

# Package ‘DMRcompare’

July 30, 2018

**Title** Compare Different R Tools for Detecting Differentially-Methylated Regions (DMRs) of a Genome

**Version** 0.0.0.9000

**Description** The accompanying package to the method comparison paper “An evaluation of supervised methods for identifying differentially methylated regions in epigenome-wide association studies” by Mallik et al. (2018), submitted to Briefings in Bioinformatics as a review article. This package contains the organized and documented R scripts necessary to replicate the multi-design-point comparative simulation study, as well as associated tables and figures.

**Depends** R (>= 3.3.0),  
ChAMPdata,  
DMRcatedata

**Imports** bumphunter,  
ChAMP,  
ChIPpeakAnno,  
data.table,  
doParallel,  
DMRcate,  
foreach,  
GenomicRanges,  
graphics,  
grDevices,  
IRanges,  
minfi,  
parallel,  
PRROC,  
stats

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**Encoding** UTF-8

**LazyData** true

**RoxygenNote** 6.0.1

**Suggests** GEOquery,  
testthat

## R topics documented:

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betaVals_mat	<i>Annotated CPG Data Set</i>
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## Description

A beta value matrix for selected methylation samples from a 450k methylation array. These beta values are trimmed to range from 0.05 to 0.95.

## Usage

```
betaVals_mat
```

## Format

A matrix containing 356603 CPGs measured on 14 subjects. The rownames are the CPG IDs, and the column names indicate the sample IDs of the subjects.

## Source

Calculated via the 1\_Aclust\_data\_import.R script in the old\_scripts sub-directory of the inst directory.

**Description**

Given a directory of best-performing results files from one of the simulation functions ([WriteDMRcateResults](#), [WriteProbeLassoResults](#), [WriteBumphunterResults](#), or results from the Comb-p method in Python), import the raw data files and construct PR-curve objects via the [pr.curve](#) function.

**Usage**

```
BuildPRcurve(bestResultsDir, delta = c(0.025, 0.05, 0.1, 0.15, 0.2, 0.3, 0.4),
  seed = c(100, 210, 330, 450, 680), beta_mat = betaVals_mat,
  AclustCPG_df = startEndCPG_df, CPGs_df = cpgLocation_df, min.cpgs = 5)
```

**Arguments**

bestResultsDir	The name of the directory where the method results from the best-performing parameter settings are stored. For the full design we have included ( <code>delta = c(0.025, 0.05, 0.1, 0.15, 0.2, 0.3, 0.4)</code> , and <code>seed = c(100, 210, 330, 450, 680)</code> ), this directory should contain 35 .RDS files per method.
delta	A treatment size corresponding to one of the simulations with completed results files in the bestResultsDir directory.
seed	A seed value corresponding to one of the simulations with completed results files in the bestResultsDir directory.
beta_mat	A beta value matrix for selected methylation samples from a 450k methylation array with CPG IDs as the row names and sample IDs as the column names. An example is given in the betaVals_mat data set.
AclustCPG_df	A data frame of beta values and CpG information for clusters of CpGs over a 450k methylation array. The rows correspond to the CPGs. The columns have information on the cluster number, chromosome, cluster start and end locations, and the beta values for each subject grouped by some clinical indicator (e.g. case v. control). An example is given in the startEndCPG_df data set.
CPGs_df	An annotation table that indicates locations of CpGs. This data frame has CPG IDs as the rows with matching chromosome and location info in the columns. Specifically, the columns are: ILMNID - the CPG ID; chr - the chromosome label; and MAPINFO - the chromosome location. An example is given in the cpgLocation_df data set. This data set is only necessary if the results directory contains Comb-p results with the specified delta and seed values.
min.cpgs	The minimum number of CPGs necessary to consider a result significant. Defaults to 5. This argument is only required if the results directory contains Comb-p results with the specified delta and seed values.

**Value**

A list of PR-curve objects, to be plotted via the [PlotPRCurve](#) function.

## Examples

```
## Not run:
BuildPRcurve(
  bestResultsDir = "best_cases_results/",
  delta = 0.4,
  seed = 100
)

## End(Not run)
```

---

cpgLocation_df	<i>CPG Locations</i>
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## Description

An annotation table that indicates locations of CpGs. This data frame has CPG IDs as the rows with matching chromosome and location info in the columns.

## Usage

```
cpgLocation_df
```

## Format

A data frame containing 485512 CPGs (rows) and three columns. The columns are:

- ILMNID : the CPG ID, as a factor
- chr : the chromosome label, as a character
- MAPINFO : the chromosome location, as an integer

## Source

Compiled via the 1\_Aclust\_data\_import.R script in the old\_scripts sub-directory of the inst directory.

