

Alinity c Alanine Aminotransferase-01

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BIENNIAL REVIEW:

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SUPERSEDES: Procedure titled _____

INTENDED USE

The Alanine Aminotransferase2 assay is used for the quantitation of alanine aminotransferase in human serum and plasma on the Alinity c system.

The Alanine Aminotransferase2 assay is to be used as an aid in the diagnosis and treatment of certain liver diseases (e.g., viral hepatitis and cirrhosis).

SUMMARY AND EXPLANATION OF THE TEST

Alanine aminotransferase (ALT) is an enzyme found abundantly in the cytosol of the hepatocyte, and its activity in the liver is about 3000 times that of serum activity. Although it is generally thought to be specific to the liver, it is also found in the kidney and in much

smaller quantities in heart and skeletal muscle cells. ALT has a plasma half-life of 47 ± 10 hours, which is longer than that of aspartate aminotransferase (AST) (17 ± 5 hours).¹

ALT rises in disease states that cause hepatocellular injury.¹

The cause of hepatocellular injury may result in varying magnitudes of elevation in ALT and AST. Borderline ALT elevation is defined as <2 times the upper limit of normal (ULN), mild ALT elevation is defined as 2 to 5 times the ULN, moderate ALT elevation is defined as 5 to 15 times the ULN, and severe ALT elevation is defined as >15 times the ULN.² Further work-up and management is determined by the magnitude of elevation in conjunction with other diagnostic factors.

PRINCIPLES OF THE PROCEDURE

The Alanine Aminotransferase2 assay is an automated clinical chemistry assay.

ALT present in the sample catalyzes the transfer of the amino group from *L*-alanine to α -ketoglutarate, forming pyruvate and *L*-glutamate. Pyruvate in the presence of NADH and lactate dehydrogenase is reduced to *L*-lactate. In this reaction, NADH is oxidized to NAD^+ . The reaction is monitored by measuring the rate of decrease in absorbance at 340 nm due to the oxidation of NADH to NAD^+ .

Methodology: NADH (without P-5'-P)

For additional information on system and assay technology, refer to the Alinity ci-series Operations Manual, Section 3.

REAGENTS

Kit Contents

Alanine Aminotransferase2 Reagent Kit 04T84

Volumes (mL) listed in the following table indicate the volume per cartridge.

REF	04T8420	04T8430
Tests per cartridge	300	780
Number of cartridges per kit	4	4
Tests per kit	1200	3120
R1	28.7 mL	67.5 mL
R2	19.3 mL	43.3 mL
R1 Active ingredients: <i>L</i> -alanine (66.820 g/L), β -NADH (0.305 g/L), lactate dehydrogenase (5.000 KU/L). Preservative: sodium azide.		
R2 Active ingredients: <i>L</i> -alanine (89.090 g/L), α -ketoglutaric acid (13.150 g/L). Preservative: ProClin 300.		


Warnings and Precautions

- **IVD**
- For *In Vitro* Diagnostic Use
- **Rx ONLY**

Safety Precautions

CAUTION: This product requires the handling of human specimens. It is recommended that all human-sourced materials and all consumables contaminated with potentially infectious materials be considered potentially infectious and handled in accordance with the OSHA Standard on Bloodborne Pathogens. Biosafety Level 2 or other appropriate regional, national, and institutional biosafety practices should be used for materials that contain, are suspected of containing, or are contaminated with infectious agents.[3](#), [4](#), [5](#), [6](#)

The following warnings and precautions apply to: R1	
Contains sodium azide.	
EUH032	Contact with acids liberates very toxic gas.
P501	Dispose of contents / container in accordance with local regulations.

The following warnings and precautions apply to: R2	
	
WARNING	Contains methylisothiazolones.
H317	May cause an allergic skin reaction.
H402*	Harmful to aquatic life.
H412	Harmful to aquatic life with long lasting effects.
Prevention	
P261	Avoid breathing mist / vapors / spray.
P272	Contaminated work clothing should not be allowed out of the workplace.
P280	Wear protective gloves / protective clothing / eye protection.
P273	Avoid release to the environment.
Response	
P302+P352	IF ON SKIN: Wash with plenty of water.
P333+P313	If skin irritation or rash occurs: Get medical advice / attention.
P362+P364	Take off contaminated clothing and wash it before reuse.
Disposal	
P501	Dispose of contents / container in accordance with local regulations.

