

Class 9: Structural Bioinformatics 1

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The main database for structural data is called the PDB (Protein Data Bank). Let's see what it contains:

Data from: <https://www.rcsb.org/stats> URL: https://bioboot.github.io/bimm143_F24/class-material/pdb_stats.csv

Read this into R

```
pdb_statistics <- read.csv("pdb_stats.csv", row.names=1)
pdb_statistics
```

	X.ray	EM	NMR	Multiple.methods	Neutron	Other
Protein (only)	167,192	15,572	12,529	208	77	32
Protein/Oligosaccharide	9,639	2,635	34	8	2	0
Protein/NA	8,730	4,697	286	7	0	0
Nucleic acid (only)	2,869	137	1,507	14	3	1
Other	170	10	33	0	0	0
Oligosaccharide (only)	11	0	6	1	0	4
Total						
Protein (only)	195,610					
Protein/Oligosaccharide	12,318					
Protein/NA	13,720					
Nucleic acid (only)	4,531					
Other	213					
Oligosaccharide (only)	22					

Q1: What percentage of structures in the PDB are solved by X-Ray and Electron Microscopy?

```
pdb_statistics$Total
```

```
[1] "195,610" "12,318" "13,720" "4,531" "213" "22"
```

I need to remove the comma and convert it to numeric in order to do math:

```
# get rid of the commas to make it integers
as.numeric(sub(",", "", pdb_statistics$Total))
```

```
[1] 195610 12318 13720 4531 213 22
```

I could turn this into a function to fix the whole table or any future table I read like this:

```
x <- pdb_statistics$Total
as.numeric(sub(",", "", x))
```

```
[1] 195610 12318 13720 4531 213 22
```

```
comma2numeric <- function(x) {
  as.numeric(sub(",", "", x))
}
```

Test it

```
comma2numeric(pdb_statistics$X.ray)
```

```
[1] 167192 9639 8730 2869 170 11
```

```
apply(pdb_statistics, 2, comma2numeric)
```

	X.ray	EM	NMR	Multiple.methods	Neutron	Other	Total
[1,]	167192	15572	12529	208	77	32	195610
[2,]	9639	2635	34	8	2	0	12318
[3,]	8730	4697	286	7	0	0	13720
[4,]	2869	137	1507	14	3	1	4531
[5,]	170	10	33	0	0	0	213
[6,]	11	0	6	1	0	4	22

Or try a different read/import function:

```
library(readr)
pdb_statistics <- read_csv("pdb_stats.csv")
```

```
sum(pdb_statistics$Total)
```

```
[1] 226414
```

```
sum(pdb_statistics$`X-ray`)/sum(pdb_statistics$Total)*100
```

```
[1] 83.30359
```

```
sum(pdb_statistics$EM)/sum(pdb_statistics$Total)*100
```

```
[1] 10.18091
```

X-ray: 83.30% Electron Microscopy: 10.18%

Q2: What proportion of structures in the PDB are protein?

```
pdb_statistics[1, "Total"]
```

```
# A tibble: 1 x 1
  Total
  <dbl>
1 195610
```

Calculating the total amount of structures.

```
sum(pdb_statistics[, "Total"])
```

```
[1] 226414
```

Dividing the total proteins by the total amount.

```
pdb_statistics[1, "Total"]/sum(pdb_statistics[, "Total"])*100
```

```
Total
1 86.39483
```

Or

```
pdb_statistics$Total[1]/sum(pdb_statistics$Total)*100
```

```
[1] 86.39483
```

Protein: 86.39%

Q3: Type HIV in the PDB website search box on the home page and determine how many HIV-1 protease structures are in the current PDB?

There are 4563 structures

Mol*

Mol* (pronounced “molstar”) is a new web-based molecular viewer that we will need to learn the basics of here.

<https://molstar.org/viewer/>

We will use PDB code: 1HSG

Q4: Water molecules normally have 3 atoms. Why do we see just one atom per water molecule in this structure?

We see only one atom per water molecule, representing oxygen. This is most likely because the structure is already complex and they want to minimize/simplify the representations of the water molecules.

Q5: There is a critical “conserved” water molecule in the binding site. Can you identify this water molecule? What residue number does this water molecule have?

This water molecule has the residue number 308.



Figure 1: A first image from molstar

Some more custom images:



