LAB 1a — EXPLORING NCBI

[Software needed: web access]

The National Center for Biotechnology Information (NCBI) maintained by the US National Library of Medicine and National Institutes of Health is one of the world's most important resources and repositories for biological data. This fantastic online resource provides an extensive network of databases cataloging an ever-growing wealth of genetic, medical, and biochemical information from all walks and crawls of life. Entire genomes, from viruses to humans, are compiled, organized, and cross-referenced within these networks, such that surfing the genome can be almost as easy as surfing the web.

But you have to know a) what you're looking *for*, and b) what you're looking *at* to get anything out of these databases. This is what this first lab is going to help you do. Note that Google and other search engines typically do not index database-driven websites, which is why it cannot be used for searching for information that is stored at NCBI (nor does it handle sequence searching well, especially in the case of protein sequences).

The primary portal for accessing data at NCBI is called *Search NCBI*. But first, let's start by visiting NCBI's website and examining the interface, which undergoes constant change.

1. Open your Web browser and go to NCBI's homepage: www.ncbi.nlm.nih.gov. This page provides links to all of NCBI databases and resources. It's worth exploring here just to get a better idea of the scope of NCBI. If you click **About the NCBI** you will be taken to a page summarizing some of these resources. You can also check out the *NCBI Handbook* (https://www.ncbi.nlm.nih.gov/books/NBK143764/) for more information.

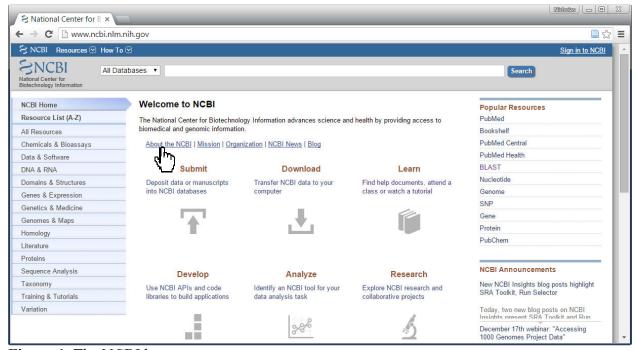


Figure 1. The NCBI homepage.

2. Now let's move to the *Search NCBI* (formerly known as *GQuery* or *Entrez*) portal – select **All Databases** from the navigation bar at the top of the NCBI start page and click "Search" beside the empty field. First, scan down the assortment of databases queried through this portal. You will notice there is everything from the biomedical literature at PubMed to nucleotide databases, taxonomy databases, protein structure databases, and expression profile databases. Let's see what happens when you do an unguided search on the site. In the "Search NCBI" box, type in *bacterial*. The output is a summary page of the number of hits in each section. A search of *bacterial* gives millions of hits – not very helpful. We need specifics.

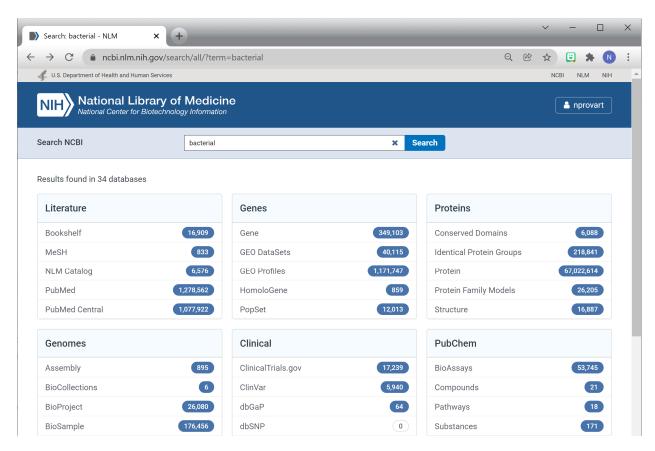


Figure 2. The *Search NCBI* portal page with *bacterial* used as a search word.

3. Usually when searching these databases, you have either a region of DNA or a protein (or protein function) of interest. For this lab you'll be using a gene from *Arabidopsis thaliana*, a small flowering plant that is like the fruit fly of the plant world as it has a comparatively rapid life cycle and requires little space to grow. The protein product of this gene is recorded under accession number NP_001318308, and it is an E3 ligase, involved in ubiquitination of proteins, which is a signal for their degradation.

- 4. Go back to the *Search NCBI* portal page and try a more focused search. Use the search terms found associated with the gene sequence we'll be using with the GenBank Field Qualifiers shown below (a full list of qualifiers is presented in Appendix 1). Try the four different searches presented below and look at the number records, specifically "Protein" records, found:
 - gene keywords
 - e.g. ubiquitin-protein ligase
 - gene keyword AND organism
 - e.g. ubiquitin-protein ligase AND Arabidopsis thaliana
 - gene keyword [PROT] AND organism [ORGN]
 - e.g. ubiquitin-protein ligase [PROT] AND Arabidopsis thaliana [ORGN]
 - accession or GI number
 - e.g. NP_001318308

That narrowed things down significantly!

Lab Quiz
Question 1
*Answer lab quiz
questions while doing lab!

Note that using parentheses can be very helpful in making sure you get exactly what you want. For example:

SMC AND (yeast [ORGN] OR Arabidopsis [ORGN])

is a very different search than

SMC AND yeast [ORGN] OR Arabidopsis [ORGN]

Also, using quotation marks can also dramatically affect your search (i.e.: 16s rRNA vs. "16s rRNA").

Finally, always capitalize the Boolean operators such as AND / OR / NOT. Ultimately, the most specific search items you can use are accession numbers.

Box 1. Accession Numbers and Version Numbers (and GI Numbers...)

