

# quicR: An R Library for Streamlined Data Handling of Real-Time Quaking Induced Conversion Assays

Gage Rowden<sup>a,b,c,\*</sup>, Peter Larsen<sup>a,b,c</sup>

<sup>a</sup>*Department of Veterinary and Biomedical Sciences, University of Minnesota, USA.*

<sup>b</sup>*Minnesota Center for Prion Research and Outreach, University of Minnesota, USA.*

<sup>c</sup>*Priogen Corp., USA.*

---

## Abstract

Real-time quaking induced conversion (RT-QuIC) has quickly become a valuable diagnostic tool for protein misfolding disorders such as Creutzfeldt-Jakob disease and Parkinson's disease. Given that the technology is relatively new, academic and industry standards for quality filtering data and high throughput analysis of results have yet to be fully established. The open source R library, quicR, was developed to provide a standardized approach to RT-QuIC data analysis. quicR provides functions, which can be easily integrated into existing R workflows, for data curation, analysis, and visualization.

*Keywords:* R package, RT-QuIC, prion, diagnostics, CJD, Parkinson's

---

## Metadata

| Nr. | Code metadata description                                       | Metadata  |
|-----|---|---|
| C1  | Current code version  | V2.1.0  |
| C2  | Permanent link to code/repository used for this code version    | <a href="https://github.com/gage1145/quicR">https://github.com/gage1145/quicR</a>   |
| C3  | Permanent link to Reproducible Capsule                          | <a href="https://github.com/gage1145/quicR/releases/tag/v2.1.0">https://github.com/gage1145/quicR/releases/tag/v2.1.0</a>     |
| C4  | Legal Code License  | GPL-3   |
| C5  | Code versioning system used                                     | git   |
| C6  | Software code languages, tools, and services used               | R   |
| C7  | Compilation requirements, operating environments & dependencies | R (>=4.1.0)   |
| C8  | If available Link to developer documentation/manual             | <a href="https://cran.r-project.org/web/packages/quicR/quicR.pdf">https://cran.r-project.org/web/packages/quicR/quicR.pdf</a> |
| C9  | Support email for questions                                     | rowde002@umn.edu  |

Table 1: Code metadata

---

\*Corresponding author.

<sup>1</sup> *E-mail address:* rowde002@umn.edu

## 1. Motivation and significance

Real-time quaking induced conversion (RT-QuIC) is a cutting-edge diagnostic assay that has garnered significant attention for its ultra-sensitive detection of misfolded protein aggregates [1, 2]. The assay works by converting a recombinant protein substrate into an amyloid aggregate in the presence of a misfolded seed [1, 3, 4, 5, 6, 7, 8, 9, 10]. The assay’s sensitivity and specificity make RT-QuIC a promising tool for diagnosing diseases such as prion disorders and other protein misfolding pathologies [11, 12, 13, 14]. However, the relatively recent development and novelty of the assay have left a gap in widely accepted academic and industry standards for data analysis and interpretation [15].

To address this gap, we introduce quicR, an open-source library, developed in R [16], dedicated to the cleaning, analysis, and visualization of RT-QuIC data. By consolidating key metrics and providing robust analytical tools, quicR aims to standardize the analysis pipeline and foster reproducibility within the field of quaking induced assays including related assays such as Nano-QuIC [17] and Micro-QuIC [18]. quicR is designed with both researchers and diagnosticians in mind, providing a user-friendly interface that integrates seamlessly with existing R workflows.

While universal diagnostic criteria for RT-QuIC have yet to be established, certain analytical metrics have emerged as valuable tools for interpreting assay results and kinetics. These include:

1. Time-to-threshold (TtT): The time required for the fluorescence signal to exceed a predefined threshold [5].
2. Rate of amyloid formation (RAF): A measure of the kinetics of aggregate growth, which provides insight into the relative quantity of misfolded seed [19].
3. Maxpoint ratio (MPR): A ratio-based metric measuring peak normalized fluorescence intensities [15].
4. Maximum slope (MS): The steepest rate of fluorescence increase, reflecting the most rapid phase of aggregation [20].

Together, these metrics enable researchers to characterize the kinetics of RT-QuIC reactions comprehensively, enhancing the rigor and reliability of diagnostic decisions.