Figure 2: The all important catalytic ASP25 amino acids

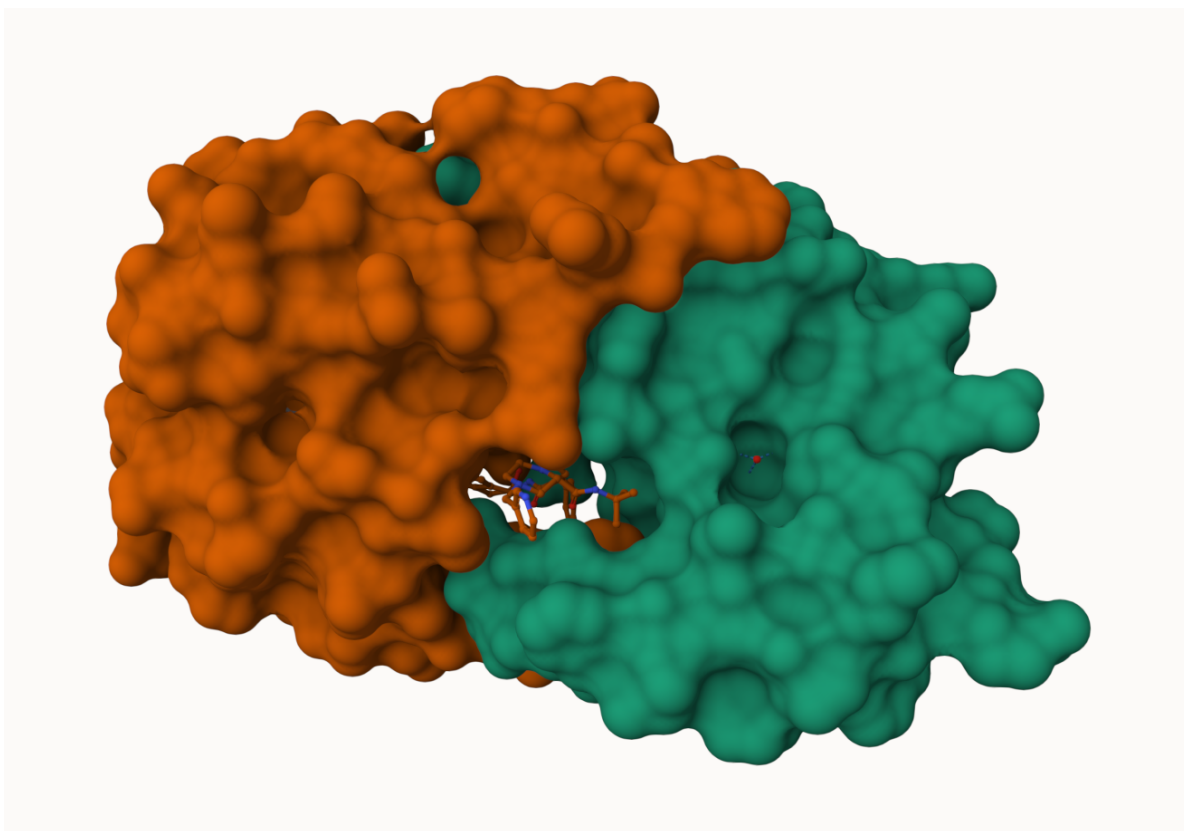


Figure 3: Surface display

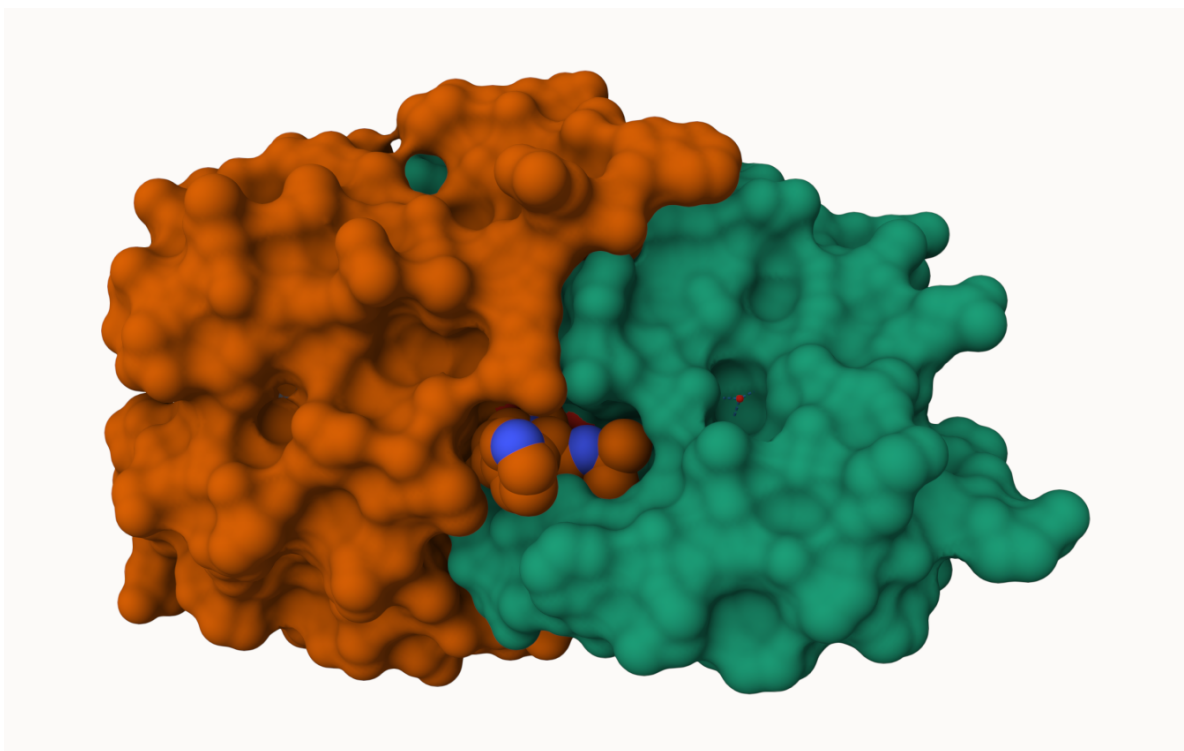
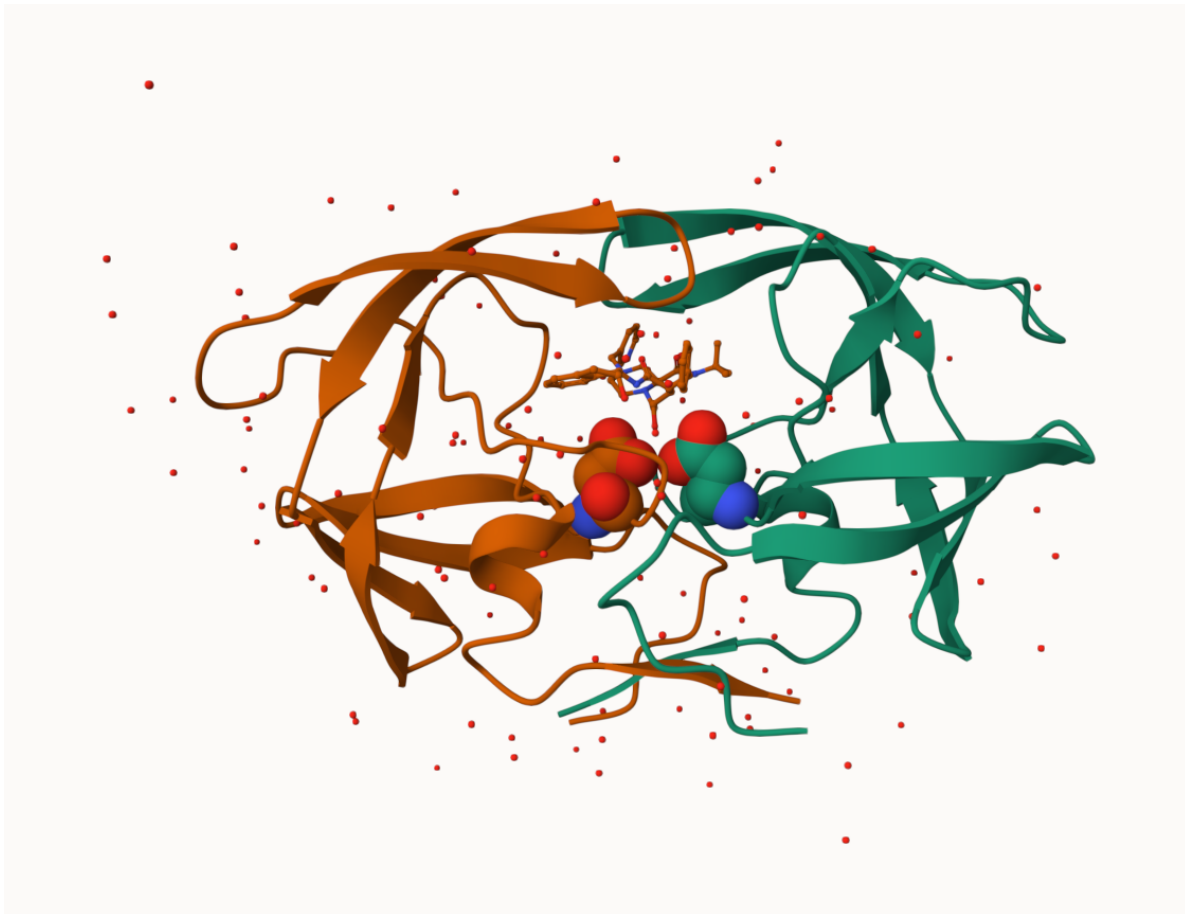