An **Accession number** is a unique identifier for a particular sequence record. An accession number is assigned to a specific record and stays with that record forever. In other words, Accession numbers track a particular record and do not change even if the information in the record is changed at the author's request (e.g., if a better annotation or more complete sequence is provided). Accession numbers are usually a combination of a letter(s) and numbers, such as a single letter followed by five digits (e.g., U12345) or two letters followed by six digits (e.g., AF123456).

Version numbers follow the Accession number and indicate the revision history of that entry starting with 1 and increasing with each revision. The standard format is *Accession.Version*. A **GI number** (GenInfo Identifier – sometimes written in lower case, "gi") was simply a series

of digits that was, until recently, assigned consecutively to each sequence record processed by NCBI. The GI system of identifiers ran in parallel to the Accession. Version system; therefore, if the DNA or protein sequence changed in any way, it would receive a new GI number.

Example: When a new entry was submitted to GenBank it was assigned an accession number (say AF000001). Since this is the first version the Accession would be appended with '.1', so it would look like AF000001.1. At the same time was given a GI number (say GI:1234567). Now imagine that the researcher who originally submitted the record wanted to update the information. The updated record would keep the same Accession number, but would increase in version number (AF000001.2). The new record would have been given a completely new GI number (say GI:9876543).

Why is this important? The *Accession number* will always give you the most up-to-date information on a record, while the *Accession.Version* will always take you to a specific record. There are times when you want the most current information, and other times when you want to point to a particular piece of information from a particular point in time (e.g. a particular record that you did an analysis with), even if more information has been subsequently added. Note that as of September 2016, NCBI started phasing out the use of GI numbers. The use of *Accession.Version* form is now recommended for accessing a particular record, instead of the GI number. GI numbers are not to be confused with Entrez Gene IDs, which are an entirely different referencing system that NCBI uses!

Box 2. NCBI Help

This is a good time to get familiar with NCBI's thorough **Help** index for future reference. With this index, you should be able to access most of the background you need for understanding how these databases work on your own (there's also an NCBI YouTube channel if you'd like to check that out too).

- 1. At the bottom left of the NCBI homepage find the "NCBI Help Manual" link. Click on it. Then access the "Entrez Help" section.
- 2. You are now in Entrez Help. The Entrez collection of databases is queried when you use the *Search NCBI* interface. Note the contents that explain everything from search options to saving sets of records.
- 3. Notice that under the section **Entrez Searching Options** some other appropriate qualifiers are given, as illustrated on the previous page.
- 5. Search for our accession number of interest (e.g. NP_001318308 from above) through the *Search NCBI* portal page. It should give you protein sequence hit in the Proteins section. Click on it so that you get its full GenBank description (you can also click on the "armadillo/beta-catenin repeat protein [Arabidopsis thaliana]" link at the top of the page as the NCBI system recognizes that you've entered a protein identifier and hence provides some summary information for that above the numerical overview of results).

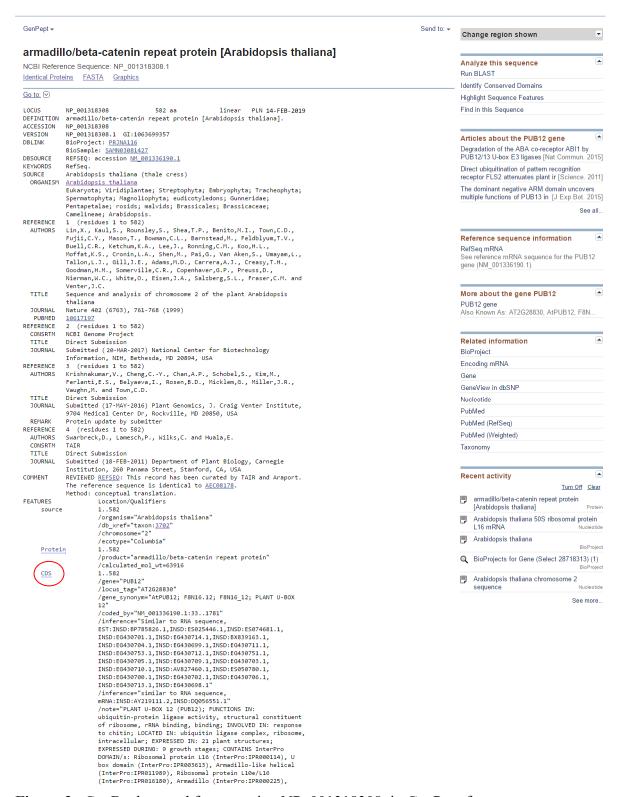


Figure 3. GenBank record for accession NP 001318308, in GenPept format.

- 6. Notice all the hyperlinks within the text. It looks messy but is in fact straightforward. For example, for taxonomic information, click on the **SOURCE ORGANISM** hyperlink (3702). Some records have links to the primary publication where this sequence was originally cited in a **PUBMED** number hyperlink (not the case in the above example, but there is a PubMed reference for the sequence). Click around on different links and see what you find.
 - a. What is the taxonomic lineage of your organism?
 - b. Has the genome of this organism been sequenced, i.e., is there a Genome Project?
 - c. If so, can you find the accession for the full sequence or one of the chromosomes?
 - > To find out much more information on the structure of the GenBank file at http://www.ncbi.nlm.nih.gov/Sitemap/samplerecord.html
- 7. Go back to the GenBank record and click on the **CDS** link, just above the actual sequence (circled in red in Figure 3 on the previous page).
 - a. Where did this take you or what happened when you did this?
- 8. Go back to the GenBank record and examine the **Related Information** section on the lower right. This gives you direct links to other databases with information on this query. Find the **Gene** link.



