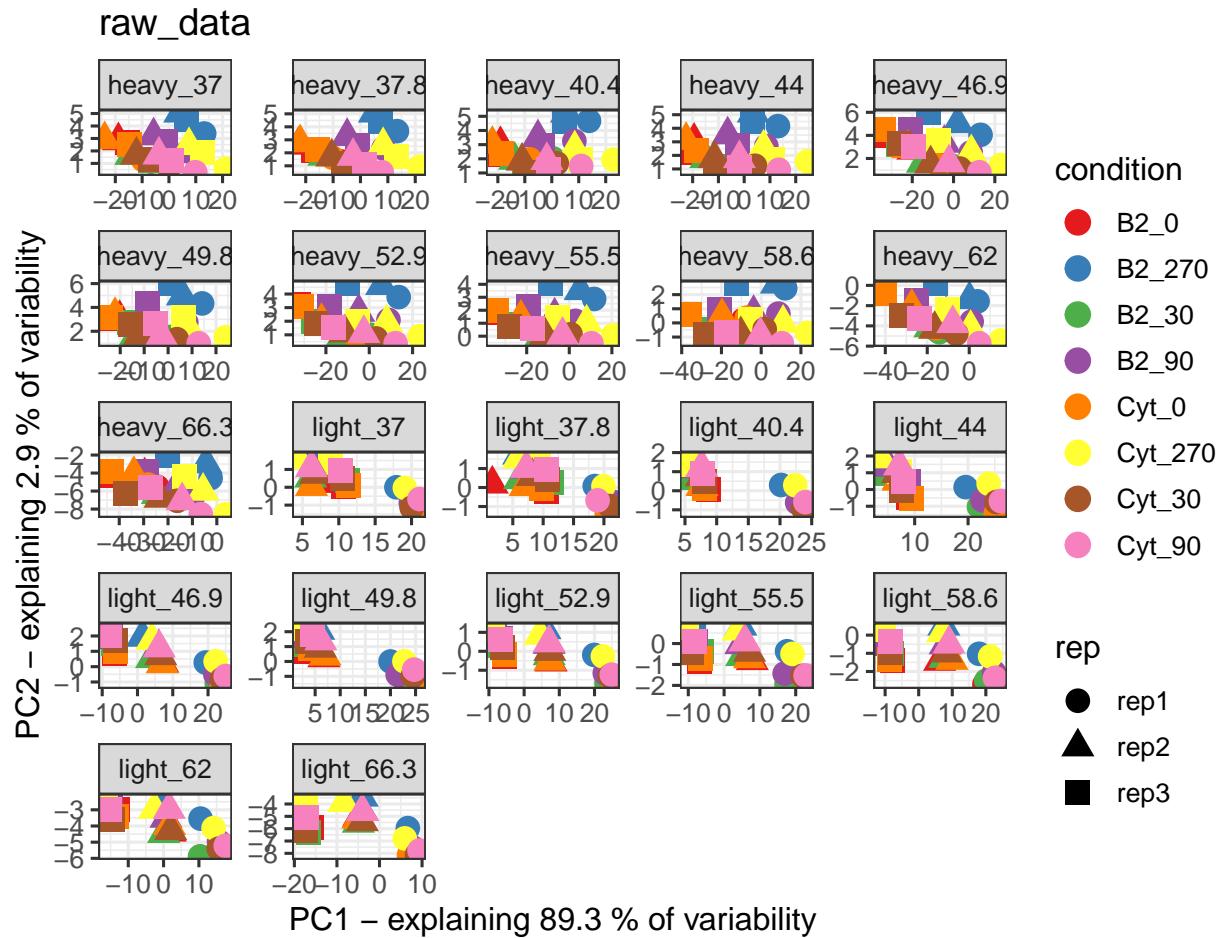
rm(s)

```
#PCA - principal component analysis
#PCA of raw_data
raw_PCA_m <- t(na.omit(exprs(raw_dataE)))
if (length(raw_PCA_m) > 50)
{
  raw_PCA <- prcomp(raw_PCA_m, scale = FALSE)
  perc_var <-
    round(100 * raw_PCA$sdev ^ 2 /
      sum(raw_PCA$sdev ^ 2), 1)
  PCA_raw_data <-
    data.frame(PC1 = raw_PCA$x[, 1],
               PC2 = raw_PCA$x[, 2],
               rep = pData(raw_dataE)$rep,
               condition = pData(raw_dataE)$condition,
               sample = pData(raw_dataE)$sample)
  print(ggplot(data = PCA_raw_data, aes(PC1, PC2)) +
    geom_point(aes(color = condition, shape = rep), size = 4) +
    guides(size = FALSE) +
    customPlot +
    ggtitle("raw_data") +
    xlab(paste("PC1 - explaining", perc_var[1], "% of variability")) +
    ylab(paste("PC2 - explaining", perc_var[2], "% of variability")) +
    facet_wrap(~ sample, scale = "free"))
  ggsave(file.path(dir_save, paste0("PCA_raw_data_", script.version, ".pdf")))
}
```

```

width = 15, height = 12)
rm(raw_PCA, perc_var, PCA_raw_data)
}

```



```

rm(raw_PCA_m)

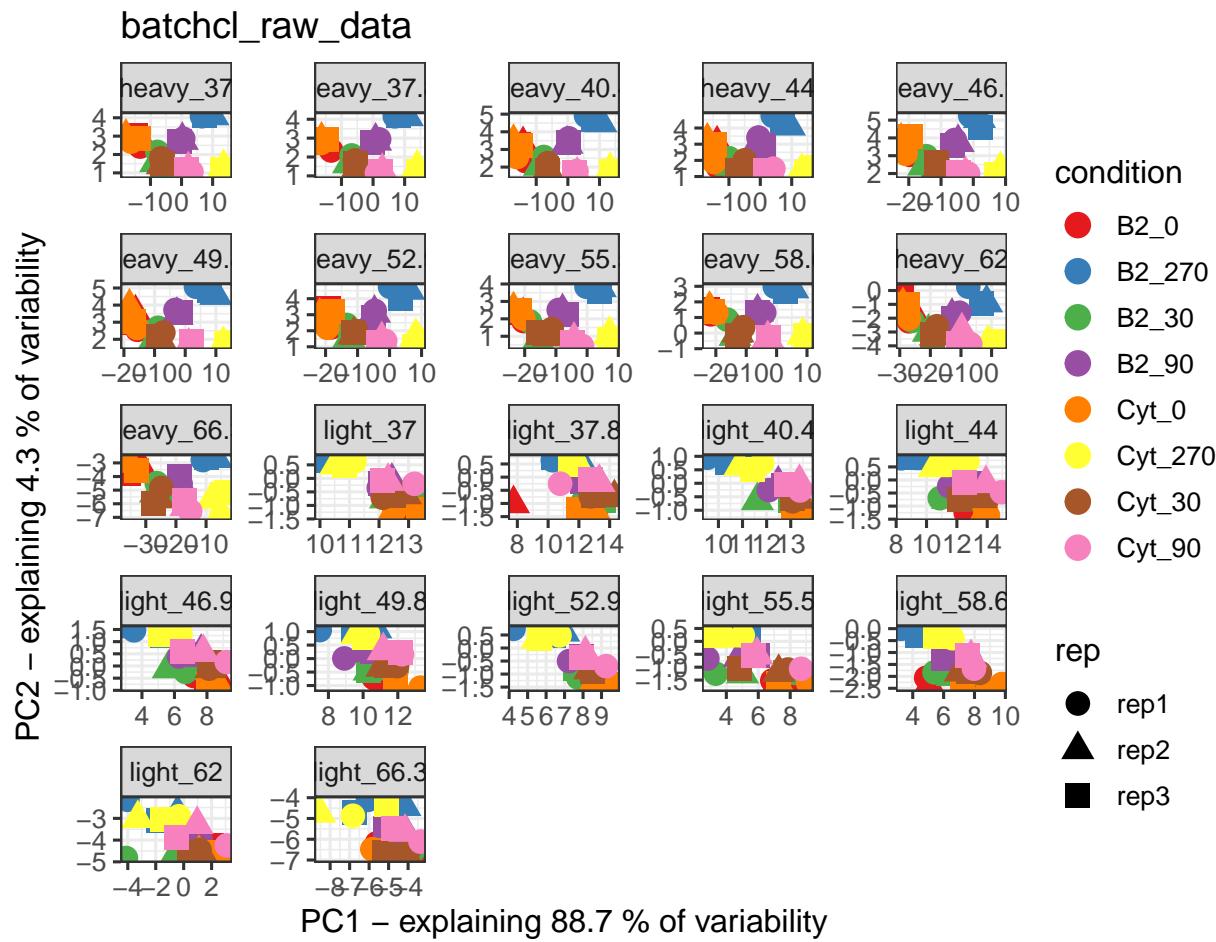
#PCA of batchcl_raw_data
batchcl_raw_PCA_m <- t(na.omit(exprs(batchcl_raw_dataE)))
if (length(batchcl_raw_PCA_m) > 50)
{
  batchcl_raw_PCA_m <- prcomp(batchcl_raw_PCA_m, scale = FALSE)
  perc_var <-
    round(100 * batchcl_raw_PCA_m$sdev ^ 2 /
      sum(batchcl_raw_PCA_m$sdev ^ 2), 1)
  PCA_batchcl_raw_data <-
    data.frame(PC1 = batchcl_raw_PCA_m$x[, 1],
               PC2 = batchcl_raw_PCA_m$x[, 2],
               rep = pData(batchcl_raw_dataE)$rep,
               condition = pData(batchcl_raw_dataE)$condition,
               sample = pData(batchcl_raw_dataE)$sample)
  print(ggplot(data = PCA_batchcl_raw_data, aes(PC1, PC2)) +
    geom_point(aes(color = condition, shape = rep), size = 4) +
    guides(size = FALSE) +
    theme_minimal())
}

```

```

    customPlot +
    ggtitle("batchcl_raw_data") +
    xlab(paste("PC1 - explaining", perc_var[1], "% of variability")) +
    ylab(paste("PC2 - explaining", perc_var[2], "% of variability")) +
    facet_wrap(~ sample, scale = "free")
ggsave(file.path(dir_save, paste0("PCA_batchcl_raw_data_", script.version, ".pdf")),
       width = 15, height = 12)
rm(batchcl_raw_PCA, perc_var, PCA_batchcl_raw_data)
}

```



```

rm(batchcl_raw_PCA_m)

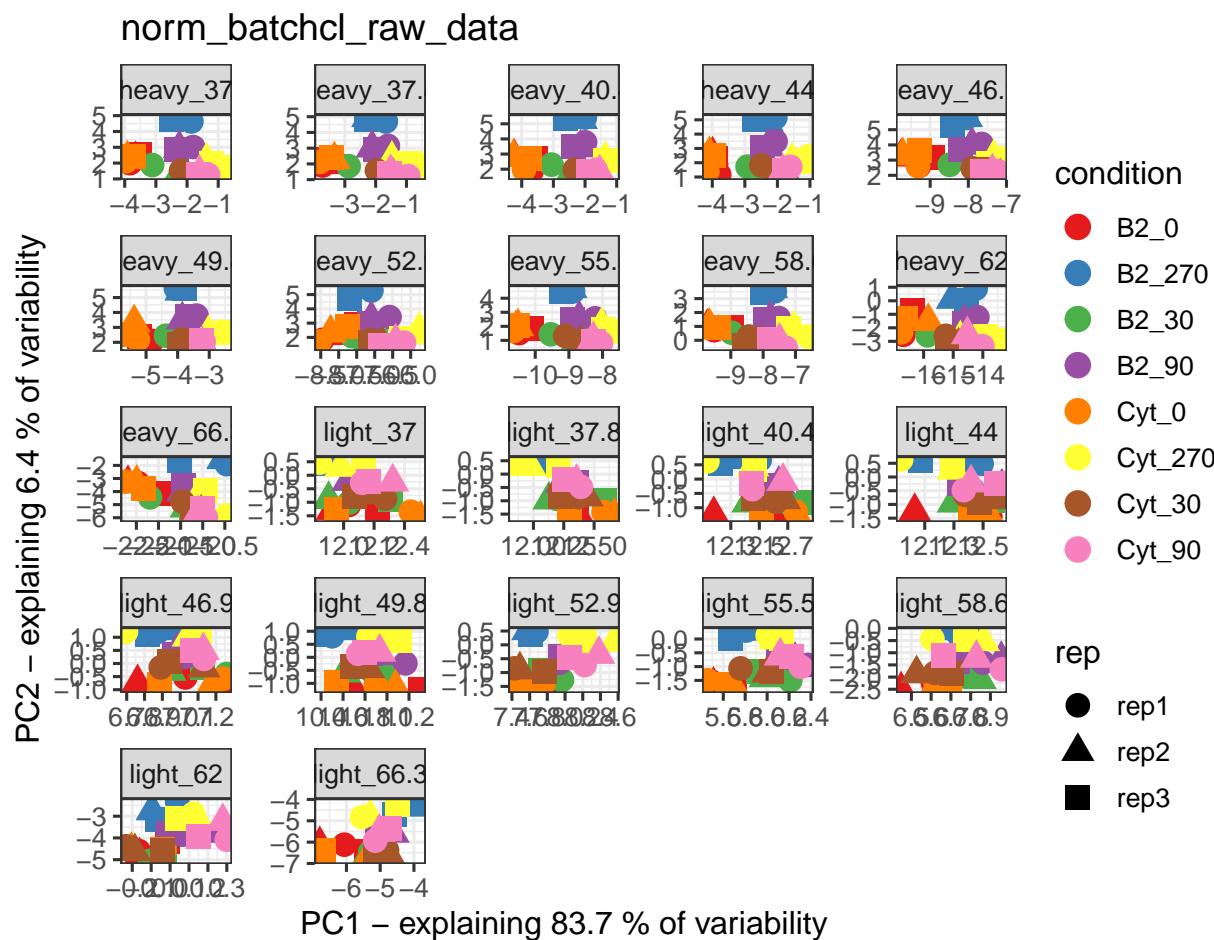
#PCA of norm_batchcl_raw_data
norm_batchcl_raw_PCA_m <- t(na.omit(exprs(norm_batchcl_raw_dataE)))
if (length(norm_batchcl_raw_PCA_m) > 50)
{
  norm_batchcl_raw_PCA <- prcomp(norm_batchcl_raw_PCA_m, scale = FALSE)
  perc_var <-
    round(100 * norm_batchcl_raw_PCA$sdev ^ 2 /
          sum(norm_batchcl_raw_PCA$sdev ^ 2), 1)
  PCA_norm_batchcl_raw_data <-
    data.frame(PC1 = norm_batchcl_raw_PCA$x[, 1],
               PC2 = norm_batchcl_raw_PCA$x[, 2],
               sample = sample)
}

```

```

    rep = pData(norm_batchcl_raw_dataE)$rep,
    condition = pData(norm_batchcl_raw_dataE)$condition,
    sample = pData(norm_batchcl_raw_dataE)$sample)
print(ggplot(data = PCA_norm_batchcl_raw_data, aes(PC1, PC2)) +
  geom_point(aes(color = condition, shape = rep), size = 4) +
  guides(size = FALSE) +
  customPlot +
  ggtitle("norm_batchcl_raw_data") +
  xlab(paste("PC1 - explaining", perc_var[1], "% of variability")) +
  ylab(paste("PC2 - explaining", perc_var[2], "% of variability")) +
  facet_wrap(~ sample, scale = "free"))
ggsave(file.path(dir_save, paste0("PCA_norm_batchcl_raw_data_", script.version, ".pdf")),
       width = 15, height = 12)
rm(norm_batchcl_raw_PCA, perc_var, PCA_norm_batchcl_raw_data)
}

```



```

rm(norm_batchcl_raw_PCA_m)

#Merge modified data back into 'cdata'
cdata_i <- as.data.frame(2^exprs(batchcl_raw_dataE))
names(cdata_i) <- paste0("batchcl_raw_signal_sum_", names(cdata_i))
cdata_i$gene_name <- rownames(cdata_i)
cdata <- left_join(cdata, cdata_i)