Figure 4: Surface display showing the ligand fitting into the compound

Q6: Generate and save a figure clearly showing the two distinct chains of HIV-protease along with the ligand. You might also consider showing the catalytic residues ASP 25 in each chain and the critical water (we recommend “Ball & Stick” for these side-chains). Add this figure to your Quarto document.



The Bio3D package

The bio3d package allows us to do all sorts of structural bioinformatics work in R.

Let's start with how it can read these PDB files:

```
library(bio3d)  
pdb <- read.pdb("1hsg")
```

Note: Accessing on-line PDB file

```
pdb
```

```
Call: read.pdb(file = "1hsg")
```

```
Total Models#: 1
```

```
Total Atoms#: 1686, XYZs#: 5058 Chains#: 2 (values: A B)
```

```
Protein Atoms#: 1514 (residues/Calpha atoms#: 198)
```

```
Nucleic acid Atoms#: 0 (residues/phosphate atoms#: 0)
```

```
Non-protein/nucleic Atoms#: 172 (residues: 128)
```

```
Non-protein/nucleic resid values: [ HOH (127), MK1 (1) ]
```

```
Protein sequence:
```

```
PQITLWQRPLVTIKIGGQLKEALLDTGADDTVLEEMSLPGRWKPKMIGGIGGFIKVRQYD  
QILIEICGHKAIGTVLVGPTPVNIIGRNLLTQIGCTLNFPQITLWQRPLVTIKIGGQLKE  
ALLDTGADDTVLEEMSLPGRWKPKMIGGIGGFIKVRQYDQILIEICGHKAIGTVLVGPTP  
VNIIGRNLLTQIGCTLNF
```

```
+ attr: atom, xyz, seqres, helix, sheet,  
      calpha, remark, call
```

```
attributes(pdb)
```

```
$names
```

```
[1] "atom" "xyz" "seqres" "helix" "sheet" "calpha" "remark" "call"
```

```
$class
```

```
[1] "pdb" "sse"
```

```
head(pdb$atom)
```

	type	eleno	elety	alt	resid	chain	resno	insert	x	y	z	o	b
1	ATOM	1	N	<NA>	PRO	A	1	<NA>	29.361	39.686	5.862	1	38.10
2	ATOM	2	CA	<NA>	PRO	A	1	<NA>	30.307	38.663	5.319	1	40.62
3	ATOM	3	C	<NA>	PRO	A	1	<NA>	29.760	38.071	4.022	1	42.64
4	ATOM	4	O	<NA>	PRO	A	1	<NA>	28.600	38.302	3.676	1	43.40
5	ATOM	5	CB	<NA>	PRO	A	1	<NA>	30.508	37.541	6.342	1	37.87
6	ATOM	6	CG	<NA>	PRO	A	1	<NA>	29.296	37.591	7.162	1	38.40
	segid	elesy	charge										
1	<NA>	N	<NA>										
2	<NA>	C	<NA>										

```
3 <NA>      C   <NA>
4 <NA>      O   <NA>
5 <NA>      C   <NA>
6 <NA>      C   <NA>
```

```
pdbseq(pdb)[25]
```

```
25
"D"
```

Q7: How many amino acid residues are there in this pdb object?

```
sum(pdb$calpha)
```

```
[1] 198
```

or

```
length(pdbseq(pdb))
```

```
[1] 198
```

198 calpha residues

Q8: Name one of the two non-protein residues?

HOH and MK1

Q9: How many protein chains are in this structure?

```
unique(pdb$atom$chain)
```

```
[1] "A" "B"
```

2 protein chains

Predicting functional motions of a single structure

Let's do a bioinformatics prediction of functional motions - i.e. the movements that one of these molecules needs to make to do its stuff.

```
adk <- read.pdb("6s36")
```

Note: Accessing on-line PDB file
PDB has ALT records, taking A only, rm.alt=TRUE

```
adk
```

```
Call: read.pdb(file = "6s36")
```

```
Total Models#: 1
```

```
Total Atoms#: 1898, XYZs#: 5694 Chains#: 1 (values: A)
```

```
Protein Atoms#: 1654 (residues/Calpha atoms#: 214)
```

```
Nucleic acid Atoms#: 0 (residues/phosphate atoms#: 0)
```

```
Non-protein/nucleic Atoms#: 244 (residues: 244)
```

```
Non-protein/nucleic resid values: [ CL (3), HOH (238), MG (2), NA (1) ]
```

```
Protein sequence:
```

```
MRIILLGAPGAGKGTQAQFIMEKYGIPQISTGDMRLAAVKSSELGKQAKDIMDAGKLV  
DELVIALVKERIAQEDCRNGFLDGFRTIPQADAMKEAGINVDYVLEFDVPDELIVDKI  
VGRRVHAPSGRVYHVKFNPVKVEGKDDVTGEELTTRKDDQEETVRKRLVEYHQMTAPLIG  
YYSKEAEAGNTKYAKVDGTPVAEVRADLEKILG
```

```
+ attr: atom, xyz, seqres, helix, sheet,  
      calpha, remark, call
```

