

Package ‘DMRcompare’

July 25, 2018

Title Compare Different R Tools for Detecting Differentially-Methylated Regions (DMRs) of a Genome

Version 0.0.0.9000

Description The accompanying package to the method comparison paper “An evaluation of supervised methods for identifying differentially methylated regions in epigenome-wide association studies” by Mallik et al. (2018), submitted to Briefings in Bioinformatics as a review article. This package contains the organized and documented R scripts necessary to replicate the multi-design-point comparative simulation study, as well as associated tables and figures.

Depends R (>= 3.3.0),
ChAMPdata,
DMRcatedata

Imports bumphunter,
ChAMP,
ChIPpeakAnno,
data.table,
doParallel,
DMRcate,
foreach,
GenomicRanges,
graphics,
grDevices,
IRanges,
minfi,
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PRROC,
stats

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testthat

R topics documented:

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betaVals_mat	<i>Annotated CPG Data Set</i>
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Description

A beta value matrix for selected methylation samples from a 450k methylation array. These beta values are trimmed to range from 0.05 to 0.95.

Usage

betaVals_mat

Format

A matrix containing 356603 CPGs measured on 14 subjects. The rownames are the CPG IDs, and the column names indicate the sample IDs of the subjects.

Source

Calculated via the 1_Aclust_data_import.R script in the old_scripts sub-directory of the inst directory.

Description

Given a directory of best-performing results files from one of the simulation functions ([WriteDMRcateResults](#), [WriteProbeLassoResults](#), [WriteBumphunterResults](#), or results from the Comb-p method in Python), import the raw data files and construct PR-curve objects via the [pr.curve](#) function.

Usage

```
BuildPRcurve(bestResultsDir, delta = c(0.025, 0.05, 0.1, 0.15, 0.2, 0.3, 0.4),
             seed = c(100, 210, 330, 450, 680), beta_mat = betaVals_mat,
             AclustCPG_df = startEndCPG_df, CPGs_df = cpgLocation_df, min.cpgs = 5)
```

Arguments

bestResultsDir	The name of the directory where the method results from the best-performing parameter settings are stored. For the full design we have included (<code>delta = c(0.025, 0.05, 0.1, 0.15, 0.2, 0.3, 0.4)</code> , and <code>seed = c(100, 210, 330, 450, 680)</code>), this directory should contain 35 .RDS files per method.
delta	A treatment size corresponding to one of the simulations with completed results files in the bestResultsDir directory.
seed	A seed value corresponding to one of the simulations with completed results files in the bestResultsDir directory.
beta_mat	A beta value matrix for selected methylation samples from a 450k methylation array with CPG IDs as the row names and sample IDs as the column names. An example is given in the betaVals_mat data set.
AclustCPG_df	A data frame of beta values and CpG information for clusters of CpGs over a 450k methylation array. The rows correspond to the CPGs. The columns have information on the cluster number, chromosome, cluster start and end locations, and the beta values for each subject grouped by some clinical indicator (e.g. case v. control). An example is given in the startEndCPG_df data set.
CPGs_df	An annotation table that indicates locations of CpGs. This data frame has CPG IDs as the rows with matching chromosome and location info in the columns. Specifically, the columns are: ILMNID - the CPG ID; chr - the chromosome label; and MAPINFO - the chromosome location. An example is given in the cpgLocation_df data set. This data set is only necessary if the results directory contains Comb-p results with the specified delta and seed values.
min.cpgs	The minimum number of CPGs necessary to consider a result significant. Defaults to 5. This argument is only required if the results directory contains Comb-p results with the specified delta and seed values.

Value

A list of PR-curve objects, to be plotted via the [PlotPRCurve](#) function.

Examples

```
## Not run:
BuildPRcurve(
  bestResultsDir = "best_cases_results/",
  delta = 0.4,
  seed = 100
)

## End(Not run)
```

cpgLocation_df	<i>CPG Locations</i>
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Description

An annotation table that indicates locations of CpGs. This data frame has CPG IDs as the rows with matching chromosome and location info in the columns.

Usage

```
cpgLocation_df
```

Format

A data frame containing 485512 CPGs (rows) and three columns. The columns are:

- ILMNID : the CPG ID, as a factor
- chr : the chromosome label, as a character
- MAPINFO : the chromosome location, as an integer

Source

Compiled via the 1_Aclust_data_import.R script in the old_scripts sub-directory of the inst directory.

