BIMM-143: INTRODUCTION TO BIOINFORMATICS

Professor Barry J. Grant

Find A Gene Final Project

Garrett Cole | A15988021 g1cole@ucsd.edu

[Q1] Tell me the name of a protein you are interested in. Include the species and the accession number. This can be a human protein or a protein from any other species as long as it's function is known. If you do not have a favorite protein, select human RBP4 or KIF11. Do not use beta globin as this is in the worked example report that I provide you with online.

Name: Malate Dehydrogenase (mitochondrial isoform 2 precursor)

Accession: NP_001269332 Species: Homo Sapiens

Functions in Catalytic Activity using both NADP+ or NAD+ as cofactors to increase the catalyzation

rate of interconversion among the acids oxaloacetate and malate

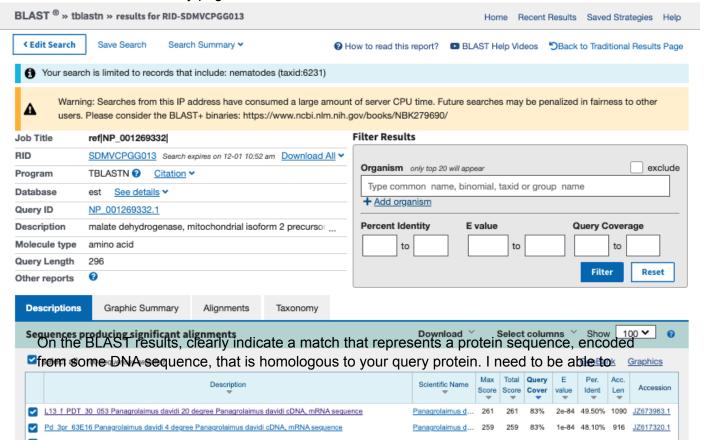
[Q2] Perform a BLAST search against a DNA database, such as a database consisting of genomic DNA or ESTs. The BLAST server can be at NCBI or elsewhere. Include details of the BLAST method used, database searched and any limits applied (e.g. Organism).

Method: TBLASTN search against nematode ESTs

Database: Expressed Sequence Tags (est)

Organism: Nematodes (TaxID: 6231)

Also include the output of that BLAST search in your document. If appropriate, change the font to Courier size 10 so that the results are displayed neatly. You can also screen capture a BLAST output (e.g. alt print screen on a PC or on a MAC press \mathbb{H}-shift-4. The pointer becomes a bulls eye. Select the area you wish to capture and release. The image is saved as a file called Screen Shot [].png in your Desktop directory). It is not necessary to print out all of the blast results if there are many pages.



inspect the pairwise alignment you have selected, including the E value and score. It should be labeled a "genomic clone" or "mRNA sequence", etc. - but include no functional annotation.

Chosen match: Accession JZ673983.1, a L13_f_PDT_30_053, 1090 base pair 20 degree cDNA from *Panagrolaimus davidi*

Alignment details:

≜ Download ✓ GenBank Graphics

L13_f_PDT_30_053 Panagrolaimus davidi 20 degree Panagrolaimus davidi cDNA, mRNA sequence Sequence ID: JZ673983.1 Length: 1090 Number of Matches: 1