Figure 4. The **Related Information** menu for NP_001318308, to the right of the record. The arrow is pointing to the "Gene" link.

- 9. Select **Gene** from the **Related Information** menu. This is a great starter resource at NCBI. Scroll through the different sections. Use them to answer the following questions.
 - a. Where is your gene's location in the genome? (Tip: hover with your cursor over the green bars in the "Genomic regions, transcripts, and products" section; the green bars represent the gene in the sequence viewer)
 - b. How many exons do you see in this gene? Tip: how many green boxes are there?
 - c. What are the names of the genes surrounding it (i.e. what is its "Genomic context")?
 - d. Does it have any conserved domains? What are they called? (Tip: use the "Related Information" link to Conserved Domains on the right of the **Gene** page)
 - e. After exploring conserved domains go back to the **Gene** page. What biological process (Gene Ontology terms) is this gene involved with (scroll down!)?

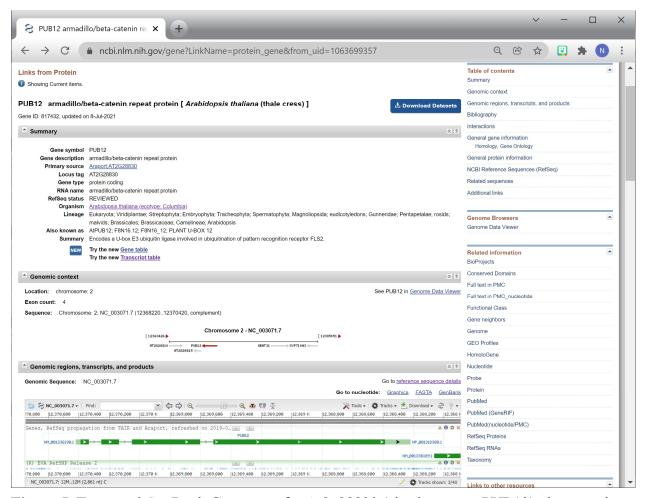


Figure 5. Truncated GenBank **Gene** page for At2g28830 (also known as PUB12), the gene that encodes NP_001318308.

- 10. On the Gene page, there are also **Additional links** to examine a gene's structure, function and phylogenetic relationships further. The navigation sidebar on the right has an "Additional links" hyperlink which will take you to the bottom of the page, where they're found for most genes. Click [+] Gene LinkOut to see them.
 - a. Click on Additional Links. What kind of information is in this section?
- ➤ Click around and explore the variety of ways that data for PUB12 are interconnected and displayed (don't worry, you can't break anything). Using the **Related Information** links can you find any publications associated with this gene? What about gene expression data? The next page shows the related "RefSeq RNA" record for the corresponding encoding mRNA (NCBI's RefSeq aims to provide canonical "reference" sequences genomic, mRNA, CDS, protein etc. for many model organisms).
 - b. Why is the length of the mRNA different from the value you can calculate from the start and stop positions in Question 9a?

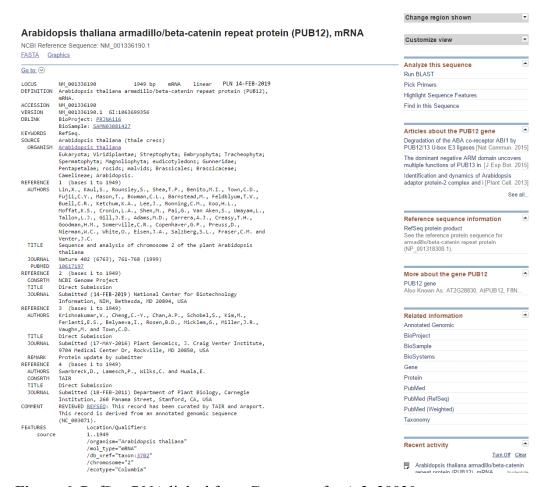


Figure 6. RefSeq RNA linked from Gene page for At2g28830.

Box 3. Helpful Hints for NCBI searches

On most NCBI search pages (except, oddly, *Search NCBI*) click on "Save Search" or "Create Alert" below the search box under the Advanced settings. Register for an account and save your search. You can also combine previous searches using the **History** tab and the search numbers listed within it, as well as save your searches by registering for a *My NCBI* account, so you don't have to keep redoing the same searches in the future.

Lab 1b — Basic BLAST (blastn)

One of the most important bioinformatic strategies used for the functional annotation of genes and genomes is to predict the function of uncharacterized genes or proteins based on their similarity to sequences with better functional annotations. BLAST is perhaps the single most important tool for finding database sequences that are similar to a query sequence of interest.

Box 4. BLAST and Homology

The Basic Local Alignment and Search Tool (BLAST; Altschul *et al.*, 1997) is a very powerful approach to identifying database sequences that share local similarity to a query sequence (see below for definitions). There is a very important chain of <u>assumptions</u> used in biological research that is generally followed when using BLAST:

- Homologous genes share sequence similarity
 - Orthologous genes have the highest similarity among multiple species
 - Orthologous genes most likely have similar functions
 - Consequently, sequences that are most similar between multiple species share similar functions

Note, it is very important to understand that these are only assumptions, and there are many reasons and instances where these assumptions prove to be false. Nevertheless, they are a reasonable starting place.