PlotOverlaps	<i>Plot Venn Diagrams of DMR Overlaps</i>
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Description

Given a directory of best-performing results files from one of the simulation functions ([WriteDMRcateResults](#), [WriteProbeLassoResults](#), [WriteBumphunterResults](#), or results from the Comb-p method in Python), call the [BuildOverlaps](#) function to import the raw data files and DMR overlap lists, then plot those Venn diagrams and save the plots to a PDF.

Usage

```
PlotOverlaps(bestResultsDir, figFileName, device = pdf,
  plotTitle = "default", delta_num = c(0.025, 0.05, 0.1, 0.15, 0.2, 0.3,
  0.4), seeds_int = c(100, 210, 330, 450, 680), totalTest_int = 3063,
  CPGs_df = cpgLocation_df, min.cpgs = 5, ...)
```

Arguments

<code>bestResultsDir</code>	The name of the directory where the method results from the best-performing parameter settings are stored. For the full design we have included (<code>delta = c(0.025, 0.05, 0.1, 0.2)</code> and <code>seed = c(100, 210, 330, 450, 680)</code>), this directory should contain 35 .RDS files per method.
<code>figFileName</code>	The name of the figure
<code>device</code>	Which graphics device should be used to save the figures? Defaults to pdf . Note that if you use a device other than PDF (jpeg for instance), you can only plot one <code>delta_num</code> and <code>seed_int</code> combination per file.
<code>plotTitle</code>	The title of the plot. This argument is passed to the makeVennDiagram function. The default value of "default" will make the plot title "Venn Diagram for mu = DELTA, rep = INDEX OF SEED".
<code>delta_num</code>	A vector of treatment sizes with values corresponding to one of the simulations with completed results files in the <code>bestResultsDir</code> directory.
<code>seeds_int</code>	A vector of random seeds with values corresponding to one of the simulations with completed results files in the <code>bestResultsDir</code> directory.
<code>totalTest_int</code>	Parameter passed to the makeVennDiagram function. This is an interger value specifying the total number of tests performed to obtain the list of peaks. It should be much larger than the number of peaks in the largest peak set.
<code>CPGs_df</code>	An annotation table that indicates locations of CpGs. This data frame has CPG IDs as the rows with matching chromosome and location info in the columns. Specifically, the columns are: <code>ILMNID</code> - the CPG ID; <code>chr</code> - the chromosome label; and <code>MAPINFO</code> - the chromosome location. An example is given in the <code>cpgLocation_df</code> data set. This data set is only necessary if the results directory contains Comb-p results with the specified <code>delta</code> and <code>seed</code> values. This is passed to the BuildOverlaps function.
<code>min.cpgs</code>	The minimum number of CPGs before we consider a result significant. Defaults to 5. This argument is only required if the results directory contains Comb-p results with the specified <code>delta</code> and <code>seed</code> values. This is passed to the BuildOverlaps function.
<code>...</code>	Dots for additional arguments to be passed to the graphics device

Value

Nothing. A PDF file of plots is created as a side effect.

Examples

```
## Not run:
PlotOverlaps(
  bestResultsDir = "best_cases_results/",
  figFileName = "best_cases_results/resultsFigures/testVenn_allDesigns2"
)

## End(Not run)
```

PlotPRCurve	<i>Plot Precision-Recall Curves</i>
-------------	-------------------------------------

Description

Given a list of PR-curve objects as returned by the [BuildPRcurve](#) function, plot the precision-recall curve for each method in a shared figure.

Usage

```
PlotPRCurve(prCurves_ls, plotTitle = "default", new = TRUE, lineWidth = 1,
  colours = NULL)
```

Arguments

prCurves_ls	A list of PR-curve objects
plotTitle	The title of the plot. The default value of "default" will make the plot title "Venn Diagram for mu = DELTA, rep = INDEX OF SEED".
new	Should the PR curves from this list form their own graph (TRUE) or be added onto a previous PR-curve figure (FALSE). Defaults to TRUE.
lineWidth	The line width of each PR curve in the plot. Defaults to 1.
colours	Optionally add your own colours for each line. Otherwise, the colours are created with the hcl function.

Value

Nothing. A plot is created as a side effect.

Examples

```
## Not run:
prCurves_0.4_100_ls <-
  BuildPRcurve(
    bestResultsDir = "best_cases_results/",
    delta = 0.4,
    seed = 100
  )

PlotPRCurve(prCurves_0.4_100_ls)

## End(Not run)
```

ProcessBumphunterResults

Process Bumphunter Results Files

Description

Given a directory of saved Bumphunter results, as written by the [WriteBumphunterResults](#) function, import and summarize these data files.