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PlotOverlaps	<i>Plot Venn Diagrams of DMR Overlaps</i>
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---

## Description

Given a directory of best-performing results files from one of the simulation functions ([WriteDMRcateResults](#), [WriteProbeLassoResults](#), [WriteBumphunterResults](#), or results from the Comb-p method in Python), call the [BuildOverlaps](#) function to import the raw data files and DMR overlap lists, then plot those Venn diagrams and save the plots to a PDF.

## Usage

```
PlotOverlaps(bestResultsDir, figFileName, device = pdf,
  plotTitle = "default", delta_num = c(0.025, 0.05, 0.1, 0.15, 0.2, 0.3,
  0.4), seeds_int = c(100, 210, 330, 450, 680), totalTest_int = 3063,
  CPGs_df = cpgLocation_df, min.cpgs = 5, ...)
```

**Arguments**

<code>bestResultsDir</code>	The name of the directory where the method results from the best-performing parameter settings are stored. For the full design we have included ( <code>delta = c(0.025, 0.05, 0.1, 0.2)</code> and <code>seed = c(100, 210, 330, 450, 680)</code> ), this directory should contain 35 .RDS files per method.
<code>figFileName</code>	The name of the figure
<code>device</code>	Which graphics device should be used to save the figures? Defaults to <a href="#">pdf</a> . Note that if you use a device other than PDF ( <a href="#">jpeg</a> for instance), you can only plot one <code>delta_num</code> and <code>seed_int</code> combination per file.
<code>plotTitle</code>	The title of the plot. This argument is passed to the <a href="#">makeVennDiagram</a> function. The default value of "default" will make the plot title "Venn Diagram for mu = DELTA, rep = INDEX OF SEED".
<code>delta_num</code>	A vector of treatment sizes with values corresponding to one of the simulations with completed results files in the <code>bestResultsDir</code> directory.
<code>seeds_int</code>	A vector of random seeds with values corresponding to one of the simulations with completed results files in the <code>bestResultsDir</code> directory.
<code>totalTest_int</code>	Parameter passed to the <a href="#">makeVennDiagram</a> function. This is an interger value specifying the total number of tests performed to obtain the list of peaks. It should be much larger than the number of peaks in the largest peak set.
<code>CPGs_df</code>	An annotation table that indicates locations of CpGs. This data frame has CPG IDs as the rows with matching chromosome and location info in the columns. Specifically, the columns are: <code>ILMNID</code> - the CPG ID; <code>chr</code> - the chromosome label; and <code>MAPINFO</code> - the chromosome location. An example is given in the <code>cpgLocation_df</code> data set. This data set is only necessary if the results directory contains Comb-p results with the specified <code>delta</code> and <code>seed</code> values. This is passed to the <a href="#">BuildOverlaps</a> function.
<code>min.cpgs</code>	The minimum number of CPGs before we consider a result significant. Defaults to 5. This argument is only required if the results directory contains Comb-p results with the specified <code>delta</code> and <code>seed</code> values. This is passed to the <a href="#">BuildOverlaps</a> function.
<code>...</code>	Dots for additional arguments to be passed to the graphics device

**Value**

Nothing. A PDF file of plots is created as a side effect.

**Examples**

```
## Not run:
PlotOverlaps(
  bestResultsDir = "best_cases_results/",
  figFileName = "best_cases_results/resultsFigures/testVenn_allDesigns2"
)

## End(Not run)
```

---

PlotPRCurve	<i>Plot Precision-Recall Curves</i>
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### Description

Given a list of PR-curve objects as returned by the [BuildPRcurve](#) function, plot the precision-recall curve for each method in a shared figure.

### Usage

```
PlotPRCurve(prCurves_ls, plotTitle = "default", new = TRUE, lineWidth = 1,
  colours = NULL)
```

### Arguments

prCurves_ls	A list of PR-curve objects
plotTitle	The title of the plot. The default value of "default" will make the plot title "Venn Diagram for mu = DELTA, rep = INDEX OF SEED".
new	Should the PR curves from this list form their own graph (TRUE) or be added onto a previous PR-curve figure (FALSE). Defaults to TRUE.
lineWidth	The line width of each PR curve in the plot. Defaults to 1.
colours	Optionally add your own colours for each line. Otherwise, the colours are created with the <a href="#">hcl</a> function.

### Value

Nothing. A plot is created as a side effect.

### Examples

```
## Not run:
prCurves_0.4_100_ls <-
  BuildPRcurve(
    bestResultsDir = "best_cases_results/",
    delta = 0.4,
    seed = 100
  )

PlotPRCurve(prCurves_0.4_100_ls)

## End(Not run)
```

---

ProcessBumphunterResults

*Process Bumphunter Results Files*


---

**Description**

Given a directory of saved Bumphunter results, as written by the `WriteBumphunterResults` function, import and summarize these data files.