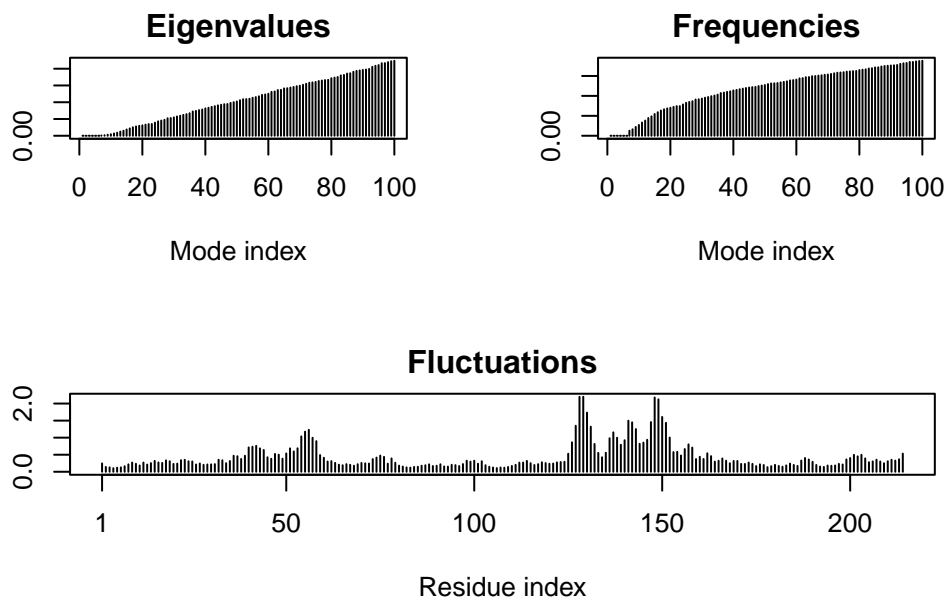
```
# Perform flexibility prediction
```

```
m <- nma(adk)
```

```
Building Hessian... Done in 0.014 seconds.
```

```
Diagonalizing Hessian... Done in 0.282 seconds.
```

```
plot(m)
```



Write out multi-model PDB file (trajectory) that we can use to make an animation of the predicted motions.

```
mktrj(m, file="adk_m7.pdb")
```

I can open this in Mol* to play the trajectory...

Comparative analysis of protein structures

```
library(bio3d)
```

Here we will find and analyze all ADK structures in the PDB database.

We will start with a single database accession id: "1ake_A"

```
id <- "1ake_A"
aa <- get.seq(id)
```

Warning in get.seq(id): Removing existing file: seqs.fasta

Fetching... Please wait. Done.

I ran these cmds in the R brain/console

```
install.packages("BiocManager") BiocManager::install("msa")
```

Q10. Which of the packages above is found only on BioConductor and not CRAN?

BioConductor = whole new set of packages purely for bioinformatics; the **msa** package is only on BioConductor

Q11. Which of the above packages is not found on BioConductor or CRAN?

Q12. True or False? Functions from the devtools package can be used to install packages from GitHub and BitBucket?

True

Q13. How many amino acids are in this sequence, i.e. how long is this sequence?

```
length(aa)
```

```
[1] 3
```

```
attributes(aa)
```

```
$names
```

```
[1] "id"    "ali"   "call"
```

```
$class
```

```
[1] "fasta"
```

```
aa$id
```

```
[1] "pdb|1AKE|A"
```

```
ncol(aa$ali)
```

```
[1] 214
```

```
aa
```

```

      1      .      .      .      .      .      .      60
pdb|1AKE|A  MRIILLGAPGAGKGTQAQFIMEKYGIPQISTGDMLRAAVKSGSELGKQAKDIMDAGKLV
      1      .      .      .      .      .      .      60

      61      .      .      .      .      .      .      120
pdb|1AKE|A  DELVIALVKERIAQEDCRNGFLLDGFRTIPQADAMKEAGINVDYVLEFDVPDELIVDRI
      61      .      .      .      .      .      .      120

      121      .      .      .      .      .      .      180
pdb|1AKE|A  VGRRVHAPSGRVYHVKNPPKVEGKDDVTGEELTTRKDDQEETVRKRLVEYHQMTAPLIG
      121      .      .      .      .      .      .      180

      181      .      .      .      214
pdb|1AKE|A  YYSKEAEAGNTKYAKVDGTPVAEVRADLEKILG
      181      .      .      .      214

```

Call:

```
read.fasta(file = outfile)
```

Class:

```
fasta
```

Alignment dimensions:

```
1 sequence rows; 214 position columns (214 non-gap, 0 gap)
```

```
+ attr: id, ali, call
```

There are 214 amino acids.

```
# Blast or hmmer search
b <- blast.pdb(aa)
```

```
Searching ... please wait (updates every 5 seconds) RID = JMWA6HTP013
```

```
.....
```

```
Reporting 85 hits
```

```
attributes(b)
```

```
$names
```