```

```

rm(cdata_i)

cdata_i <- as.data.frame(2^exprs(norm_batchcl_raw_dataE))
names(cdata_i) <- paste0("norm_batchcl_raw_signal_sum_", names(cdata_i))
cdata_i$gene_name <- rownames(cdata_i)
cdata <- left_join(cdata, cdata_i)
rm(cdata_i)

write.csv(cdata, file = file.path(dir_save, paste0("Full_dataset_", script.version, ".csv")))

#Create tidy data
mdata <- NULL

mdata_i <- tidy(raw_dataE)
mdata_i <- mdata_i %>%
  mutate(value = 2 ^ value)
names(mdata_i)[1] <- "gene_name"
names(mdata_i)[2] <- "ID"
mdata_i <- left_join(mdata_i, conditions)
mdata_i$measurement <- "raw_signal_sum"
mdata <- bind_rows(mdata, mdata_i)
rm(mdata_i)

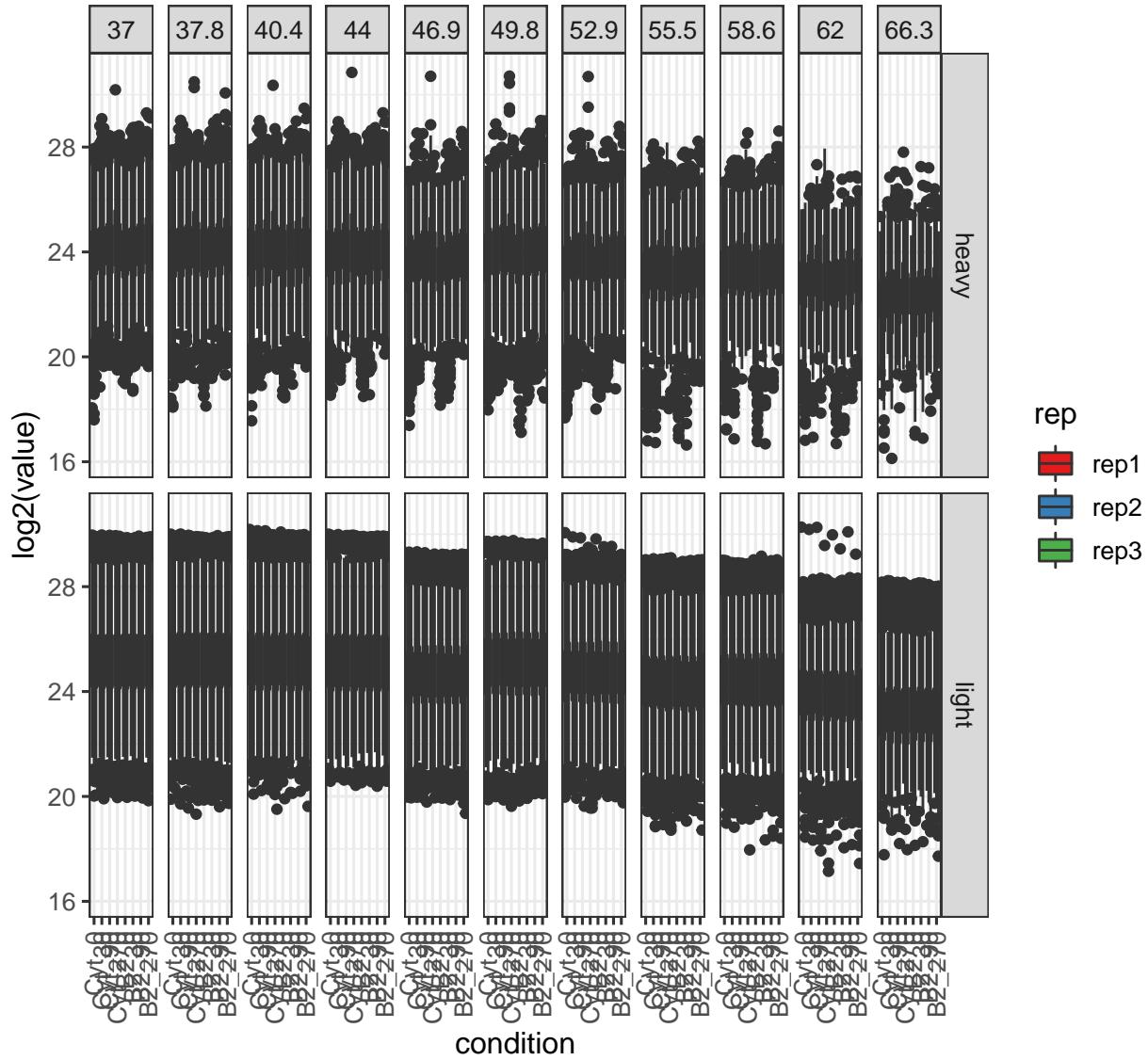
mdata_i <- tidy(batchcl_raw_dataE)
mdata_i <- mdata_i %>%
  mutate(value = 2 ^ value)
names(mdata_i)[1] <- "gene_name"
names(mdata_i)[2] <- "ID"
mdata_i <- left_join(mdata_i, conditions)
mdata_i$measurement <- "batchcl_raw_signal_sum"
mdata <- bind_rows(mdata, mdata_i)
rm(mdata_i)

mdata_i <- tidy(norm_batchcl_raw_dataE)
mdata_i <- mdata_i %>%
  mutate(value = 2 ^ value)
names(mdata_i)[1] <- "gene_name"
names(mdata_i)[2] <- "ID"
mdata_i <- left_join(mdata_i, conditions)
mdata_i$measurement <- "norm_batchcl_raw_signal_sum"
mdata <- bind_rows(mdata, mdata_i)
rm(mdata_i)

mdata$condition <- factor(mdata$condition, ordered = TRUE, levels = c("Cyt_0", "Cyt_30", "Cyt_90", "Cyt"))
conditions$condition <- factor(conditions$condition, ordered = TRUE, levels = as.character(unique(conditions$condition)))
mdata$measurement <- factor(mdata$measurement, ordered = TRUE, levels = c("raw_signal_sum", "batchcl_raw"))

#Data transformation overview
ggplot(data = subset(mdata, grepl("norm_batchcl_raw", measurement)), aes(condition, log2(value))) +
  geom_boxplot(aes(fill = rep)) +
  customPlot +
  theme(axis.text.x = element_text(angle = 90, hjust = 1, vjust = 0.5)) +
  facet_grid(SILAC ~ Temperature)

```



```
ggsave(file.path(dir_save, paste0("Normalization_overview_", script.version, ".pdf")), width = 25, height = 15)
#Calculation of fold changes
```

Calculation of ctrl.ratios

```
fc.data <- as.data.frame(2^exprs(norm_batchcl_raw_dataE))
initial.names <- names(fc.data)
comps <- read.csv("stat_comparisons.csv")
fc.metadata <- NULL
for (i in seq_along(comps$ratio.name))
{
  IDs <- grep(paste0("[0-9]_", comps$nominator[i], "_rep"), conditions$ID, value = TRUE)
  if (length(IDs) == 0)
  {
    IDs <- grep(paste0(comps$nominator[i], "_rep"), fc.metadata$new.ID, value = TRUE)
  }
}
```

```

for (j in seq_along(IDs))
{
  nominator <- IDs[j]
  denominator <- gsub(comps$nominator[i], comps$denominator[i], nominator)
  ratio.name <- paste0(gsub(comps$nominator[i], comps$ratio.name[i], nominator),
                        ".ctrl.ratio")
  if (nominator %in% conditions$ID & denominator %in% conditions$ID)
  {
    fc.metadata <- bind_rows(fc.metadata,
                             data.frame(ID = nominator,
                                         new.ID = ratio.name,
                                         condition = comps$ratio.name[i]))
  }else
  {
    fc.metadata <- bind_rows(fc.metadata,
                             data.frame(ID = subset(fc.metadata, new.ID == nominator)$ID,
                                         new.ID = ratio.name,
                                         condition = comps$ratio.name[i]))
  }
  if (denominator %in% names(fc.data))
  {
    fc.data[, ratio.name] <-
      fc.data[, as.character(nominator)] /
      fc.data[, as.character(denominator)]
  }else
  {
    print(paste(denominator, "was not a column name in fc.data"))
    stop()
  }
}
rm(j, IDs)
}
rm(i)
fc.data <- fc.data %>%
  dplyr::select(ends_with(".ctrl.ratio"))
fc.data$gene_name <- rownames(fc.data)
cdata <- left_join(cdata, fc.data)
fc.metadata <- left_join(fc.metadata,
                         conditions[, c("ID",
                                        "rep",
                                        "Temperature",
                                        "construct",
                                        "time",
                                        "SILAC",
                                        "sample",
                                        "col.name",
                                        "file")])
fc.metadata$ID <- fc.metadata$new.ID
fc.metadata$new.ID <- NULL
mfc.data <- fc.data %>%
  group_by(gene_name) %>%
  gather(key = ID, value = value, -gene_name) %>%
  mutate(measurement = "ctrl.ratio")

```

```

mfc.data <- left_join(mfc.data, fc.metadata)
mfc.data <- mfc.data[, names(mdata)]
mdata <- bind_rows(mdata, mfc.data)
rm(fc.data, mfc.data, fc.metadata)

```

Calculation of temp.ratios

```

fc.data <- as.data.frame(exprs(norm_batchcl_raw_dataE))
for (i in seq_along(conditions$ID))
{
  sort(conditions$Temperature)[1]
  fc.data[, paste0(conditions$ID[i], ".temp.ratio")] <-
    fc.data[, as.character(conditions$ID[i])] /
    fc.data[, as.character(gsub(conditions$Temperature[i],
                                sort(conditions$Temperature)[1],
                                conditions$ID[i]))]
}
rm(i)
fc.data <- fc.data %>%
  dplyr::select(ends_with(".temp.ratio"))
fc.data$gene_name <- rownames(fc.data)
cdata <- left_join(cdata, fc.data)
fc.metadata <- data.frame(ID = gsub(".temp.ratio", "", names(fc.data)),
                           new.ID = names(fc.data))
fc.metadata <- left_join(fc.metadata, conditions)
fc.metadata$ID <- fc.metadata$new.ID
fc.metadata$new.ID <- NULL
mfc.data <- fc.data %>%
  group_by(gene_name) %>%
  gather(key = ID, value = value, -gene_name) %>%
  mutate(measurement = "temp.ratio")
mfc.data <- left_join(mfc.data, fc.metadata)
mfc.data <- mfc.data[, names(mdata)]
mdata <- bind_rows(mdata, mfc.data)
rm(fc.data, mfc.data, fc.metadata)

```

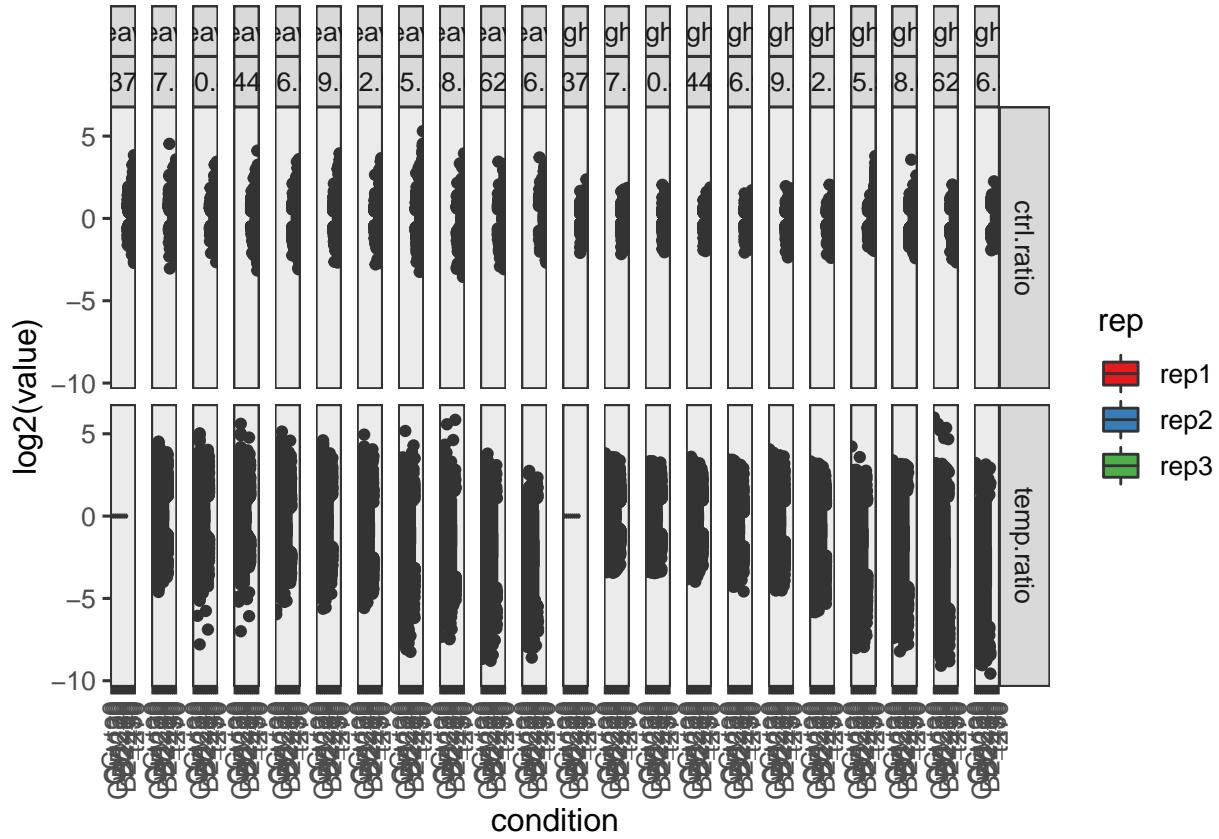
#Factorize tidy data variables

```

mdata$condition <- factor(mdata$condition, ordered = TRUE, levels = unique(append(as.character(levels(c(
  mdata$measurement <- factor(mdata$measurement, ordered = TRUE, levels = c("raw_signal_sum", "batchcl_ra

ggplot(data = subset(mdata, grepl("ratio", measurement)), aes(condition, log2(value))) +
  geom_boxplot(aes(fill = rep)) +
  customPlot +
  theme(axis.text.x = element_text(angle = 90, hjust = 1, vjust = 0.5)) +
  facet_grid(measurement ~ SILAC + Temperature)

```



```

ggsave(file.path(dir_save, paste0("Normalization_overview_ratios_", script.version, ".pdf")), width = 40)

#LIMMA analysis
##Creating limma data set
limma.score.data <- mdata %>%
  dplyr::filter(measurement == "ctrl.ratio", !is.na(value)) %>%
  mutate(ratio = value) %>%
  dplyr::select(gene_name, condition, Temperature, SILAC, rep, ratio)
limma.score.data.length <- mdata %>%
  dplyr::filter(measurement == "ctrl.ratio",
                Temperature %in% sort(unique(conditions$Temperature))[1:2],
                !is.na(value)) %>%
  group_by(gene_name, SILAC, condition, rep) %>%
  summarise(No.of.abundance.Ts = n())
limma.score.data <- left_join(limma.score.data, limma.score.data.length)
rm(limma.score.data.length)
limma.score.data.length <- mdata %>%
  dplyr::filter(measurement == "ctrl.ratio",
                !is.na(value)) %>%
  group_by(gene_name, SILAC, condition, rep) %>%
  summarise(No.of.Ts.total = n())
limma.score.data <- left_join(limma.score.data, limma.score.data.length)
rm(limma.score.data.length)
rep.cut.off <- 2
#allowing only proteins that have at least 2/3 of all replicates present for at least one of the two lo

```

```

score.data.length <- limma.score.data %>%
  dplyr::filter(No.of.abundance.Ts >= 1,
                Temperature %in% sort(unique(conditions$Temperature))[1:2],
                !is.na(ratio)) %>%
  group_by(gene_name, SILAC, condition, Temperature) %>%
  summarise(rep = n()) %>%
  dplyr::filter(rep >= rep.cut.off) %>%
  group_by(gene_name, SILAC, condition) %>%
  summarise(Freq = n()) %>%
  dplyr::filter(Freq >= 1) %>%
  dplyr::select(gene_name, SILAC, condition) %>%
  mutate(abundance.score.valid = TRUE)
limma.score.data <- left_join(limma.score.data, score.data.length)
rm(score.data.length)
limma.score.data$abundance.score.valid[limma.score.data$No.of.abundance.Ts < 1] <- NA
limma.score.data$abundance.score.valid[is.na(limma.score.data$No.of.abundance.Ts)] <- NA
#allowing only Temperature for the stability score calculation for which data points of at least 2/3rd
score.data.length <- limma.score.data %>%
  dplyr::filter(No.of.abundance.Ts >= 1,
                abundance.score.valid,
                No.of.Ts.total >= 5,
                !is.na(ratio)) %>%
  group_by(gene_name, SILAC, condition, Temperature) %>%
  summarise(rep = n()) %>%
  dplyr::filter(rep >= rep.cut.off) %>%
  mutate(stability.score.valid = TRUE) %>%
  dplyr::select(gene_name, SILAC, condition, Temperature, stability.score.valid)
# score.data.length <- mdata %>%
#   dplyr::filter(measurement == "ctrl.ratio",
#                 !is.na(value)) %>%
#   group_by(gene_name, SILAC, condition, Temperature) %>%
#   summarise(rep = n()) %>%
#   dplyr::filter(rep >= rep.cut.off) %>%
#   mutate(stability.score.valid = TRUE) %>%
#   dplyr::select(gene_name, SILAC, condition, Temperature, stability.score.valid)

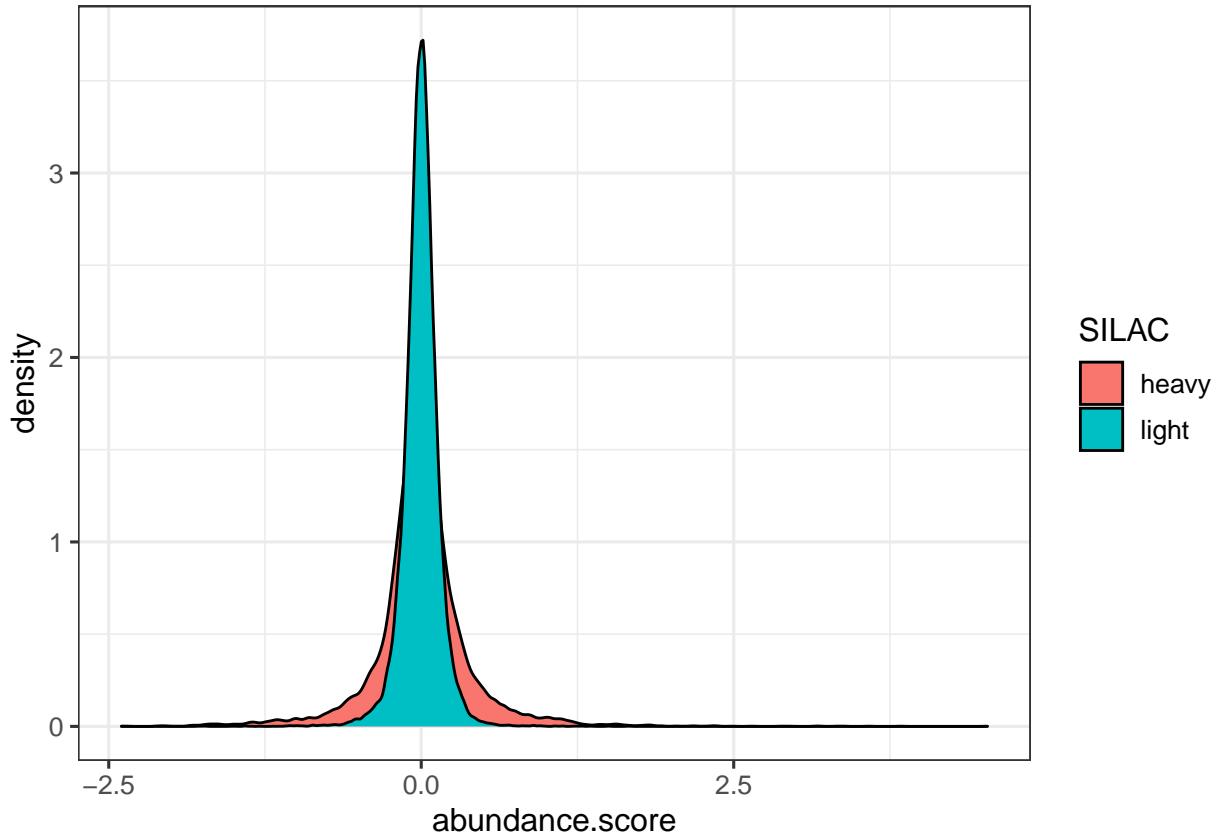
limma.score.data <- left_join(limma.score.data, score.data.length)
limma.score.data$stability.score.valid[limma.score.data$No.of.Ts.total < 5] <- NA
score.data.length <- limma.score.data %>%
  group_by(gene_name, condition, SILAC, rep) %>%
  dplyr::filter(abundance.score.valid,
                stability.score.valid) %>%
  summarise(n = n()) %>%
  dplyr::filter(n >= 5) %>%
  group_by(gene_name, condition, SILAC) %>%
  summarise(n = n()) %>%
  dplyr::filter(n >= 2) %>%
  dplyr::select(gene_name, condition, SILAC) %>%
  mutate(stability.score.valid.rep = TRUE)
limma.score.data <- left_join(limma.score.data, score.data.length)
rm(score.data.length)
limma.score.data$stability.score.valid <- limma.score.data$stability.score.valid & limma.score.data$sta
limma.score.data$stability.score.valid.rep <- NULL