* Not applicable where regulation EC 1272/2008 (CLP) has been implemented.

Follow recommendations and content in the Safety Data Sheet to determine the safe disposal of this product.

For the most current hazard information, see the product Safety Data Sheet.

Safety Data Sheets are available at www.corelaboratory.abbott and within lab Alinity SDS folder.

For a detailed discussion of safety precautions during system operation, refer to the Alinity ci-series Operations Manual, Section 8.

Reagent Handling

- Upon receipt, place reagent cartridges in an upright position for 1 hour before use to allow bubbles that may have formed to dissipate.
- If a reagent cartridge is dropped, place in an upright position for 1 hour before use to allow bubbles that may have formed to dissipate.
- Reagents are susceptible to the formation of foam and bubbles. Bubbles may interfere with the detection of the reagent level in the cartridge and cause insufficient reagent aspiration that may adversely affect results.

For a detailed discussion of reagent handling precautions during system operation, refer to the Alinity ci-series Operations Manual, Section 7.

Reagent Storage

	Storage Temperature	Maximum Storage Time	Additional Storage Instructions
Unopened	2 to 8°C	Until expiration date	Store in upright position.
Onboard	System Temperature	30 days	
Opened	2 to 8°C	Until expiration date	Store in upright position. Do not reuse original reagent caps or replacement caps due to the risk of contamination and the potential to compromise reagent performance.

Reagents may be stored on or off the system. If removed from the system, store reagents with new replacement caps in an upright position at 2 to 8°C. For reagents stored off the system, it is recommended that they be stored in their original trays or boxes to ensure they remain upright.

For information on unloading reagents, refer to the Alinity ci-series Operations Manual, Section 5.

Indications of Reagent Deterioration

Deterioration of the reagents may be indicated when:

- a calibration error occurs
- a control value is out of the specified range

Associated test results are invalid, and samples must be retested. Assay recalibration may be necessary.

For troubleshooting information, refer to the Alinity ci-series Operations Manual, Section 10.

INSTRUMENT PROCEDURE

The Alanine Aminotransferase2 assay file must be installed on the Alinity c system prior to performing the assay.

The Alinity ci-series system software version 3.2.0 or higher must be installed on the Alinity c system prior to performing the assay.

For detailed information on assay file installation and viewing and editing assay parameters, refer to the Alinity ci-series Operations Manual, Section 2.

For information on printing assay parameters, refer to the Alinity ci-series Operations Manual, Section 5.

For a detailed description of system procedures, refer to the Alinity ci-series Operations Manual.

Alternate Result Units

Edit assay parameter "Result Units" to select an alternate unit.

Conversion formula:

(Concentration in Default result unit) x (Conversion factor) = (Concentration in Alternate result unit)

Default Result Unit	Conversion Factor	Alternate Result Unit
U/L	0.01667	μkat/L

SPECIMEN COLLECTION AND PREPARATION FOR ANALYSIS

Specimen Types

The specimen types listed below were verified for use with this assay.

Other specimen types and collection tube types have not been verified with this assay.

Specimen Types	Collection Tubes
Serum	Serum
	Serum separator
Plasma	Dipotassium EDTA
	Lithium heparin
	Lithium heparin separator
	Sodium heparin

- Liquid anticoagulants may have a dilution effect resulting in lower concentration values for individual specimens.

Specimen Conditions

Do not use:

- heat-inactivated specimens
- pooled specimens
- grossly hemolyzed specimens
- specimens with obvious microbial contamination
- specimens with fungal growth
- For accurate results, serum and plasma specimens should be free of fibrin, red blood cells, and other particulate matter. Serum specimens from patients receiving anticoagulant or thrombolytic therapy may contain fibrin due to incomplete clot formation.
- To prevent cross contamination, use of disposable pipettes or pipette tips is recommended.

Preparation for Analysis

- Follow the tube manufacturer's processing instructions for collection tubes. Gravity separation is not sufficient for specimen preparation.
- Specimens should be free of bubbles. Remove bubbles with an applicator stick before analysis. Use a new applicator stick for each specimen to prevent cross contamination.