In addition to analytical tools, quicR provides flexible and customizable visualization capabilities. Leveraging the powerful ggplot2 library [21], quicR enables users to generate high-quality, publication-ready figures. These visualizations can be further customized using the intuitive ‘+’ syntax of ggplot2, allowing for tailored presentations of RT-QuIC data.

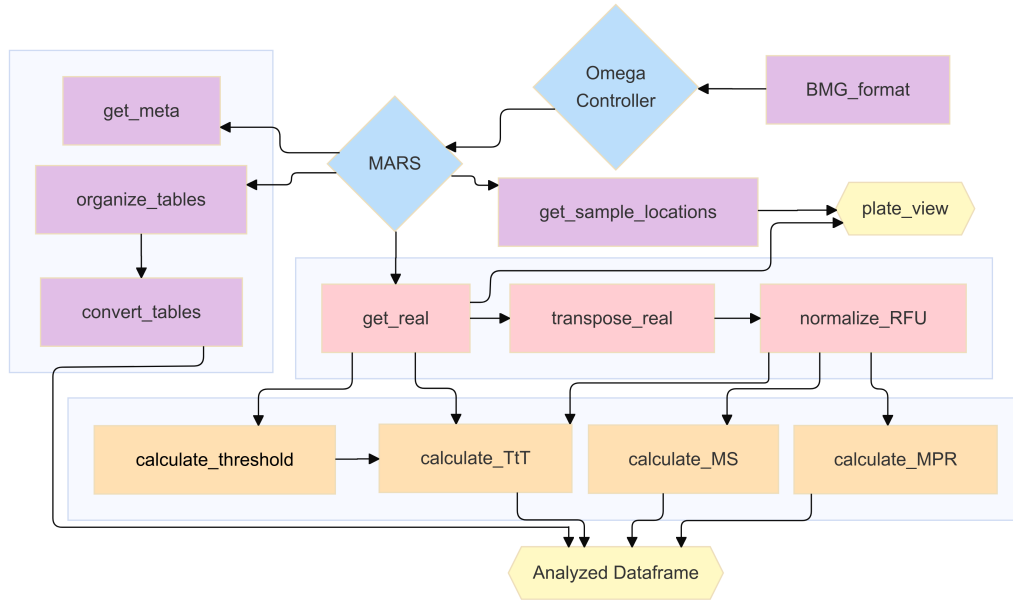
By combining standardized metrics, advanced visualization tools, and a commitment to open source science, quicR serves as a foundational resource for the growing RT-QuIC community. Its goal is to empower researchers to analyze and present their data with clarity, consistency, and cohesion.

## 2. Software description

### 2.1. Software architecture

quicR was developed to address the growing need for efficient data conversion, analysis, and visualization of RT-QuIC data. With a focus on usability and reproducibility, the package is designed to standardize workflows and ensure compatibility across multiple laboratories. Its primary input format is data exported as Excel workbooks from the proprietary MARS software (BMG Labtech, Ortenberg, Germany), providing seamless integration with existing experimental workflows. See Figure 1 for a detailed workflow.

Figure 1: Workflow hierarchy of the quicR package. Blue nodes indicate steps where BMG software is needed. Purple nodes indicate functions dedicated to handling metadata. Red nodes are functions that acquire and manipulate raw data. Orange nodes are functions which calculate some metric. Finally, yellow nodes represent data analysis endpoints.



### 2.2. Software functionalities

*Present the major functionalities of the software.*

### *2.3. Sample code snippets analysis (optional)*

## **3. Illustrative examples**

## **4. Impact**

## **5. Conclusions**

quicR offers a powerful solution for the cleaning, analysis, and visualization of RT-QuIC data, addressing critical needs in a rapidly evolving field. By enabling consistent data handling and interpretation, quicR lays the groundwork for improved diagnostic consistency and reproducibility. The package's open-source nature ensures that it will continue to evolve, integrating new insights and technologies as they emerge.

As RT-QuIC technology advances, tools like quicR will play a pivotal role in bridging the gap between assay development and practical application. By equipping researchers with reliable, standardized tools, quicR not only supports the study of prion and protein misfolding disorders but also serves as a model for the development of software solutions in other diagnostic fields.