Definitions:

- **Similar sequences** sequences that share a significant number of residues (nucleotides or amino acids). Sequences can be similar due to homology or simply by chance. The higher the similarity between sequences, the more likely they are to be homologous.
- **Homologous sequences** sequences that are related through common ancestry. Homology is qualitative two sequences either are, or are not related through common ancestry. Homologous sequences can vary greatly in their level of *similarity* from 100% to 0%.
- Orthologous sequences sequences that are related through a past speciation event. Orthologous sequences are assumed to share common functions.
- **Paralogous sequences** sequences that are related through a past gene duplication event. Genes often diverge in function after duplicating; therefore, paralogous sequences are not assumed to share a common function.
- Query sequence your sequence; the sequence you are interested in finding more about.
- **High Scoring Segment Pair** (HSP) 'hits' to the database. A subsequence match between your query sequence and a database sequence returned by BLAST.
- Local alignment a sequence alignment that extends only across part of the sequence.
- Global alignment a sequence alignment that extends across the entire sequence (from end to end).
- 1. First, we need a query sequence for the search. Let's start with our given gene again, but this time we'll use the nucleotide sequence corresponding to the protein sequence, not the protein sequence. First try finding the gene's DNA sequence using Search NCBI tool again.
 - On the Search NCBI Portal page, search "All Databases" for your given protein sequence again using the Accession number. Using the protein from the first part of this lab, we would search for NP 001318308.

• The first page that comes up is the summary page. Once you're on this page you can move to the database of interest. In this case you probably don't have hits in too many databases since you had a very specific search.

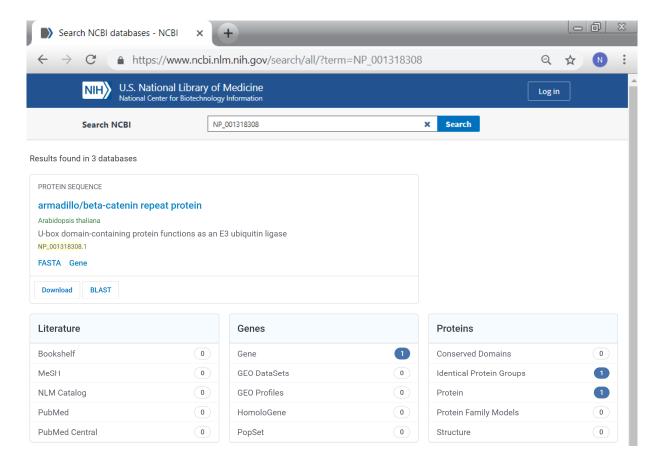


Figure 7. *Search NCBI* portal queried for NP_001318308 (partial view), with Gene results highlighted (numbers of results may differ slightly depending on when you're accessing NCBI).

• Try clicking the **Gene** link. Does the Gene page give you the gene sequence alone? What do you get instead? Note the context specific link menus that pop up when you hover over the graphic of the gene with your mouse pointer. You can click on the green boxes denoting the exons of the gene to get links to various sequences and analyses associated with the gene. Note that the green track is a composite of the mRNA and CDS tracks – click on either the NM_ or NP_ number to see the deconvolution of the green track (Figure 8).

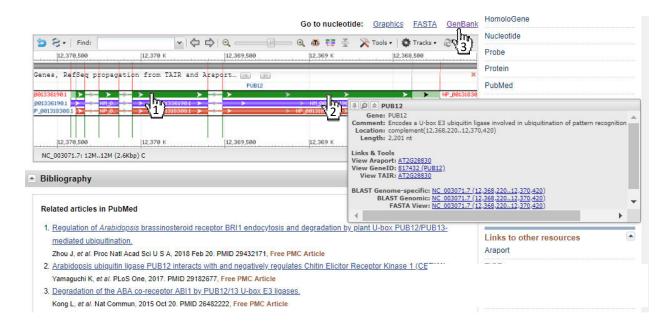


Figure 8. Part of the Gene page for NP_001318308, showing pop-up to sequence links. 1. Click the green bars to make mRNA and protein tracks appear; 2. hover over the mRNA track to see info panel; 3. Click "Genbank" link to see Genbank record for the genomic region for this gene.

- Click on the **RefSeq RNAs** link in the "Related information" panel on the right. This takes you to the mRNA that encodes the protein you have been looking at (we are accessing the same record you accessed in Step 10 of the first part of the lab). Notice the feature list in the record. One Feature in the GenBank record is **gene**, and corresponds to base position 1 1949 on this record. Another features is the coding sequence (**CDS**), which corresponds to base position 33 1781.
 - a. Given your biology background knowledge, why do you think these are different?
- Above the Sequence Viewer panel, click on the "Go to nucleotide: Genbank" link (see Step 3. in Figure 8 above). You will be taken you to the genomic region that encodes the mRNA you were just looking at. Notice how the gene feature corresponds to positions 1–2201, while the mRNA feature corresponds to positions 1–86, 170–286, 370–819, and 906–2201 and the CDS feature corresponds to nucleotide positions 33–86, 170–286, 370–819, and 906–2033. You may have remarked that the sequence from the chromosome has been reverse complemented.
 - b. Again, why are these different? Tip: recall the Central Dogma of Molecular Biology!



Figure 9. GenBank record for NM 001336190 mRNA.

- Let's return the mRNA record we were previously working with (NM_001336190, as on page 11, accessed through RefSeq RNAs link). Click on the **CDS** link. Now you are looking at the information for the coding sequence, as opposed to the whole gene or protein (highlighted in brown).
- Using the "Display: FASTA" option in the grey bar at the bottom of the page generate a FASTA-formatted version of the CDS.
- Now you have the sequence in the most basic and easily managed format **FASTA** format. FASTA format is simply a header line that starts with a '>' followed by text describing the sequence, and then the actual sequence beginning on the next line. The sequence can be either DNA or protein, and may be continuous (scrolling off the page), or cut into more manageable lengths typically ranging between 60-80 residues.