**Usage**

```
ProcessBumphunterResults(resultsDir, beta_mat, AclustCPG_df, verbose = TRUE)
```

**Arguments**

<code>resultsDir</code>	The name of the directory where the Bumphunter method results are stored. This should match the directory name supplied to the <code>resultsDir</code> argument of the <code>WriteBumphunterResults</code> function.
<code>beta_mat</code>	A beta value matrix for methylation samples from a 450k methylation array with CpG IDs as the row names and sample IDs as the column names. An example is given in the <code>betaVals_mat</code> data set.
<code>AclustCPG_df</code>	A data frame of beta values and CpG information for clusters of CpGs over a 450k methylation array. The rows correspond to the CPGs. The columns have information on the cluster number, chromosome, cluster start and end locations, and the beta values for each subject grouped by some clinical indicator (e.g. case v. control). An example is given in the <code>startEndCPG_df</code> data set. This data set also has information on true status of the clusters, via variable <code>status</code> , with values "positive" or "negative", indicating whether treatment effect was added to the cluster.
<code>verbose</code>	Should the function print progress messages? Defaults to TRUE.

**Value**

A data frame of model performance measures for the Bumphunter method under each of the given parameter combinations applied to the data generated with different treatment effects

**Examples**

```
## Not run:
data("betaVals_mat")
data("startEndCPG_df")

bumphunterRes_df <- ProcessBumphunterResults(
  resultsDir = "DMRcate_results/",
  beta_mat = betaVals_mat,
  AclustCPG_df = startEndCPG_df
)

## End(Not run)
```

---

ProcessCombpResults     *Extract and Process Comb-p Results Files*


---

## Description

Given a directory of saved Comb-p results, as .RDS files, import, standardize, and summarize these data files.

## Usage

```
ProcessCombpResults(resultsDir, beta_mat, AclustCPG_df, cpgLocation_df,
  dmr.sig.threshold = 0.05, min.cpgs = 5, verbose = TRUE)
```

## Arguments

resultsDir	The name of the directory where the Comb-p method results are stored.
beta_mat	A beta value matrix for methylation samples from a 450k methylation array with CpG IDs as the row names and sample IDs as the column names. An example is given in the betaVals_mat data set.
AclustCPG_df	A data frame of beta values and CpG information for clusters of CpGs over a 450k methylation array. The rows correspond to the CPGs. The columns have information on the cluster number, chromosome, cluster start and end locations, and the beta values for each subject grouped by some clinical indicator (e.g. case v. control). An example is given in the startEndCPG_df data set. This data set also has information on true status of the clusters, via variable status, with values "positive" or "negative", indicating whether treatment effect was added to the cluster.
cpgLocation_df	An annotation table that indicates locations of CpGs. This data frame has CpG IDs as the rows with matching chromosome and location info in the columns. Specifically, the columns are: ILMNID - the CPG ID; chr - the chromosome label; and MAPINFO - the chromosome location. An example is given in the cpgLocation_df data set.
dmr.sig.threshold	Regions with DMR p-value less than dmr.sig.threshold are selected for the output.
min.cpgs	Minimum number of CpGs. Regions with at least min.cpgs are selected for the output. Defaults to 5.
verbose	Should the function print progress messages? Defaults to TRUE.

## Value

A data frame of model performance measures for the Comb-p method under each of the given parameter combinations applied to the data generated with different treatment effects

## Examples

```
## Not run:
data("betaVals_mat")
data("startEndCPG_df")
data("cpgLocation_df")
```



```

combpRes_df <- ProcessCombpResults(
  resultsDir = "DMRcate_results/",
  beta_mat = betaVals_mat,
  AclustCPG_df = startEndCPG_df,
  cpgLocation_df = cpgLocation_df
)

## End(Not run)

```

---

## ProcessDMRcateResults *Process DMRcate Results Files*

---

### Description

Given a directory of saved DMRcate results, as written by the [WriteDMRcateResults](#) function, import and summarize these data files.

### Usage

```
ProcessDMRcateResults(resultsDir, beta_mat, AclustCPG_df, verbose = TRUE)
```

### Arguments

resultsDir	The name of the directory where the DMRcate method results are stored. This should match the directory name supplied to the resultsDir argument of the <a href="#">WriteDMRcateResults</a> function.
beta_mat	A beta value matrix for methylation samples from a 450k methylation array with CpG IDs as the row names and sample IDs as the column names. An example is given in the betaVals_mat data set.
AclustCPG_df	A data frame of beta values and CpG information for clusters of CpGs over a 450k methylation array. The rows correspond to the CPGs. The columns have information on the cluster number, chromosome, cluster start and end locations, and the beta values for each subject grouped by some clinical indicator (e.g. case v. control). An example is given in the startEndCPG_df data set. This data set also has information on true status of the clusters, via variable status, with values "positive" or "negative", indicating whether treatment effect was added to the cluster.
verbose	Should the function print progress messages? Defaults to TRUE.