Range 1	l: 52 t	945 GenBank Graphics		▼	Next Match A	Previous N
Score		Expect Method	Identities	Positives	Gaps	Frame
261 bits	s(667)	2e-84 Compositional matrix adjust	148/299(49%)	183/299(61%)	52/299(17%	6) +1
\uery	20	SAQNNAKVAVLGASGGIGQPLSLI SA+N + KVA+LGASGGIGQPL LI				76
bjct	52	SARNTSSAPKVALLGASGGIGQPLGLI				231
uery	77	VKGYLGPEQLPDCLKGCDVVVIPAGVI V + G L+G D+VVIPAGVI				136
bjct	232	VTAHTGXXXXHSALEGADIVVIPAGVI	RKPGMTRDDLFN	VNAGIVRDLAEA	AAKACPKAF	411
uery	137	ICVIANP++I NP			DPARVNVPV D ++ +PV	154
bjct	412	VAIITNPVNSTVPIAAEVYKNNGVYDI	KRIFGVTTLDVV	RSQAFIAELKKL	DVSKTVIPV	591
uery	155	IGGHAGKTIIPLISQCTPKVDFPQDQI IGGH+G TIIPL+SQC P F +-				214
bjct	592	IGGHSGVTIIPLLSQCQPSAQFSDSE:				771
uery	215	RFVFSLVDAMNGKEGVVECSFVKSQE	TE-CTYFSTPLLL + YFSTPL L		NLGIGKVS NL K+S	267
bjct	772	RFVDALISGLQGKK-TVQCAYVQSDV				945

In general, [Q2] is the most difficult for students because it requires you to have a "feel" for how to interpret BLAST results. You need to distinguish between a perfect match to your query (i.e. a sequence that is not "novel"), a near match (something that might be "novel", depending on the results of [Q4]), and a non-homologous result. If you are having trouble finding a novel gene try restricting your search to an organism that is poorly annotated.

[Q3] Gather information about this "novel" **protein**. At a minimum, show me the protein sequence of the "novel" protein as displayed in your BLAST results from [Q2] as FASTA format (you can copy and paste the aligned sequence subject lines from your BLAST result page if necessary) or translate your novel DNA sequence using a tool called EMBOSS Transeq at the EBI. Don't forget to translate all six reading frames; the ORF (open reading frame) is likely to be the longest sequence without a stop codon. It may not start with a methionine if you don't have the complete coding region. Make sure the sequence you provide includes a header/subject line and is in traditional FASTA format.

Chosen Sequence:

>52-945_1 L13_f_PDT_30_053 Panagrolaimus davidi 20 degree Panagrolaimus davidi cDNA, mRNA sequence SARNTSSAPKVALLGASGGIGQPLGLLLKTNPKVASLALYDVANTAGVGADLSHIDTHAQ VTAHTGXXXXHSALEGADIVVIPAGVPRKPGMTRDDLFNVNAGIVRDLAEAAAKACPKAF VAIITNPVNSTVPIAAEVYKNNGVYDPKRIFGVTTLDVVRSQAFIAELKKLDVSKTVIPV IGGHSGVTIIPLLSQCQPSAQFSDSEIEKLTARIQDAGTEVVKAKAGAGSATLSMAFAGA RFVDALISGLQGKKTVQCAYVQSDVVKGVDYFSTPLELEPNGVEKFLKTVNLXFMKIS

Here, tell me the name of the novel protein, and the species from which it derives. It is very unlikely (but still definitely possible) that you will find a novel gene from an organism such as S. cerevisiae, human or mouse, because those genomes have already been thoroughly annotated. It is more likely that you will discover a new gene in a genome that is currently being sequenced, such as bacteria or plants or protozoa.

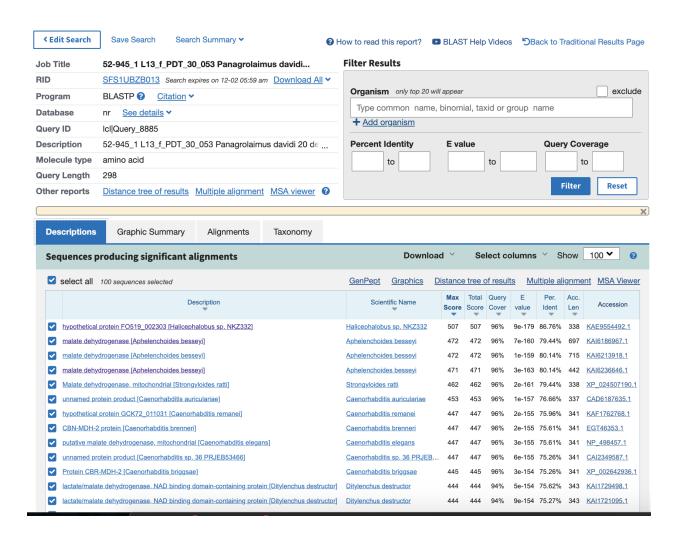
Name: Malate Dehydrogenase Species: *Panagrolaimus*