## **Acknowledgements**

Special thanks to Beni Altmann at The Comprehensive R Archive Network (CRAN) for help during the submission process to CRAN. We thank Tiffany Wolf and Marc Schwabenlander for their support through the Minnesota Center for Prion Research and Outreach. We would like to acknowledge Suzanne Stone and Sarah Gresch for maintaining lab operations.

## **References**

- [1] J. M. Wilham, C. D. Orrú, R. A. Bessen, R. Atarashi, K. Sano, B. Race, K. D. Meade-White, L. M. Taubner, A. Timmes, B. Caughey, Rapid end-point quantitation of prion seeding activity with sensitivity comparable to bioassays, *PLOS Pathog* 6 (12 2010). doi:10.1371/journal.ppat.1001217.
- [2] R. Atarashi, K. Sano, K. Satoh, N. Nishida, Real-time quaking-induced conversion: A highly sensitive assay for prion detection, *Prion* 5 (2011) 150–153. doi:10.4161/pri.5.3.16893.  
URL <https://www.ncbi.nlm.nih.gov/pmc/articles/PMC3226039/>
- [3] C. D. Orrú, J. M. Wilham, L. D. Raymond, F. Kuhn, B. Schroeder, A. J. Raeber, B. Caughey, Prion disease blood test using immunoprecipitation and improved quaking-induced conversion, *mBio* 2 (3) (2012). doi:10.1128/mBio.00078-11.

- [4] C. D. Orrù, B. R. Groveman, A. G. Hughson, M. Manca, L. D. Raymond, G. J. Raymond, K. J. Campbell, K. J. Anson, A. Kraus, B. Caughey, RT-QuIC assays for prion disease detection and diagnostics, in: *Methods in Molecular Biology*, Vol. 1658, Humana Press Inc., 2017, pp. 185–203.
- [5] C. D. Orrù, B. R. Groveman, A. G. Hughson, G. Zanusso, M. B. Coulthart, B. Caughey, Rapid and sensitive rt-quic detection of human creutzfeldt-jakob disease using cerebrospinal fluid, *mBio* 6 (1) (2015). doi:10.1128/mBio.02451-14.  
URL [/pmc/articles/PMC4313917//pmc/articles/PMC4313917/?report=abstracthttps://www.ncbi.nlm.nih.gov/pmc/articles/PMC4313917/](https://www.ncbi.nlm.nih.gov/pmc/articles/PMC4313917/)
- [6] M. Bongianni, A. Ladogana, S. Capaldi, S. Klotz, S. Baiardi, A. Cagnin, D. Perra, M. Fiorini, A. Poggi, G. Legname, T. Cattaruzza, F. Janes, M. Tabaton, B. Ghetti, S. Monaco, G. G. Kovacs, P. Parchi, M. Pocchiari, G. Zanusso,  $\alpha$ -Synuclein RT-QuIC assay in cerebrospinal fluid of patients with dementia with lewy bodies, *Annals of Clinical and Translational Neurology* 6 (10) (2019) 2120–2126.
- [7] R. P. Dassanayake, C. D. Orrù, A. G. Hughson, B. Caughey, T. Graça, D. Zhuang, S. A. Madsen-Bouterse, D. P. Knowles, D. A. Schneider, Sensitive and specific detection of classical scrapie prions in the brains of goats by real-time quaking-induced conversion, *J. Gen. Virol.* 97 (3) (2016) 803–812.
- [8] S. Hwang, M. H. West Greenlee, A. Balkema-Buschmann, M. H. Groschup, E. M. Nicholson, J. J. Greenlee, Real-time quaking-induced conversion detection of bovine spongiform encephalopathy prions in a subclinical steer, *Frontiers in Veterinary Science* 4 (JAN) (2018) 19.
- [9] B. R. Groveman, C. D. Orrù, A. G. Hughson, L. D. Raymond, G. Zanusso, B. Ghetti, K. J. Campbell, J. Safar, D. Galasko, B. Caughey, Rapid and ultra-sensitive quantitation of disease-associated  $\alpha$ -synuclein seeds in brain and cerebrospinal fluid by  $\alpha$ Syn RT-QuIC, *Acta Neuropathologica Communications* 6 (1) (2018) 7.
- [10] M. A. Metrick, 2nd, N. d. C. Ferreira, E. Saijo, A. Kraus, K. Newell, G. Zanusso, M. Vendruscolo, B. Ghetti, B. Caughey, A single ultrasensitive assay for detection and discrimination of tau aggregates of alzheimer and pick diseases, *Acta Neuropathol Commun* 8 (1) (2020) 22.