To ensure consistency in results, recentrifuge specimens prior to testing if

- they contain fibrin, red blood cells, or other particulate matter.

NOTE: If fibrin, red blood cells, or other particulate matter are observed, mix by low speed vortex or by inverting 10 times prior to recentrifugation.

Prepare frozen specimens as follows:

- Frozen specimens must be completely thawed before mixing.
- Mix thawed specimens thoroughly by low speed vortex or by inverting 10 times.
- Visually inspect the specimens. If layering or stratification is observed, mix until specimens are visibly homogeneous.
- If specimens are not mixed thoroughly, inconsistent results may be obtained.
- Recentrifuge specimens.

Recentrifugation of Specimens

- Transfer specimens to a centrifuge tube and centrifuge.
- Transfer clarified specimen to a sample cup or secondary tube for testing. For centrifuged specimens with a lipid layer, transfer only the clarified specimen and not the lipemic material.

Specimen Storage

Specimen Type	Temperature	Maximum Storage Time
Serum/Plasma	Room temperature (20 to 25°C)	3 days ⁷
	2 to 8°C	7 days ⁷
	-20°C	60 days ⁸

Avoid multiple freeze/thaw cycles.⁸

For additional information on sample handling and processing, refer to CLSI GP44-A4.[9](#) The storage information provided here is based on references.

Stored specimens must be inspected for particulates. If present, mix with a low speed vortex or by inversion and centrifuge the specimen to remove particulates prior to testing.

Specimen Shipping

Package and label specimens in compliance with applicable state, federal, and international regulations covering the transport of clinical specimens and infectious substances.

Do not exceed the storage limitations listed above.

PROCEDURE

Materials Provided

04T84 Alanine Aminotransferase2 Reagent Kit

Materials Required but not Provided

- Alanine Aminotransferase2 assay file
- 04V6201 Consolidated Chemistry Calibrator, if using the Calibration method
- Controls containing alanine aminotransferase
- Saline (0.85% to 0.90% NaCl) for specimen dilution

For information on materials required for operation of the instrument, refer to the Alinity ci-series Operations Manual, Section 1.

For information on materials required for maintenance procedures, refer to the Alinity ci-series Operations Manual, Section 9.

Assay Procedure


For a detailed description of how to run an assay, refer to the Alinity ci-series Operations Manual, Section 5.

- If using primary or aliquot tubes, refer to the Alinity ci-series Operations Manual, Section 4 to ensure sufficient specimen is present.
- Minimum sample cup volume is calculated by the system and printed on the Order List report. To minimize the effects of evaporation, verify adequate sample cup volume is present prior to running the test.

Minimum sample volume requirements:

- Sample volume for single test: 5.3 µL.

NOTE: This amount does not include the dead volume plus the additional over-aspiration volume. For total sample volume requirements, refer to the Alinity ci-series Operations Manual, Section 4.

- Refer to the Consolidated Chemistry Calibrator package insert  04V6201 and/or commercially available control material package insert for preparation and usage.
- For general operating procedures, refer to the Alinity ci-series Operations Manual, Section 5.
- For optimal performance, it is important to perform routine maintenance as described in the Alinity ci-series Operations Manual, Section 9. Perform maintenance more frequently when required by laboratory procedures.

Sample Dilution Procedures

Samples with an alanine aminotransferase value exceeding 3258 U/L (54.31 µkat/L) are flagged with the code "> 3258 U/L" ("> 54.31 µkat/L") and may be diluted with either the Automated Dilution Protocol or the Manual Dilution Procedure.

Automated Dilution Protocol

The system performs a 1:5 dilution of the sample and automatically calculates the concentration by multiplying the result by the dilution factor.

For details on configuring automated dilutions, refer to the Alinity ci-series Operations Manual, Section 2.

Manual Dilution Procedure

Dilute the sample with saline (0.85% to 0.90% NaCl).

The operator must enter the manual dilution factor in the Specimen or Control tab of the Create Order screen. The system will use this dilution factor to automatically calculate the concentration of the sample and report the result.

If the operator does not enter the manual dilution factor, the result must be manually multiplied by the appropriate manual dilution factor before reporting the result. If a diluted sample result is less than 2 U/L (0.03 μ kat/L), do not report the result. Rerun using an appropriate dilution.

