

Worksheet-7a

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```
#Worksheet7a #Gabby
```

```
install.packages("Hmisc")
install.packages("pastecs")
```

```
#1. Create a data frame for the table below
```

```
Student <- seq(1:10)
PreTest <- c(55,54,47,57,51,61,57,54,63,58)
PostTest <- c(61,60,56,63,56,63,59,56,62,61)
```

```
DF <- data.frame(Student,PreTest,PostTest)
DF
```

```
#a. Compute the descriptive statistics using different packages (Hmisc and pastecs).
```

```
#Write the codes and its result.
```

```
library(Hmisc)
library(pastecs)
```

```
describe(DF)
```

```
stat.desc(DF)
```

```
#2. The Department of Agriculture was studying the effects of several levels of a #fertilizer on the growth of a plant. For some analyses, it might be useful to convert #the fertilizer levels to an ordered factor.
```

```
DepartmentofAgriculture <- c(10,10,10,20,20,50,10,
                             20,10,50,20,50,20,10)
```

```
#a. Write the codes and describe the result.
```

```
In_Ord <- sort(DepartmentofAgriculture, decreasing = FALSE)
In_Ord
```

```
#3. Abdul Hassan, president of Floor Coverings Unlimited, has asked you to study #the exercise levels undertaken by 10 subjects were "l", "n", "n", "i", "l", "l", "n", "n", "i", "l"; n=none, l=light, i=intense
```

```
Subjects <- c("l","n","n","i","l","l","n","n","i","l")
```

#a. What is the best way to represent this in R?

#DATAFRAME

```
out <- data.frame(Subjects)
out
```

#4. Sample of 30 tax accountants from all the states and territories of Australia and #their individual state of origin is specified by a character vector of state mnemonics #as:

```
state <- c("tas", "sa", "qld", "nsw", "nsw", "nt", "wa", "wa", "qld",
          "vic", "nsw", "vic", "qld", "qld", "sa", "tas", "sa", "nt",
          "wa", "vic", "qld", "nsw", "nsw", "wa", "sa", "act", "nsw",
          "vic", "vic", "act")
state
```

#a. Apply the factor function and factor level. Describe the results.

```
hello <- function(state)
hello
```

#5. From #4 - continuation:

• Suppose we have the incomes of the same tax accountants in another vector (in incomes <- c(60, 49, 40, 61, 64, 60, 59, 54, 62, 69, 70, 42, 56, 61, 61, 61, 58, 51, 48, 65, 49, 49, 41, 48, 52, 46, 59, 46, 58, 43))

#a. Calculate the sample mean income for each state we can now use the special #function tapply():

```
Calc <- tapply(state, incomes, mean)
Calc
```

#b. Copy the results and interpret.

40 41 42 43 46 48 49 51 52 54 56 58 59 60 61 62 64 65 69 70

#NA NA NA NA NA NA NA NA NA NA NA NA NA NA NA NA NA NA NA

#6. Calculate the standard errors of the state income means (refer again to number 3)

```
Calc_ST.n <- length(Calc)
Calc_1.sd <- sd(Calc)
Calc_Final.se <- Calc_1.sd/sqrt(Calc_ST.n)
Calc_Final.se
```

#a. What is the standard error? Write the codes. #NA #b. Interpret the result. #the result is not available because some variables are character type so it won't be able to get the standard error. #7. Use the titanic dataset.

```
data("Titanic")
```

```
head<- data.frame(Titanic)
```

#a. subset the titanic dataset of those who survived and not survived. Show the #codes and its result.

```
head_subset <- subset(head, select = "Survived")
head_subset
```

#8. The data sets are about the breast cancer Wisconsin. The samples arrive periodically as Dr. Wolberg reports his clinical cases. The database therefore reflects this #chronological grouping of the data. You can create this dataset in Microsoft Excel.

#a. describe what is the dataset all about. #The dataset s all about Breast Cancer.

#b. Import the data from MS Excel. Copy the codes.

```
library("readxl")
```

```
DATA <- read_excel("C:\\Users\\User\\Desktop\\Gabby1\\Worksheets\\Worksheet#7\\Breast_Cancer.xlsx")
DATA
```

#c. Compute the descriptive statistics using different packages. Find the values of:

#c.1 Standard error of the mean for clump thickness.

```
Clump <- length(DATA$'CL. thickness')
Clump_A <- sd(DATA$'CL. thickness')
Clump_B <- Clump_A/sqrt(DATA$'CL. thickness')
Clump_B
```

#c.2 Coefficient of variability for Marginal Adhesion.

```
COV <- sd(DATA$'Marg. Adhesion') / mean(DATA$'Marg. Adhesion')* 100
COV
```

#c.3 Number of null values of Bare Nuclei.

```
Null_Values <- subset(DATA,'Bare. Nuclei' == "NA")
```

#c.4 Mean and standard deviation for Bland Chromatin

```
mean(DATA$'Bl. Chromatin')
sd(DATA$'Bl. Chromatin')
```

#c.5 Confidence interval of the mean for Uniformity of Cell Shape

#Calculate the mean

```
Calc_Mean <- mean(DATA$'Cell Shape')
Calc_Mean
```

#Calculate the standard error of the mean

```
SE_M <- length(DATA$'Cell Shape')
SD_B <- sd(DATA$'Cell Shape')
Ans_1 <- SD_B/sqrt(SE_M)
Ans_1
```

```

#Find the t-score that corresponds to the confidence level

D = 0.05
numE = SE_M - 1
numF = qt(p = D/ 2, df = numE,lower.tail = F)
numF

#Constructing the confidence interval

numG <- numF * numC

#Lower

numH <- Calc_Mean - numG

#Upper

numI <- Calc_Mean + numG

c(numH,numI)

#d. How many attributes?

attributes(DATA)

#e. Find the percentage of respondents who are malignant. Interpret the results.

P_R <- subset(DATA, Class == "malignant")
P_R

#There 17 respondents who are malignant. #And there are total of 49 respondent.
#Getting the percentage

17 / 49 * 100

#9. Export the data abalone to the Microsoft excel file. Copy the codes.

install.packages("AppliedPredictiveModeling")

library("AppliedPredictiveModeling")
data("abalone")
View(abalone)
head(abalone)
summary(abalone)

#Exporting the data abalone to the Microsoft excel file install.packages("xlsxjars")

library(xlsx)
write.xlsx("abalone", "C:\\Users\\User\\Desktop\\Gabby1\\Worksheets\\Worksheet#7\\abalone.xlsx")

```