 $>NM_001336190.1:33-1781$ Arabidopsis thaliana armadillo/beta-catenin repeat protein (PUB12), mRNA

AAGTGATGGTGAAATTTCAGAAAGTGACTTCTCTATTGGAACAAGCTTTAAGTATAATCCCTTATGAGAA TCTGGAAATTTCAGATGAACTTAAAGAACAGGTGGAGCTTGTTTTAGTTCAGTTAAGAAGATCGTTAGGA AAACGCGGTGGCGATGTATGATGATGATGATTGTATAAGGATGTTCTATCTCTTTATAGTGGTAGAGGTA GTGTAATGGAGTCTGATATGGTTAGGAGAGTGGCGGAGAAGCTTCAGTTGATGACTATAACTGACCTTAC GCAAGAGTCATTGGCTTTACTTGACATGGTTAGTTCTAGTGGTGATGATCCTGGTGAAAGTTTTGAG AAGATGTCTATGGTTCTTAAGAAGATTAAGGACTTTGTGCAAACTTATAATCCTAACTTGGATGATGCTC CATTGAGACTGAAATCATCGCTTCCGAAGTCGCGAGATGATCGAGATATGCTAATTCCGCCTGAAGA GTTCCGTTGTCCAATATCTCTAGAATTGATGACTGATCCAGTTATTGTTTCTTCAGGGCAGACTTATGAA CGTGAGTGCATTAAGAAGTGGCTTGAAGGAGGACACTTGACGTGTCCAAAGACGCAAGAAACGCTGACAA GCGATATCATGACACCAAACTATGTTCTAAGAAGCCTTATAGCTCAATGGTGTGAGTCCAATGGCATCGA ACCTCCAAAGCGTCCCAACATATCTCAACCGAGTAGTAAGGCCTCATCTTCGTCGTCAGCCCCTGATGAT GAACATAACAAGATTGAAGAACTTCTACTTAAGCTCACATCGCAACAGCCTGAAGACCGAAGATCTGCTG CAGGAGAAATCCGTCTTCTAGCAAAACAAACAATCATAACCGAGTCGCCATTGCTGCCTCAGGCGCGAT CCCTCTTCTGGTGAATCTCCTCACGATATCTAATGACTCTCGGACTCAAGAACACGCTGTGACATCGATT CTTAACCTCTCGATATGTCAAGAGAACAAGGGAAGATTGTTTATTCATCTGGAGCAGTTCCAGGTATTG TTCATGTGCTTCAGAAAGGTAGCATGGAAGCTAGAGAAAACGCAGCTACACTTTTCAGCCTCTCGGT TATAGACGAGAACAAAGTGACAATAGGTGCCGCAGGAGCGATCCCGCCTCTTGTGACCTTGCTGAGCGAA GGATCACAGAGAGGCAAAAAAGACGCGGCAACTGCTCTGTTTAATCTCTGCATATTTCAAGGAAACAAAG GAAAAGCTGTGAGAGCCGGTTTAGTTCCCGTGCTAATGAGGTTACTAACAGAACCCGAAAGCGGAATGGT TGATGAATCACTCTCGATATTAGCCATACTATCGAGTCATCCGGACGGGAAATCAGAGGTTGGAGCCGCT GATGCAGTTCCAGTTCTGGTAGATTTTATAAGAAGCGGGTCACCGCGGAACAAAGAAAACTCAGCTGCGG TTTAATAGAAATGGCTGAGAATGGTACTGACAGAGGAAAACGCAAAGCGGCACAGTTACTTAACCGCTTT AGCCGTTTTAACGACCAGCAGAAACACCTCTGGTTTAGGTTTGGAAGATCAAATCTCCCTAATCTGA

Figure 10. Sequence in FASTA text format.

2. Let's do some BLASTing! Use the "Run BLAST" link in the "Analyze This Sequence" part of the webpage. [Or open a new tab or window in your browser and go back to the NCBI home page (www.ncbi.nlm.nih.gov), then select **BLAST** from the Resources dropdown along the top, under the DNA&RNA subsection].

There are lots of options here. We will discuss some of these next lab, but right now let's work with the simplest. Since our sequence is a nucleotide sequence, we want to do a *nucleotide blast*.

• On the BLAST page, note that under the **Enter Query Sequence** section, the NCBI

system has automatically entered the **accession number** (but you can also enter **a GI number**, or **FASTA sequence**) and **subrange** (we'll be searching with just the coding sequence part of the mRNA sequence). You could also copy-and-paste the FASTA formatted CDS sequence you found as in Figure 10 into the query box *without* defining a subrange – you should be clear on the difference between an mRNA sequence and coding sequence at this point...

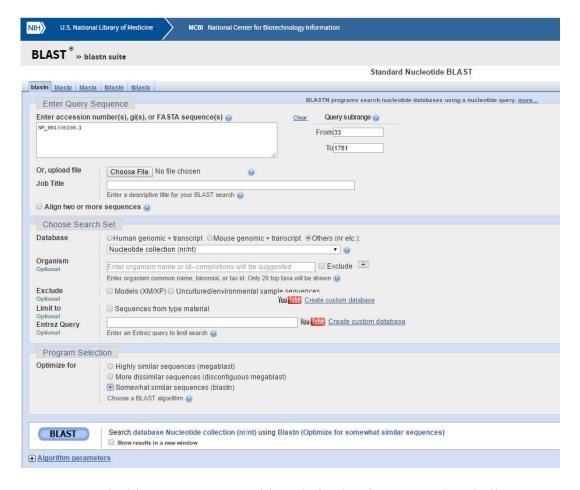


Figure 11. The blastn query page, with optimization for "Somewhat similar sequences (blastn)" selected.