- [11] M. Fiorini, G. Iselle, D. Perra, M. Bongiani, S. Capaldi, L. Sacchetto, S. Ferrari, A. Mombello, S. Vascellari, S. Testi, S. Monaco, G. Zanusso, High diagnostic accuracy of rt-quic assay in a prospective study of patients with suspected scjd, *International Journal of Molecular Sciences* 21 (3) (2020). doi:10.3390/ijms21030880.  
URL <https://www.mdpi.com/1422-0067/21/3/880>
- [12] A. Franceschini, S. Baiardi, A. G. Hughson, N. McKenzie, F. Moda, M. Rossi, S. Capellari, A. Green, G. Giaccone, B. Caughey, P. Parchi, High diagnostic value of second generation csf rt-quic across the wide spectrum of cjd prions, *Sci Rep* (9 2017). doi:10.1038/s41598-017-10922-w.
- [13] C. Picasso-Risso, M. D. Schwabenlander, G. Rowden, M. Carstensen, J. C. Bartz, P. A. Larsen, T. M. Wolf, Assessment of Real-Time Quaking-Induced conversion (RT-QuIC) assay, immunohistochemistry and ELISA for detection of chronic wasting disease under field conditions in White-Tailed deer: A bayesian approach, *Pathogens* 11 (5) (2022).
- [14] C. L. Holz, J. R. Darish, K. Straka, N. Grosjean, S. Bolin, M. Kiupel, S. Sreevatsan, Evaluation of Real-Time Quaking-Induced conversion, ELISA, and immunohistochemistry for chronic wasting disease diagnosis, *Front Vet Sci* 8 (2021) 824815.
- [15] G. R. Rowden, C. Picasso-Risso, M. Li, M. D. Schwabenlander, T. M. Wolf, P. A. Larsen, Standardization of data analysis for rt-quic-based detection of chronic wasting disease, *Pathogens* 12 (2023) 309. doi:10.3390/PATHOGENS12020309/S1.  
URL <https://www.mdpi.com/2076-0817/12/2/309/html><https://www.mdpi.com/2076-0817/12/2/309>
- [16] R Core Team, R: A Language and Environment for Statistical Computing, R Foundation for Statistical Computing, Vienna, Austria (2024).  
URL <https://www.R-project.org/>
- [17] P. R. Christenson, M. Li, G. Rowden, P. A. Larsen, S.-H. Oh, Nanoparticle-enhanced rt-quic (nano-quic) diagnostic assay for misfolded proteins, *Nano Letters* 23 (9) (2023) 4074–4081, pMID: 37126029. arXiv:<https://doi.org/10.1021/acs.nanolett.3c01001>, doi:10.1021/acs.nanolett.3c01001.  
URL <https://doi.org/10.1021/acs.nanolett.3c01001>

- [18] D. J. Lee, P. R. Christenson, G. Rowden, N. C. Lindquist, P. A. Larsen, S.-H. Oh, Rapid on-site amplification and visual detection of misfolded proteins via microfluidic quaking-induced conversion (micro-quic), *npj Biosensing* 1 (6) (7 2024). doi:10.1038/s44328-024-00006-x.  
URL <https://www.nature.com/articles/s44328-024-00006-x#citeas>
- [19] N. J. Gallups, A. S. Harms, ‘seeding’ the idea of early diagnostics in synucleinopathies, *Brain* 145 (2022) 418–419. doi:10.1093/BRAIN/AWAC062.  
URL <https://dx.doi.org/10.1093/brain/awac062>
- [20] D. M. Henderson, K. A. Davenport, N. J. Haley, N. D. Denkers, C. K. Mathiason, E. A. Hoover, Quantitative assessment of prion infectivity in tissues and body fluids by real-time quaking-induced conversion, *Journal of General Virology* 96 (2015) 210–219. doi:10.1099/vir.0.069906-0.
- [21] H. Wickham, *ggplot2: Elegant Graphics for Data Analysis*, Springer-Verlag New York, 2016.  
URL <https://ggplot2.tidyverse.org>