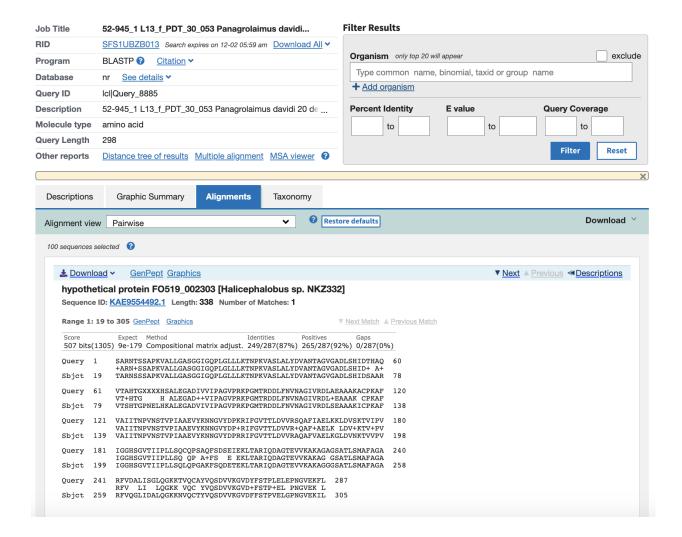
[Q4] Prove that this gene, and its corresponding protein, are novel. For the purposes of this project, "novel" is defined as follows. Take the protein sequence (your answer to [Q3]), and use it as a query in a blastp search of the nr database at NCBI.

- If there is a match with 100% amino acid identity to a protein in the database, from the same species, then your protein is NOT novel (even if the match is to a protein with a name such as "unknown"). Someone has already found and annotated this sequence, and assigned it an accession number.
- If the top match reported has less than 100% identity, then it is likely that your protein is novel, and you have succeeded.
- If there is a match with 100% identity, but to a different species than the one you started with, then you have likely succeeded in finding a novel gene.
- If there are no database matches to the original query from [Q1], this indicates that you have partially succeeded: yes, you may have found a new gene, but no, it is not actually homologous to the original query. You should probably start over.

Details:

A BLASTP search against the NR database produced a top hit from the Halicephalobus (Panagrolaimidae) species. Output details below:





[Q5] Generate a multiple sequence alignment with your novel protein, your original query protein, and a group of other members of this family from different species. A typical number of proteins to use in a multiple sequence alignment for this assignment purpose is a minimum of 5 and a maximum of 20 - although the exact number is up to you. Include the multiple sequence alignment in your report. Use Courier font with a size appropriate to fit page width.

Side-note: Indicate your sequence in the alignment by choosing an appropriate name for each sequence in the input unaligned sequence file (i.e. edit the sequence file so that the species, or short common, names (rather than accession numbers) display in the output alignment and in the subsequent answers below). The goal in this step is to create an interesting an alignment for building a phylogenetic tree that illustrates species divergence.

Labeled Sequences for Alignment:

Panagrolaimus davidi (Antarctic Nematode):

> Antartic_MDH2 | 52-945_1 L13_f_PDT_30_053 Panagrolaimus davidi 20 degree Panagrolaimus davidi cDNA, mRNA sequence

SARNTSSAPKVALLGASGGIGQPLGLLLKTNPKVASLALYDVANTAGVGADLSHIDTHAQVTAHTGXXXXHSAL EGADIVVIPAGVPRKPGMTRDDLFNVNAGIVRDLAEAAAKACPKAFVAIITNPVNSTVPIAAEVYKNNGVYDPK RIFGVTTLDVVRSQAFIAELKKLDVSKTVIPVIGGHSGVTIIPLLSQCQPSAQFSDSEIEKLTARIQDAGTEVV KAKAGAGSATLSMAFAGARFVDALISGLQGKKTVQCAYVQSDVVKGVDYFSTPLELEPNGVEKFLKTVNLXFMK IS