- Scan the sections of the page. You have quite a bit of control over how the algorithm runs (particularly if you click [+] **Algorithm parameters** near the bottom.
- We want to query the full NCBI database; the NCBI linking system has automatically changed the default **Database** (which is Human) to *Other* and *Nucleotide collection* (*nr/nt*) because our sequence is non-human. The nr database is the non-redundant collection of sequences in GenBank.
- Change the **Program Selected / Optimized for** to **Somewhat similar sequences (blastn)**.
- Note all the small question mark icons around the page. Click any one of these to find
 out more about the associated parameter. For example, by clicking the question mark in
 the **Program Selection** section you get a very brief summary of the different methods.
 By clicking **more** you jump to a new page with full documentation for the algorithms.

a. When would you want to use megaBLAST? What about discontinuous megaBLAST? (if you have time, try each to see how your results differ)

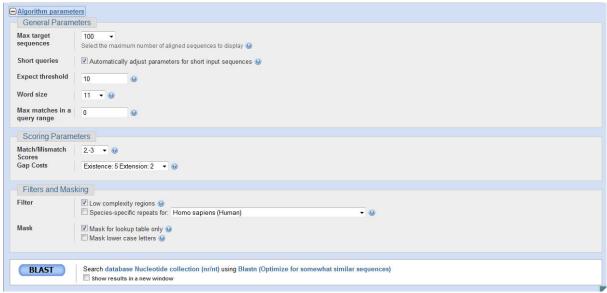


Figure 12. Algorithm parameters for blastn.

- Open the Algorithm Parameters near the bottom.
 - b. What is the **Expect threshold**?
 - c. What would happen if you decreased it? Increased it?
 - d. What would be the effect of increasing the **Word size**?
 - e. Why is there a **Low complexity regions** filter? Should we keep it on?
- Make sure you have your query sequence entered in the input box, and check the box next to Show results in a new window near the BLAST button. Now (finally) click the BLAST button.
- While BLAST is running or after the search is complete you can choose to adjust the format of the search results by clicking on the **Format options** link. We won't do this right now, as the defaults usually work fine.

Box 5. How Good is My Hit?

The quality of a BLAST HSP is quantified in a number of different ways. It is important that you understand the differences between these metrics and use the appropriate one.

- Identity the extent to which two sequences are invariant. A very poor measure since it doesn't take into account the subtleties of sequence relationships (e.g., a small region of a highly conserved domain within two sequences that are otherwise very poorly conserved).
- Bit score the alignment score (S). A very precise measure that is normalized over the particular score system employed. Suffers from the disadvantage of being dependent on the length of the query.



• E value – the expect value. A value that is based on the number of different alignments with scores at least as good as that observed, which are expected to occur simply by chance. The lower the E value, the more significant the score. This is by far the best metric to use since results of different searches in the same database can be readily compared. Note that E value is dependent on the size of the database (n) and the length of the query sequence (m). The same sequence searched on different databases containing identical hit sequences would result in different E values being reported.

$$E = mn2^{-S}$$

We'll go into greater detail about this calculation in next week's class.

- 3. The Results page is broken up into sections.
 - At the very top is the job summary, which simply shows details about your query and the
 database searched. You can find more details about your search by clicking Search
 Summary.
 - a. How many sequences are in the nr database?
 - b. What sequences are not included in the nr database? (Trick question: this information is actually available by clicking on the question mark beside the Database option on the input page!)

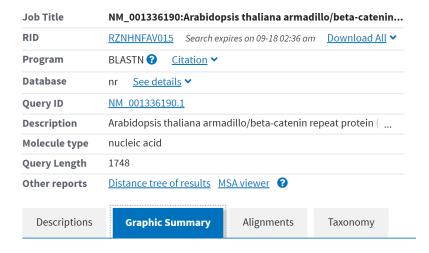


Figure 13. Blastn output Search Summary.

- Explore the **Graphic Summary** tab. Scroll your mouse over the coloured bars.
 - c. What do the coloured bars mean?
 - *d.* How does the colour code work?
 - e. What information is displayed when you hover on an entry?
 - f. What do you notice about the significance values as you move down the graphical summary?
 - g. What is the genus and species of the top (best) hit?
 - h. What happens if you click on one of the entries?

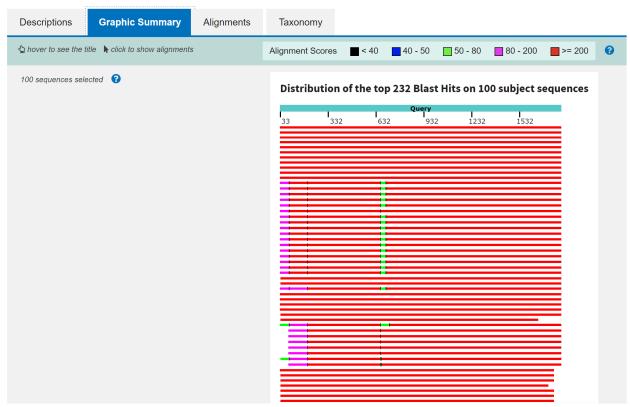


Figure 14. Blastn output Graphic Summary.