NOTE: The default Low Linearity value of the assay file corresponds to the lower limit of the reportable interval of 2 U/L (0.03 μ kat/L). To flag values using the lower limit of the analytical measuring interval of 7 U/L (0.12 μ kat/L), the operator must edit the Low Linearity value.


For detailed information on editing the result settings of assay parameters, refer to the Alinity ci-series Operations Manual, Section 2.

For detailed information on ordering dilutions, refer to the Alinity ci-series Operations Manual, Section 5.

Calibration

For instructions on performing a calibration, refer to the Alinity ci-series Operations Manual, Section 5.

Calibration can be performed using one of 2 methods:

- Calibration method, using the Consolidated Chemistry Calibrator  04V6201. For the Calibration method, use assay file ALT2.
- Calibration Factor method, using a fixed calibration factor value to calculate the result. For the Calibration Factor method, use assay file ALT2F.

Calibration is stable for approximately 30 days (720 hours) but is required with each change in reagent lot. Verify calibration with at least 2 levels of controls according to the established quality control requirements for your laboratory. If control results fall outside acceptable ranges, recalibration may be necessary.

This assay may require recalibration after maintenance to critical parts or subsystems or after service procedures have been performed.

Quality Control Procedures

- At least 2 levels of controls (low and high) are to be run every 24 hours.
- If quality control results do not meet the acceptance criteria defined by your laboratory, sample results may be suspect. Follow the established quality control procedures to troubleshoot. Recalibration may be necessary. For troubleshooting information, refer to the Alinity ci-series Operations Manual, Section 10.
- Review quality control results and acceptance criteria following a change of reagent or calibrator lot using lot to lot verification.

Controls should be used according to the guidelines and recommendations of the control manufacturer. Concentration ranges provided in the control package insert should be used only for guidance.

For any control material in use, the laboratory should ensure that the matrix of the control material is suitable for use in the assay per the assay package insert.

Quality Control Guidance

Refer to “Basic QC Practices” by James O. Westgard, Ph.D. for guidance on laboratory quality control practices.[10](#)

Verification of Assay Claims

For protocols to verify package insert claims, refer to Verification of Assay Claims in the Alinity ci-series Operations Manual.

RESULTS

Calculation

Calibration method

The Alanine Aminotransferase2 (ALT2) assay utilizes the Linear data reduction method to generate a calibration and results.

Calibration Factor method

The Alanine Aminotransferase2 (ALT2F) assay utilizes the Factor data reduction method to generate a calibration and results.

The calibration factor for the Alanine Aminotransferase2 is 8615.

The Alanine Aminotransferase2 assay is traceable to the IFCC (International Federation of Clinical Chemistry) reference method.[11](#) The assigned values for the calibrator and the calibration factor are traceable to the standardization.

For additional information, refer to the Alinity ci-series Operations Manual, Section 6.

Flags

Some results may contain information in the Flags field. For a description of the flags that may appear in this field, refer to the Alinity ci-series Operations Manual, Section 5.

Reportable Interval

Based on representative data for the limit of quantitation (LoQ) and the limit of detection (LoD), the ranges over which results can be reported are provided below according to the definitions from CLSI EP34, 1st ed.[12](#)

	U/L	μkat/L
Analytical Measuring Interval (AMI) ^a	7 - 3258	0.12 - 54.31
Extended Measuring Interval (EMI) ^b	3258 - 16 290	54.31 - 271.55
Reportable Interval ^c	2 - 16 290	0.03 - 271.55

^a AMI: The AMI extends from the LoQ to the upper limit of quantitation (ULoQ). This is determined by the range of values in U/L (μkat/L) that demonstrated acceptable performance for linearity, imprecision, and bias.

^bEMI: The EMI extends from the ULoQ to the ULoQ × dilution factor.

^cThe reportable interval extends from the LoD to the upper limit of the EMI.

NOTE: The default Low Linearity value of the assay file corresponds to the lower limit of the reportable interval.