```

```

abundance.score <- limma.score.data %>%
  subset(Temperature %in% sort(unique(limma.score.data$Temperature))[1:2]) %>%
  subset(abundance.score.valid) %>%
  group_by(gene_name, SILAC, condition, rep) %>%
  summarise(abundance.score = mean(log2(ratio), na.rm = TRUE))
limma.score.data <- left_join(limma.score.data, abundance.score)
limma.score.data <- limma.score.data %>%
  mutate(stability = log2(ratio) - abundance.score)
limma.score.data$stability[!limma.score.data$stability.score.valid] <- NA
limma.score.data$stability[is.na(limma.score.data$stability.score.valid)] <- NA
stability.score <- limma.score.data %>%
  dplyr::filter(stability.score.valid & abundance.score.valid) %>%
  group_by(gene_name, SILAC, condition, rep) %>%
  summarise(stability.score = sum(stability, na.rm = TRUE))
score.count.table <- limma.score.data %>%
  dplyr::select(gene_name, condition, SILAC, rep, No.of.abundance.Ts, No.of.Ts.total) %>%
  unique()
limma.score.data <- left_join(score.count.table, abundance.score)
limma.score.data <- left_join(limma.score.data, stability.score)
# limma.score.data <- left_join(abundance.score, stability.score)
limma.score.data <- limma.score.data %>%
  mutate(ID = paste0(SILAC, "_", condition, "_", rep))
rm(abundance.score, stability.score)
# limma.score.data <- limma.score.data %>%
#   group_by(gene_name, SILAC, condition, rep) %>%
#   dplyr::select(gene_name, SILAC, condition, No.of.abundance.Ts, No.of.Ts.total,
#                 abundance.score.valid, stability.score.valid,
#                 rep, abundance.score, stability.score)
# limma.score.data <- unique(limma.score.data)
# limma.score.data <- limma.score.data %>%
#   mutate(ID = paste0(SILAC, "_", condition, "_", rep)) %>%
#   group_by(gene_name, ID, SILAC, condition, rep) %>%
#   dplyr::select(gene_name, ID, SILAC, condition, rep, No.of.abundance.Ts, No.of.Ts.total,
#                 abundance.score.valid, stability.score.valid,
#                 abundance.score, stability.score) %>%
#   filter(No.of.abundance.Ts >= 1, No.of.Ts.total >= 4)
ggplot(data = limma.score.data, aes(abundance.score)) +
  geom_density(aes(fill = SILAC)) +
  theme_bw(base_size = 12)

```



```

limma.score.data$abundance.score[limma.score.data$SILAC == "heavy"] <- scale(limma.score.data$abundance
limma.score.data$stability.score[limma.score.data$SILAC == "heavy"] <- scale(limma.score.data$stability

limma.score.data$abundance.score[limma.score.data$SILAC == "light"] <- scale(limma.score.data$abundance
limma.score.data$stability.score[limma.score.data$SILAC == "light"] <- scale(limma.score.data$stability
write.csv(limma.score.data,file.path(dir_save, paste0("raw_score_data_",script.version,".csv")))

##Correlation of rep1 to rep2 to rep3
rep.cor.data <- limma.score.data %>%
  dplyr::filter(abundance.score.valid, stability.score.valid)
rep.cor.data <- rep.cor.data %>%
  ungroup() %>%
  select(gene_name, ID, SILAC, abundance.score, stability.score) %>%
  unique()
rep.cor.data$rep <- gsub(".+_\rep","rep",rep.cor.data$ID)
rep.cor.data$ID <- gsub("_\rep.+","",rep.cor.data$ID)

crep.cor.data <- rep.cor.data %>%
  group_by(gene_name, SILAC, ID, rep) %>%
  gather(key = score.name, value = score, -gene_name, -ID, -rep, -SILAC) %>%
  group_by(gene_name, SILAC, score.name) %>%
  spread(key = rep, value = score) %>%
  mutate(time = gsub("(heavy_|(light_)", "", ID))

ggplot(data = crep.cor.data, aes(rep1, rep2)) +
  geom_abline() +

```

```

geom_vline(xintercept = 0) +
geom_hline(yintercept = 0) +
geom_point(alpha = 0.1) +
facet_grid(ID ~ score.name, scale = "free") +
geom_text(data = subset(crep.cor.data, abs(rep2 - rep1) > 3), aes(label = gene_name), size = 2,check_
geom_point(data = subset(crep.cor.data, abs(rep2 - rep1) > 3), size = 2, colour = "red") +
geom_smooth(method = "loess") +
theme_bw(base_size = 12)
ggsave(file.path(dir_save, paste0("Rep1_vs_rep2_score_correlation_", script.version, ".pdf")),width = 12, height = 8)

ggplot(data = crep.cor.data, aes(rep1, rep3)) +
geom_abline() +
geom_vline(xintercept = 0) +
geom_hline(yintercept = 0) +
geom_point(alpha = 0.1) +
facet_grid(ID ~ score.name, scale = "free") +
geom_text(data = subset(crep.cor.data, abs(rep3 - rep1) > 3), aes(label = gene_name), size = 2,check_
geom_point(data = subset(crep.cor.data, abs(rep3 - rep1) > 3), size = 2, colour = "red") +
geom_smooth(method = "loess") +
theme_bw(base_size = 12)
ggsave(file.path(dir_save, paste0("Rep1_vs_rep3_score_correlation_", script.version, ".pdf")),width = 12, height = 8)

ggplot(data = crep.cor.data, aes(rep2, rep3)) +
geom_abline() +
geom_vline(xintercept = 0) +
geom_hline(yintercept = 0) +
geom_point(alpha = 0.1) +
facet_grid(ID ~ score.name, scale = "free") +
geom_text(data = subset(crep.cor.data, abs(rep3 - rep2) > 3), aes(label = gene_name), size = 2,check_
geom_point(data = subset(crep.cor.data, abs(rep3 - rep2) > 3), size = 2, colour = "red") +
geom_smooth(method = "loess") +
theme_bw(base_size = 12)
ggsave(file.path(dir_save, paste0("Rep2_vs_rep3_score_correlation_", script.version, ".pdf")),width = 12, height = 8)

write.csv(crep.cor.data,file.path(dir_save, paste0("Data_replicate_score_correlation_",script.version, ".csv")))

##Constructing expression sets for abundance and stability data

##abundance
abundance.data <- limma.score.data %>%
  group_by(gene_name) %>%
  dplyr::select(gene_name, ID, abundance.score) %>%
  unique() %>%
  spread(ID, abundance.score)
abundance.weights <- limma.score.data %>%
  group_by(gene_name) %>%
  dplyr::select(gene_name, ID, No.of.abundance.Ts) %>%
  unique() %>%
  spread(ID, No.of.abundance.Ts)
abundance.weights <- base::as.data.frame(abundance.weights)
rownames(abundance.weights) <- abundance.weights$gene_name
abundance.weights$gene_name <- NULL
abundance.weights <- as.matrix(abundance.weights)
abundance.data <- base::as.data.frame(abundance.data)

```

```

rownames(abundance.data) <- abundance.data$gene_name
abundance.data$gene_name <- NULL
abundance.metadata <- data.frame(ID = names(abundance.data))
abundance.metadata <- left_join(abundance.metadata,
                                unique(limma.score.data[, c("ID", "SILAC", "condition", "rep")]))
abundance.metadata$condition <- paste(abundance.metadata$SILAC, abundance.metadata$condition, sep = "_")
rownames(abundance.metadata) <- abundance.metadata$ID
colnames(abundance.data) <- abundance.metadata$ID
abundance.data <- as.matrix(abundance.data)
abundance.dataE <- ExpressionSet(assayData = abundance.data,
                                   phenoData = AnnotatedDataFrame(abundance.metadata))
validObject(abundance.dataE)

## [1] TRUE
rm(abundance.data, abundance.metadata)

##stability
stability.data <- limma.score.data %>%
  group_by(gene_name) %>%
  dplyr::select(gene_name, ID, stability.score) %>%
  unique() %>%
  spread(ID, stability.score)
stability.weights <- limma.score.data %>%
  group_by(gene_name) %>%
  dplyr::select(gene_name, ID, No.of.Ts.total) %>%
  unique() %>%
  spread(ID, No.of.Ts.total)
stability.weights <- base::as.data.frame(stability.weights)
rownames(stability.weights) <- stability.weights$gene_name
stability.weights$gene_name <- NULL
stability.data <- base::as.data.frame(stability.data)
rownames(stability.data) <- stability.data$gene_name
stability.data$gene_name <- NULL
stability.metadata <- data.frame(ID = names(stability.data))
stability.metadata <- left_join(stability.metadata,
                                unique(limma.score.data[, c("ID", "SILAC", "condition", "rep")]))
stability.metadata$condition <- paste(stability.metadata$SILAC, stability.metadata$condition, sep = "_")
rownames(stability.metadata) <- stability.metadata$ID
colnames(stability.data) <- stability.metadata$ID
stability.data <- as.matrix(stability.data)
stability.dataE <- ExpressionSet(assayData = stability.data,
                                   phenoData = AnnotatedDataFrame(stability.metadata))
validObject(stability.dataE)

## [1] TRUE
rm(stability.data, stability.metadata)

##Limma evaluation of stability and abundance scores
limma_results <- NULL

##Construction of design matrix and defining conditions that will be compared

```

```

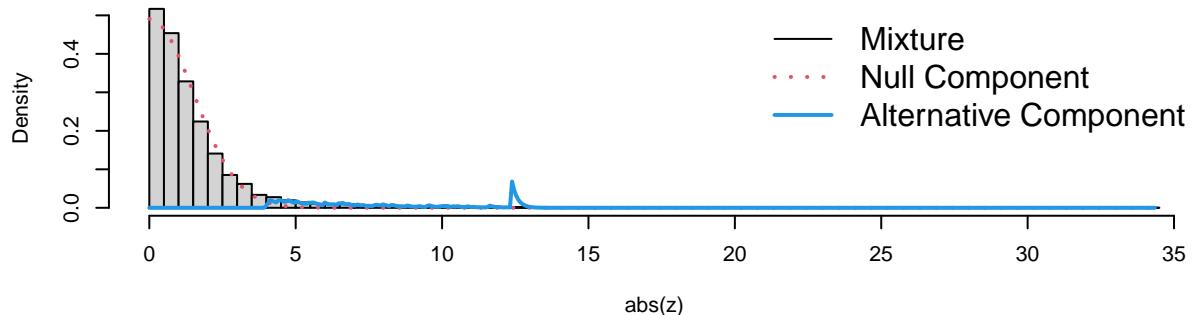
for (to_condition in unique(gsub("_rep.+","", limma.score.data$ID)))
{
  limma_data = abundance.dataE
  limma_data <- limma_data[, limma_data$condition == to_condition]
  condition <- factor(pData(limma_data)$condition, ordered = FALSE)
  replicate <- factor(pData(limma_data)$rep, ordered = FALSE)
  abundance.weights.sub <- abundance.weights[, grepl(paste0("^", to_condition, "_rep"),
                                                       colnames(abundance.weights))]
  mut_comparison <- eBayes(lmFit(limma_data, weights = abundance.weights.sub^2))
  res <- limma::topTable(mut_comparison, sort.by = "t",
                         coef = 1, number = Inf)
  res$gene_name <- rownames(res)
  names(res)[grep("P.Value", names(res))] <- "pvalue.limma"
  names(res)[grep("adj.P.Val", names(res))] <- "fdr.limma"
  res <- subset(res,!is.na(logFC))
  res$pvalue.fdrtool <- NA
  res$qval.fdrtool <- NA
  res$lfdr.fdrtool <- NA
  res$comparison <- paste0("abundance - ", to_condition)
  res$comparison2 <- to_condition
  res$score <- "abundance"
  limma_results <- rbind(limma_results, res)

  limma_data = stability.dataE
  limma_data <- limma_data[, limma_data$condition == to_condition]
  condition <- factor(pData(limma_data)$condition, ordered = FALSE)
  replicate <- factor(pData(limma_data)$rep, ordered = FALSE)
  condition_d <- model.matrix(~1)
  stability.weights.sub <- stability.weights[, grepl(paste0("^", to_condition, "_rep"),
                                                       colnames(stability.weights))]
  mut_comparison <- eBayes(lmFit(limma_data, weights = stability.weights.sub^2))
  res <- limma::topTable(mut_comparison, sort.by = "t",
                         coef = 1, number = Inf)
  res$gene_name <- rownames(res)
  names(res)[grep("P.Value", names(res))] <- "pvalue.limma"
  names(res)[grep("adj.P.Val", names(res))] <- "fdr.limma"
  res <- subset(res,!is.na(logFC))
  res$pvalue.fdrtool <- NA
  res$qval.fdrtool <- NA
  res$lfdr.fdrtool <- NA
  res$comparison <- paste0("stability - ", to_condition)
  res$comparison2 <- to_condition
  res$score <- "stability"
  limma_results <- rbind(limma_results, res)
}
rm(to_condition)

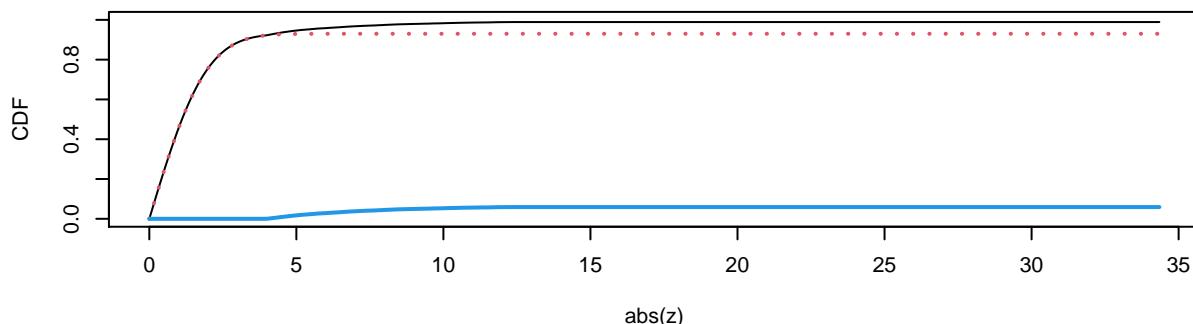
fdr_res_abundance <- NULL
try(fdr_res_abundance <- fdrtool(limma_results$t[limma_results$score == "abundance"], plot = TRUE, verbose = TRUE))

```

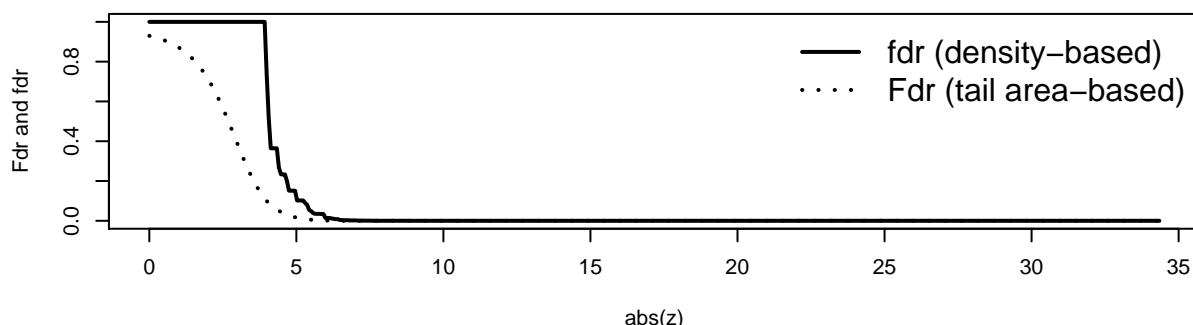
Type of Statistic: z-Score (sd = 1.512, eta0 = 0.9301)



Density (first row) and Distribution Function (second row)