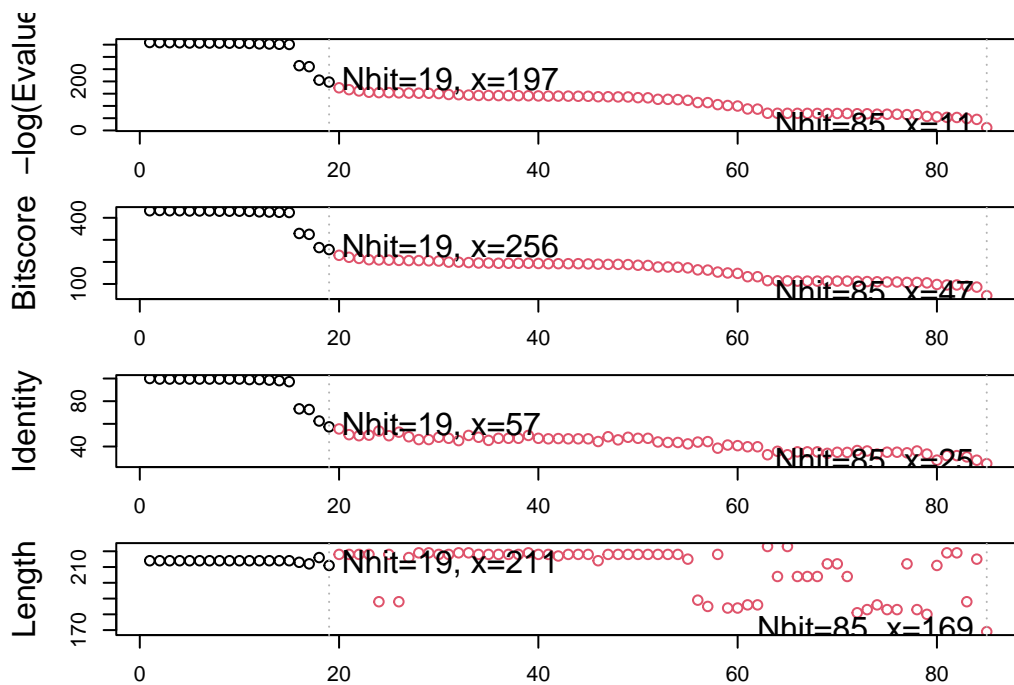
```
[1] "hit.tbl" "raw"      "url"
```

```
$class
[1] "blast"
```

```
# Plot a summary of search results
hits <- plot(b)
```

```
* Possible cutoff values:    197 11
    Yielding Nhits:         19 85
```

```
* Chosen cutoff value of:    197
    Yielding Nhits:         19
```



```
# List out some 'top hits'
hits$ pdb.id
```

```
[1] "1AKE_A" "8BQF_A" "4X8M_A" "6S36_A" "8Q2B_A" "8RJ9_A" "6RZE_A" "4X8H_A"
[9] "3HPR_A" "1E4V_A" "5EJE_A" "1E4Y_A" "3X2S_A" "6HAP_A" "6HAM_A" "4K46_A"
[17] "4NP6_A" "3GMT_A" "4PZL_A"
```



```
# Download related PDB files
files <- get.pdb(hits$pdb.id, path="pdbs", split=TRUE, gzip=TRUE)
```

```
Warning in get.pdb(hits$pdb.id, path = "pdbs", split = TRUE, gzip = TRUE):
pdbs/1AKE.pdb.gz exists. Skipping download
```

```
Warning in get.pdb(hits$pdb.id, path = "pdbs", split = TRUE, gzip = TRUE):
pdbs/8BQF.pdb.gz exists. Skipping download
```

```
Warning in get.pdb(hits$pdb.id, path = "pdbs", split = TRUE, gzip = TRUE):
pdbs/4X8M.pdb.gz exists. Skipping download
```

```
Warning in get.pdb(hits$pdb.id, path = "pdbs", split = TRUE, gzip = TRUE):
pdbs/6S36.pdb.gz exists. Skipping download
```

```
Warning in get.pdb(hits$pdb.id, path = "pdbs", split = TRUE, gzip = TRUE):
pdbs/8Q2B.pdb.gz exists. Skipping download
```

```
Warning in get.pdb(hits$pdb.id, path = "pdbs", split = TRUE, gzip = TRUE):
pdbs/8RJ9.pdb.gz exists. Skipping download
```

```
Warning in get.pdb(hits$pdb.id, path = "pdbs", split = TRUE, gzip = TRUE):
pdbs/6RZE.pdb.gz exists. Skipping download
```

```
Warning in get.pdb(hits$pdb.id, path = "pdbs", split = TRUE, gzip = TRUE):
pdbs/4X8H.pdb.gz exists. Skipping download
```

```
Warning in get.pdb(hits$pdb.id, path = "pdbs", split = TRUE, gzip = TRUE):
pdbs/3HPR.pdb.gz exists. Skipping download
```

```
Warning in get.pdb(hits$pdb.id, path = "pdbs", split = TRUE, gzip = TRUE):
pdbs/1E4V.pdb.gz exists. Skipping download
```

```
Warning in get.pdb(hits$pdb.id, path = "pdbs", split = TRUE, gzip = TRUE):
pdbs/5EJE.pdb.gz exists. Skipping download
```

```
Warning in get.pdb(hits$pdb.id, path = "pdbs", split = TRUE, gzip = TRUE):
pdbs/1E4Y.pdb.gz exists. Skipping download
```

Warning in get.pdb(hits\$pdb.id, path = "pdbs", split = TRUE, gzip = TRUE):
pdbs/3X2S.pdb.gz exists. Skipping download

Warning in get.pdb(hits\$pdb.id, path = "pdbs", split = TRUE, gzip = TRUE):
pdbs/6HAP.pdb.gz exists. Skipping download

Warning in get.pdb(hits\$pdb.id, path = "pdbs", split = TRUE, gzip = TRUE):
pdbs/6HAM.pdb.gz exists. Skipping download

Warning in get.pdb(hits\$pdb.id, path = "pdbs", split = TRUE, gzip = TRUE):
pdbs/4K46.pdb.gz exists. Skipping download

Warning in get.pdb(hits\$pdb.id, path = "pdbs", split = TRUE, gzip = TRUE):
pdbs/4NP6.pdb.gz exists. Skipping download

Warning in get.pdb(hits\$pdb.id, path = "pdbs", split = TRUE, gzip = TRUE):
pdbs/3GMT.pdb.gz exists. Skipping download

Warning in get.pdb(hits\$pdb.id, path = "pdbs", split = TRUE, gzip = TRUE):
pdbs/4PZL.pdb.gz exists. Skipping download

		0%
====		5%
=====		11%
=====		16%
=====		21%
=====		26%
=====		32%
=====		37%
=====		42%



Next we will use the `pdbaln()` function to align and also optionally fit (i.e. superpose) the identified PDB structures.

```
# Align related PDBs
pdbs <- pdbaln(files, fit = TRUE, exefile="msa")
```

Reading PDB files:

```
pdbs/split_chain/1AKE_A.pdb
pdbs/split_chain/8BQF_A.pdb
pdbs/split_chain/4X8M_A.pdb
pdbs/split_chain/6S36_A.pdb
pdbs/split_chain/8Q2B_A.pdb
pdbs/split_chain/8RJ9_A.pdb
pdbs/split_chain/6RZE_A.pdb
pdbs/split_chain/4X8H_A.pdb
pdbs/split_chain/3HPR_A.pdb
pdbs/split_chain/1E4V_A.pdb
pdbs/split_chain/5EJE_A.pdb
```

```

pdb/split_chain/1E4Y_A.pdb
pdb/split_chain/3X2S_A.pdb
pdb/split_chain/6HAP_A.pdb
pdb/split_chain/6HAM_A.pdb
pdb/split_chain/4K46_A.pdb
pdb/split_chain/4NP6_A.pdb
pdb/split_chain/3GMT_A.pdb
pdb/split_chain/4PZL_A.pdb
    PDB has ALT records, taking A only, rm.alt=TRUE
.   PDB has ALT records, taking A only, rm.alt=TRUE
..  PDB has ALT records, taking A only, rm.alt=TRUE
.   PDB has ALT records, taking A only, rm.alt=TRUE
.   PDB has ALT records, taking A only, rm.alt=TRUE
.   PDB has ALT records, taking A only, rm.alt=TRUE
..  PDB has ALT records, taking A only, rm.alt=TRUE
..  PDB has ALT records, taking A only, rm.alt=TRUE
.... PDB has ALT records, taking A only, rm.alt=TRUE
.   PDB has ALT records, taking A only, rm.alt=TRUE
....