### Value

A data frame of model performance measures for the DMRcate method under each of the given parameter combinations applied to the data generated with different treatment effects

### Examples

```

## Not run:
data("betaVals_mat")
data("startEndCPG_df")

dmrcateRes_df <- ProcessDMRcateResults(

```

```

    resultsDir = "DMRcate_results/",
    beta_mat = betaVals_mat,
    AclustCPG_df = startEndCPG_df
  )

  ## End(Not run)

```

---

## ProcessProbeLassoResults

*Process ProbeLasso Results Files*

---

### Description

Given a directory of saved ProbeLasso results, as written by the [WriteProbeLassoResults](#) function, import and summarize these data files.

### Usage

```
ProcessProbeLassoResults(resultsDir, beta_mat, AclustCPG_df, verbose = TRUE)
```

### Arguments

resultsDir	The name of the directory where the ProbeLasso method results are stored. This should match the directory name supplied to the resultsDir argument of the <a href="#">WriteProbeLassoResults</a> function.
beta_mat	A beta value matrix for methylation samples from a 450k methylation array with CpG IDs as the row names and sample IDs as the column names. An example is given in the betaVals_mat data set.
AclustCPG_df	A data frame of beta values and CpG information for clusters of CpGs over a 450k methylation array. The rows correspond to the CPGs. The columns have information on the cluster number, chromosome, cluster start and end locations, and the beta values for each subject grouped by some clinical indicator (e.g. case v. control). An example is given in the startEndCPG_df data set. This data set also has information on true status of the clusters, via variable status, with values "positive" or "negative", indicating whether treatment effect was added to the cluster.
verbose	Should the function print progress messages? Defaults to TRUE.

### Value

A data frame of model performance measures for the ProbeLasso method under each of the given parameter combinations applied to the data generated with different treatment effects

### Examples

```

## Not run:
data("betaVals_mat")
data("startEndCPG_df")

probeLassoRes_df <- ProcessProbeLassoResults(
  resultsDir = "DMRcate_results/",
  beta_mat = betaVals_mat,

```

```

        AcLustCPG_df = startEndCPG_df
    )

    ## End(Not run)

```

RunBumphunter

*Return Results from the bumphunter Function*

## Description

A wrapper function for the Bumphunter method as implemented in the bumphunter package, called internally by the [WriteBumphunterResults](#) function.

## Usage

```

RunBumphunter(betaVals_mat, labels_fct = factor(c(rep("Tumor", 7),
  rep("Normal", 7))), chromos_char, chromPosit_num, cpgLocation_df,
  pickCutoffQ_num, maxGap_int, B_int = 10, numCores = detectCores() - 1,
  dmr.sig.threshold = 0.05, min.cpgs = 5)

```

## Arguments

betaVals_mat	A matrix of beta values returned in the first entry of the output from the SimulatedData function, ordered by the CpGs. Note this dataset includes all CpGs on the array.
labels_fct	A factor vector of subject class labels. These should match the observations contained in the columns of the betaVals_mat matrix. Defaults to seven "Tumor" followed by seven "Normal" samples.
chromos_char	A character vector for the chromosomes on which the CpGs are located
chromPosit_num	A numeric vector for the locations of the CpGs
cpgLocation_df	An annotation table that indicates locations of CpGs. This data frame has CpG IDs as the rows with matching chromosome and location info in the columns. Specifically, the columns are: ILMNID - the CpG ID; chr - the chromosome label; and MAPINFO - the chromosome location. An example is given in the cpgLocation_df data set.
pickCutoffQ_num	The quantile used for picking the cutoff using the permutation distribution, passed to the <a href="#">bumphunter</a> function.
maxGap_int	The maximum location gap, passed to the <a href="#">bumphunter</a> function.
B_int	An integer denoting the number of resamples to use when computing null distributions, passed to the <a href="#">bumphunter</a> function.
numCores	The number of computing cores for parallel execution, passed to the <a href="#">registerDoParallel</a> function. Defaults to one less than the number of cores available on your machine, as detected via the <a href="#">detectCores</a> function.
dmr.sig.threshold	Regions with DMR p-value less than dmr.sig.threshold are selected for the output.
min.cpgs	Minimum number of CpGs. Regions with at least min.cpgs are selected for the output. Defaults to 5.

**Value**

A list of two elements: a data frame of bumphunter results and the computing time for the bumphunter method.

**Examples**

```
# Called internally by the WriteBumpHunterResults() function.
## Not run:
data("betaVals_mat")
data("cpgLocation_df")
data("startEndCPG_df")

treat_ls <- SimulateData(beta_mat = betaVals_mat,
                        Aclusters_df = startEndCPG_df,
                        delta_num = 0.4,
                        seed_int = 100)
class_fct <- factor(c(rep("Tumor", 7), rep("Normal", 7)))

RunBumpHunter(
  betaVals_mat = treat_ls$simBetaVals_df,
  labels_fct = class_fct,
  cpgLocation_df = cpgLocation_df,
  pickCutoffQ_num = 0.95,
  maxGap_int = 250
)

## End(Not run)
```

RunDMRcate

*Return Results from the dmr cate Function***Description**

A wrapper function for the DMRcate method from the DMRcate package, called internally by the [WriteDMRcateResults](#) function.