Usage

```
ProcessBumphunterResults(resultsDir, beta_mat, AclustCPG_df, verbose = TRUE)
```

Arguments

resultsDir	The name of the directory where the Bumphunter method results are stored. This should match the directory name supplied to the resultsDir argument of the WriteBumphunterResults function.
beta_mat	A beta value matrix for selected methylation samples from a 450k methylation array with CPG IDs as the row names and sample IDs as the column names. An example is given in the betaVals_mat data set.
AclustCPG_df	A data frame of beta values and CpG information for clusters of CpGs over a 450k methylation array. The rows correspond to the CPGs. The columns have information on the cluster number, chromosome, cluster start and end locations, and the beta values for each subject grouped by some clinical indicator (e.g. case v. control). An example is given in the startEndCPG_df data set.
verbose	Should the function print progress messages? Defaults to TRUE.

Value

A data frame of model fit statistics for the Bumphunter method under each of the given parameter combinations to the data generated for each design configuration

Examples

```
## Not run:
data("betaVals_mat")
data("startEndCPG_df")

bumphunterRes_df <- ProcessBumphunterResults(
  resultsDir = "DMRcate_results/",
  beta_mat = betaVals_mat,
  AclustCPG_df = startEndCPG_df
)

## End(Not run)
```

ProcessCombpResults *Extract and Process Comb-p Results Files*

Description

Given a directory of saved Comb-p results, as .RDS files, import, standardize, and summarize these data files.

Usage

```
ProcessCombpResults(resultsDir, beta_mat, AclustCPG_df, cpgLocation_df,
  dmr.sig.threshold = 0.05, min.cpgs = 5, verbose = TRUE)
```

Arguments

resultsDir	The name of the directory where the Comb-p method results are stored.
beta_mat	A beta value matrix for selected methylation samples from a 450k methylation array with CPG IDs as the row names and sample IDs as the column names. An example is given in the betaVals_mat data set.
AclustCPG_df	A data frame of beta values and CpG information for clusters of CpGs over a 450k methylation array. The rows correspond to the CPGs. The columns have information on the cluster number, chromosome, cluster start and end locations, and the beta values for each subject grouped by some clinical indicator (e.g. case v. control). An example is given in the startEndCPG_df data set.
cpgLocation_df	An annotation table that indicates locations of CpGs. This data frame has CPG IDs as the rows with matching chromosome and location info in the columns. Specifically, the columns are: ILMNID - the CPG ID; chr - the chromosome label; and MAPINFO - the chromosome location. An example is given in the cpgLocation_df data set.
dmr.sig.threshold	Significance level to select regions (with dmr.pval less than the specified value) passed to the internal MergeDMRsWithCPGs function.
min.cpgs	The minimum number of CPGs necessary to consider a result significant. Defaults to 5.
verbose	Should the function print progress messages? Defaults to TRUE.

Value

A data frame of model fit statistics for the Comb-p method under each of the given parameter combinations to the data generated for each design configuration

Examples

```
## Not run:
data("betaVals_mat")
data("startEndCPG_df")
data("cpgLocation_df")

combpRes_df <- ProcessCombpResults(
  resultsDir = "DMRcate_results/",
```



```

    beta_mat = betaVals_mat,
    AclustCPG_df = startEndCPG_df,
    cpgLocation_df = cpgLocation_df
  )

## End(Not run)

```

ProcessDMRcateResults *Process DMRcate Results Files*

Description

Given a directory of saved DMRcate results, as written by the [WriteDMRcateResults](#) function, import and summarize these data files.

Usage

```
ProcessDMRcateResults(resultsDir, beta_mat, AclustCPG_df, verbose = TRUE)
```

Arguments

resultsDir	The name of the directory where the DMRcate method results are stored. This should match the directory name supplied to the resultsDir argument of the WriteDMRcateResults function.
beta_mat	A beta value matrix for selected methylation samples from a 450k methylation array with CPG IDs as the row names and sample IDs as the column names. An example is given in the betaVals_mat data set.
AclustCPG_df	A data frame of beta values and CpG information for clusters of CpGs over a 450k methylation array. The rows correspond to the CPGs. The columns have information on the cluster number, chromosome, cluster start and end locations, and the beta values for each subject grouped by some clinical indicator (e.g. case v. control). An example is given in the startEndCPG_df data set.
verbose	Should the function print progress messages? Defaults to TRUE.

Value

A data frame of model fit statistics for the DMRcate method under each of the given parameter combinations to the data generated for each design configuration

Examples

```

## Not run:
data("betaVals_mat")
data("startEndCPG_df")

dmrcateRes_df <- ProcessDMRcateResults(
  resultsDir = "DMRcate_results/",
  beta_mat = betaVals_mat,
  AclustCPG_df = startEndCPG_df
)

## End(Not run)

```

ProcessProbeLassoResults

Process ProbeLasso Results Files

Description

Given a directory of saved ProbeLasso results, as written by the [WriteProbeLassoResults](#) function, import and summarize these data files.