```

Extracting sequences

```

pdb/seq: 1   name: pdb/split_chain/1AKE_A.pdb
    PDB has ALT records, taking A only, rm.alt=TRUE
pdb/seq: 2   name: pdb/split_chain/8BQF_A.pdb
    PDB has ALT records, taking A only, rm.alt=TRUE
pdb/seq: 3   name: pdb/split_chain/4X8M_A.pdb
pdb/seq: 4   name: pdb/split_chain/6S36_A.pdb
    PDB has ALT records, taking A only, rm.alt=TRUE
pdb/seq: 5   name: pdb/split_chain/8Q2B_A.pdb
    PDB has ALT records, taking A only, rm.alt=TRUE
pdb/seq: 6   name: pdb/split_chain/8RJ9_A.pdb
    PDB has ALT records, taking A only, rm.alt=TRUE
pdb/seq: 7   name: pdb/split_chain/6RZE_A.pdb
    PDB has ALT records, taking A only, rm.alt=TRUE
pdb/seq: 8   name: pdb/split_chain/4X8H_A.pdb
pdb/seq: 9   name: pdb/split_chain/3HPR_A.pdb
    PDB has ALT records, taking A only, rm.alt=TRUE
pdb/seq: 10  name: pdb/split_chain/1E4V_A.pdb
pdb/seq: 11  name: pdb/split_chain/5EJE_A.pdb
    PDB has ALT records, taking A only, rm.alt=TRUE
pdb/seq: 12  name: pdb/split_chain/1E4Y_A.pdb
pdb/seq: 13  name: pdb/split_chain/3X2S_A.pdb

```

```

pdb/seq: 14  name: pdbs/split_chain/6HAP_A.pdb
pdb/seq: 15  name: pdbs/split_chain/6HAM_A.pdb
PDB has ALT records, taking A only, rm.alt=TRUE
pdb/seq: 16  name: pdbs/split_chain/4K46_A.pdb
PDB has ALT records, taking A only, rm.alt=TRUE
pdb/seq: 17  name: pdbs/split_chain/4NP6_A.pdb
pdb/seq: 18  name: pdbs/split_chain/3GMT_A.pdb
pdb/seq: 19  name: pdbs/split_chain/4PZL_A.pdb

```

pdbs

```

1 . . . 40
[Truncated_Name:1] 1AKE_A.pdb -----MRIILLGAPGAGKGTQAQFIMEKYGIPQIS
[Truncated_Name:2] 8BQF_A.pdb -----MRIILLGAPGAGKGTQAQFIMEKYGIPQIS
[Truncated_Name:3] 4X8M_A.pdb -----MRIILLGAPGAGKGTQAQFIMEKYGIPQIS
[Truncated_Name:4] 6S36_A.pdb -----MRIILLGAPGAGKGTQAQFIMEKYGIPQIS
[Truncated_Name:5] 8Q2B_A.pdb -----MRIILLGAPGAGKGTQAQFIMEKYGIPQIS
[Truncated_Name:6] 8RJ9_A.pdb -----MRIILLGAPGAGKGTQAQFIMEKYGIPQIS
[Truncated_Name:7] 6RZE_A.pdb -----MRIILLGAPGAGKGTQAQFIMEKYGIPQIS
[Truncated_Name:8] 4X8H_A.pdb -----MRIILLGAPGAGKGTQAQFIMEKYGIPQIS
[Truncated_Name:9] 3HPR_A.pdb -----MRIILLGAPGAGKGTQAQFIMEKYGIPQIS
[Truncated_Name:10] 1E4V_A.pdb -----MRIILLGAPVAGKGTQAQFIMEKYGIPQIS
[Truncated_Name:11] 5EJE_A.pdb -----MRIILLGAPGAGKGTQAQFIMEKYGIPQIS
[Truncated_Name:12] 1E4Y_A.pdb -----MRIILLGALVAGKGTQAQFIMEKYGIPQIS
[Truncated_Name:13] 3X2S_A.pdb -----MRIILLGAPGAGKGTQAQFIMEKYGIPQIS
[Truncated_Name:14] 6HAP_A.pdb -----MRIILLGAPGAGKGTQAQFIMEKYGIPQIS
[Truncated_Name:15] 6HAM_A.pdb -----MRIILLGAPGAGKGTQAQFIMEKYGIPQIS
[Truncated_Name:16] 4K46_A.pdb -----MRIILLGAPGAGKGTQAQFIMAKFGIPQIS
[Truncated_Name:17] 4NP6_A.pdb -----NAMRIILLGAPGAGKGTQAQFIMEKFGIPQIS
[Truncated_Name:18] 3GMT_A.pdb -----MRLILLGAPGAGKGTQANFIKEKFGIPQIS
[Truncated_Name:19] 4PZL_A.pdb TENLYFQSNAMRIILLGAPGAGKGTQAKIIEQKYNIAHIS
                        **^*****  *  *^ *  **
1 . . . 40

41 . . . 80
[Truncated_Name:1] 1AKE_A.pdb TGDMLRAAVKSGSELGKQAKDIMDAGKLVDELVIALVKE
[Truncated_Name:2] 8BQF_A.pdb TGDMLRAAVKSGSELGKQAKDIMDAGKLVDELVIALVKE
[Truncated_Name:3] 4X8M_A.pdb TGDMLRAAVKSGSELGKQAKDIMDAGKLVDELVIALVKE
[Truncated_Name:4] 6S36_A.pdb TGDMLRAAVKSGSELGKQAKDIMDAGKLVDELVIALVKE
[Truncated_Name:5] 8Q2B_A.pdb TGDMLRAAVKSGSELGKQAKDIMDAGKLVDELVIALVKE
[Truncated_Name:6] 8RJ9_A.pdb TGDMLRAAVKSGSELGKQAKDIMDAGKLVDELVIALVKE
[Truncated_Name:7] 6RZE_A.pdb TGDMLRAAVKSGSELGKQAKDIMDAGKLVDELVIALVKE