Halicephalobus (Panagrolaimidae):

> Halicephalobus_MDH2 | KAE9554492.1:19-305 hypothetical protein F0519_002303 [Halicephalobus sp. NKZ332]

TARNSSSAPKVALLGASGGIGQPLGLLLKTNPKVASLALYDVANTAGVGADLSHIDSAARVTSHTGPNELHKAL EGADVIVIPAGVPRKPGMTRDDLFNVNAGIVRDLSEAAAKICPKAFVAIITNPVNSTVPIAAEVYKNNGVYDPR RIFGVTTLDVVRAQAFVAELKGLDVNKTVVPVIGGHSGVTIIPLLSQLQPGAKFSQDETEKLTARIQDAGTEVV KAKAGGGSATLSMAFAGARFVOGLIDALOGKKNVOCTYVOSDVVKGVDFFSTPVELGPNGVEKIL

Camelus Ferus (Camel):

> Camel_MDH2 | 165-1130_1 PREDICTED: Camelus ferus malate dehydrogenase 2
(MDH2), transcript variant X1, mRNA

FSTSAQNNAKVAVLGASGGIGQPLSLLLKNSPLVSRLTLYDIAHTPGVAADLSHIETRATVKGYLGPEQLPDCL KGCDVVVIPAGVPRKPGMTRDDLFNTNATIVATLTAACAQHCPEAMICIISNPVNSTIPITAEVFKKHGVYNPD KIFGVTTLDIVRANTFVAELKGLDPARVNVPVIGGHAGKTIIPVISQCTPKVDFPQDQLTTLTGRIQEAGTEVV KAKAGAGSATLSMAYAGARFVFSLLDAMNGKEGVVECSFVKSQETDCPYFSTPLLLGKKGIEKNLGIGKISPFE EKMIAEAIPELKASIKKGEEFVKSMK

Apodemus Sylvaticus (Wood Mouse):

> Mouse_MDH2 | 167-1132_1 PREDICTED: Apodemus sylvaticus malate dehydrogenase 2 (LOC127672705), mRNA

FSTSAQNNAKVAVLGASGGIGQPLSLLLKNSPLVSRLTLYDIAHTPGVAADLSHIETRANVKGYLGPEQLPDCL KGCDVVVIPAGVPRKPGMTRDDLFNTNATIVATLTAACAQHCPEAMICIIANPVNSTIPITAEVFKKHGVYNPN KIFGVTTLDIVRANTFVAELKGLDPARVNVPVIGGHAGKTIIPLISQCTPKVDFPQDQLATLTGRIQEAGTEVV KAKAGAGSATLSMAYAGARFVFSLVDAMNGKEGVVECSFVQSKETECTYFSTPLLLGKKGLEKNLGIGKITPFE EKMIAEAIPELKASIKKGEDFVKNMK

Puma concolor (Cougar):

> Cougar_MDH2 | 138-977_1 PREDICTED: Puma concolor malate dehydrogenase 2 (MDH2), transcript variant X2, mRNA

FSTSAQNNAKVAVLGASGGIGQPLSLLLKNSPLVSRLTLYDIAHTPGVAADLSHIETRAAVKGYLGPEQLPDCL KGCDVVVIPAGVPRKPGMTRDDLFNTNASIVATLTAACAQHCPEAMICIISNPGLDPARVNVPVIGGHAGKTII PLISQCTPKVDLPQDQLTAVTGRIQEAGTEVVKAKAGAGSATLSMAYAGARFVFSLVDAINGKEGVVECSFVKS OETDCPYFSTPLLLGKKGIEKNLGIGKISPFEEKMIAEALPELKASIKKGEEFVKNMK