Usage

```
ProcessProbeLassoResults(resultsDir, beta_mat, AclustCPG_df, verbose = TRUE)
```

Arguments

resultsDir	The name of the directory where the ProbeLasso method results are stored. This should match the directory name supplied to the resultsDir argument of the WriteProbeLassoResults function.
beta_mat	A beta value matrix for selected methylation samples from a 450k methylation array with CPG IDs as the row names and sample IDs as the column names. An example is given in the betaVals_mat data set.
AclustCPG_df	A data frame of beta values and CpG information for clusters of CpGs over a 450k methylation array. The rows correspond to the CPGs. The columns have information on the cluster number, chromosome, cluster start and end locations, and the beta values for each subject grouped by some clinical indicator (e.g. case v. control). An example is given in the startEndCPG_df data set.
verbose	Should the function print progress messages? Defaults to TRUE.

Value

A data frame of model fit statistics for the ProbeLasso method under each of the given parameter combinations to the data generated for each design configuration

Examples

```
## Not run:
data("betaVals_mat")
data("startEndCPG_df")

probeLassoRes_df <- ProcessProbeLassoResults(
  resultsDir = "DMRcate_results/",
  beta_mat = betaVals_mat,
  AclustCPG_df = startEndCPG_df
)

## End(Not run)
```

RunBumphunter

*Return Results from the bumphunter Function***Description**

A wrapper function for the Bumphunter method as implemented in the bumphunter package, called internally by the [WriteBumphunterResults](#) function.

Usage

```
RunBumphunter(betaVals_mat, labels_fct = factor(c(rep("Tumor", 7),
  rep("Normal", 7))), chromos_char, chromPosit_num, cpgLocation_df,
  pickCutoffQ_num, maxGap_int, B_int = 10, numCores = detectCores() - 1,
  dmr.sig.threshold = 0.05, min.cpgs = 5)
```

Arguments

betaVals_mat	A matrix of beta values returned in the second entry of the output from the SimulateData function, ordered by the CPGs.
labels_fct	A factor vector of subject class labels. These should match the observations contained in the columns of the betaVals_mat matrix. Defaults to factor(c(rep("Tumor", 7), rep("Normal", 7))).
chromos_char	A character vector with the chromosomes of each location
chromPosit_num	A numeric vector representing the chromosomal position
cpgLocation_df	An annotation table that indicates locations of CpGs. This data frame has CPG IDs as the rows with matching chromosome and location info in the columns. Specifically, the columns are: ILMNID - the CPG ID; chr - the chromosome label; and MAPINFO - the chromosome location. An example is given in the cpgLocation_df data set.
pickCutoffQ_num	The quantile used for picking the cutoff using the permutation distribution, passed to the bumphunter function.
maxGap_int	The maximum location gap, passed to the bumphunter function. This will be used to define the clusters of locations that are to be analyzed together via the clusterMaker function.
B_int	An integer denoting the number of resamples to use when computing null distributions, passed to the bumphunter function.
numCores	The number of computing cores for parallel execution, passed to the registerDoParallel function. Defaults to one less than the number of cores available on your machine, as detected via the detectCores function.
dmr.sig.threshold	Significance level to select regions (with dmr.pval less than the specified value) passed to the internal StandardizeOutput function.
min.cpgs	The minimum number of CPGs before we consider a result significant, passed to the internal StandardizeOutput function. Defaults to 5.

Value

A list of two elements: a data frame of bumphunter results that have been standardized by the [StandardizeOutput](#) function and the computing time for the bumphunter method.

Examples

```
# Called internally by the WriteBumphunterResults() function.
## Not run:
data("betaVals_mat")
data("cpgLocation_df")
data("startEndCPG_df")

treat_ls <- SimulateData(beta_mat = betaVals_mat,
                        AclustCPG_df = startEndCPG_df,
                        delta_num = 0.4,
                        seed_int = 100)
class_fct <- factor(c(rep("Tumor", 7), rep("Normal", 7)))

RunBumphunter(
  betaVals_mat = treat_ls$simBetaVals_df,
  labels_fct = class_fct,
  cpgLocation_df = cpgLocation_df,
  pickCutoffQ_num = 0.95,
  maxGap_int = 250
)

## End(Not run)
```

RunDMRcate

Return Results from the dmr cate Function

Description

A wrapper function for the DMRcate method from the DMRcate package, called internally by the [WriteDMRcateResults](#) function.