```

[Truncated_Name:8] 4X8H_A.pdb	TGDMRLRAAVKSGSELGKQAKDIMDAGKLVDELVIALVKE
[Truncated_Name:9] 3HPR_A.pdb	TGDMRLRAAVKSGSELGKQAKDIMDAGKLVDELVIALVKE
[Truncated_Name:10] 1E4V_A.pdb	TGDMRLRAAVKSGSELGKQAKDIMDAGKLVDELVIALVKE
[Truncated_Name:11] 5EJE_A.pdb	TGDMRLRAAVKSGSELGKQAKDIMDACKLVDELVIALVKE
[Truncated_Name:12] 1E4Y_A.pdb	TGDMRLRAAVKSGSELGKQAKDIMDAGKLVDELVIALVKE
[Truncated_Name:13] 3X2S_A.pdb	TGDMRLRAAVKSGSELGKQAKDIMDCGKLVDELVIALVKE
[Truncated_Name:14] 6HAP_A.pdb	TGDMRLRAAVKSGSELGKQAKDIMDAGKLVDELVIALVRE
[Truncated_Name:15] 6HAM_A.pdb	TGDMRLRAAIKSGSELGKQAKDIMDAGKLVDEIIIALVKE
[Truncated_Name:16] 4K46_A.pdb	TGDMRLRAAIKAGTELGKQAKSVIDAGQLVSDDIILGLVKE
[Truncated_Name:17] 4NP6_A.pdb	TGDMRLRAAIKAGTELGKQAKAVIDAGQLVSDDIILGLIKE
[Truncated_Name:18] 3GMT_A.pdb	TGDMRLRAAVKAGTPLGVEAKTYMDEGKLVPSLIIGLVKE
[Truncated_Name:19] 4PZL_A.pdb	TGDMIRETIKSGSALGQELKKVLDAGELVSDEFIIVKVD
	****~* ~* *~ ** * ~* ** * ~ ~~~~~
	41 . . . 80
	81 . . . 120
[Truncated_Name:1] 1AKE_A.pdb	RIAQEDCRNGFLLDGFPR TIPQADAMKEAGINVDYVLEFD
[Truncated_Name:2] 8BQF_A.pdb	RIAQE----GFLLDGFPR TIPQADAMKEAGINVDYVIEFD
[Truncated_Name:3] 4X8M_A.pdb	RIAQEDCRNGFLLDGFPR TIPQADAMKEAGINVDYVLEFD
[Truncated_Name:4] 6S36_A.pdb	RIAQEDCRNGFLLDGFPR TIPQADAMKEAGINVDYVLEFD
[Truncated_Name:5] 8Q2B_A.pdb	RIAQEDCRNGFLLDGFPR TIPQADAMKEAGINVDYVLEFD
[Truncated_Name:6] 8RJ9_A.pdb	RIAQEDCRNGFLLAGFPR TIPQADAMKEAGINVDYVLEFD
[Truncated_Name:7] 6RZE_A.pdb	RIAQEDCRNGFLLDGFPR TIPQADAMKEAGINVDYVLEFD
[Truncated_Name:8] 4X8H_A.pdb	RIAQEDCRNGFLLDGFPR TIPQADAMKEAGINVDYVLEFD
[Truncated_Name:9] 3HPR_A.pdb	RIAQEDCRNGFLLDGFPR TIPQADAMKEAGINVDYVLEFD
[Truncated_Name:10] 1E4V_A.pdb	RIAQEDCRNGFLLDGFPR TIPQADAMKEAGINVDYVLEFD
[Truncated_Name:11] 5EJE_A.pdb	RIAQEDCRNGFLLDGFPR TIPQADAMKEAGINVDYVLEFD
[Truncated_Name:12] 1E4Y_A.pdb	RIAQEDCRNGFLLDGFPR TIPQADAMKEAGINVDYVLEFD
[Truncated_Name:13] 3X2S_A.pdb	RIAQEDSRNGFLLDGFPR TIPQADAMKEAGINVDYVLEFD
[Truncated_Name:14] 6HAP_A.pdb	RICQEDSRNGFLLDGFPR TIPQADAMKEAGINVDYVLEFD
[Truncated_Name:15] 6HAM_A.pdb	RICQEDSRNGFLLDGFPR TIPQADAMKEAGINVDYVLEFD
[Truncated_Name:16] 4K46_A.pdb	RIAQDDCAKGFLDGFPR TIPQADGLKEVGVVVDYVIEFD
[Truncated_Name:17] 4NP6_A.pdb	RIAQADCEKGFLDGFPR TIPQADGLKEMGINVDYVIEFD
[Truncated_Name:18] 3GMT_A.pdb	RLKEADCANGYLFDFPR TIPQADAMKEAGVAIDYVLEID
[Truncated_Name:19] 4PZL_A.pdb	RISKNDCCNNGFLLDGVPR TIPQAQELDKLGVNIDYIVEVD
	*~ *~* * ***** ** ^ *~ ^**~* *
	81 . . . 120
	121 . . . 160
[Truncated_Name:1] 1AKE_A.pdb	VPDELIVDRIVGRRVHAPSGRVYHV KFNPPKVEGKDDVTG
[Truncated_Name:2] 8BQF_A.pdb	VPDELIVDRIVGRRVHAPSGRVYHV KFNPPKVEGKDDVTG
[Truncated_Name:3] 4X8M_A.pdb	VPDELIVDRIVGRRVHAPSGRVYHV KFNPPKVEGKDDVTG
[Truncated_Name:4] 6S36_A.pdb	VPDELIVDKIVGRRVHAPSGRVYHV KFNPPKVEGKDDVTG