- The **Descriptions** tab lists the following:
 - **Description** [hyperlinked to corresponding **Alignment(s)** in Alignments section]
 - Max Score the alignment bit score
 - **Total Score** another alignment bit score which may differ from the **Max Score** if your query matched a single database entry in multiple regions.
 - Query Coverage what percent of the guery had similarity to the database hit.
 - **E-value** probably the best measure of hit quality. Smaller numbers mean better hits, with 0.0 being the best value possible.
 - **Identity** the highest identity found between guery and HSP.
 - Accession linked to the indicated sequence at NCBI
 - i. How many sequence matches are listed for this query sequence? How are they ordered? (you can sort these segments in other ways, like by identity, score, and query start position.)
 - j. What happens if you click the Accession hotlink?
 - k. What happens if you click the Alignments hotlink?

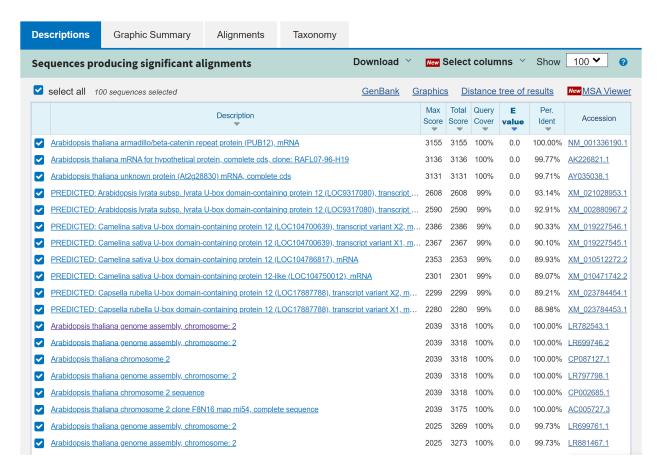


Figure 15. Blastn output descriptions (some default columns removed in image).

Lab Quiz Question 3

- Finally we can explore the actual HSP **Alignments** in the Alignments tab.
 - Compare the information presented for the first HSP alignment to the first entry in the graphical summary and HSP summary.
 - As you scroll down the alignments, you will see the alignment quality drop that is, the e-value increases.
 - *l.* What do the vertical bars (|) represent between the **Query** and the **Sbjct** (database sequence)?
 - m. What does **Strand=Plus/Plus**, **Strand=Plus/Minus** mean? Hint: are genes always in the same direction on a piece of chromosomal DNA?
- Go back to the top of the page and click **Formatting options**. Change the **Alignment View** to **Query-anchored with dots for identities**. Click **Reformat** and score down to the HSP alignment section.
 - n. Describe the difference between this format and the previous format. Can you imagine cases where the different formats might be most useful?
 - o. Play with these format options to get a feel for what they mean.

- Return the formatting to the original Pairwise format. Go back to the graphical summary. If there are any low-scoring segments (i.e.: green or blue-coded blocks), click on one.
 - n. What is its E-value?
 - o. Does it have a high percent identity? If so, why would BLAST give it such a poor E-value?
 - p. Do you think these hits are homologous? Why or why not?

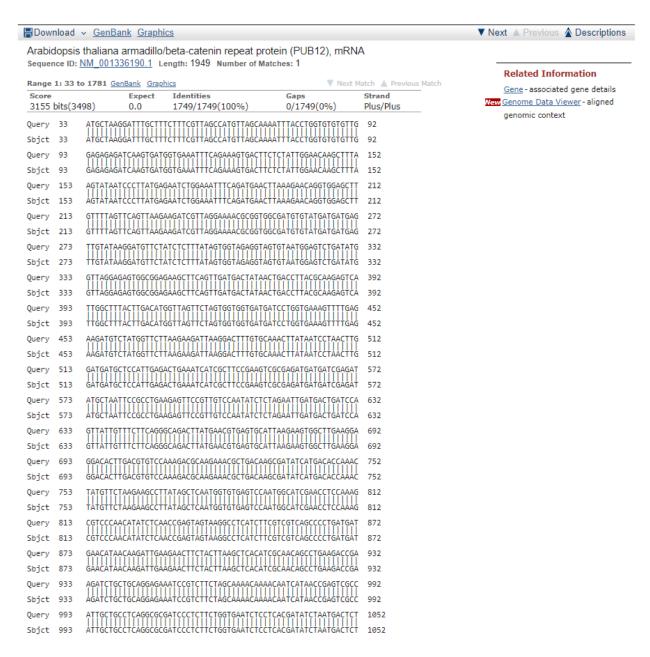


Figure 16. Blastn output alignments.

End of Lab!

Lab 1 Objectives

By the end of Lab 1 (comprising the lab including its boxes, and the lecture), you should:

- know how to search for records at NCBI, both using search terms or identifiers (first part of lab) and Search NCBI / GQuery, or using a nucleotide sequence and BLAST;
- know the difference between a GenBank accession number, a version number, and a GI number;
- understand the difference between the nucleotide sequence database part of GenBank and the protein sequence part of it;
- know the parts of a GenBank record and be able to switch between sequence formats (e.g. to FASTA format);
- be familiar with the interconnectedness of various NCBI databases and be able to call up linked records with ease;
- be able to use nucleotide BLAST (Blastn) to search GenBank, and be able to interpret the output what does the E-value tell you etc.?;
- understand the meaning of homologous, orthologous, and paralogous sequences;
- be able to use the Help function to address any question you may have with regards to the NCBI interface (if you have any questions on background material, check in with the forums for this course on Coursera!).

Do not hesitate to post any questions you might have to the Forum section of the Coursera website for this course if you do not understand any of the above after reading the relevant material.

Further Reading

Chapter 2 "Information Organization and Sequence Databases" in *Concepts in Bioinformatics and Genomics* by Jamil Momand and Alison McCurdy, Oxford University Press, 2017. pp 21-37.