Usage

```
RunDMRcate(betaVals_mat, labels_fct = factor(c(rep("Tumor", 7), rep("Normal",
7))), cpgLocation_df, lambda_int, C_int, nCores = 1,
dmr.sig.threshold = 0.05, min.cpgs = 5, genome = "hg19")
```

Arguments

betaVals_mat	A matrix of beta values returned in the second entry of the output from the SimulateData function.
labels_fct	A factor vector of subject class labels. These should match the observations contained in the columns of the betaVals_mat matrix. Defaults to factor(c(rep("Tumor", 7), rep("Normal", 7))).
cpgLocation_df	An annotation table that indicates locations of CpGs. This data frame has CPG IDs as the rows with matching chromosome and location info in the columns. Specifically, the columns are: ILMNID - the CPG ID; chr - the chromosome label; and MAPINFO - the chromosome location. An example is given in the cpgLocation_df data set.
lambda_int	Gaussian kernel bandwidth for smoothed-function estimation in the called dmrcate function.

C_int	Scaling factor for bandwidth in the internal call to the dmrcate function
nCores	How many cores should be used to perform calculations? Defaults to 1. Note that this function should be called from within the WriteDMRcateResults function, which is already written in parallel. Further note that the DMRcate package (as of version 1.16.0), does not support parallelization in Windows environments.
dmr.sig.threshold	Significance level to select regions (with dmr.pval less than the specified value) passed to the internal StandardizeOutput function.
min.cpgs	The minimum number of CPGs necessary to consider a result significant, passed to the internal StandardizeOutput function. Defaults to 5.
genome	Reference genome for annotating DMRs with promoter overlaps, passed to the extractRanges function. Can be one of "hg19", "hg38", or "mm10". Defaults to "hg19".

Value

A list of two elements: a data frame of dmrcate results that have been standardized by the [StandardizeOutput](#) function and the computing time for the DMRcate method.

Examples

```
# Called internally by the WriteDMRcateResults() function.
## Not run:
data("betaVals_mat")
data("cpgLocation_df")
data("startEndCPG_df")

treat_ls <- SimulateData(beta_mat = betaVals_mat,
                        AclustCPG_df = startEndCPG_df,
                        delta_num = 0.4,
                        seed_int = 100)
class_fct <- factor(c(rep("Tumor", 7), rep("Normal", 7)))

RunDMRcate(
  betaVals_mat = treat_ls$simBetaVals_df,
  labels_fct = class_fct,
  cpgLocation_df = cpgLocation_df,
  lambda_int = 500, C_int = 5
)

## End(Not run)
```

Description

A wrapper function for the ProbeLasso method, called internally by the [WriteProbeLassoResults](#) function. This function calls the [champ.DMR](#) function to perform the ProbeLasso method calculations.

Usage

```
RunProbeLasso(betaVals_mat, labels_fct = factor(c(rep("Tumor", 7),
  rep("Normal", 7))), cpgLocation_df, adjPvalProbe_num, meanLassoRadius_int,
  minDmrSep_int, nCores = 1, dmr.sig.threshold = 0.05, min.cpgs = 5)
```

Arguments

<code>betaVals_mat</code>	A matrix of beta values returned in the second entry of the output from the <code>SimulateData</code> function
<code>labels_fct</code>	A factor vector of subject class labels. These should match the observations contained in the columns of the <code>betaVals_mat</code> matrix. Defaults to <code>factor(c(rep("Tumor", 7), rep("Normal", 7)))</code>
<code>cpgLocation_df</code>	An annotation table that indicates locations of CpGs. This data frame has CPG IDs as the rows with matching chromosome and location info in the columns. Specifically, the columns are: <code>ILMNID</code> - the CPG ID; <code>chr</code> - the chromosome label; and <code>MAPINFO</code> - the chromosome location. An example is given in the <code>cpgLocation_df</code> data set.
<code>adjPvalProbe_num</code>	The minimum threshold of significance for probes to be included in DMRs, passed to the champ.DMR function.
<code>meanLassoRadius_int</code>	Radius around each DMP to detect DMR, passed to the champ.DMR function.
<code>minDmrSep_int</code>	The minimum separation (bp) between neighbouring DMRs, passed to the champ.DMR function.
<code>nCores</code>	How many cores should be used to perform calculations? Defaults to 1. Note that this function should be called from within the WriteProbeLassoResults function, which is already written in parallel. If this function is executed directly (not from within this function), then this argument is passed to the <code>cores</code> argument of the champ.DMR function.
<code>dmr.sig.threshold</code>	Significance level to select regions (with <code>dmr.pval</code> less than the specified value) passed to the internal StandardizeOutput function
<code>min.cpgs</code>	The minimum number of CPGs necessary to consider a result significant, passed to the internal StandardizeOutput function. Defaults to 5.