LIMITATIONS OF THE PROCEDURE

- Results should be used in conjunction with other data; e.g., symptoms, results of other tests, and clinical impressions.
- Avoid hemolyzed samples due to potential interference. Specimens with hemoglobin levels greater than 150 mg/dL may cause falsely elevated results with the Alanine Aminotransferase2 assay. Refer to the SPECIFIC PERFORMANCE CHARACTERISTICS, Analytical Specificity, Interference section of this package insert.
- Specimens with Sulfasalazine levels greater than 50 mg/L may cause falsely depressed results with the Alanine Aminotransferase2 assay. Refer to the SPECIFIC PERFORMANCE CHARACTERISTICS, Analytical Specificity, Interference section of this package insert.
- Substances that demonstrated interference with the Alanine Aminotransferase2 assay are listed in the SPECIFIC PERFORMANCE CHARACTERISTICS, Analytical Specificity, Interference section of this package insert.
- Potential interference has not been evaluated for substances other than those described in the SPECIFIC PERFORMANCE CHARACTERISTICS, Analytical Specificity, Interference section of this package insert.

EXPECTED VALUES

Manufacturere provided refrence range adopted, effort made to verify locally.

Reference Range¹³

	Range (U/L)	Range ^a (µkat/L)
Adult Male	< 45	< 0.75
Adult Female	< 34	< 0.57

^aAlternate result units were calculated by Abbott.

SPECIFIC PERFORMANCE CHARACTERISTICS

Representative performance data are provided in this section. Results obtained in individual laboratories may vary.

The Alinity c system and the ARCHITECT c System utilize the same reagents and sample/reagent ratios.

Unless otherwise specified, all studies were performed on the Alinity c system.

Unless otherwise specified, the study results provided in this package insert were generated using the Calibration method.

Precision

Within-Laboratory Precision

A study was performed based on guidance from CLSI EP05-A3.¹⁴ Testing was conducted using 3 lots of the Alanine Aminotransferase2 reagents, 3 lots of the Consolidated Chemistry Calibrator, 1 lot of commercially available controls, and 1 instrument. Two controls and 4 serum panels were tested in a minimum of 2 replicates, twice per day for at least 20 days on 3 reagent lot/calibrator lot combinations, where a unique reagent lot and a unique calibrator lot are paired. The performance from a representative combination is shown in the following table.

Sample	n	Mean (U/L)	Within-Run (Repeatability)		Within-Laboratory ^a	
			SD	%CV	SD (Range ^b)	%CV (Range ^b)
Control Level 1	126	33	0.5	1.6	0.9 (0.9 - 0.9)	2.7 (2.6 - 2.7)
Control Level 2	126	101	0.5	0.5	1.8 (1.8 - 1.9)	1.8 (1.8 - 1.9)
Panel A	126	8	0.5	6.7	0.6 (0.5 - 0.6)	7.1 (6.7 - 7.5)
Panel B	126	18	0.5	3.1	0.5 (0.5 - 0.8)	3.1 (3.1 - 4.4)
Panel C	126	362	4.1	1.1	4.2 (2.1 - 4.2)	1.2 (0.6 - 1.2)
Panel D	126	2902	7.5	0.3	14.9 (14.9 - 27.1)	0.5 (0.5 - 0.9)

^aIncludes within-run, between-run, and between-day variability.

^b Minimum and maximum SD or %CV across the 3 reagent lot/calibrator lot combinations.

Sample	n	Mean (μ kat/L)	Within-Run (Repeatability)		Within-Laboratory ^a	
			SD	%CV	SD (Range ^b)	%CV (Range ^b)

Sample	n	Mean (μ kat/L)	Within-Run (Repeatability)		Within-Laboratory ^a	
			SD	%CV	SD (Range ^b)	%CV (Range ^b)
Control Level 1	126	0.55	0.009	1.6	0.015 (0.014 - 0.016)	2.7 (2.5 - 2.8)
Control Level 2	126	1.68	0.009	0.6	0.032 (0.031 - 0.032)	1.9 (1.9 - 1.9)
Panel A	126	0.13	0.008	6.0	0.008 (0.008 - 0.009)	6.4 (6.4 - 6.8)
Panel B	126	0.29	0.008	2.7	0.008 (0.008 - 0.013)	2.8 (2.8 - 4.5)
Panel C	126	6.03	0.068	1.1	0.070 (0.035 - 0.070)	1.2 (0.6 - 1.2)
Panel D	126	48.38	0.125	0.3	0.249 (0.249 - 0.451)	0.5 (0.5 - 0.9)

^aIncludes within-run, between-run, and between-day variability.