Erinaceus Eeuropaeus (European Hedgehog):

> HedgeHog_MDH2 | 172-1011_1 PREDICTED: Erinaceus europaeus malate dehydrogenase 2 (MDH2), transcript variant X2, mRNA FSTSTQNNAKVAVLGASGGIGQPLSLLLKNSPLVSRLTLYDIAHTPGVAADLSHIETRANVKGYLGPEQLPDCL KGCDVVVVPAGVPRKPGMTRDDLFNTNATIVATLAAACAQHCPEAMICIIANPGLDPARVNVPVIGGHAGKTII PLISQCTPKVDLPQDKLTALTGRIQEAGTEVVQAKAGAGSATLSMAYAGARFVFSLVDAMNGKEGVVECSFVKS QETDCTYFSTPLLLGRKGLEKNLGIGKVTPFEEKMISEAIPELKASIKKGEEFVKNMK

Lipotes vexillifer (Yangtze River Dolphin):

> Dolphin_MDH2 | 49-888_1 PREDICTED: Lipotes vexillifer malate dehydrogenase 2, NAD (mitochondrial) (MDH2), transcript variant X2, mRNA FSTSAQNNAKVAVLGASGGIGQPLSLLLKNSPLVSRLTLYDIAHTPGVAADLSHIETRATVKGYLGPEQLPDCL KGCDVVVIPAGVPRKPGMTRDDLFNTNATIVATLTAACAQHCPEAMICIISNPGLDPARVSVPVIGGHAGKTII PLASQCTPKVDFPQDQLTTLIGRIQEAGTEVVKAKAGAGSATLSMAYAGARFVFSLVDAMNGKEGVVECSFVKS QETDCPFFSTPLLLGKKGIEKNLGIGKISPFEEKMIAEAIPELKASIKKGEEFVKNMK

Myotis lucifugus (Little Brown Bat):

> Bat_MDH2 | 171-1136_1 PREDICTED: Myotis lucifugus malate dehydrogenase 2 (MDH2), transcript variant X1, mRNA

FSTSAQNNAKVAVLGASGGIGQPLSLLLKNSPLVSRLTLYDIAHTPGVAADLSHIETRASVKGYLGPEQLPDCL KGCDLVVIPAGVPRKPGMTRDDLFNTNATIVANLTAACAQNCPEAMICVIANPVNSTIPITSEVFKKHGVYNPN KIFGVTTLDVVRANAFVAELKGLDPARVNVPVIGGHAGKTIIPLISQCTPKVEFPQDQLTTLTGRIQEAGTEVV KAKAGAGSATLSMAYAGARFVFSLLDAINGKEGVVECSFVKSQETDCSYFSTPLLLGKKGIEKNLGIGKISSFE EKMIAEAIPELKASIKKGEDFVKNMK

Carlito syrichta (Philippine tarsier):

> Tarsier_MDH2 | 194-1159_1 PREDICTED: Carlito syrichta malate dehydrogenase 2 (MDH2), transcript variant X1, mRNA

FGTSAQNNAKVAVLGASGGIGQPLSLLLKNSPLVSRLTLYDIAHTPGVAADLSHIETRATVKGYLGPEQLPDCL KGCDVVVIPAGVPRKPGMTRDDLFNTNATIVATLAAACAQHCPEAMICIIANPVNSTIPITAEVFKKHGVYNPN KVFGVTTLDIVRANTFVAELKGLDPARVNVPVIGGHAGKTIIPLISQCTPKVDFPQDQLTALTGRIQEAGTEVV KAKAGAGSATLSMAYAGARFVFSLVDAMNGKEGVVECSFVKSQETDCTYFSTPLLLGKKGLEKNLGIGKVSSFE EKMITEAMPELKASIKKGEEFVKNMK

Alignment (Obtained using MUSCLE via EBI):

CLUSTAL multiple sequence alignment by MUSCLE (3.8)