**Usage**

```
RunDMRcate(betaVals_mat, labels_fct = factor(c(rep("Tumor", 7), rep("Normal",
7))), cpgLocation_df, lambda_int, C_int, nCores = 1,
dmr.sig.threshold = 0.05, min.cpgs = 5, genome = "hg19")
```

**Arguments**

betaVals_mat	A matrix of beta values returned in the first entry of the output from the SimulateData function, ordered by the CpGs. Note this dataset includes all CpGs on the array.
labels_fct	A factor vector of subject class labels. These should match the observations contained in the columns of the betaVals_mat matrix. Defaults to seven "Tumor" followed by seven "Normal" samples.

<code>cpgLocation_df</code>	An annotation table that indicates locations of CpGs. This data frame has CpG IDs as the rows with matching chromosome and location info in the columns. Specifically, the columns are: <code>ILMNID</code> - the CpG ID; <code>chr</code> - the chromosome label; and <code>MAPINFO</code> - the chromosome location. An example is given in the <code>cpgLocation_df</code> data set.
<code>lambda_int</code>	Gaussian kernel bandwidth for smoothed-function estimation in the called <a href="#">dmrcate</a> function.
<code>C_int</code>	Scaling factor for bandwidth in the internal call to the <a href="#">dmrcate</a> function
<code>nCores</code>	How many cores should be used to perform calculations? Defaults to 1. Note that this function should be called from within the <a href="#">WriteDMRcateResults</a> function, which is already written in parallel. Further note that the DMRcate package (as of version 1.16.0), does not support parallelization in Windows environments.
<code>dmr.sig.threshold</code>	Regions with DMR p-value less than <code>dmr.sig.threshold</code> are selected for the output
<code>min.cpgs</code>	Minimum number of CpGs. Regions with at least <code>min.cpgs</code> are selected for the output. Defaults to 5.
<code>genome</code>	Reference genome for annotating DMRs, passed to the <a href="#">extractRanges</a> function in DMRcate. Can be one of "hg19", "hg38", or "mm10". Defaults to "hg19".

## Value

A list of two elements: a data frame of `dmrcate` results and the computing time for the DMRcate method.

## Examples

```
# Called internally by the WriteDMRcateResults() function.
## Not run:
data("betaVals_mat")
data("cpgLocation_df")
data("startEndCPG_df")

treat_ls <- SimulateData(beta_mat = betaVals_mat,
                        Aclusters_df = startEndCPG_df,
                        delta_num = 0.4,
                        seed_int = 100)
class_fct <- factor(c(rep("Tumor", 7), rep("Normal", 7)))

RunDMRcate(
  betaVals_mat = treat_ls$simBetaVals_df,
  labels_fct = class_fct,
  cpgLocation_df = cpgLocation_df,
  lambda_int = 500, C_int = 5
)

## End(Not run)
```

RunProbeLasso

*Return Results from the champ.DMR Function***Description**

A wrapper function for the ProbeLasso method, called internally by the [WriteProbeLassoResults](#) function. This function calls the [champ.DMR](#) function to perform the ProbeLasso method calculations.

**Usage**

```
RunProbeLasso(betaVals_mat, labels_fct = factor(c(rep("Tumor", 7),
  rep("Normal", 7))), cpGLocation_df, adjPvalProbe_num, meanLassoRadius_int,
  minDmrSep_int, nCores = 1, dmr.sig.threshold = 0.05, min.cpgs = 5)
```

**Arguments**

- |                     |  |
|---------------------|--|
| betaVals_mat        | A matrix of beta values returned in the first entry of the output from the SimulateData function, ordered by the CpGs. Note this dataset includes all CpGs on the array.   |
| labels_fct          | A factor vector of subject class labels. These should match the observations contained in the columns of the betaVals_mat matrix. Defaults to seven "Tumor" followed by seven "Normal" samples.  |
| cpGLocation_df      | An annotation table that indicates locations of CpGs. This data frame has CPG IDs as the rows with matching chromosome and location info in the columns. Specifically, the columns are: ILMNID - the CpG ID; chr - the chromosome label; and MAPINFO - the chromosome location. An example is given in the cpGLocation_df data set.  |
| adjPvalProbe_num    | The minimum threshold of significance for probes to be included in DMRs, passed to the <a href="#">champ.DMR</a> function.   |
| meanLassoRadius_int | Radius around each DMP to detect DMR, passed to the <a href="#">champ.DMR</a> function.  |
| minDmrSep_int       | The minimum separation (bp) between neighbouring DMRs, passed to the <a href="#">champ.DMR</a> function.   |
| nCores              | How many cores should be used to perform calculations? Defaults to 1. Note that this function should be called from within the <a href="#">WriteProbeLassoResults</a> function, which is already written in parallel. If this function is executed directly (not from within this function), then this argument is passed to the cores argument of the <a href="#">champ.DMR</a> function. |
| dmr.sig.threshold   | Regions with DMR p-value less than dmr.sig.threshold are selected for the output   |
| min.cpgs            | Minimum number of CpGs. Regions with at least min.cpgs are selected for the output. Defaults to 5.   |

**Value**

A list of two elements: a data frame of champ.DMR results and the computing time for the ProbeLasso method.