SF Altschul, TL Madden, AA Schaffer, J Zhang, Z Zhang, W Miller, and DJ Lipman (1997) Gapped BLAST and PSI-BLAST: a new generation of protein database search programs. Nucl. Acids Res. 25: 3389-3402.

NM Luscombe, D Greenbaum, M Gerstein (2001) What is bioinformatics? An introduction and overview. Yearkbook of Medical Informatics 2001:83.

CA Kerfeld, KM Scott (2011) Using BLAST to Teach "E-value-tionary" Concepts. PLoS Biol 9(2): e1001014. http://dx.doi.org/10.1371/journal.pbio.1001014.

Appendix 1: GenBank Field Qualifiers

From http://www.ncbi.nlm.nih.gov/books/NBK3837/#EntrezHelp.Entrez Searching Options

Accession [ACCN]

Contains the unique accession number of the sequence or record, assigned to the nucleotide, protein, structure, genome record, or PopSet by a sequence database builder. The Structure database accession index contains the PDB IDs but not the MMDB IDs.

All Fields [ALL]

Contains all terms from all searchable database fields in the database.

Author Name [AUTH]

Contains all authors from all references in the database records. The format is last name space first initial(s), without punctuation (e.g., marley jf).

EC/RN Number [ECNO]

Number assigned by the Enzyme Commission or Chemical Abstract Service (CAS) to designate a particular enzyme or chemical, respectively.

Feature Key [FKEY]

Contains the biological features assigned or annotated to the nucleotide sequences and defined in the DDBJ/EMBL/GenBank Feature Table (http://www.ncbi.nlm.nih.gov/projects/collab/FT/index.html). Not available for the Protein or Structure databases.

Filter [FILT]

Contains predetermined or filtered subsets of the various databases. These subsets or filters are created by grouping records that are commonly linked to other GQuery databases or within the same database. For example, the PopSet database Filter index includes PopSet all, PopSet medline, PopSet nucleotide, and PopSet protein. The PopSet medline filter includes all PopSet records with links to PubMed; the PopSet nucleotide filter includes all PopSet records with links to the nucleotide database; and, the PopSet protein filter includes all PopSet records with links to the protein database. The PopSet all filter includes all PopSet records.

Gene Name [GENE]

Contains the standard and common names of genes found in the database records. This field is not available in Structure database.

Issue [ISS]

Contains the issue number of the journal in which the data were published.

Journal Name [JOUR]

Contains the name of the journal in which the data were published. Journal names are indexed in the database in abbreviated form (e.g., J Biol Chem). Journals are also indexed by their by ISSNs. Browse the index if you do not know the ISSN or are not sure how a particular journal name is abbreviated.

Keyword [KYWD]

Contains special index terms from the controlled vocabularies associated with the GenBank, EMBL, DDBJ, SWISS-Prot, PIR, PRF, or PDB databases. Browse the Keyword indexes of the individual databases to become familiar with these vocabularies. A Keyword index is not available in the Structure database.

Modification Date [MDAT]

Contains the date that the most recent modification to that record is indexed in GQuery, in the format YYYY/MM/DD (e.g., 1999/08/05). A year alone, (e.g., 1999) will retrieve all records modified for that year; a year and month (e.g., 1999/03) retrieves all records modified for that month that are indexed in GQuery.

Molecular Weight [MOLWT]

Molecular weight of a protein, in Daltons (Da), calculated by the method described in the Searching by Molecular

Weight section of the GQuery help document. Note that molecular weight must be entered as a fixed 6 digit field, filled with leading zeros (not letter O), e.g., 002002 [MOLWT]

Organism [ORGN]

Contains the scientific and common names for the organisms associated with protein and nucleotide sequences.

Page Number [PAGE]

Contains the number of the first journal page of the article in which the data were published.

Primary Accession [PACC]

Contains the primary accession number of the sequence or record, assigned to the nucleotide, protein, structure, genome record, or PopSet by a sequence database builder. A Primary Accession index is not available in the Structure database.

Properties [PROP]

Contains properties of the nucleotide or protein sequence. For example, the Nucleotide database's Properties index includes molecule types, publication status, molecule locations, and GenBank divisions. A Properties index is not available in the Structure database.

Protein Name [PROT]

Contains the standard names of proteins found in database records. Common names may not be indexed in this field so it is best to also consider All Fields or Text Words. A Protein Name index is not available in the Structure database.

Publication Date [PDAT]

Contains the date that records are released into GQuery, in the format YYYY/MM/DD (e.g., 1999/08/05). It is the date the entry first appeared in GenBank explicitly indexed in GQuery. A year alone, (e.g., 1999) will retrieve all records for that year; a year and month (e.g., 1999/03) will retrieve all records released into GenBank for that month.

SeqID String [SQID]

Contains the special string identifier, similar to a FASTA identifier, for a given sequence. A SeqID String index is not available in the Structure database.

Sequence Length [SLEN]

Contains the total length of the sequence. Sequence Length indexes are not available in the Structure or PopSet databases.

Substance Name [SUBS]

Contains the names of any chemicals associated with this record from the CAS registry and the MEDLINE Name of Substance field. Substance Name indexes are not available in the Genome or PopSet databases.

Text Word [WORD]

Contains all of the "free text" associated with a record.

Title Word [TITL]

Includes only those words found in the definition line of a record. The definition line summarizes the biology of the sequence and is carefully constructed by database staff. A standard definition line will include the organism, product name, gene symbol, molecule type and whether it is a partial or complete cds. Title Word indexes are not available in the Structure or PopSet databases.

Uid [UID]

Contains the Medline unique identifier for records that contain published references that are linked to PubMed. The Uid index is not browsable.

Volume [VOL]

Contains the volume number of the journal in which the data were published.