Bat_MDH2
HedgeHog_MDH2
Cougar_MDH2
Dolphin_MDH2
Camel_MDH2
Mouse_MDH2
Human_MDH2
Tarsier_MDH2
Antartic_MDH2
Halicephalobus MDH2

FSTSAQNNAKVAVLGASGGIGQPLSLLLKNSPLVSRLTLYDIAHTPGVAADLSHIETRAS
FSTSTQNNAKVAVLGASGGIGQPLSLLLKNSPLVSRLTLYDIAHTPGVAADLSHIETRAN
FSTSAQNNAKVAVLGASGGIGQPLSLLLKNSPLVSRLTLYDIAHTPGVAADLSHIETRAA
FSTSAQNNAKVAVLGASGGIGQPLSLLLKNSPLVSRLTLYDIAHTPGVAADLSHIETRAT
FSTSAQNNAKVAVLGASGGIGQPLSLLLKNSPLVSRLTLYDIAHTPGVAADLSHIETRAT
FSTSAQNNAKVAVLGASGGIGQPLSLLLKNSPLVSRLTLYDIAHTPGVAADLSHIETRAN
---SAQNNAKVAVLGASGGIGQPLSLLLKNSPLVSRLTLYDIAHTPGVAADLSHIETKAA
FGTSAQNNAKVAVLGASGGIGQPLSLLLKNSPLVSRLTLYDIAHTPGVAADLSHIETRAT
SARNTSSAPKVALLGASGGIGQPLGLLLKTNPKVASLALYDVANTAGVGADLSHIDTHAQ
TARNSSSAPKVALLGASGGIGQPLGLLLKTNPKVASLALYDVANTAGVGADLSHIDSAAR

Bat_MDH2
HedgeHog_MDH2
Cougar_MDH2
Dolphin_MDH2
Camel_MDH2
Mouse_MDH2
Human_MDH2
Tarsier_MDH2
Antartic_MDH2
Halicephalobus MDH2

Bat_MDH2
HedgeHog_MDH2
Cougar_MDH2
Dolphin_MDH2
Camel_MDH2
Mouse_MDH2
Human_MDH2
Tarsier_MDH2
Antartic_MDH2
Halicephalobus MDH2

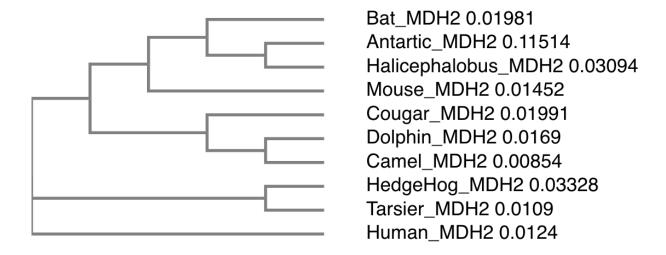
Bat_MDH2
HedgeHog_MDH2
Cougar_MDH2
Dolphin_MDH2
Camel_MDH2
Mouse_MDH2
Human_MDH2
Tarsier_MDH2
Antartic_MDH2
Halicephalobus MDH2

Bat_MDH2
HedgeHog_MDH2
Cougar_MDH2
Dolphin_MDH2
Camel_MDH2
Mouse_MDH2
Human_MDH2
Tarsier_MDH2
Antartic_MDH2
Halicephalobus MDH2