**Examples**

```
# Called internally by the WriteProbeLassoResults() function.
## Not run:
data("betaVals_mat")
data("cpgLocation_df")
data("startEndCPG_df")

treat_ls <- SimulateData(beta_mat = betaVals_mat,
                        Aclusters_df = startEndCPG_df,
                        delta_num = 0.4,
                        seed_int = 100)
class_fct <- factor(c(rep("Tumor", 7), rep("Normal", 7)))

RunProbeLasso(
  betaVals_mat = treat_ls$simBetaVals_df,
  labels_fct = class_fct,
  cpgLocation_df = cpgLocation_df,
  adjPvalProbe_num = 0.05,
  meanLassoRadius_int = 1000,
  minDmrSep_int = 1000
)

## End(Not run)
```

---

SimulateData	<i>Simulate Differentially Methylated Regions (DMRs) in Methylation Data</i>
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---

**Description**

Generate a methylation dataset where treatment effects are added to beta values in one group of samples for some randomly selected co-methylated clusters.

**Usage**

```
SimulateData(beta_mat, Aclusters_df, delta_num, seed_int, betaCols_idx = 9:22,
             numEx_int = 7, numClusters_int = 500)
```

**Arguments**

beta_mat	A beta value matrix for methylation samples from a 450k methylation array with CpG IDs as the row names and sample IDs as the column names. An example is given in the betaVals_mat data set.
Aclusters_df	A data frame of beta values and CpG information for clusters of CpGs over a 450k methylation array. The rows correspond to the CPGs. The columns have information on the cluster number, chromosome, cluster start and end locations, and the beta values for each subject grouped by some clinical indicator (e.g. case v. control). An example is given in the startEndCPG_df data set. This data set can be generated by the file /inst/1_Aclust_data_import.R
delta_num	The treatment size: a non-negative real number to add to the beta values within randomly-selected clusters for the first numEx_int samples. This artificially creates differentially- methylated regions (DMRs).

seed_int	The seed value passed to the <a href="#">Random</a> function to enable reproducible results
betaCols_idx	The column numbers of the Aclusters_df data frame in which beta values for each subject are stored.
numEx_int	The number of samples in the first group. This function assumes that samples in one group are contiguous columns of the Aclusters_df data frame.
numClusters_int	The total number of randomly selected clusters, for which the treatment effect, delta_num, is then added

### Value

A list with two elements:

- simBetaVals\_df : A data frame of beta values after treatment effects were added, used for input for different DMR-finding methods. Note this dataset includes all the CpGs on the array.
- simAclusters\_df : A data frame of the methylation values only for clusters identified by Aclust and indicator variable for whether treatment effects were added. Note this has only CpGs mapped to all the clusters found by the Aclust method.

### Examples

```
## Not run:
data("startEndCPG_df")
data("betaVals_mat")

SimulateData(beta_mat = betaVals_mat,
             Aclusters_df = startEndCPG_df,
             delta_num = 0.4,
             seed_int = 12345)

## End(Not run)
```

---

startEndCPG_df	<i>Annotated CPG Data Set</i>
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---

### Description

A matrix of beta values for clusters of CpGs over a 450k methylation array.

### Usage

```
startEndCPG_df
```

### Format

A data frame containing 20361 CPG locations measured on 14 subjects. The rows are the CPG IDs. The first eight columns are the metadata for the CPGs, including: Clusternumber, the CPG ID (cpg), chromosome (CHR), chromosome location (MAPINFO), chromosome start position (start\_position), and chromosome end position (end\_position). The remaining 14 columns are the beta values for the subjects. The column names for the subjects indicate from which phenotypic group the subjects were drawn; for example, the 9744-Tumor column indicates that this subject was from the case group.



**Source**

Calculated via the 1\_Aclust\_data\_import.R script in the old\_scripts sub-directory of the inst directory.

---

WriteBumphunterResults

*Calculate and Save Bumphunter Method Results for Specified Design Points*

---

**Description**

Given a set of design points (treatment effect size to be added and number of repetitions), simulate methylation data with DMRs and then apply the bumphunter method to them. Write the results to a file.