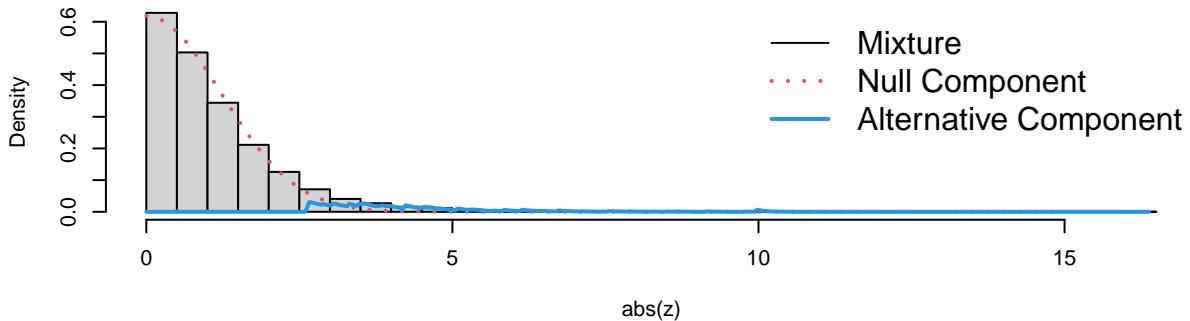
(Local) False Discovery Rate



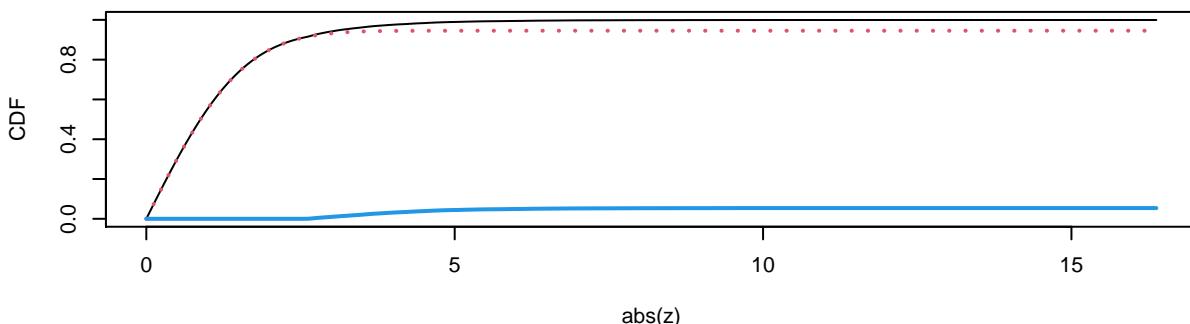
```

if (length(fdr_res_abundance) > 0)
{
  limma_results$pvalue.fdrtool[limma_results$score == "abundance"] <- fdr_res_abundance$pval
  limma_results$qval.fdrtool[limma_results$score == "abundance"] <- fdr_res_abundance$qval
  limma_results$lfdr.fdrtool[limma_results$score == "abundance"] <- fdr_res_abundance$lfdr
}
rm(fdr_res_abundance)
fdr_res_stability <- NULL
try(fdr_res_stability <- fdrtool(limma_results$t[limma_results$score == "stability"]),
  plot = TRUE, verbose = TRUE)
  
```

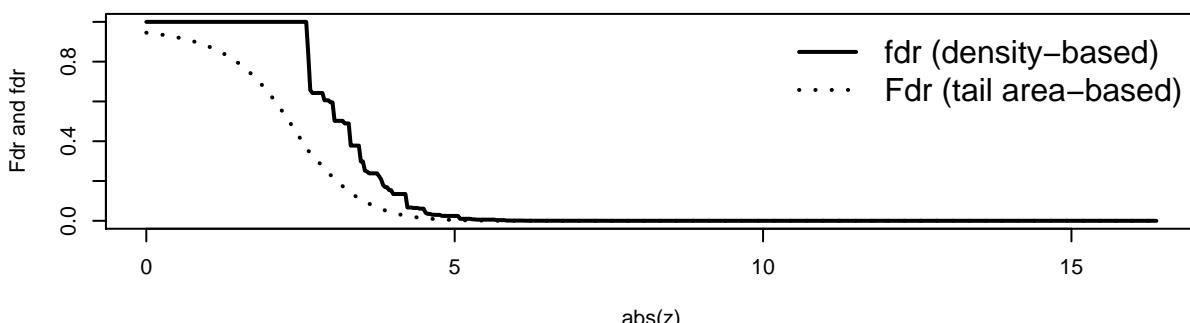
Type of Statistic: z-Score (sd = 1.22, eta0 = 0.9454)



Density (first row) and Distribution Function (second row)



(Local) False Discovery Rate



```

if (length(fdr_res_stability) > 0)
{
  limma_results$pvalue.fdrtool[limma_results$score == "stability"] <- fdr_res_stability$pval
  limma_results$qval.fdrtool[limma_results$score == "stability"] <- fdr_res_stability$qval
  limma_results$lfdr.fdrtool[limma_results$score == "stability"] <- fdr_res_stability$lfdr
}
rm(fdr_res_stability)

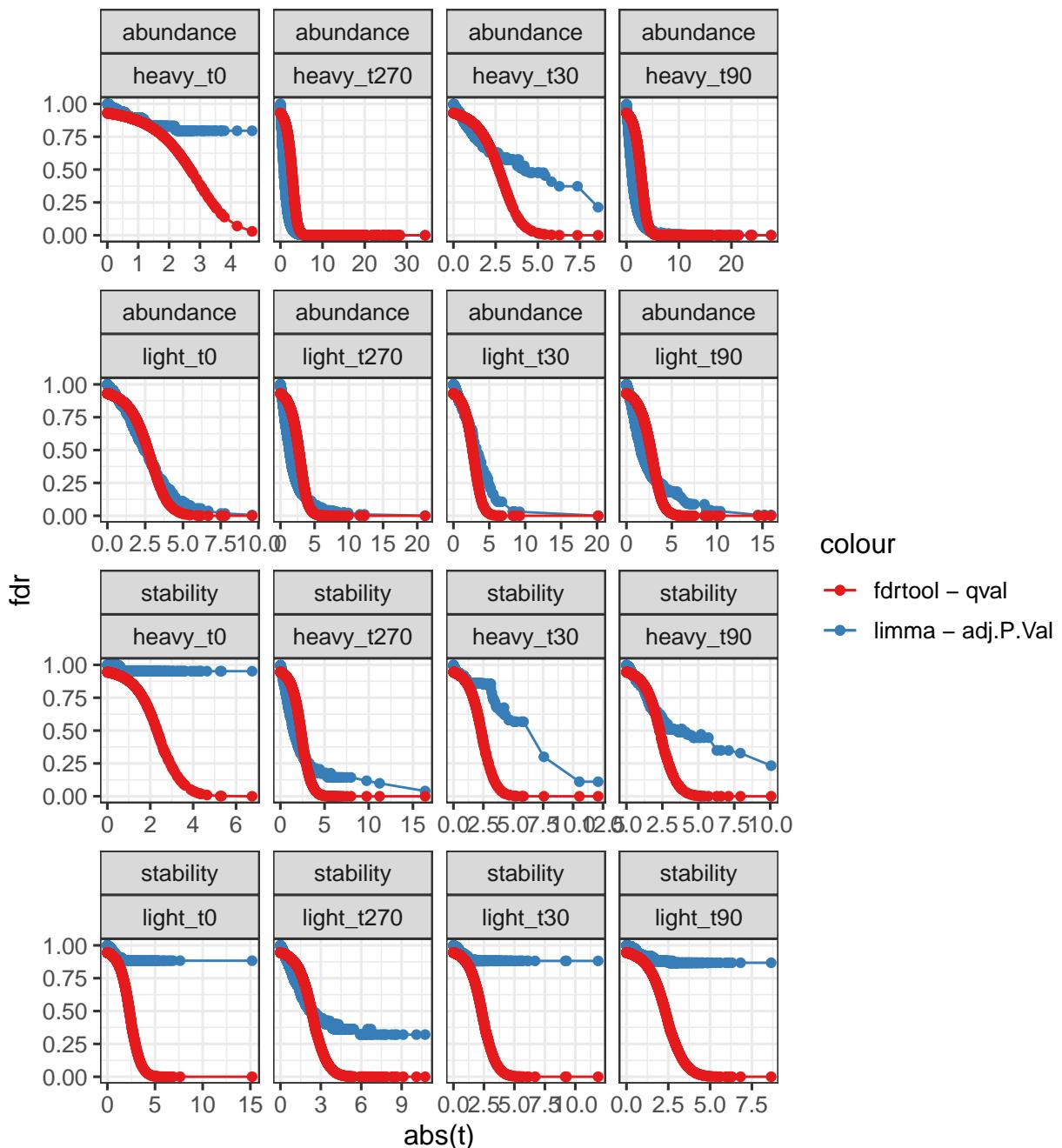
##Limma and fdrtool statistics comparison
ggplot(data = limma_results, aes(abs(t))) +
  geom_line(aes(y = fdr.limma, colour = "limma - adj.P.Val")) +

```

```

geom_line(aes(y = qval.fdrtool, colour = "fdrtool - qval")) +
geom_point(aes(y = fdr.limma, colour = "limma - adj.P.Val")) +
geom_point(aes(y = qval.fdrtool, colour = "fdrtool - qval")) +
facet_wrap(score ~ comparison2, scale = "free_x") + ylab("fdr") +
theme_bw(base_size = 12) +
scale_colour_brewer(palette = "Set1")

```



```

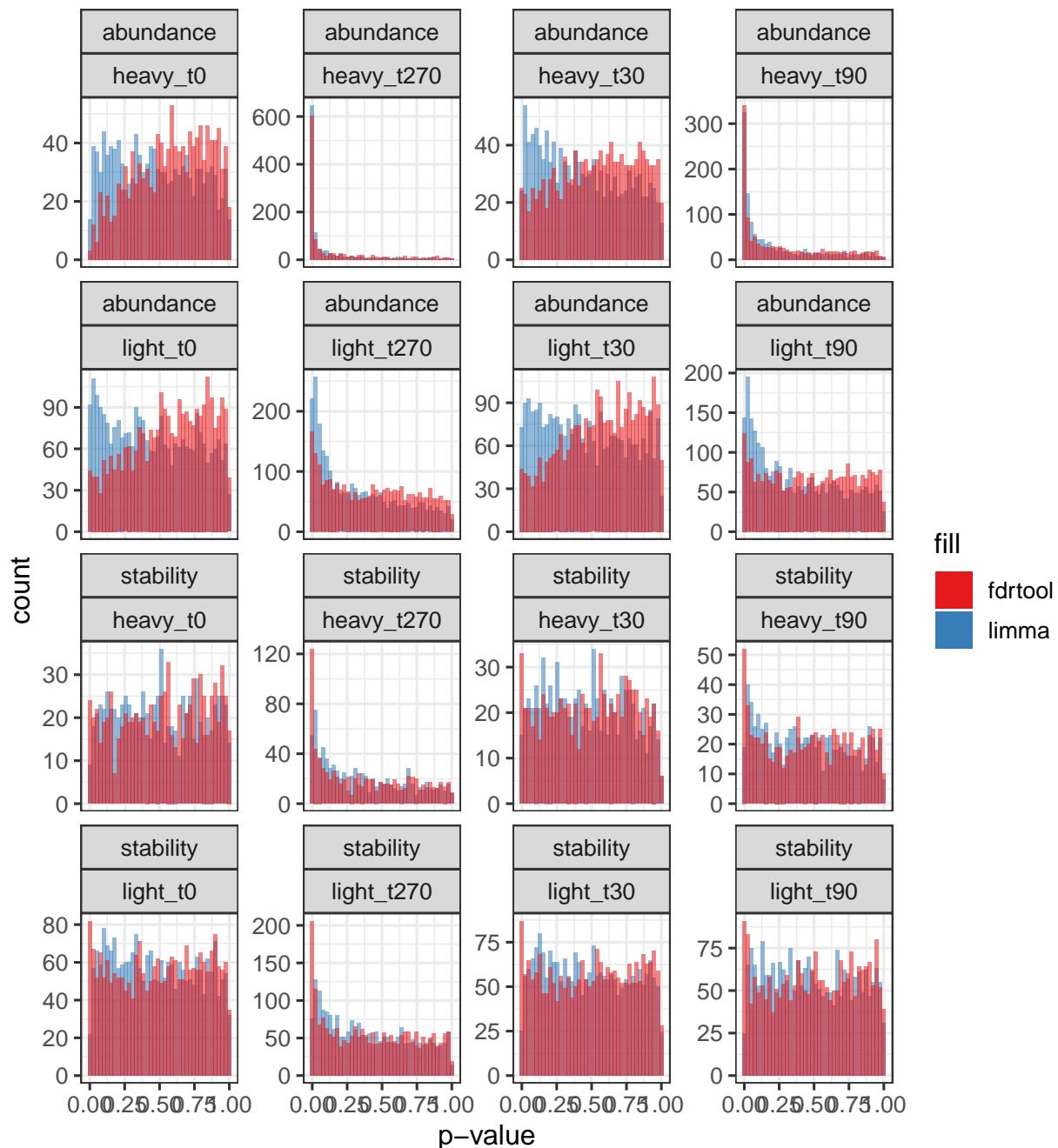
ggplot(data = limma_results) +
  geom_histogram(aes(pvalue.limma, alpha = 0.5, fill = "limma"), bins = 40) +

```

```

facet_wrap(score ~ comparison2, scale = "free_y") +
geom_histogram(aes(pvalue.fdrtool, alpha = 0.5, fill = "fdrtool"), bins = 40) +
guides(alpha = FALSE) +
xlab("p-value") +
theme_bw(base_size = 12) +
scale_fill_brewer(palette = "Set1")

```



```

ggsave(file.path(dir_save, paste0("p-value_histogram_limma_vs_fdrtool_", script.version, ".pdf")), width
##Hit annotation

```

```

limma_results$score.value <- limma_results$logFC

limma_results$hit_annotation_method <- NA
fdr_hit_threshold <- 0.01
fdr_candidate_threshold = 0.05
score_hit_threshold <- 3
score_candidate_threshold <- 2
limma_results$pvalue <- NA
limma_results$fdr <- NA
for (comp in unique(limma_results$comparison))
{
  limma_hits <-
    nrow(limma_results[limma_results$comparison == comp &
                      limma_results$fdr.limma <= fdr_hit_threshold,])
  fdrtool_hits <-
    nrow(limma_results[limma_results$comparison == comp &
                      limma_results$qval.fdrtool <= fdr_hit_threshold,])
  if (limma_hits >= fdrtool_hits)
  {
    limma_results$hit_annotation_method[limma_results$comparison == comp] <- "limma"
  }
  if (fdrtool_hits > limma_hits)
  {
    limma_results$hit_annotation_method[limma_results$comparison == comp] <- "fdrtool"
  }
  rm(limma_hits,fdrtool_hits)
}
table(limma_results$hit_annotation_method)

##
## fdrtool      limma
## 26043      2464

limma_results$hit_annotation_method <- "fdrtool"

limma_results$pvalue[limma_results$hit_annotation_method == "limma"] <-
  limma_results$pvalue.limma[limma_results$hit_annotation_method == "limma"]
limma_results$fdr[limma_results$hit_annotation_method == "limma"] <-
  limma_results$fdr.limma[limma_results$hit_annotation_method == "limma"]

limma_results$pvalue[limma_results$hit_annotation_method == "fdrtool"] <-
  limma_results$pvalue.fdrtool[limma_results$hit_annotation_method == "fdrtool"]
limma_results$fdr[limma_results$hit_annotation_method == "fdrtool"] <-
  limma_results$qval.fdrtool[limma_results$hit_annotation_method == "fdrtool"]

limma_results$hit <- with(limma_results,
                         ifelse(fdr <= fdr_hit_threshold & abs(score.value) >=
                                score_hit_threshold,T,F))
limma_results$hit_annotation <- with(limma_results,
                                      ifelse(fdr <= fdr_hit_threshold & abs(score.value) >=
                                             score_hit_threshold,"hit",
                                             ifelse(fdr <= fdr_candidate_threshold & abs(score.value) >=
                                                    score_candidate_threshold,
                                                       "candidate","no hit")))

```

```

limma_results$hit_annotation <- factor(limma_results$hit_annotation, ordered = TRUE,
                                         levels = c("hit", "candidate", "no hit"))

with(limma_results, table(comparison, hit_annotation))

##                                     hit_annotation
## comparison                  hit candidate no hit
## abundance - heavy_t0        0      0    1232
## abundance - heavy_t270     88     117   1027
## abundance - heavy_t30       1      0    1231
## abundance - heavy_t90       5      37   1190
## abundance - light_t0        2      16   2765
## abundance - light_t270     23     32   2728
## abundance - light_t30       4      13   2766
## abundance - light_t90       8      15   2760
## stability - heavy_t0       1      4    830
## stability - heavy_t270     3      9    824
## stability - heavy_t30       0      6    830
## stability - heavy_t90       1      2    833
## stability - light_t0       1      9    2266
## stability - light_t270     4     23   2249
## stability - light_t30       1      3   2272
## stability - light_t90       1      4   2271

##Save limma results
write.csv(limma_results, file.path(dir_save, paste0("Limma_results_", script.version, ".csv")), row.names = FALSE)

##Merge limma significances with raw score data
limma.cp.data <- limma.score.data %>%
  group_by(gene_name, ID, SILAC, condition, rep) %>%
  gather(key = score, value = score.value, -gene_name, -ID, -No.of.abundance.Ts,
         -No.of.Ts.total,
         -condition, -rep, -SILAC)
limma.sub <- limma_results[, c("gene_name", "fdr", "comparison2", "score")] %>%
  unique()
limma.sub$score <- paste0(limma.sub$score, ".score")
limma.sub$condition <- limma.sub$comparison2
limma.sub$comparison2 <- NULL
limma.sub$SILAC <- gsub("_+", "", limma.sub$condition)
limma.sub$condition <- gsub(".+_", "", limma.sub$condition)
limma.cp.data <- left_join(limma.cp.data, limma.sub)
limma.cp.data$fdr.star <-
  as.character(cut(as.numeric(as.character(limma.cp.data$fdr)), ,
                   breaks = c(0, 0.001, 0.01, 0.05, 1),
                   include.lowest = TRUE))
limma.cp.data$fdr.star[is.na(limma.cp.data$fdr.star)] <- ""
limma.cp.data$fdr.star <- recode(limma.cp.data$fdr.star,
                                    "(0.05,1]" = "",
                                    "(0.01,0.05]" = "*",
                                    "(0.001,0.01]" = "**",
                                    "[0,0.001]" = "***")

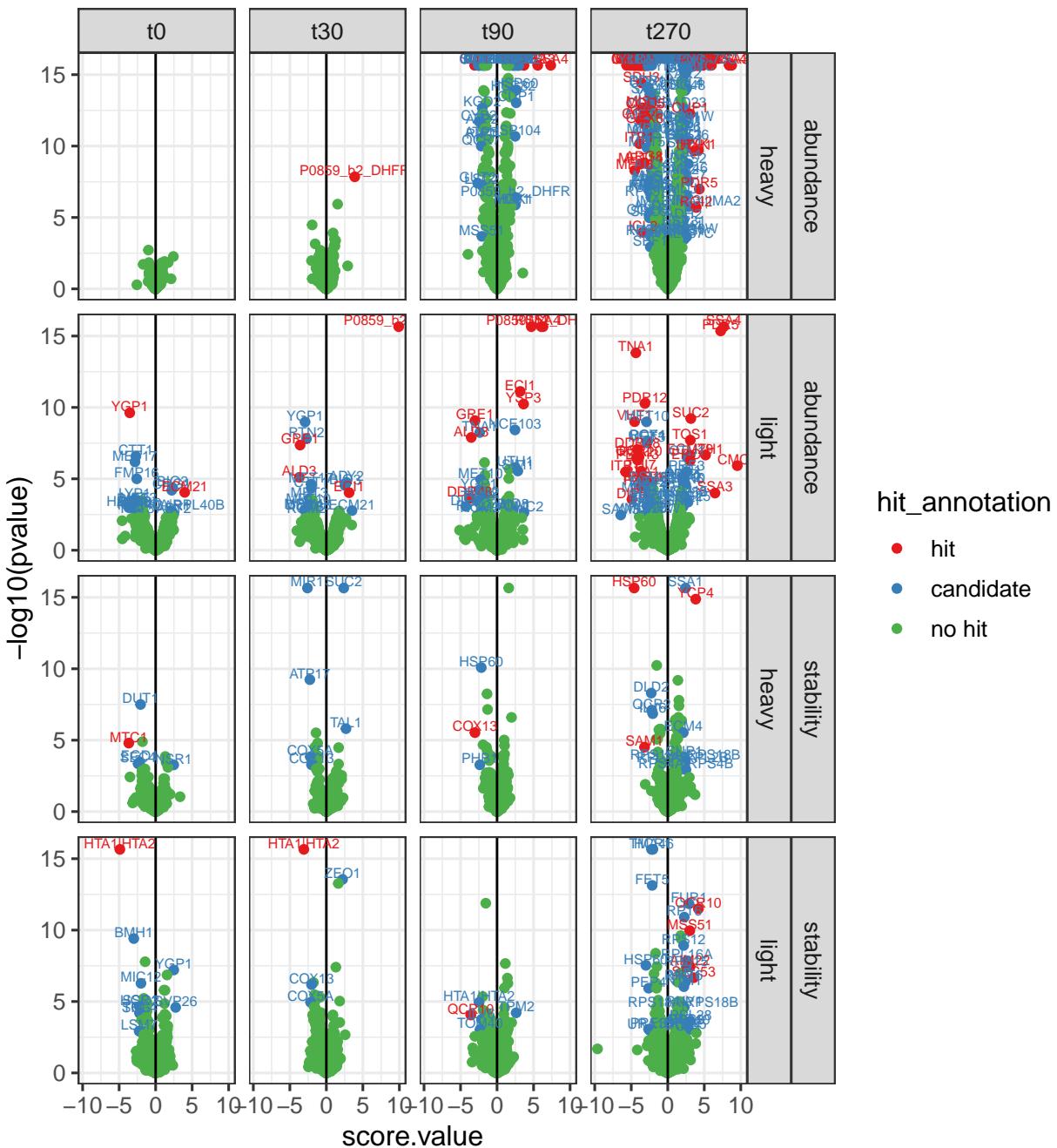
##Volcano-Plot