Value

A list of two elements: a data frame of `champ.DMR` results that have been standardized by the [StandardizeOutput](#) function and the computing time for the ProbeLasso method.

Examples

```
# Called internally by the WriteProbeLassoResults() function.
## Not run:
data("betaVals_mat")
data("cpgLocation_df")
data("startEndCPG_df")

treat_ls <- SimulateData(beta_mat = betaVals_mat,
  AclustCPG_df = startEndCPG_df,
  delta_num = 0.4,
  seed_int = 100)
```

```

class_fct <- factor(c(rep("Tumor", 7), rep("Normal", 7)))

RunProbeLasso(
  betaVals_mat = treat_ls$simBetaVals_df,
  labels_fct = class_fct,
  cpgLocation_df = cpgLocation_df,
  adjPvalProbe_num = 0.05,
  meanLassoRadius_int = 1000,
  minDmrSep_int = 1000
)

## End(Not run)

```

SimulateData

*Simulate Differences in Methylation Data***Description**

Given a randomly selected subset of clusters, add some constant value to each beta value in one observation class

Usage

```

SimulateData(beta_mat, AclustCPG_df, delta_num, seed_int, betaCols_idx = 9:22,
  numEx_int = 7, numClusters_int = 500)

```

Arguments

beta_mat	A beta value matrix for selected methylation samples from a 450k methylation array with CPG IDs as the row names and sample IDs as the column names. An example is given in the betaVals_mat data set.
AclustCPG_df	A data frame of beta values and CpG information for clusters of CpGs over a 450k methylation array. The rows correspond to the CPGs. The columns have information on the cluster number, chromosome, cluster start and end locations, and the beta values for each subject grouped by some clinical indicator (e.g. case v. control). An example is given in the startEndCPG_df data set.
delta_num	The treatment size: a non-negative real number to add to the beta values within randomly-selected clusters for a single class of subjects. This artificially creates differentially-methylated regions (DMRs).
seed_int	The seed value passed to the Random function to enable reproducible results
betaCols_idx	The column numbers of the AclustCPG_df data frame in which beta values for each subject are stored. This function assumes that the subject columns are grouped by their class.
numEx_int	The number of samples in the first group. Once again, this function assumes that these samples are contiguous columns of the AclustCPG_df data frame.
numClusters_int	The total number of clusters to randomly select to be inflated by the treatment amount, delta_num

Value

A list with two elements:

- `simBetaVals_df` A data frame of beta values after treatment effects were added, used for input for different DMR-finding methods. Note this is whole-genome data.
- `simAclusters_df` A data frame of the methylation values only for Aclust and annotation for whether treatment effects were added. Note this has only CPGs mapped to all the clusters found by the Aclust method.

Examples

```
## Not run:
data("startEndCPG_df")
data("betaVals_mat")

SimulateData(beta_mat = betaVals_mat,
             AclustCPG_df = startEndCPG_df,
             delta_num = 0.4,
             seed_int = 12345)

## End(Not run)
```

startEndCPG_df	<i>Annotated CPG Data Set</i>
----------------	-------------------------------

Description

A matrix of beta values for clusters of CpGs over a 450k methylation array.

Usage

```
startEndCPG_df
```

Format

A data frame containing 20361 CPG locations measured on 14 subjects. The rows are the CPG IDs. The first eight columns are the metadata for the CPGs, including: Clusternumber, the CPG ID (cpg), chromosome (CHR), chromosome location (MAPINFO), chromosome start position (start_position), and chromosome end position (end_position). The remaining 14 columns are the beta values for the subjects. The column names for the subjects indicate from which phenotypic group the subjects were drawn; for example, the 9744-Tumor column indicates that this subject was from the case group.

Source

Calculated via the `1_Aclust_data_import.R` script in the `old_scripts` sub-directory of the `inst` directory.

WriteBumphunterResults*Calculate and Save Bumphunter Method Results for Specified Design Points*

Description

Given a set of design points, simulate appropriate DMR data and apply the bumphunter method to them (with parameters also within the design). Write the results to a file.