^b Minimum and maximum SD or %CV across the 3 reagent lot/calibrator lot combinations.

Accuracy

A study was performed to estimate the bias of the Alanine Aminotransferase2 assay relative to standard reference material (ERM-AD454k/IFCC).

Calibration Method

Testing was conducted using 3 lots of the Alanine Aminotransferase2 reagents, 1 lot of the Consolidated Chemistry Calibrator, and 1 instrument. The bias ranged from 4.3% to 5.4% across all reagent lots.

Calibration Factor Method

Testing was conducted using 3 lots of the Alanine Aminotransferase2 reagents and 1 instrument. The bias ranged from -0.6% to 2.0% across all reagent lots.

Lower Limits of Measurement

A study was performed based on guidance from CLSI EP17-A2.[15](#) Testing was conducted using 3 lots of the Alanine Aminotransferase2 reagents on each of 2 instruments over a minimum of 3 days. The limit of blank (LoB), limit of detection (LoD), and limit of quantitation (LoQ) values are summarized below. These representative data support the lower limit of the analytical measuring interval.

	U/L	μkat/L
LoB ^a	1	0.02
LoD ^b	2	0.03
LoQ ^c	7	0.12

^aThe LoB represents the 95th percentile from $n \geq 60$ replicates of zero-analyte samples.

^bThe LoD represents the lowest concentration at which the analyte can be detected with 95% probability based on $n \geq 60$ replicates of low-analyte level samples.

^c The LoQ presented in the table is in alignment with the low end of the AMI for the Alanine Aminotransferase2 assay on the ARCHITECT c System. The observed LoQ on the Alinity c system was 4 U/L (0.07 μkat/L). This LoQ was generated using the Calibration method and is defined as the lowest concentration at which a maximum allowable precision of 20 %CV was met and was determined from $n \geq 60$ replicates of low-analyte level samples.

Linearity

A study was performed based on guidance from CLSI EP06-A.[16](#)

This assay is linear across the analytical measuring interval of 7 to 3258 U/L (0.12 to 54.31 μkat/L).

Analytical Specificity

Interference

These studies were performed on the ARCHITECT c System.

Potentially Interfering Endogenous Substances

A study was performed based on guidance from CLSI EP07, 3rd ed.[17](#) Each substance was tested at 2 levels of the analyte (approximately 30 U/L and 150 U/L).

No significant interference (interference within $\pm 10\%$) was observed at the following concentrations.

No Significant Interference (Interference within $\pm 10\%$)		
Potentially Interfering Substance	Interferent Level	
	Default Units	Alternate Units
Bilirubin - conjugated	60 mg/dL	712 $\mu\text{mol/L}$
Bilirubin - unconjugated	60 mg/dL	1026 $\mu\text{mol/L}$
Hemoglobin	150 mg/dL	1.5 g/L
Total protein	15 g/dL	150 g/L
Triglycerides	1500 mg/dL	17 mmol/L

Interference beyond $\pm 10\%$ (based on 95% Confidence Interval [CI]) was observed at the concentration shown below for the following substance.

Interference beyond $\pm 10\%$ (based on 95% Confidence Interval [CI])					
Potentially Interfering Substance	Interferent Level		Analyte Level		% Interference (95% CI)
	Default Units	Alternate Units	Default Units	Alternate Units	
Hemoglobin	250 mg/dL	2.5 g/L	30 U/L	0.50 $\mu\text{kat/L}$	13% (11%, 15%)

Potentially Interfering Exogenous Substances

A study was performed based on guidance from CLSI EP07, 3rd ed.[17](#) Each substance was tested at 2 levels of the analyte (approximately 30 U/L and 150 U/L).

No significant interference (interference within $\pm 10\%$) was observed at the following concentrations.