**Usage**

```
WriteBumphunterResults(beta_mat, CPGs_df, Aclusters_df, parallel = TRUE,
  numCores = detectCores() - 2, deltas_num = c(0, 0.025, 0.05, 0.1, 0.15,
  0.2, 0.3, 0.4), seeds_int = c(100, 210, 330, 450, 680),
  cutoffQ_num = c(0.9, 0.95, 0.99), maxGap_int = c(200, 250, 500, 750,
  1000), resultsDir = "DMRcate_compare/", verbose = TRUE)
```

**Arguments**

beta_mat	A beta value matrix for methylation samples from a 450k methylation array with CpG IDs as the row names and sample IDs as the column names. An example is given in the betaVals_mat data set.
CPGs_df	Annotation table that indicates locations of CpGs. This data frame has CpG IDs as the rows with matching chromosome and location info in the columns. Specifically, the columns are: ILMNID - the CPG ID; chr - the chromosome label; and MAPINFO - the chromosome location. An example is given in the cpgLocation_df data set.
Aclusters_df	A data frame of beta values and CpG information for clusters of CpGs over a 450k methylation array. The rows correspond to the CPGs. The columns have information on the cluster number, chromosome, cluster start and end locations, and the beta values for each subject grouped by some clinical indicator (e.g. case v. control). An example is given in the startEndCPG_df data set. This data set can be generated by the file /inst/1_Aclust_data_import.R
parallel	Should computing be completed over multiple computing cores? Defaults to TRUE.
numCores	If parallel, how many cores should be used? Defaults to two less than the number of available cores (as calculated by the <a href="#">detectCores</a> function). These cores are used internally by the <a href="#">bumphunter</a> function.
deltas_num	A vector of treatment sizes: non-negative real numbers to add to the beta values within randomly-selected clusters for a single class of subjects. This artificially creates differentially-methylated regions (DMRs).
seeds_int	A vector of seed values passed to the <a href="#">Random</a> function to enable reproducible results

cutoffQ_num	A vector of quantiles used for picking the cutoff using the permutation distribution, passed through the call to the internal <a href="#">RunBumphunter</a> call to <a href="#">bumphunter</a> .
maxGap_int	A vector of maximum location gaps, passed to the <a href="#">bumphunter</a> function. These will be used to define the clusters of locations that are to be analyzed together via the <a href="#">clusterMaker</a> function.
resultsDir	Where should the results be saved? Defaults to Bumphunter_compare/.
verbose	Should the function print progress messages? Defaults to TRUE.

### Details

This function creates matrices of all combinations of design points and all combinations of parameters. For each combination, this function executes the internal [RunBumphunter](#) function and saves the results as a compressed .RDS file.

### Value

Saves output files in the specified results directory.

### Examples

```
## Not run:
data("betaVals_mat")
data("cpgLocation_df")
data("startEndCPG_df")

WriteBumphunterResults(
  beta_mat = betaVals_mat,
  CPGs_df = cpgLocation_df,
  Aclusters_df = startEndCPG_df
)

## End(Not run)
```

---

WriteDMRcateResults	<i>Calculate and Save DMRcate Method Results for Specified Design Points</i>
---------------------	--

---

### Description

Given a set of design points (treatment effect size to be added and number of repetitions), simulate methylation data with DMRs and then apply the dmr\_cate method to them. Write the results to a file.

### Usage

```
WriteDMRcateResults(beta_mat, CPGs_df, Aclusters_df, parallel = TRUE,
  numCores = detectCores() - 2, deltas_num = c(0, 0.025, 0.05, 0.1, 0.15,
  0.2, 0.3, 0.4), seeds_int = c(100, 210, 330, 450, 680),
  lambdas_num = c(200, 250, 500, 750, 1000), Cs_int = 1:5,
  resultsDir = "DMRcate_compare/", verbose = !parallel)
```

**Arguments**

beta_mat	A beta value matrix for methylation samples from a 450k methylation array with CpG IDs as the row names and sample IDs as the column names. An example is given in the betaVals_mat data set.
CPGs_df	An annotation table that indicates locations of CpGs. This data frame has CpG IDs as the rows with matching chromosome and location info in the columns. Specifically, the columns are: ILMNID - the CpG ID; chr - the chromosome label; and MAPINFO - the chromosome location. An example is given in the cpgLocation_df data set.
Aclusters_df	A data frame of beta values and CpG information for clusters of CpGs over a 450k methylation array. The rows correspond to the CPGs. The columns have information on the cluster number, chromosome, cluster start and end locations, and the beta values for each subject grouped by some clinical indicator (e.g. case v. control). An example is given in the startEndCPG_df data set. This data set can be generated by the file /inst/1_Aclust_data_import.R
parallel	Should computing be completed over multiple computing cores? Defaults to TRUE.
numCores	If parallel, how many cores should be used? Defaults to two less than the number of available cores (as calculated by the <a href="#">detectCores</a> function).
deltas_num	A vector of treatment sizes: non-negative real numbers to add to the beta values within randomly-selected clusters for a single class of subjects. This artificially creates differentially-methylated regions (DMRs).
seeds_int	A vector of seed values passed to the <a href="#">Random</a> function to enable reproducible results
lambdas_num	A vector of Gaussian kernel bandwidths for smoothed- function estimation in the called <a href="#">dmrcate</a> function
Cs_int	A vector of scaling factors for bandwidth in the internal call to the <a href="#">dmrcate</a> function
resultsDir	Where should the results be saved? Defaults to DMRcate_compare/.
verbose	Should the function print progress messages? Defaults to TRUE only if parallel = FALSE. See the internal <a href="#">RunDMRcate</a> function for more details about parallel computing with DMRcate.