Usage

```
WriteBumphunterResults(beta_mat, CPGs_df, Aclusters_df, parallel = TRUE,
  numCores = detectCores() - 2, deltas_num = c(0, 0.025, 0.05, 0.1, 0.15,
  0.2, 0.3, 0.4), seeds_int = c(100, 210, 330, 450, 680),
  cutoffQ_num = c(0.9, 0.95, 0.99), maxGap_int = c(200, 250, 500, 750,
  1000), resultsDir = "DMRcate_compare/", verbose = TRUE)
```

Arguments

beta_mat	A beta value matrix for selected methylation samples from a 450k methylation array with CPG IDs as the row names and sample IDs as the column names. An example is given in the betaVals_mat data set.
CPGs_df	An annotation table that indicates locations of CpGs. This data frame has CPG IDs as the rows with matching chromosome and location info in the columns. Specifically, the columns are: ILMNID - the CPG ID; chr - the chromosome label; and MAPINFO - the chromosome location. An example is given in the cpgLocation_df data set.
Aclusters_df	A data frame of beta values and CpG information for clusters of CpGs over a 450k methylation array. The rows correspond to the CPGs. The columns have information on the cluster number, chromosome, cluster start and end locations, and the beta values for each subject grouped by some clinical indicator (e.g. case v. control). An example is given in the startEndCPG_df data set.
parallel	Should computing be completed over multiple computing cores? Defaults to TRUE.
numCores	If parallel, how many cores should be used? Defaults to two less than the number of available cores (as calculated by the detectCores function). These cores are used internally by the bumphunter function.
deltas_num	A vector of treatment sizes: non-negative real numbers to add to the beta values within randomly-selected clusters for a single class of subjects. This artificially creates differentially-methylated regions (DMRs).
seeds_int	A vector of seed values passed to the Random function to enable reproducible results
cutoffQ_num	A vector of quantiles used for picking the cutoff using the permutation distribution, passed through the call to the internal RunBumphunter call to bumphunter .
maxGap_int	A vector of maximum location gaps, passed to the bumphunter function. These will be used to define the clusters of locations that are to be analyzed together via the clusterMaker function.
resultsDir	Where should the results be saved? Defaults to DMRcate_compare/.
verbose	Should the function print progress messages? Defaults to TRUE.

Details

This function creates matrices of all combinations of design points and all combinations of parameters. For each combination, this function executes the internal [RunBumphunter](#) function and saves the results as a compressed .RDS file.

Value

Nothing. Saves output to a file in the specified results directory.

Examples

```
## Not run:
data("betaVals_mat")
data("cpgLocation_df")
data("startEndCPG_df")

WriteBumphunterResults(
  beta_mat = betaVals_mat,
  CPGs_df = cpgLocation_df,
  Aclusters_df = startEndCPG_df
)

## End(Not run)
```

WriteDMRcateResults	<i>Calculate and Save DMRcate Method Results for Specified Design Points</i>
---------------------	--

Description

Given a set of design points, simulate appropriate DMR data and apply the dmr cate method to them (with parameters also within the design). Write the results to a file.

Usage

```
WriteDMRcateResults(beta_mat, CPGs_df, Aclusters_df, parallel = TRUE,
  numCores = detectCores() - 2, deltas_num = c(0, 0.025, 0.05, 0.1, 0.15,
  0.2, 0.3, 0.4), seeds_int = c(100, 210, 330, 450, 680),
  lambdas_num = c(200, 250, 500, 750, 1000), Cs_int = 1:5,
  resultsDir = "DMRcate_compare/", verbose = !parallel)
```

Arguments

beta_mat	A beta value matrix for selected methylation samples from a 450k methylation array with CPG IDs as the row names and sample IDs as the column names. An example is given in the betaVals_mat data set.
CPGs_df	An annotation table that indicates locations of CpGs. This data frame has CPG IDs as the rows with matching chromosome and location info in the columns. Specifically, the columns are: ILMNID - the CPG ID; chr - the chromosome label; and MAPINFO - the chromosome location. An example is given in the cpgLocation_df data set.

<code>Aclusters_df</code>	A data frame of beta values and CpG information for clusters of CpGs over a 450k methylation array. The rows correspond to the CPGs. The columns have information on the cluster number, chromosome, cluster start and end locations, and the beta values for each subject grouped by some clinical indicator (e.g. case v. control). An example is given in the <code>startEndCPG_df</code> data set.
<code>parallel</code>	Should computing be completed over multiple computing cores? Defaults to TRUE.
<code>numCores</code>	If <code>parallel</code> , how many cores should be used? Defaults to two less than the number of available cores (as calculated by the <code>detectCores</code> function).
<code>deltas_num</code>	A vector of treatment sizes: non-negative real numbers to add to the beta values within randomly-selected clusters for a single class of subjects. This artificially creates differentially-methylated regions (DMRs).
<code>seeds_int</code>	A vector of seed values passed to the <code>Random</code> function to enable reproducible results
<code>lambdas_num</code>	A vector of Gaussian kernel bandwidths for smoothed- function estimation in the called <code>dmrcate</code> function
<code>Cs_int</code>	A vector of scaling factors for bandwidth in the internal call to the <code>dmrcate</code> function
<code>resultsDir</code>	Where should the results be saved? Defaults to <code>DMRcate_compare/</code> .
<code>verbose</code>	Should the function print progress messages? Defaults to TRUE only if <code>parallel = FALSE</code> . See the internal <code>RunDMRcate</code> function for more details about parallel computing with DMRcate.