```

```

limma_results$sample <- gsub(" - .+", "", limma_results$comparison)
limma_results$comparison <- gsub(".+ - ", "", limma_results$comparison)
limma_results$time <- gsub(".+_", "", limma_results$comparison)
limma_results$time <- factor(limma_results$time, ordered = T, levels = c("t0", "t30", "t90", "t270"))
limma_results$SILAC <- gsub("_.+", "", limma_results$comparison)
qplot(score.value, -log10(pvalue), data = limma_results, colour = hit_annotation) +
  geom_vline(aes(xintercept = 0)) +
  facet_grid(sample + SILAC ~ time) +
  geom_text(aes(label = gene_name),
            data = subset(limma_results, hit_annotation != "no hit"),
            vjust = 0, nudge_y = 0.1, size = 2, check_overlap = FALSE) +
  theme_bw(base_size = 12) +
  scale_colour_brewer(palette = "Set1")

```



```
ggsave(file.path(dir_save, paste0("Volcano_plot_", script.version, ".pdf")), width = 15, height = 15)

pdf(file.path(dir_save, paste0("Volcano_plot_single_pages_", script.version, ".pdf")), width = 12, height = 10)
for (comp in unique(limma_results$comparison2))
{
  print(comp)
  sub <- subset(limma_results, comparison2 == comp)
  print(qplot(score.value, -log10(pvalue), data = sub, colour = hit_annotation) +
    geom_vline(aes(xintercept = 0)) +
    facet_grid(comparison2 ~ score + hit_annotation_method) +
    theme_minimal() +
    xlab("Score") +
    ylab("-log10(p-value)"))
}
```

```

geom_text(aes(label = gene_name),
          data = subset(sub, hit_annotation != "no hit"),
          vjust = 0, nudge_y = 0.1, size = 2, check_overlap = FALSE) +
theme_bw(base_size = 12) +
scale_colour_brewer(palette = "Set1"))
}

## [1] "light_t0"
## [1] "heavy_t0"
## [1] "light_t30"
## [1] "heavy_t30"
## [1] "light_t90"
## [1] "heavy_t90"
## [1] "light_t270"
## [1] "heavy_t270"
dev.off()

## pdf
## 2
rm(sub)

##Correlation of limma results - abundance vs stability
cor.data <- limma_results %>%
  group_by(gene_name, comparison2) %>%
  dplyr::select(gene_name, comparison2, score.value, score) %>%
  spread(key = score, value = score.value)
cor.data_i <- limma_results %>%
  group_by(gene_name, comparison2) %>%
  dplyr::select(gene_name, comparison2, hit_annotation, score) %>%
  mutate(hit_annotation = gsub("enriched ", "", hit_annotation)) %>%
  spread(key = score, value = hit_annotation)
names(cor.data_i)[-c(1, 2)] <- paste0("hit_", names(cor.data_i)[-c(1, 2)])
cor.data <- left_join(cor.data, cor.data_i)
rm(cor.data_i)
cor.data$comparison <- cor.data$comparison2
cor.data$comparison2 <- NULL
cor.data <- cor.data %>%
  group_by(gene_name, comparison)
cor.data <- na.omit(cor.data)
cor.data$hit <- paste(cor.data$hit_abundance, "abundance -", cor.data$hit_stability, "stability")
cor.data$hit <- factor(cor.data$hit, ordered = TRUE,
                       levels = c("hit abundance - hit stability",
                                 "hit abundance - candidate stability",
                                 "candidate abundance - hit stability",
                                 "candidate abundance - candidate stability",
                                 "hit abundance - no hit stability",
                                 "no hit abundance - hit stability",
                                 "candidate abundance - no hit stability",
                                 "no hit abundance - candidate stability",

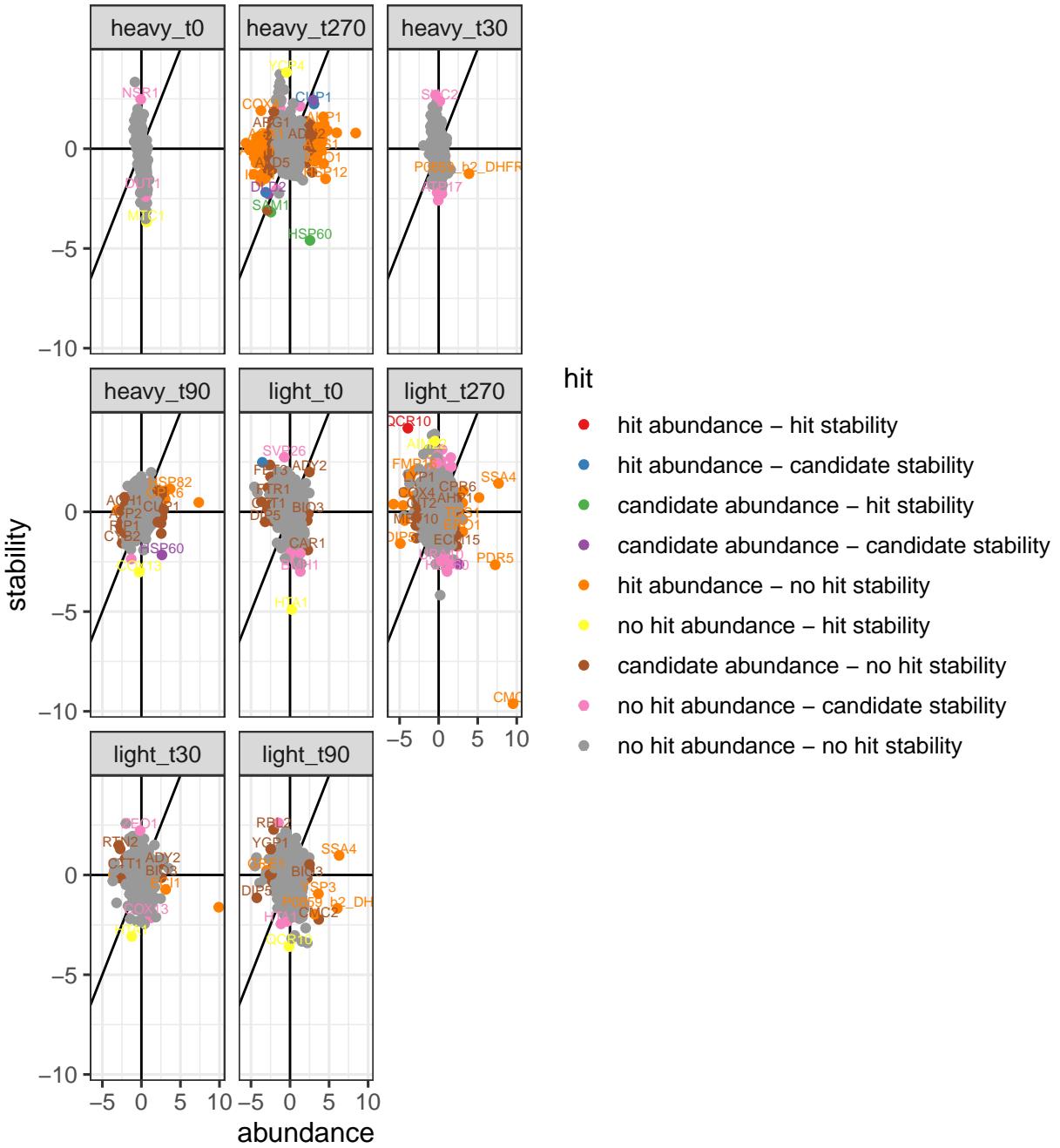
```

```

                    "no hit abundance - no hit stability"))
no.of.comparisons <- length(unique(cor.data$comparison))
gr.width <- 6 + ceiling(sqrt(no.of.comparisons)) * 7
gr.height <- floor(sqrt(no.of.comparisons)) * 7

ggplot(data = cor.data, aes(abundance, stability, colour = hit)) +
  geom_abline() +
  geom_vline(xintercept = 0) +
  geom_hline(yintercept = 0) +
  geom_point() +
  facet_wrap(~ comparison) +
  geom_text(aes(label = gsub("[|].+", "", gene_name)),
            data = subset(cor.data, hit != "no hit abundance - no hit stability"),
            vjust = 0, nudge_y = 0.1, size = 2, check_overlap = TRUE) +
  customPlot

```



```

ggsave(file.path(dir_save, paste0("Fold_change_correlation_abundance_vs_stability_", script.version, ".pdf"))

pdf(file.path(dir_save, paste0("Fold_change_correlation_abundance_vs_stability_single_pages_", script.v
for(comp in unique(cor.data$comparison))
{
  print(comp)
  sub <- subset(cor.data, comparison == comp)
  print(ggplot(data = sub, aes(abundance, stability, colour = hit)) +
    geom_abline() +
    geom_vline(xintercept = 0) +
    geom_hline(yintercept = 0) +
    geom_point())
}

```

```

    geom_point() +
    facet_wrap(~ comparison) +
    geom_text(aes(label = gsub("[|].+", "", gene_name)),
              data = subset(sub, hit != "no hit abundance - no hit stability"),
              vjust = 0, nudge_y = 0.1, size = 2, check_overlap = TRUE) +
    customPlot)
}

## [1] "heavy_t0"
## [1] "heavy_t270"
## [1] "heavy_t30"
## [1] "heavy_t90"
## [1] "light_t0"
## [1] "light_t270"
## [1] "light_t30"
## [1] "light_t90"
dev.off()

## pdf
## 2
rm(sub, comp)

write.csv(cor.data, file = file.path(dir_save, paste0("Fold_change_correlation_data_abundance_vs_stabi"))

##Correlation of limma results
cor.data <- limma_results %>%
  group_by(gene_name, score) %>%
  dplyr::select(gene_name, comparison2, score.value, score) %>%
  spread(key = comparison2, value = score.value)
cor.data_i <- limma_results %>%
  group_by(gene_name, score) %>%
  dplyr::select(gene_name, comparison2, hit_annotation, score) %>%
  mutate(hit_annotation = gsub("enriched ", "", hit_annotation)) %>%
  spread(key = comparison2, value = hit_annotation)
names(cor.data_i)[-c(1, 2)] <- paste0("hit_", names(cor.data_i)[-c(1, 2)])
cor.data <- left_join(cor.data, cor.data_i)
cor.data <- as.data.frame(cor.data)
rm(cor.data_i)
conds <- gsub("hit_", "", names(cor.data)[grep("hit_", names(cor.data))])
comparisons <- data.frame(x = c("heavy_t0", "heavy_t30", "heavy_t90", "heavy_t270", "light_t30", "light_t90",
                                 y = c("light_t0", "light_t30", "light_t90", "light_t270", "light_t0", "light_t90"))
mcor.data <- NULL
for (i in seq_along(comparisons$x))
{
  mcor.data_i <- data.frame(gene_name = cor.data$gene_name,
                            score = cor.data$score,
                            x = cor.data[, as.character(comparisons$x[i])],
                            y = cor.data[, as.character(comparisons$y[i])],
                            hit_x = cor.data[, paste0("hit_", comparisons$x[i])],

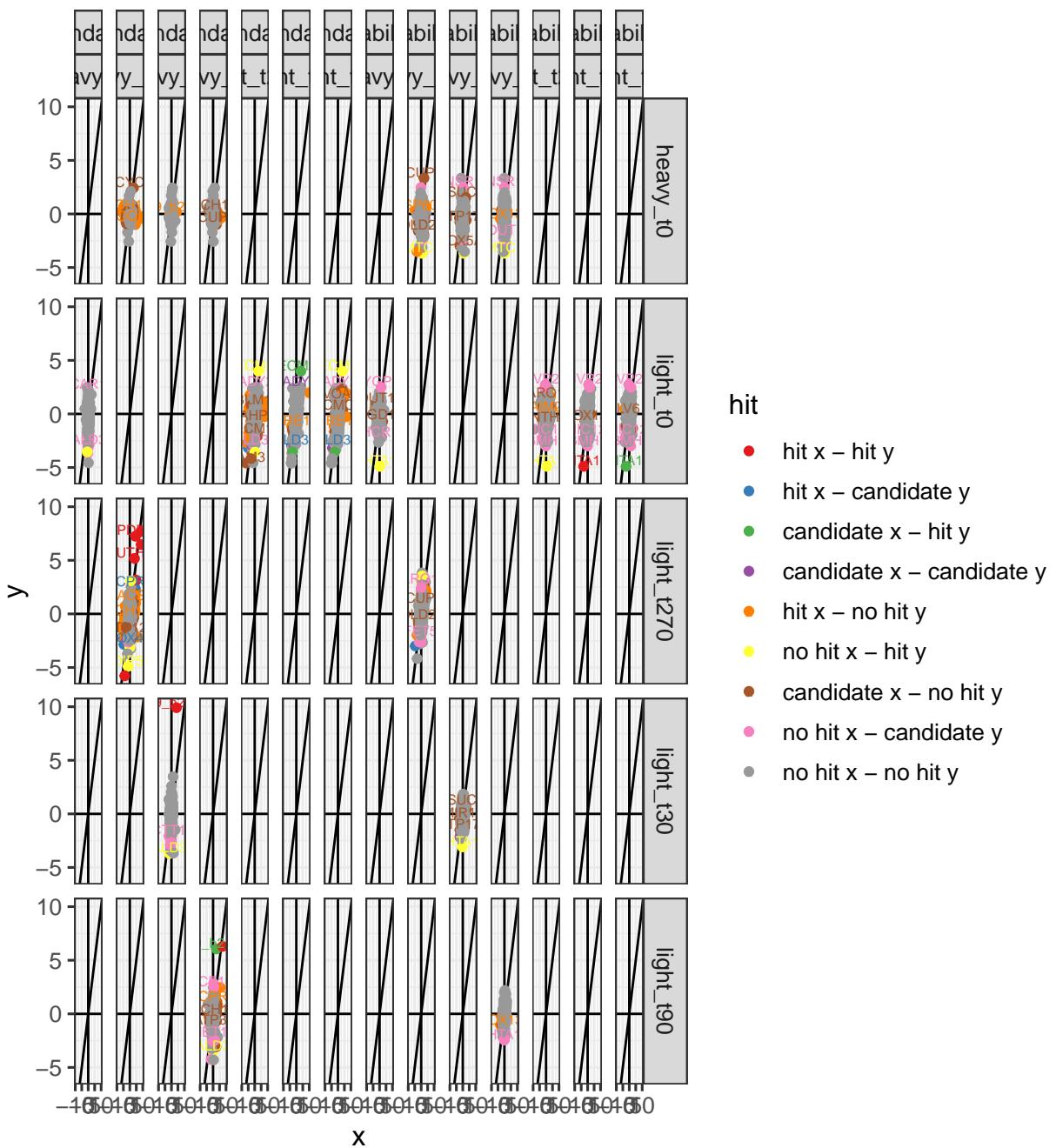
```

```

    hit_y = cor.data[, paste0("hit_", comparisons$y[i])],
    x.name = as.character(comparisons$x[i]),
    y.name = as.character(comparisons$y[i]),
    comparison = paste(comparisons$x[i], "vs", comparisons$y[i]))
mcor.data <- bind_rows(mcor.data, mcor.data_i)
rm(mcor.data_i)
}
cor.data <- mcor.data
rm(mcor.data)
cor.data <- na.omit(cor.data)
cor.data$hit <- paste(cor.data$hit_x, "x -", cor.data$hit_y, "y")
cor.data$hit <- factor(cor.data$hit, ordered = TRUE,
                       levels = c("hit x - hit y",
                                  "hit x - candidate y",
                                  "candidate x - hit y",
                                  "candidate x - candidate y",
                                  "hit x - no hit y",
                                  "no hit x - hit y",
                                  "candidate x - no hit y",
                                  "no hit x - candidate y",
                                  "no hit x - no hit y"))
no.of.comparisons <- length(unique(paste(cor.data$score, cor.data$comparison)))
gr.width <- 6 + ceiling(sqrt(no.of.comparisons)) * 7
gr.height <- floor(sqrt(no.of.comparisons)) * 7

ggplot(data = cor.data, aes(x, y, colour = hit)) +
  geom_abline() +
  geom_vline(xintercept = 0) +
  geom_hline(yintercept = 0) +
  geom_point() +
  facet_grid(y.name ~ score + x.name) +
  geom_text(aes(label = gsub("[|].+", "", gene_name)),
            data = subset(cor.data, hit != "no hit x - no hit y"),
            vjust = 0, nudge_y = 0.1, size = 2, check_overlap = TRUE) +
  customPlot