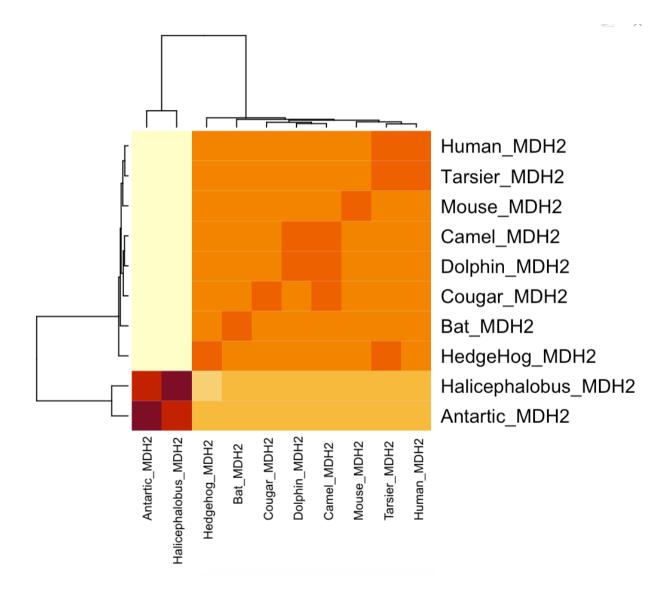
Bat_MDH2
HedgeHog_MDH2
Cougar_MDH2
Dolphin_MDH2
Camel_MDH2
Mouse_MDH2
Human_MDH2
Tarsier_MDH2
Antartic_MDH2
Halicephalobus_MDH2

IAEAIPELKASIKKGEDFVKNMK
ISEAIPELKASIKKGEEFVKNMK
IAEALPELKASIKKGEEFVKNMK
IAEAIPELKASIKKGEEFVKNMK
IAEAIPELKASIKKGEEFVKNMK
IAEAIPELKASIKKGEEFVKNMK
ITEAMPELKASIKKGEEFVKNMK

[Q6] Create a phylogenetic tree, using either a parsimony or distance-based approach. Bootstrapping and tree rooting are optional. Use "simple phylogeny" online from the EBI or any respected phylogeny program (such as MEGA, PAUP, or Phylip). Paste an image of your Cladogram or tree output in your report.



[Q7] Generate a sequence identity based **heatmap** of your aligned sequences using R. If necessary convert your sequence alignment to the ubiquitous FASTA format (Seaview can read in clustal format and "Save as" FASTA format for example). Read this FASTA format alignment into R with the help of functions in the **Bio3D package**. Calculate a sequence identity matrix (again using a function within the Bio3D package). Then generate a heatmap plot and add to your report. Do make sure your labels are visible and not cut at the figure margins.



[Q8] Using R/Bio3D (or an online blast server if you prefer), search the main protein structure database for the most similar atomic resolution structures to your aligned Sequences.

List the top 3 unique hits (i.e. not hits representing different chains from the same structure) along with their Evalue and sequence identity to your query. Please also add annotation details of these structures. For example include the annotation terms PDB identifier (structureId), Method used to solve the structure (experimentalTechnique), resolution (resolution), and source organism (source).

HINT: You can use a single sequence from your alignment or generate a consensus sequence from your alignment using the Bio3D function consensus(). The Bio3D functions blast.pdb(), plot.blast() and pdb.annotate() are likely to be of most relevance

for completing this task. Note that the results of blast.pdb() contain the hits PDB identifier (or pdb.id) as well as Evalue and identity. The results of pdb.annotate() contain the other annotation terms noted above.

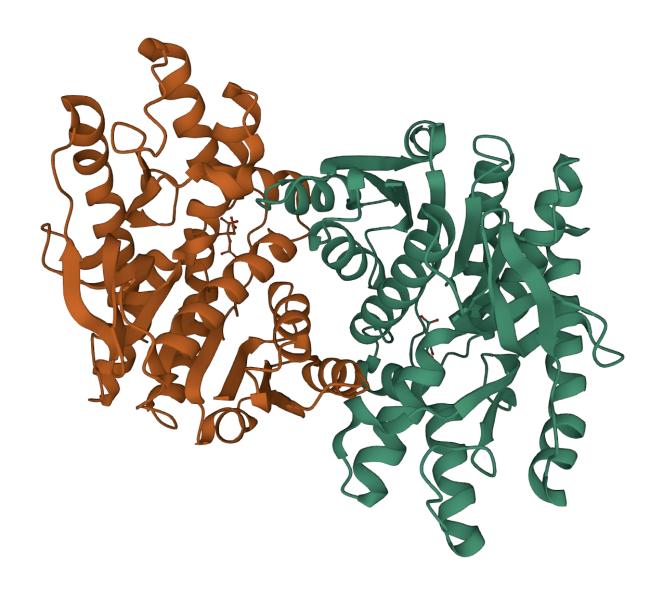
Note that if your consensus sequence has lots of gap positions then it will be better to use an original sequence from the alignment for your search of the PDB. In this case you could chose the sequence with the highest identity to all others in your alignment by calculating the row-wise maximum from your sequence identity matrix.