Details

This function creates matrices of all combinations of design points and all combinations of parameters. For each combination, this function executes the internal `RunDMRcate` function and saves the results as a compressed .RDS file.

Value

Nothing. Saves output to a file in the specified results directory.

Examples

```
## Not run:
data("betaVals_mat")
data("cpgLocation_df")
data("startEndCPG_df")

WriteDMRcateResults(
  beta_mat = betaVals_mat,
  CPGs_df = cpgLocation_df,
  Aclusters_df = startEndCPG_df
)

## End(Not run)
```

WriteProbeLassoResults

Calculate and Save ProbeLasso Method Results for Specified Design Points

Description

Given a set of design points, simulate appropriate DMR data and apply the ProbeLasso method (via the [champ.DMR](#) function) to them (with parameters also within the design). Write the results to a file.

Usage

```
WriteProbeLassoResults(beta_mat, CPGs_df, Aclusters_df, parallel = TRUE,
  numCores = detectCores() - 2, deltas_num = c(0, 0.025, 0.05, 0.1, 0.15,
  0.2, 0.3, 0.4), seeds_int = c(100, 210, 330, 450, 680),
  pVals_num = c(0.001, 0.01, 0.05, 0.1), aveLassoRad_int = c(375, 700,
  1000), minDmrSep_int = c(200, 250, 500, 750, 1000),
  resultsDir = "DMRcate_compare/", verbose = !parallel)
```

Arguments

beta_mat	A beta value matrix for selected methylation samples from a 450k methylation array with CPG IDs as the row names and sample IDs as the column names. An example is given in the betaVals_mat data set.
CPGs_df	An annotation table that indicates locations of CpGs. This data frame has CPG IDs as the rows with matching chromosome and location info in the columns. Specifically, the columns are: ILMNID - the CPG ID; chr - the chromosome label; and MAPINFO - the chromosome location. An example is given in the cpgLocation_df data set.
Aclusters_df	A data frame of beta values and CpG information for clusters of CpGs over a 450k methylation array. The rows correspond to the CPGs. The columns have information on the cluster number, chromosome, cluster start and end locations, and the beta values for each subject grouped by some clinical indicator (e.g. case v. control). An example is given in the startEndCPG_df data set.
parallel	Should computing be completed over multiple computing cores? Defaults to TRUE.
numCores	If parallel, how many cores should be used? Defaults to two less than the number of available cores (as calculated by the detectCores function).
deltas_num	A vector of treatment sizes: non-negative real numbers to add to the beta values within randomly-selected clusters for a single class of subjects. This artificially creates differentially-methylated regions (DMRs).
seeds_int	A vector of seed values passed to the Random function to enable reproducible results
pVals_num	A vector of the minimum thresholds of significance for probes to be includede in DMRs, passed through the RunProbeLasso function to the champ.DMR function.
aveLassoRad_int	A vector of radii around each differential methylation position to detect DMR, passed to the champ.DMR function.

<code>minDmrSep_int</code>	A vector of the minimum separation (bp) values between neighbouring DMRs, passed to the <code>champ.DMR</code> function.
<code>resultsDir</code>	Where should the results be saved? Defaults to <code>DMRcate_compare/</code> .
<code>verbose</code>	Should the function print progress messages? Defaults to <code>TRUE</code> only if <code>parallel = FALSE</code> .

Details

This function creates matrices of all combinations of design points and all combinations of parameters. For each combination, this function executes the internal `RunProbeLasso` function and saves the results as a compressed .RDS file.

Value

Nothing. Saves output to a file in the specified results directory.

Examples

```
## Not run:
data("betaVals_mat")
data("cpgLocation_df")
data("startEndCPG_df")

WriteProbeLassoResults(
  beta_mat = betaVals_mat,
  CPGs_df = cpgLocation_df,
  Aclusters_df = startEndCPG_df
)

## End(Not run)
```

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