```



```

ggsave(file.path(dir_save, paste0("Fold_change_correlation_", script.version, ".pdf")), width = gr.width)

pdf(file.path(dir_save, paste0("Fold_change_correlation_single_pages_", script.version, ".pdf")), width =
for(comp in unique(cor.data$score))
{
  print(comp)
  sub <- subset(cor.data, score == comp)
  print(ggplot(data = sub, aes(x, y, colour = hit)) +
    geom_abline() +
    geom_vline(xintercept = 0) +
    geom_hline(yintercept = 0) +

```

```

    geom_point() +
    facet_grid(y.name ~ score + x.name) +
    geom_text(aes(label = gsub("[|].+", "", gene_name)),
              data = subset(sub, hit != "no hit x - no hit y"),
              vjust = 0, nudge_y = 0.1, size = 2, check_overlap = TRUE) +
    customPlot)
}

## [1] "abundance"
## [1] "stability"
dev.off()

## pdf
## 2
rm(sub, comp)

write.csv(cor.data, file = file.path(dir_save, paste0("Fold_change_correlation_data_", script.version, ".csv")))

#Create Excel-Sheet for data browsing

data.sum <- mdata %>%
  dplyr::filter(measurement == "ctrl.ratio") %>%
  mutate(condition = paste(SILAC, condition, sep = "_")) %>%
  group_by(gene_name, condition, Temperature) %>%
  summarise(median_ratio = mean(value, na.rm = TRUE)) %>%
  spread(key = condition, value = median_ratio)
data.sum_i <- mdata %>%
  dplyr::filter(measurement == "ctrl.ratio") %>%
  mutate(condition = paste(SILAC, condition, sep = "_")) %>%
  group_by(gene_name, condition, Temperature) %>%
  mutate(log2.value = log2(value)) %>%
  summarise(log2.range = diff(range(log2.value, na.rm = TRUE))) %>%
  group_by(gene_name, Temperature) %>%
  mutate(condition = paste0("log2.range_", condition)) %>%
  spread(key = condition, value = log2.range)
data.sum <- left_join(data.sum, data.sum_i)
data.sum_i <- mdata %>%
  dplyr::filter(measurement == "ctrl.ratio", !is.na(value)) %>%
  group_by(gene_name, SILAC, Temperature) %>%
  dplyr::select(gene_name, SILAC, Temperature, rep) %>%
  unique() %>%
  na.omit() %>%
  summarise(found.in.replicates = n()) %>%
  ungroup() %>%
  spread(key = SILAC, value = found.in.replicates)
names(data.sum_i)[c(3,4)] <- paste0("found.in.replicates_", names(data.sum_i)[c(3,4)])
data.sum <- right_join(data.sum_i, data.sum)
data.sum$found.in.replicates_heavy[is.na(data.sum$found.in.replicates_heavy)] <- 0
data.sum$found.in.replicates_light[is.na(data.sum$found.in.replicates_light)] <- 0
data.sum_i <- mdata %>%
  dplyr::filter(measurement == "ctrl.ratio", !is.na(value)) %>%
  group_by(gene_name, SILAC) %>%
  dplyr::select(gene_name, SILAC, Temperature) %>%

```

```

unique() %>%
na.omit() %>%
summarise(found.in.Temperatures = n()) %>%
spread(key = SILAC, value = found.in.Temperatures)
names(data.sum_i)[c(2,3)] <- paste0("found.in.Temperatures_", names(data.sum_i)[c(2,3)])
data.sum <- right_join(data.sum_i, data.sum)
feature_data <- fData(raw_dataE)
data.sum <- right_join(feature_data[, c("gene_name", "protein_id", "description", "found.in.ms.runs")], data.sum)
data.sum_i <- feature_data %>%
dplyr::select(gene_name, starts_with("max.qupm_")) %>%
group_by(gene_name) %>%
gather(key = max.qupm.tag, value = max.qupm, -gene_name) %>%
mutate(Temperature = gsub(".+_", "", max.qupm.tag)) %>%
mutate(Temperature = as.numeric(as.character(Temperature))) %>%
dplyr::select(gene_name, Temperature, max.qupm)
data.sum <- left_join(data.sum, data.sum_i)
data.sum_i <- feature_data %>%
dplyr::select(gene_name, starts_with("average.top3_")) %>%
group_by(gene_name) %>%
gather(key = average.top3.tag, value = average.top3, -gene_name) %>%
mutate(Temperature = gsub(".+_", "", average.top3.tag)) %>%
mutate(Temperature = as.numeric(as.character(Temperature))) %>%
dplyr::select(gene_name, Temperature, average.top3)
data.sum <- left_join(data.sum, data.sum_i)
data.sum$path <- file.path(".", "plots_combined",
                           paste0(gsub("\\\\|", "-", data.sum$gene_name), ".pdf"))
write.csv(data.sum, file.path(dir_save, paste0("Final_results_", script.version, ".csv")),
          row.names = FALSE)

hit_classifications <- limma_results[,c("gene_name", "comparison2", "score", "hit_annotation")]
hit_classifications <- hit_classifications %>%
group_by(gene_name) %>%
mutate(key = paste0(score, "_", comparison2)) %>%
dplyr::select(gene_name, key, hit_annotation) %>%
spread(key = key, value = hit_annotation)
data.sum <- left_join(data.sum, hit_classifications)
write.csv(data.sum, file.path(dir_save, paste0("Final_results_classified_", script.version, ".csv")),
          row.names = FALSE)

limma_spread <- limma_results[, c("gene_name", "comparison2", "score", "score.value", "fdr", "hit_annotation")]
mutate(comparison = comparison2) %>%
dplyr::select(-comparison2) %>%
group_by(gene_name, comparison, score) %>%
gather(key = key, value = value, -gene_name, -comparison, -score) %>%
mutate(key = gsub(".value", "", key)) %>%
mutate(new.key = paste0(key, "_", comparison)) %>%
group_by(gene_name, score) %>%
dplyr::select(gene_name, score, new.key, value) %>%
spread(key = new.key, value = value)
limma_spread <- limma_spread[, c("gene_name", "score",
                                grep("^score_", names(limma_spread), value = TRUE),
                                grep("^fdr_", names(limma_spread), value = TRUE)),

```

```

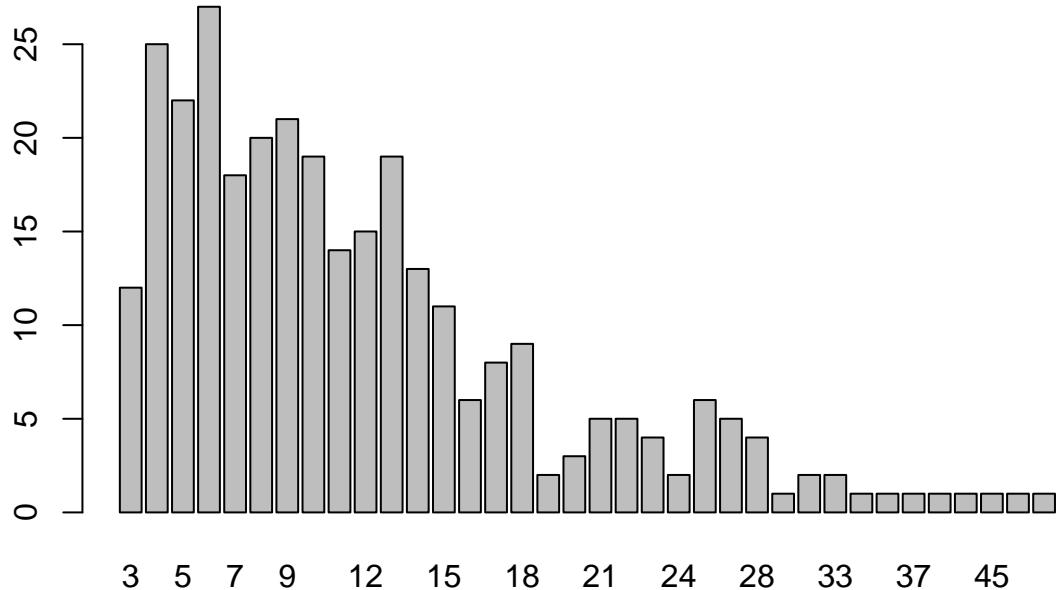
grep("hit_annotation_", names(limma_spread), value = TRUE)])
limma_spread <- right_join(
  unique(data.sum[, c("gene_name", "protein_id", "description", "found.in.ms.runs", "path")]),
  limma_spread)

write.csv(limma_spread, file.path(dir_save, paste0("Limma_results_classified_", script.version, ".csv")),
          row.names = FALSE)

#Clustering of data
##Preparation of cluster data
# stability hits
dir.hits <- "data_analysis_results_V1/stab_hits_categories/"

hits <- unique(subset(limma_results, hit_annotation %in% c("hit", "candidate"))$gene_name)
# hits <- unique(subset(limma_results, hit_annotation %in% c("hit", "candidate") & score == "stability"))
barplot(table(cdata$max.qupm[cdata$gene_name %in% hits]))

```



```

# hits <- hits[hits %in% cdata$gene_name[cdata$max.qupm >= 10]]

# old.data <- read.csv("old_full_data.csv")
# old.data <- subset(old.data, gene_name %in% mdata$gene_name)
# old.data$median.value <- log2(old.data$median.value)
# hits <- unique(append(hits, as.character(unique(old.data$gene_name)))))

m_clust.data <- limma_results %>%
  mutate(condition = comparison2) %>%

```

```

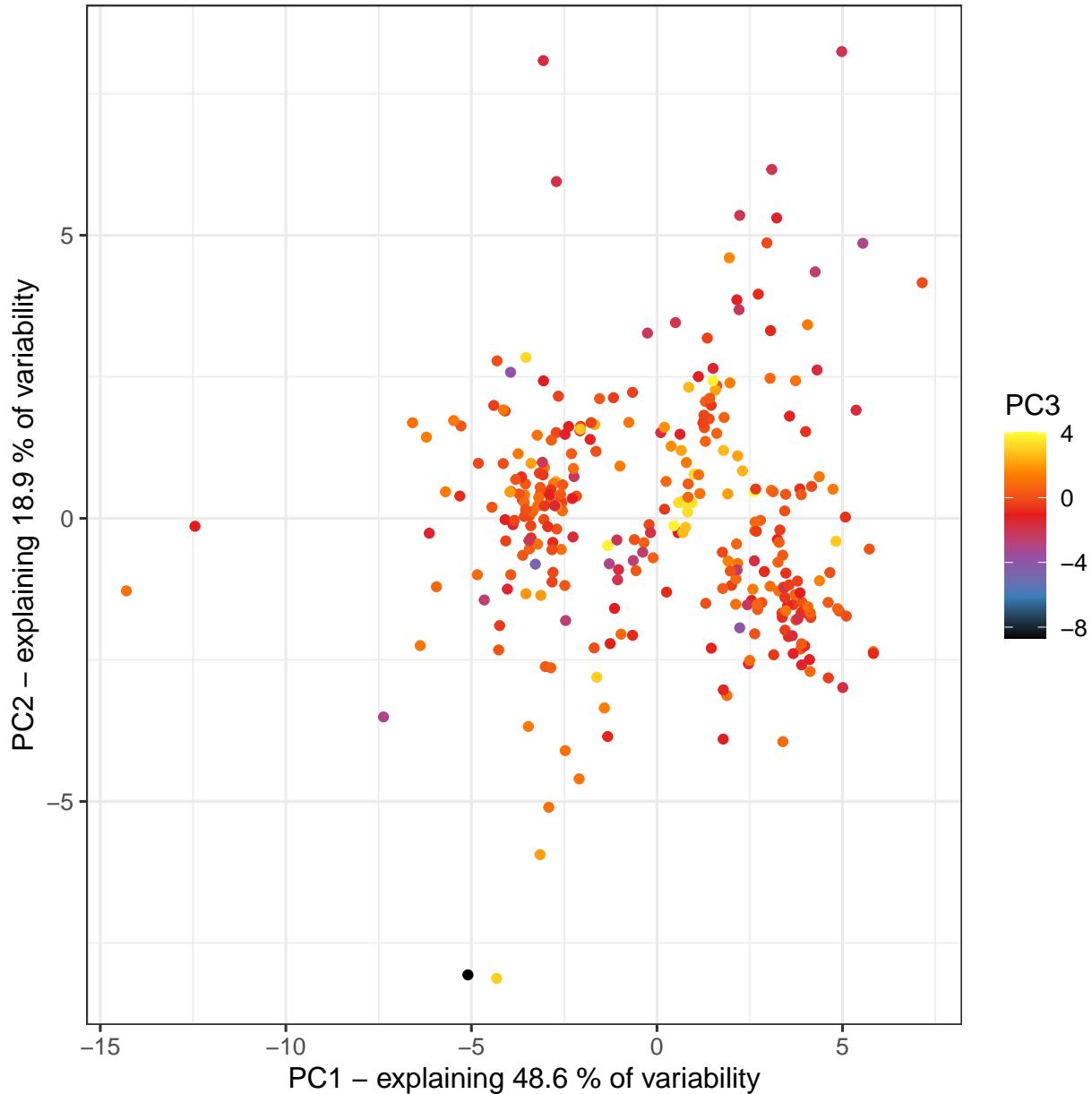
dplyr::select(-comparison2) %>%
group_by(gene_name, score, condition) %>%
dplyr::filter(gene_name %in% hits) %>%
summarise(median.value = median(score.value, na.rm = TRUE))
# m_clust.data <- bind_rows(m_clust.data, old.data)
clust.data_m <- m_clust.data %>%
mutate(key = paste(score, condition, sep = "_")) %>%
ungroup() %>%
dplyr::select(gene_name, key, median.value) %>%
group_by(gene_name) %>%
spread(key = key, value = median.value) %>%
as.data.frame()
rownames(clust.data_m) <- clust.data_m$gene_name
m_clust.data <- m_clust.data %>%
dplyr::filter(gene_name %in% clust.data_m$gene_name)
clust.data_m$gene_name <- NULL
clust.data_m <- clust.data_m %>%
as.matrix()
clust.data_m[is.na(clust.data_m)] <- 0
m_clust.data$SILAC <- gsub("_.+", "", m_clust.data$condition)
m_clust.data$condition <- gsub(".+_", "", m_clust.data$condition)
m_clust.data$condition <- factor(m_clust.data$condition, ordered = TRUE, levels = c("t0", "t30", "t90"))

##PCA of clustering data
clust.data_PCA <- prcomp(clust.data_m, scale = FALSE)
perc_var <-
round(100 * clust.data_PCA$sdev ^ 2 /
      sum(clust.data_PCA$sdev ^ 2), 1)
PCA_clust.data <-
data.frame(PC1 = clust.data_PCA$x[, 1],
           PC2 = clust.data_PCA$x[, 2],
           PC3 = clust.data_PCA$x[, 3])
ggplot(data = PCA_clust.data, aes(PC1, PC2, colour = PC3)) +
geom_point() +
theme_bw(base_size = 12) +
scale_colour_gradientn(colours = c("black", "#377eb8", "#984ea3",
                                  "#e41a1c", "#ff7f00", "#ffff33")) +
ggtitle("PCA clustering data",
        subtitle = paste("PC3 - explaining", perc_var[3], "% of variability")) +
xlab(paste("PC1 - explaining", perc_var[1], "% of variability")) +
ylab(paste("PC2 - explaining", perc_var[2], "% of variability"))

```

PCA clustering data

PC3 – explaining 9.7 % of variability



```

ggsave(file.path(dir_save, paste0("PCA_clustering_data_", script.version, ".pdf")),
       width = 7, height = 5)
rm(clust.data_PCA, perc_var, PCA_clust.data)

##Set number of clusters
cluster.number <- 10

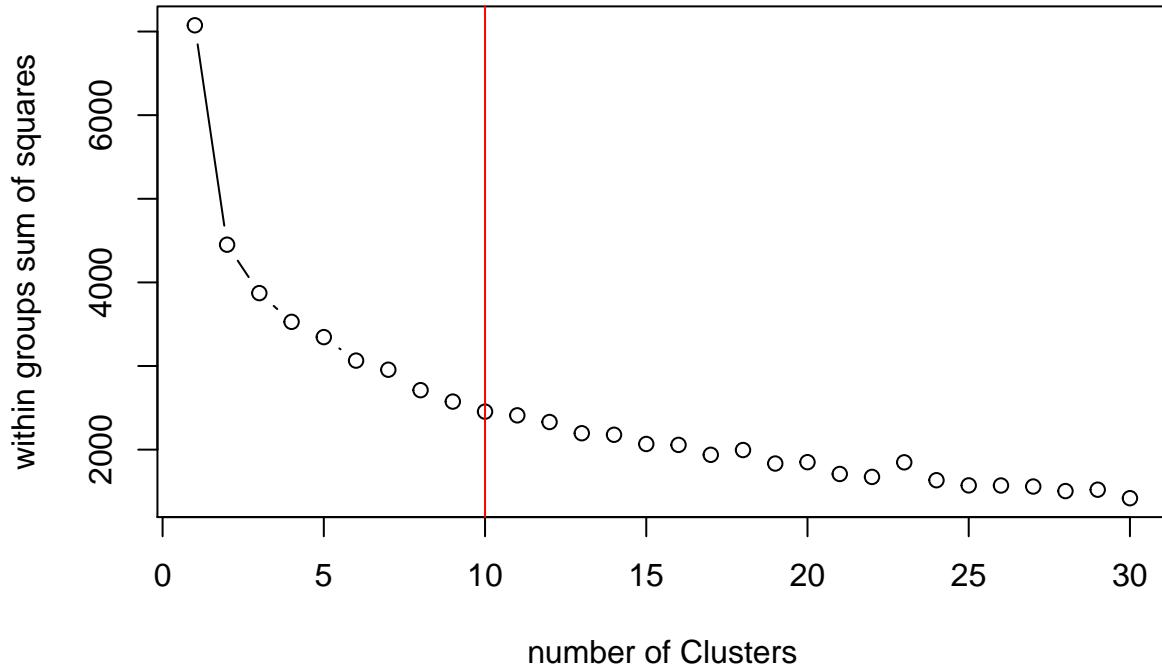
##K-means clustering
# plot within groups sum of squares for different cluster numbers
wss <- (nrow(clust.data_m) - 1) * sum(apply(clust.data_m, 2, var))
for (i in 2:(cluster.number*3)) wss[i] <- sum(kmeans(clust.data_m,