**Details**

This function creates matrices of all combinations of design points and all combinations of parameters. For each combination, this function executes the internal [RunDMRcate](#) function and saves the results as a compressed .RDS file.

**Value**

Saves output files in the specified results directory.

**Examples**

```
## Not run:
data("betaVals_mat")
data("cpgLocation_df")
data("startEndCPG_df")
```

```

WriteDMRcateResults(
  beta_mat = betaVals_mat,
  CPGs_df = cpgLocation_df,
  Aclusters_df = startEndCPG_df
)

## End(Not run)

```

---

## WriteProbeLassoResults

*Calculate and Save ProbeLasso Method Results for Specified Design Points*

---

### Description

Given a set of design points (treatment effect size to be added and number of repetitions), simulate simulate methylation data with DMRs and apply the ProbeLasso method (via the [champ.DMR](#) function) to them. Write the results to a file.

### Usage

```

WriteProbeLassoResults(beta_mat, CPGs_df, Aclusters_df, parallel = TRUE,
  numCores = detectCores() - 2, deltas_num = c(0, 0.025, 0.05, 0.1, 0.15,
  0.2, 0.3, 0.4), seeds_int = c(100, 210, 330, 450, 680),
  pVals_num = c(0.001, 0.01, 0.05, 0.1), aveLassoRad_int = c(375, 700,
  1000), minDmrSep_int = c(200, 250, 500, 750, 1000),
  resultsDir = "DMRcate_compare/", verbose = !parallel)

```

### Arguments

beta_mat	A beta value matrix for methylation samples from a 450k methylation array with CpG IDs as the row names and sample IDs as the column names. An example is given in the betaVals_mat data set. #'
CPGs_df	An annotation table that indicates locations of CpGs. This data frame has CpG IDs as the rows with matching chromosome and location info in the columns. Specifically, the columns are: ILMNID - the CPG ID; chr - the chromosome label; and MAPINFO - the chromosome location. An example is given in the cpgLocation_df data set.
Aclusters_df	A data frame of beta values and CpG information for clusters of CpGs over a 450k methylation array. The rows correspond to the CPGs. The columns have information on the cluster number, chromosome, cluster start and end locations, and the beta values for each subject grouped by some clinical indicator (e.g. case v. control). An example is given in the startEndCPG_df data set. This data set can be generated by the file /inst/1_Aclust_data_import.R
parallel	Should computing be completed over multiple computing cores? Defaults to TRUE.
numCores	If parallel, how many cores should be used? Defaults to two less than the number of available cores (as calculated by the <a href="#">detectCores</a> function).
deltas_num	A vector of treatment sizes: non-negative real numbers to add to the beta values within randomly-selected clusters for a single class of subjects. This artificially creates differentially-methylated regions (DMRs).

seeds_int	A vector of seed values passed to the <a href="#">Random</a> function to enable reproducible results
pVals_num	A vector of the minimum thresholds of significance for probes to be includede in DMRs, passed through the <a href="#">RunProbeLasso</a> function to the <a href="#">champ.DMR</a> function.
aveLassoRad_int	A vector of radii around each differential methylation position to detect DMR, passed to the <a href="#">champ.DMR</a> function.
minDmrSep_int	A vector of the minimum seperation (bp) values between neighbouring DMRs, passed to the <a href="#">champ.DMR</a> function.
resultsDir	Where should the results be saved? Defaults to DMRcate_compare/.
verbose	Should the function print progress messages? Defaults to TRUE only if parallel = FALSE.

### Details

This function creates matrices of all combinations of design points and all combinations of parameters. For each combination, this function executes the internal [RunProbeLasso](#) function and saves the results as a compressed .RDS file.

### Value

Saves output files in the specified results directory.

### Examples

```
## Not run:
data("betaVals_mat")
data("cpgLocation_df")
data("startEndCPG_df")

WriteProbeLassoResults(
  beta_mat = betaVals_mat,
  CPGs_df = cpgLocation_df,
  Aclusters_df = startEndCPG_df
)

## End(Not run)
```

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