ID	Technique	Resolution	Source	Evalue	Identity
1MLD	X-RAY DIFFRACTION	1.83 Å	Sus scrofa (Wild Boar)	0	63%
4E0B	X-RAY DIFFRACTION	2.17 Å	Vibrio vulnificus (Bacteria)	0	60%
1SMK	X-RAY DIFFRACTION	2.50 Å	Citrullus lanatus (Watermelon)	0	56%

[Q9] Generate a molecular figure of one of your identified PDB structures using the NGL viewer online (or VMD/PyMol). You can optionally highlight conserved residues that are likely to be functional. Please use a white or transparent background for your figure (i.e. not the default black).

Based on sequence similarity. How likely is this structure to be similar to your "novel" protein?

Based on the sequence similarity, I believe the structure to be somewhat similar to my "novel" protein since the sequence similarity is greater than 60%.



[Q10] Perform a "Target" search of ChEMBEL (https://www.ebi.ac.uk/chembl/) with your novel sequence. Are there any Target Associated Assays and ligand efficiency data reported that may be useful starting points for exploring potential inhibition of your novel protein?

The "Target" search via ChEMBEL outputted 2 Functional Assay (CHEMBL614281, CHEMBL2366649) and 5 Binding Assay (CHEMBL2095180, CHEMBL2189156, CHEMBL2242736, CHEMBL2326, CHEMBL2216)

Only three Binding Assay had ligand efficiency data, which is reported as follows:

CHEMBL2095180 | BEI: 27 | SEI: 6.57 CHEMBL2326 | BEI: 27.69 | SEI: 9.20 CHEMBL2216 | BEI: 35.08 | SEI: 6.78

Scoring Rubric:

[45 total points available]

Od (4 m sinds)		
Q1 (4 points) Protein name	1	1
Species	1	1
Accession number	1	1
Function known	0.5	1
		
Q2 (6 points)		
Blast method	1	1
Database searched	1	1
Limits applied	1	1
Search output list (top hits)	1	1
Alignment of choice	1	1
Evalue and other alignment stats	1	1
Q3 (3 points)	4	4
Protein sequence of choice matches Subject above Name in header	1	1
	1	1 1
Species	I	1
Q4 (3 point)		
Blastp output list with identities & Evalue	1	1
Top alignment shown with alignment statistics	1	1
Results indicates a "novel" gene found	1	1
ŭ		
Q5 (3 points)		
MSA labeled with useful names	1	1
MSA trimmed appropriately (i.e. no gap overhangs)	1	1
Pasted MSA fits report page width (i.e. font, format)	1	1
Q6 (1 point)		
Figure illustrates sequence clustering pattern	1	1
07 (40 int-)		
Q7 (10 points)	-	_
Heatmap figure included in report	5	5
Heatmap is legible (i.e. no labels obscured)	5	5
Q8 (10 points)		
PDB identifiers from multiple species reported	5	5
Annotation of PDB source, resolution and technique	5	4
tatatan ar r 22 addres, recordion and teeningue	.	•

Final Score:	44.5	1	45 = 98.8%
Evidence of ChEMBEL searches	1		1
Q10 (1 point)			
Figure of high resolution (i.e. not just snapshot)	1		1
Uses white background for molecular figure	1		1
Structure figure provided	2		2
Q9 (4 points)			
Annotation of Evalue and Sequence Identity	1		1