```

```

centers = i)$withinss)
plot(1:(cluster.number*3), wss, type = "b", xlab = "number of Clusters",
      ylab = "within groups sum of squares")
abline(v = cluster.number, col = "red")

```



```

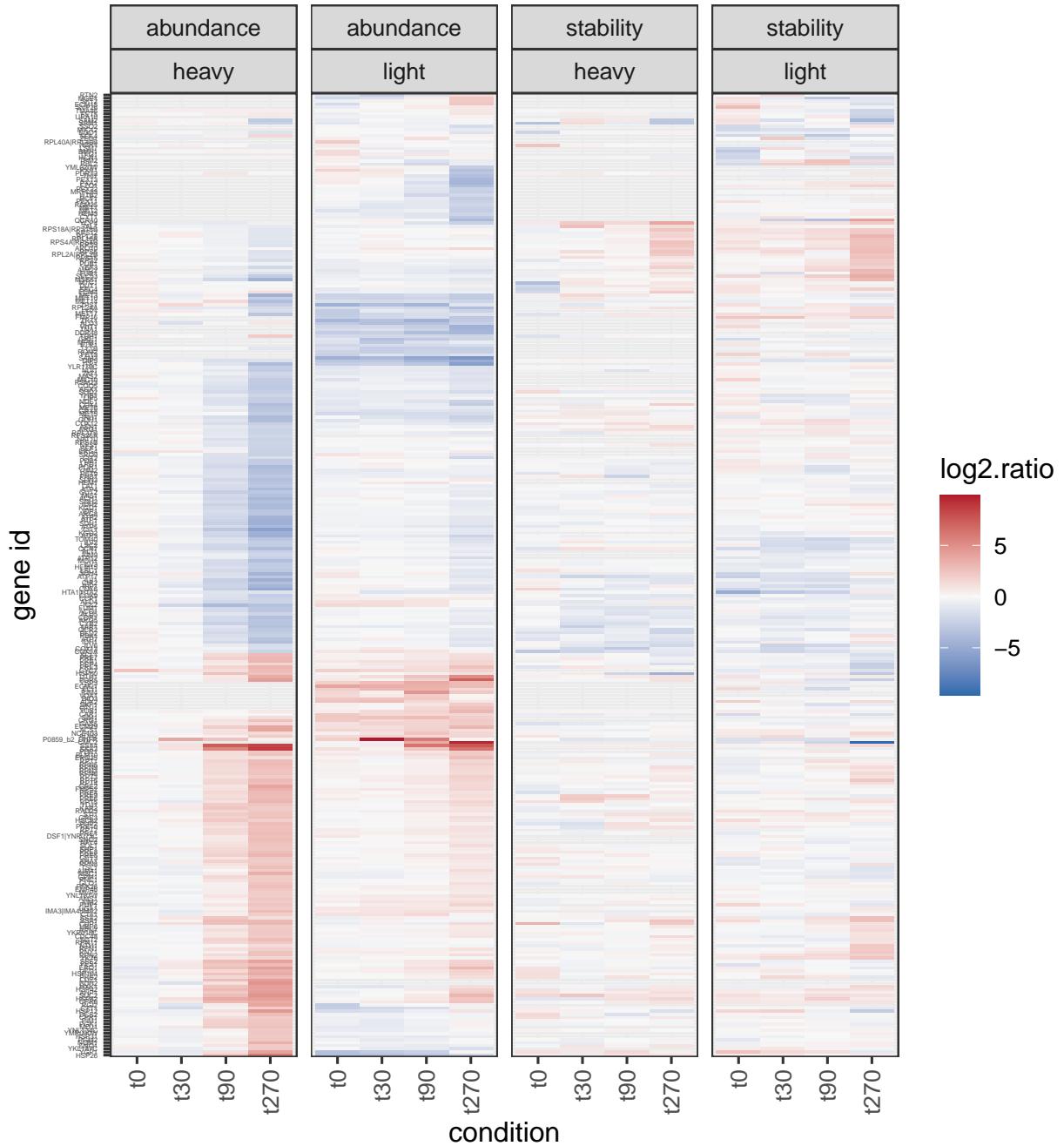
kmeans.fit <- kmeans(clust.data_m, cluster.number) # 5 cluster solution
cluster.groups <- data.frame(gene_name = names(kmeans.fit$cluster),
                                kmeans.cluster.group = kmeans.fit$cluster)
rm(wss, kmeans.fit)

##Hierarchical clustering
d <- dist(clust.data_m, method = "euclidean")
hclust.fit <- hclust(d, method = "ward.D2")
cluster.groups_hclust <-
  data.frame(gene_name = names(cutree(hclust.fit, k = cluster.number)),
             hclust.cluster.group = cutree(hclust.fit, k = cluster.number))
cluster.groups <- left_join(cluster.groups, cluster.groups_hclust)
rm(cluster.groups_hclust)
m_clust.data <- left_join(m_clust.data, cluster.groups)
m_clust.data$gene_name <- factor(m_clust.data$gene_name, ordered = TRUE,
                                    levels = hclust.fit$labels[hclust.fit$order])
rm(d, hclust.fit)

##Plot clustering results

```

```
gr.width <- 2 + ncol(clust.data_m) * 0.1 + max(nchar(rownames(clust.data_m))) * 0.1 + 4
gr.height <- 3 + nrow(clust.data_m) * 0.03
ggplot(data = m_clust.data, aes(condition, gene_name)) +
  geom_tile(aes(fill = median.value)) +
  scale_fill_gradient2(low = "#2166ac", high = "#b2182b", midpoint = 0,
                       mid = "#f7f7f7", name = "log2.ratio") +
  facet_grid(~ score + SILAC, scale = "free_x") +
  theme_bw(base_size = 12) +
  theme(axis.text.x = element_text(angle = 90, hjust = 1, vjust = 0.5),
        axis.text.y = element_text(size = 3)) +
  ylab("gene id")
```

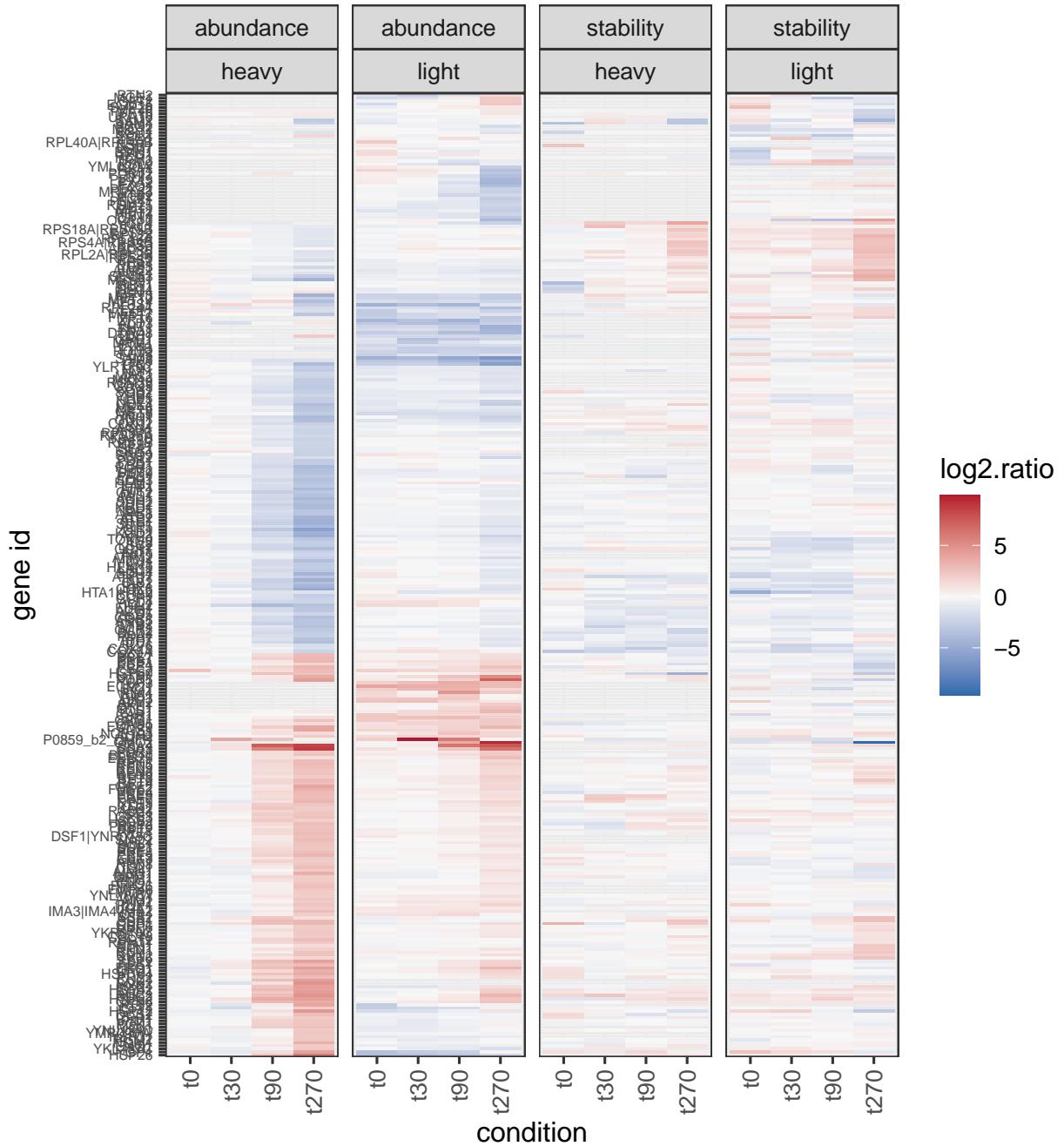


```

ggsave(file.path(dir_save, paste0("Heatmap_hits_", script.version, ".pdf")), width = gr.width, height = gr.height)

ggplot(data = m_clust.data, aes(condition, gene_name)) +
  geom_tile(aes(fill = median.value)) +
  scale_fill_gradient2(low = "#2166ac", high = "#b2182b", midpoint = 0,
                       mid = "#f7f7f7", name = "log2.ratio") +
  facet_grid(~ score + SILAC, scale = "free_x") +
  theme_bw(base_size = 12) +
  theme(axis.text.x = element_text(angle = 90, hjust = 1, vjust = 0.5),
        axis.text.y = element_text(size = 6)) +
  ylab("gene id")

```



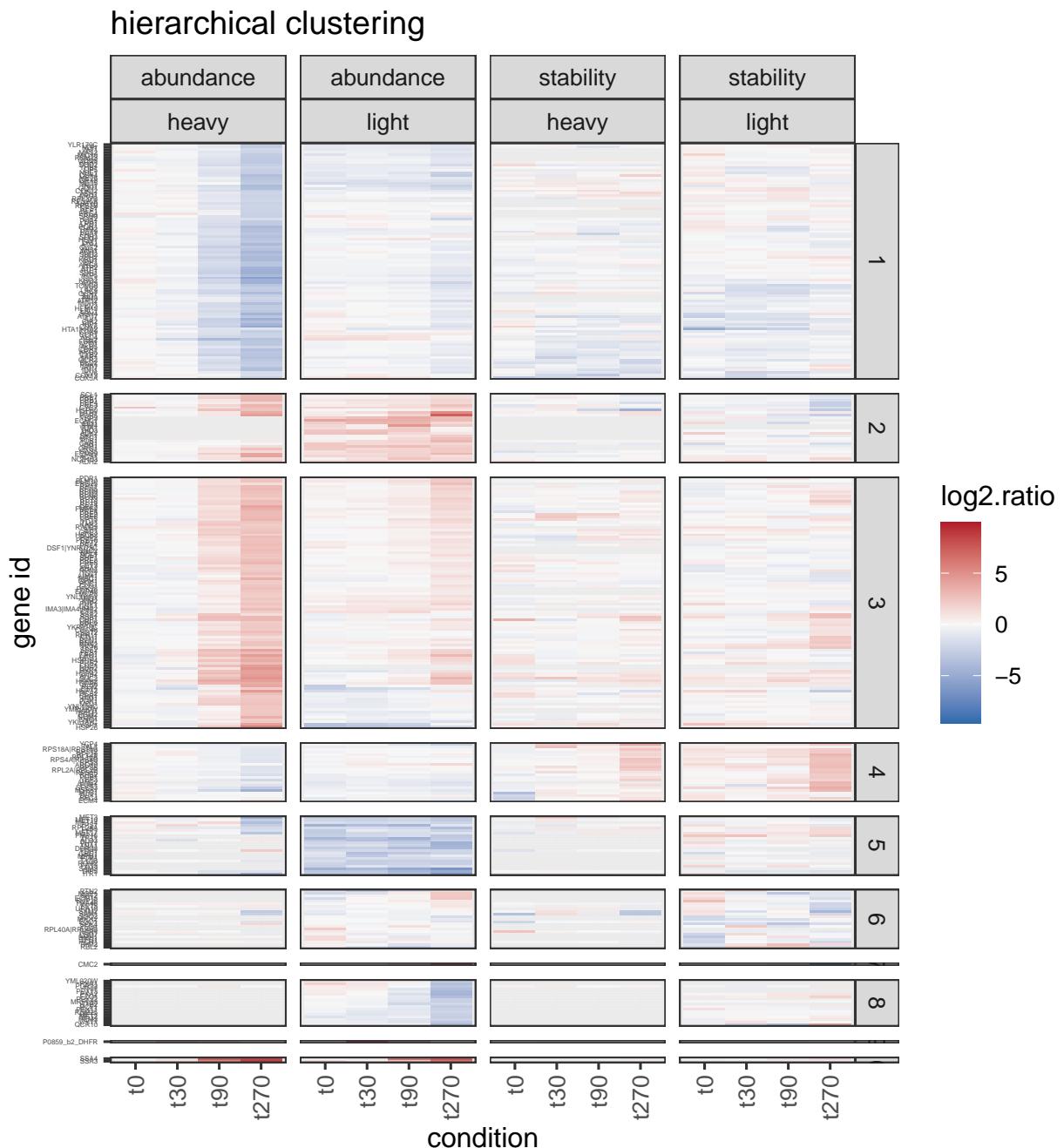
```

ggsave(file.path(dir_save, paste0("Stability_hits_Heatmap_hits_", script.version, ".pdf")), width = gr.

#Clustering heatmaps
gr.height <- 2.5 + nrow(clust.data_m) * 0.04 + cluster.number * 0.1
ggplot(data = m_clust.data, aes(condition, gene_name)) +
  geom_tile(aes(fill = median.value)) +
  scale_fill_gradient2(low = "#2166ac", high = "#b2182b", midpoint = 0,
                       mid = "#f7f7f7", name = "log2.ratio") +
  facet_grid(hclust.cluster.group ~ score + SILAC, scale = "free", space = "free") +
  theme_bw(base_size = 12) +
  theme(axis.text.x = element_text(angle = 90, hjust = 1, vjust = 0.5),
        axis.ticks.x = element_text(angle = 90, hjust = 1))

```

```
    axis.text.y = element_text(size = 3)) +  
ylab("gene id") +  
ggtitle("hierarchical clustering")
```



```

ggsave(file.path(dir_save, paste0("Clustering_heatmap_hits_hclust_", cluster.number, "_cluster_", script
    width = gr.width, height = gr.height))

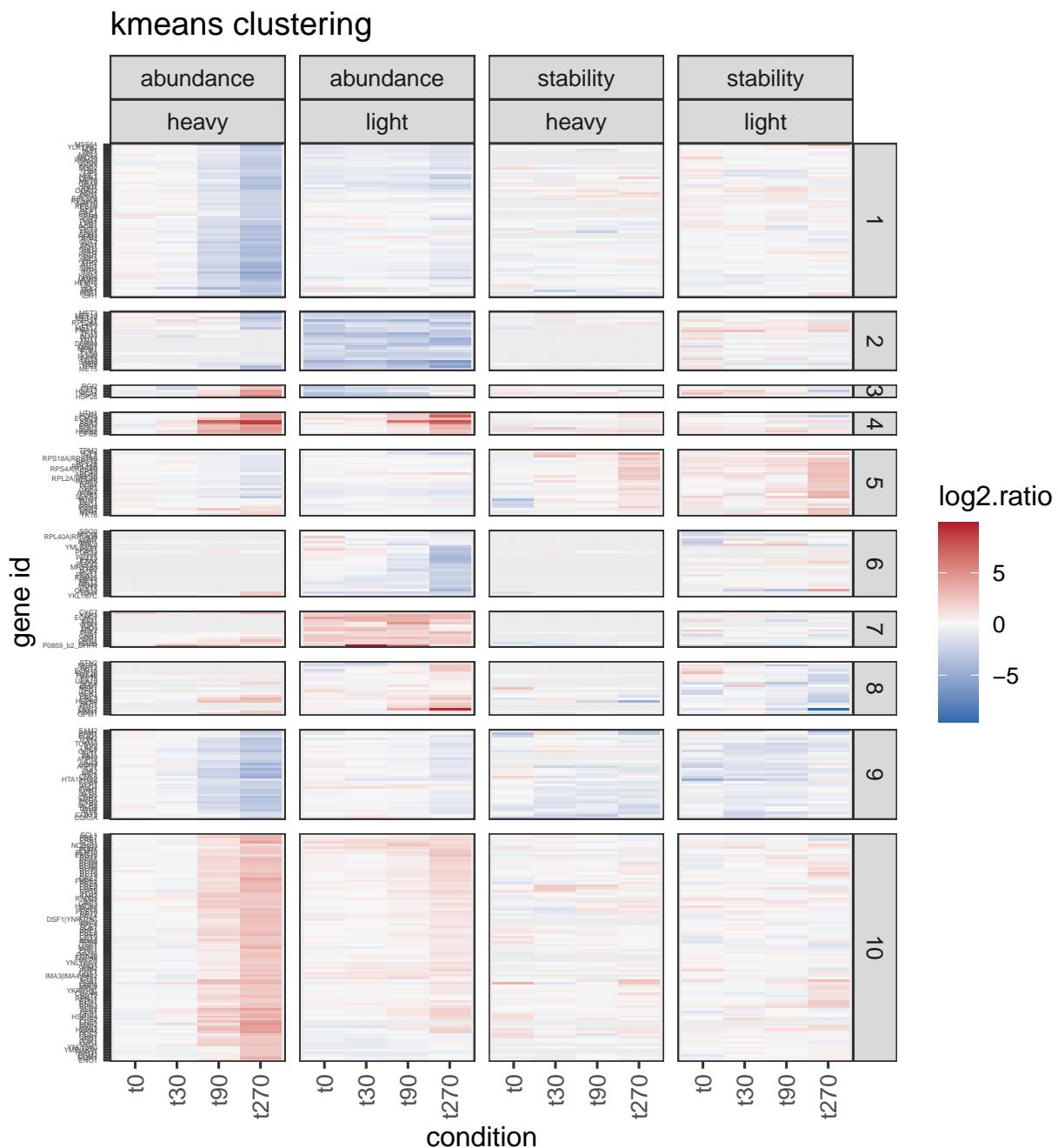
ggplot(data = m_clust.data, aes(condition, gene_name)) +
  geom_tile(aes(fill = median.value)) +
  scale_fill_gradient2(low = "#2166ac", high = "#b2182b", midpoint = 0,
                      mid = "#ff7f7f7", name = "log2.ratio") +