No Significant Interference (Interference within $\pm 10\%$)		
Potentially Interfering Substance	Interferent Level	
	Default Units	Alternate Units
3-methyl-(triazene-1-yl)imidazole-4-carboxamide (MTIC)	0.6 mg/L	3.6 $\mu\text{mol/L}$
5-amino-4-imidazolecarboxamide (AIC)	3 mg/L	24 $\mu\text{mol/L}$
Acetaminophen	160 mg/L	1059 $\mu\text{mol/L}$
Acetylcysteine	150 mg/L	920 $\mu\text{mol/L}$
Acetylsalicylic acid	30 mg/L	167 $\mu\text{mol/L}$
Aminosalicylic acid (p-Aminosalicylic acid)	100 mg/dL	6540 $\mu\text{mol/L}$
Ampicillin-Na	80 mg/L	215 $\mu\text{mol/L}$
Ascorbic acid	60 mg/L	341 $\mu\text{mol/L}$
Biotin	4250 ng/mL	17 $\mu\text{mol/L}$
Ca-dobesilate	60 mg/L	143 $\mu\text{mol/L}$
Cefoxitin	6600 mg/L	15 mmol/L
Chlordiazepoxide	1 mg/dL	33 $\mu\text{mol/L}$
Cyclosporine	2 mg/L	1.7 $\mu\text{mol/L}$
Doxycycline	20 mg/L	45 $\mu\text{mol/L}$
Furosemide	2 mg/dL	60 $\mu\text{mol/L}$
Hydroxocobalamin (Cyanokit)	1250 mg/L	929 $\mu\text{mol/L}$
Ibuprofen	220 mg/L	1067 $\mu\text{mol/L}$
Isoniazid	6 mg/dL	437 $\mu\text{mol/L}$
Levodopa	8 mg/L	41 $\mu\text{mol/L}$
Methotrexate	140 mg/dL	3080 $\mu\text{mol/L}$
Methyldopa	25 mg/L	118 $\mu\text{mol/L}$
Metronidazole	130 mg/L	759 $\mu\text{mol/L}$
Phenylbutazone	330 mg/L	1069 $\mu\text{mol/L}$

No Significant Interference (Interference within $\pm 10\%$)

Potentially Interfering Substance	Interferent Level	
	Default Units	Alternate Units
Rifampicin	50 mg/L	61 $\mu\text{mol/L}$
Sodium heparin	4 U/mL	N/A
Sulfapyridine	300 mg/L	1203 $\mu\text{mol/L}$
Sulfasalazine	50 mg/L	126 $\mu\text{mol/L}$
Suramin	50 mg/dL	386 $\mu\text{mol/L}$
Temozolomide	20 mg/L	103 $\mu\text{mol/L}$
Theophylline (1,3-dimethylxanthine)	60 mg/L	333 $\mu\text{mol/L}$
Vigabatrin	11 mg/dL	852 $\mu\text{mol/L}$

N/A= Not Applicable

Interference beyond $\pm 10\%$ (based on 95% Confidence Interval [CI]) was observed at the concentrations shown below for the following substances.

Potentially Interfering Substance	Interferent Level		Analyte Level		% Interference (95% CI)
	Default Units	Alternate Units	Default Units	Alternate Units	
Hydroxocobalamin (Cyanokit)	1430 mg/L ^a	1062 $\mu\text{mol/L}$	30 U/L	0.50 $\mu\text{kat/L}$	-12% (-14%, -11%)
Sulfasalazine	75 mg/L	188 $\mu\text{mol/L}$	30 U/L	0.50 $\mu\text{kat/L}$	-14% (-16%, -12%)

^a The Hydroxocobalamin (Cyanokit) interferent level presented in the table was generated using the Calibration Factor method.

Interferences from medication or endogenous substances may affect results.[18](#)

Method Comparison

A study was performed based on guidance from CLSI EP09-A3[19](#) using the Passing-Bablok regression method.

Alanine Aminotransferase2 vs Alanine Aminotransferase on the ARCHITECT c System

	n	Units	Correlation Coefficient	Intercept	Slope	Concentration Range
Serum	113	U/L	1.00	1	1.05	7 - 3264
		(μ kat/L)		(0.02)	(1.06)	(0.12 - 54.41)

Alanine Aminotransferase2 on the Alinity c system vs Alanine Aminotransferase2 on the ARCHITECT c System

	n	Units	Correlation Coefficient	Intercept	Slope	Concentration Range
Serum	109	U/L	1.00	0	0.99	8 - 3075
		(μ kat/L)		(0.01)		(0.13 - 51.25)

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