```

```

facet_grid(kmeans.cluster.group ~ score + SILAC, scale = "free", space = "free") +
theme_bw(base_size = 12) +
theme(axis.text.x = element_text(angle = 90, hjust = 1, vjust = 0.5),
      axis.text.y = element_text(size = 3)) +
ylab("gene id") +
ggtitle("kmeans clustering")

```



```

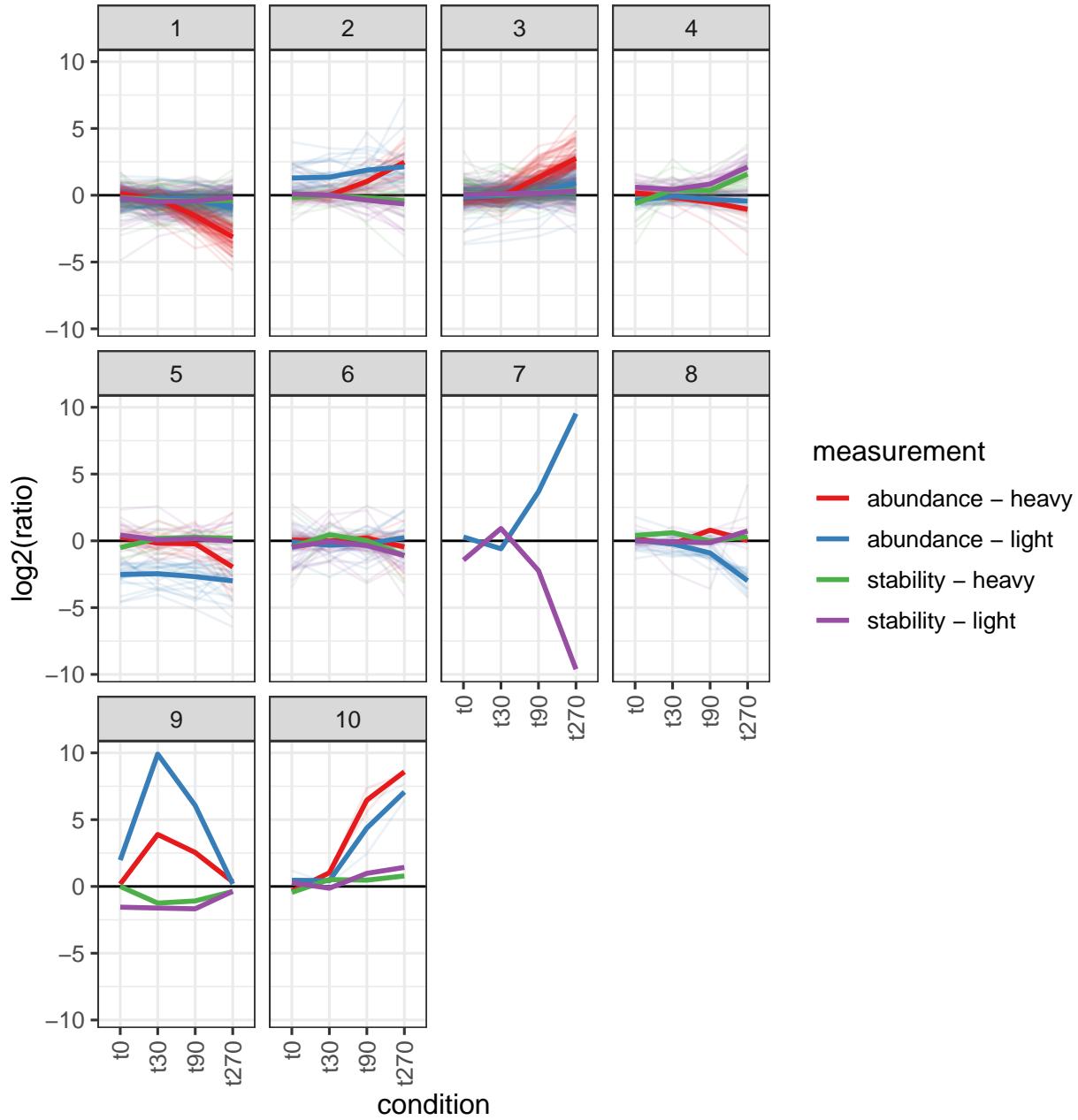
ggsave(file.path(dir_save, paste0("Clustering_heatmap_hits_kmeans_", cluster.number, "_cluster_",
                                 width = gr.width, height = gr.height))

#Line plots

```

```
gr.width <- 4 + ceiling(sqrt(cluster.number)) * 5
gr.height <- 3 + floor(sqrt(cluster.number)) * 5
m_clust.data$measurement <- paste(m_clust.data$score, m_clust.data$SILAC, sep = " - ")
ggplot(data = m_clust.data, aes(condition, median.value, colour = measurement, fill = measurement)) +
  geom_hline(yintercept = 0) +
  geom_line(aes(group = paste(gene_name, score, hclust.cluster.group, SILAC)), alpha = 0.1) +
  stat_summary(fun.data = "mean_se", geom = "smooth", aes(group = paste(hclust.cluster.group, score, SILAC))) +
  facet_wrap(~ hclust.cluster.group) +
  customPlot +
  theme(axis.text.x = element_text(angle = 90, hjust = 1, vjust = 0.5)) +
  ggtitle("hierarchical clustering") +
  ylab("log2(ratio)")
```

hierarchical clustering



```

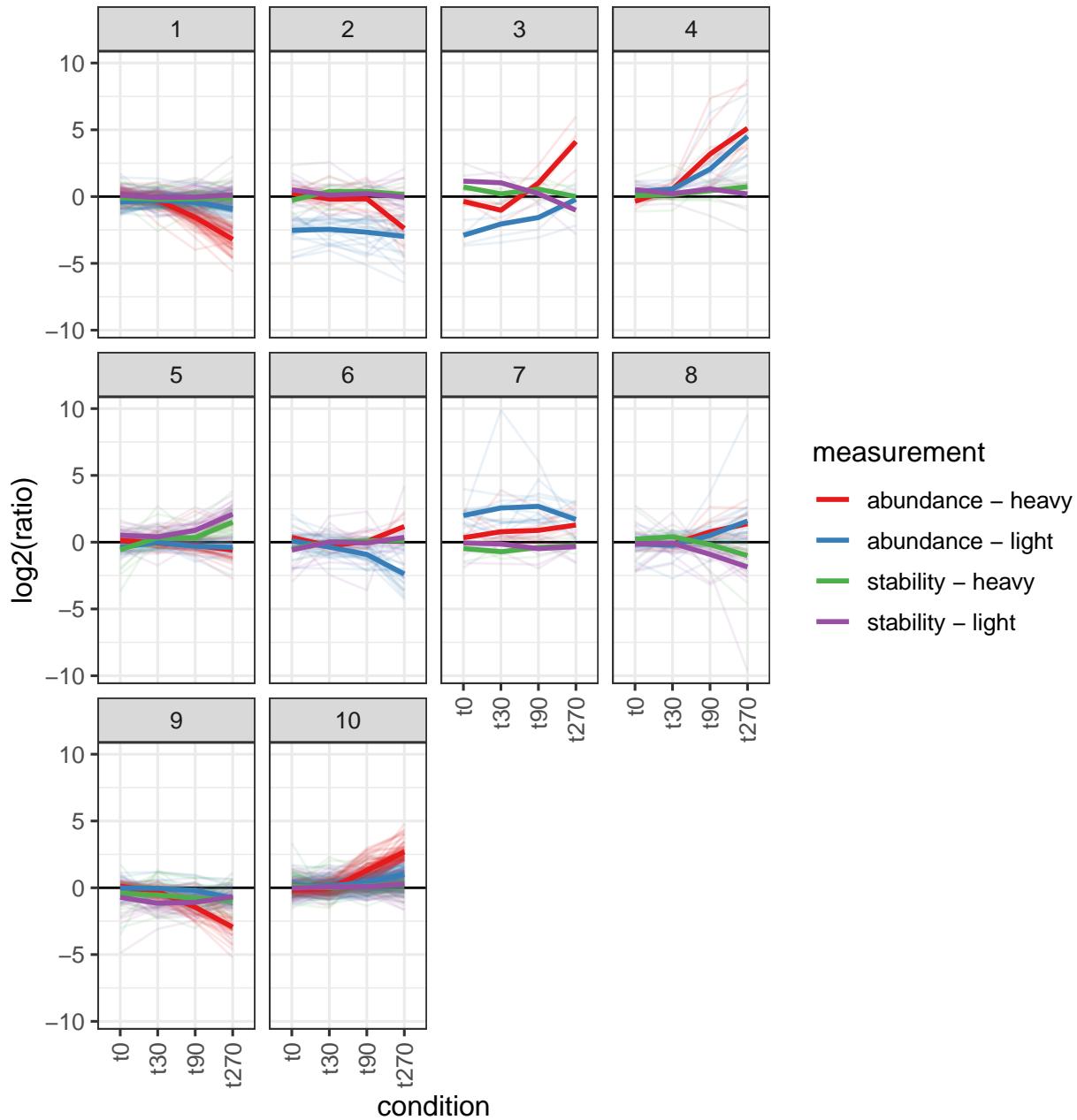
ggsave(file.path(dir_save, paste0("Clustering_line_plot_hclust_", cluster.number, "_cluster_", script.v
                                 width = gr.width, height = gr.height))

ggplot(data = m_clust.data, aes(condition, median.value, colour = measurement, fill = measurement)) +
  geom_hline(yintercept = 0) +
  geom_line(aes(group = paste(gene_name, score, kmeans.cluster.group, SILAC)), alpha = 0.1) +
  stat_summary(fun.data = "mean_se", geom = "smooth", aes(group = paste(kmeans.cluster.group, score, SI
  facet_wrap(~ kmeans.cluster.group) +
  customPlot +
  theme(axis.text.x = element_text(angle = 90, hjust = 1, vjust = 0.5)) +
  ggtitle("kmeans clustering"))

```

```
ylab("log2(ratio)")
```

kmeans clustering



```
ggsave(file.path(dir_save, paste0("Clustering_line_plot_kmeans_", cluster.number, "_cluster_", script.version, ".pdf")),
       width = gr.width, height = gr.height)
write.csv(cluster.groups, file.path(dir_save, paste0("Cluster_results_", cluster.number, "cluster_", script.version, ".csv")))
rm(gr.width, gr.height)
```

```
#Save workspace
```

```
write.csv(mdata, file = file.path(dir_save, paste0("Tidy_data_all_", script.version, ".csv")))
write.csv(limma.cp.data, file = file.path(dir_save, paste0("Tidy_score_data_", script.version, ".csv")))
```

```

save.image(file.path(dir_save, paste0("Workspace_", script.version, ".RData")))
# load(file.path(dir_save, paste0("Workspace_", script.version, ".RData")))

#Save individual plots for each protein

library(gridExtra)
dir_save.plots = file.path(dir_save, "plots_combined")
if (!dir.exists(file.path(getwd(), dir_save.plots)))
{
  dir.create(file.path(getwd(), dir_save.plots))
}
h=15
w=15
cols <- c("#e41a1c", "#377eb8", "#4daf4a", "#984ea3", "#ff7f00", "#ffff33", "#a65628",
          "#f781bf", "#999999", "#1b9e77", "#d95f02", "#7570b3", "#e7298a", "#66a61e",
          "#e6ab02", "#a6761d")

limits.limma = quantile(limma.cp.data$score.value, probs = c(0.001, 0.999), na.rm = TRUE)
for (gene in sort(as.character(unique(subset(cdata)$gene_name))))
{
  gene.name <- gsub("\\|", "-", gene)
  f.name <- file.path(dir_save.plots, paste0(gene.name, ".pdf"))
  if (!file.exists(f.name))
  {
    pdf(width = w, height = h, file = f.name)
    sub <- subset(mdata, gene_name == gene)
    sub.cdata <- subset(cdata, gene_name == gene)
    average.top3 = sub.cdata$average.top3
    max.qupm <- sub.cdata$max.qupm
    ptitle <- paste(gene, "-", "average.top3:", average.top3, "-", "max.qupm:", max.qupm)
    subtitle <- paste(sub.cdata$protein_id, "-", sub.cdata$description)

    try(p1 <- ggplot(data = subset(sub, measurement == "norm_batchcl_raw_signal_sum"), aes(factor(Tempe
      geom_tile(aes(fill=log2(value))) +
      scale_fill_gradientn(colours = c("#377eb8",
                                         "#984ea3",
                                         "#e41a1c",
                                         "#ff7f00",
                                         "#ffff33"), name = "log2(norm signal_sum)") +
      xlab("Temperature") +
      facet_grid(SILAC ~ rep, scale = "free_x", space = "free") +
      theme_bw(base_size = 12) +
      theme(legend.position="bottom") +
      theme(axis.text.x = element_text(angle = 90, hjust = 1, vjust = 0.5)))
    try(p2 <- ggplot(data = subset(sub, measurement == "norm_batchcl_raw_signal_sum"), aes(Temperature,
      stat_summary(fun.data = "mean_se", geom = "smooth") +
      theme_bw(base_size = 12) +
      theme(axis.text.x = element_text(angle = 90, hjust = 1)) +
      scale_fill_manual(values = cols) +
      scale_colour_manual(values = cols) +
      facet_grid(time ~ SILAC) +
      ylab("log2(norm signal_sum)") +
      stat_summary(fun.data = "mean_se", geom = "line", size = 2))
    try(p3 <- ggplot(data = subset(sub, measurement == "ctrl.ratio"),

```

```

        aes(factor(Temperature), condition)) +
    geom_tile(aes(fill=log2(value))) +
    scale_fill_gradient2(low = "#2166ac", high = "#b2182b", midpoint = 0, mid = "#f7f7f7", name =
    xlab("Temperature") +ylab("condition") +
    facet_grid(SILAC ~ rep, scale = "free_x", space = "free") +
    theme_bw(base_size = 12) +
    theme(axis.text.x = element_text(angle = 90, hjust = 1,vjust = 0.5), legend.position="bottom")
try(p4 <- ggplot(data = subset(sub, measurement == "temp.ratio"), aes(Temperature, log2(value),fill=
    stat_summary(fun.data = "mean_se",geom = "smooth") +
    theme_bw(base_size = 12) +
    theme(axis.text.x = element_text(angle = 90, hjust = 1)) +
    ylab("log2(temp.ratio)") +
    scale_fill_manual(values = cols) +
    facet_grid(time ~ SILAC) +
    scale_colour_manual(values = cols) +
    stat_summary(fun.data = "mean_se", geom = "line", size = 2))

if (gene%in%limma.cp.data$gene_name)
{
  sub <- subset(limma.cp.data,gene_name ==gene)
  sub$condition <- factor(sub$condition, ordered = TRUE, levels = levels(mdata$condition))
  new.limits <- limits.limma
  if (min(sub$score.value,na.rm = TRUE)<new.limits[1])
  {
    new.limits[1] <- min(sub$score.value,na.rm = TRUE)
  }
  if (max(sub$score.value,na.rm = TRUE)>new.limits[2])
  {
    new.limits[2] <- max(sub$score.value,na.rm = TRUE)
  }
# try(p5 <- ggplot(data = subset(sub, grepl("_vs_control", condition) & !grepl("^control_", condition)),
#   geom_tile() +
#   coord_polar() +
#   scale_fill_gradient2(low = "#2166ac",high = "#b2182b",
#   midpoint = 0,mid = "#f7f7f7",
#   limits = new.limits) +
#   theme_bw(base_size = 12) +
#   xlab("replicate") +
#   geom_text(aes(label = fdr.star)))
try(p6 <- ggplot(data = subset(sub, !grepl("^control", condition)), aes(x=condition,score.value, y=score),
    geom_boxplot(fill = "darkgrey") +
    geom_point(aes(shape = rep)) +
    theme_bw(base_size = 12) +
    scale_fill_brewer(palette = "Set1") +
    coord_cartesian(ylim = new.limits) +
    geom_text(aes(label = fdr.star,y = new.limits[2]),show.legend = FALSE) +
    facet_grid(. ~ score + SILAC) +
    geom_hline(yintercept = 0) +
    theme(axis.text.x = element_text(angle = 90, hjust = 1,vjust = 0.5)))
# try(suppressWarnings( print(grid.arrange(p1, p2, p3, p4, p6, ncol = 2, top = pttitle, bottom = subtitle,
#   try(suppressWarnings( print(grid.arrange(p1, p2, p3, p4, p6, layout_matrix = rbind(c(1,2), c(3,4),
} else{
  try(suppressWarnings( print(grid.arrange(p1, p2, p3, p4, ncol = 2, top = pttitle, bottom = subtitle,

```

```
    }
    dev.off()
    rm(sub,min.p.value,new.limits,max.qupm,average.top3)
}
}
```