[Truncated_Name:5]8Q2B_A.pdb	VPDELIVDRIVGRRVHAPSGRVYHVKNPPKVEGKDDVTG
[Truncated_Name:6]8RJ9_A.pdb	VPDELIVDRIVGRRVHAPSGRVYHVKNPPKVEGKDDVTG
[Truncated_Name:7]6RZE_A.pdb	VPDELIVDAIVGRRVHAPSGRVYHVKNPPKVEGKDDVTG
[Truncated_Name:8]4X8H_A.pdb	VPDELIVDRIVGRRVHAPSGRVYHVKNPPKVEGKDDVTG
[Truncated_Name:9]3HPR_A.pdb	VPDELIVDRIVGRRVHAPSGRVYHVKNPPKVEGKDDGTG
[Truncated_Name:10]1E4V_A.pdb	VPDELIVDRIVGRRVHAPSGRVYHVKNPPKVEGKDDVTG
[Truncated_Name:11]5EJE_A.pdb	VPDELIVDRIVGRRVHAPSGRVYHVKNPPKVEGKDDVTG
[Truncated_Name:12]1E4Y_A.pdb	VPDELIVDRIVGRRVHAPSGRVYHVKNPPKVEGKDDVTG
[Truncated_Name:13]3X2S_A.pdb	VPDELIVDRIVGRRVHAPSGRVYHVKNPPKVEGKDDVTG
[Truncated_Name:14]6HAP_A.pdb	VPDELIVDRIVGRRVHAPSGRVYHVKNPPKVEGKDDVTG
[Truncated_Name:15]6HAM_A.pdb	VPDELIVDRIVGRRVHAPSGRVYHVKNPPKVEGKDDVTG
[Truncated_Name:16]4K46_A.pdb	VADSVIVERMAGRAHLASGRTYHNVPNPKVEGKDDVTG
[Truncated_Name:17]4NP6_A.pdb	VADDVIVERMAGRAHLPSGRTYHVVPNPKVEGKDDVTG
[Truncated_Name:18]3GMT_A.pdb	VPFSEIIERMSGRRTHPASGRTYHVKNPPKVEGKDDVTG
[Truncated_Name:19]4PZL_A.pdb	VADNLLIERITGRIHPASGRTYHTKFNNPKVADKDDVTG
	* ^^^ ^ *** * *** ** ^***** *** **
121	. . . 160
	161 . . . 200
[Truncated_Name:1]1AKE_A.pdb	EELTTRKDDQEETVRKRLVEYHQMTAPLIGYYSKEAEAGN
[Truncated_Name:2]8BQF_A.pdb	EELTTRKDDQEETVRKRLVEYHQMTAPLIGYYSKEAEAGN
[Truncated_Name:3]4X8M_A.pdb	EELTTRKDDQEETVRKRLVEWHQMTAPLIGYYSKEAEAGN
[Truncated_Name:4]6S36_A.pdb	EELTTRKDDQEETVRKRLVEYHQMTAPLIGYYSKEAEAGN
[Truncated_Name:5]8Q2B_A.pdb	EELTTRKADQEETVRKRLVEYHQMTAPLIGYYSKEAEAGN
[Truncated_Name:6]8RJ9_A.pdb	EELTTRKDDQEETVRKRLVEYHQMTAPLIGYYSKEAEAGN
[Truncated_Name:7]6RZE_A.pdb	EELTTRKDDQEETVRKRLVEYHQMTAPLIGYYSKEAEAGN
[Truncated_Name:8]4X8H_A.pdb	EELTTRKDDQEETVRKRLVEYHQMTAALIGYYSKEAEAGN
[Truncated_Name:9]3HPR_A.pdb	EELTTRKDDQEETVRKRLVEYHQMTAPLIGYYSKEAEAGN
[Truncated_Name:10]1E4V_A.pdb	EELTTRKDDQEETVRKRLVEYHQMTAPLIGYYSKEAEAGN
[Truncated_Name:11]5EJE_A.pdb	EELTTRKDDQEECVRKRLVEYHQMTAPLIGYYSKEAEAGN
[Truncated_Name:12]1E4Y_A.pdb	EELTTRKDDQEETVRKRLVEYHQMTAPLIGYYSKEAEAGN
[Truncated_Name:13]3X2S_A.pdb	EELTTRKDDQEETVRKRLCEYHQMTAPLIGYYSKEAEAGN
[Truncated_Name:14]6HAP_A.pdb	EELTTRKDDQEETVRKRLVEYHQMTAPLIGYYSKEAEAGN
[Truncated_Name:15]6HAM_A.pdb	EELTTRKDDQEETVRKRLVEYHQMTAPLIGYYSKEAEAGN
[Truncated_Name:16]4K46_A.pdb	EDLVIREDDKEETVLARLG VYHNQTAPLIAYYGKEAEAGN
[Truncated_Name:17]4NP6_A.pdb	EDLVIREDDKEETVRARLN VYHTQTAPLIEYYGKEAAAGK
[Truncated_Name:18]3GMT_A.pdb	EPLVQRDDDKEETVKKRLDVYEAQTKPLITYYGDWARRGA
[Truncated_Name:19]4PZL_A.pdb	EPLITRTDDNEDTVKQRLSVYHAQTAKLIDFYRNFSSNT
	* * * * ^ * ** ^ * ** ^*
161	. . . 200
	201 . . . 227
[Truncated Name:1]1AKE A.pdb	T--KYAKVDGTPVAEVRADLEKILG-

[Truncated_Name:2]8BQF_A.pdb	T--KYAKVDGTPVAEVRADLEKIL--
[Truncated_Name:3]4X8M_A.pdb	T--KYAKVDGTPVAEVRADLEKILG-
[Truncated_Name:4]6S36_A.pdb	T--KYAKVDGTPVAEVRADLEKILG-
[Truncated_Name:5]8Q2B_A.pdb	T--KYAKVDGTPVAEVRADLEKILG-
[Truncated_Name:6]8RJ9_A.pdb	T--KYAKVDGTPVAEVRADLEKILG-
[Truncated_Name:7]6RZE_A.pdb	T--KYAKVDGTPVAEVRADLEKILG-
[Truncated_Name:8]4X8H_A.pdb	T--KYAKVDGTPVAEVRADLEKILG-
[Truncated_Name:9]3HPR_A.pdb	T--KYAKVDGTPVAEVRADLEKILG-
[Truncated_Name:10]1E4V_A.pdb	T--KYAKVDGTPVAEVRADLEKILG-
[Truncated_Name:11]5EJE_A.pdb	T--KYAKVDGTPVAEVRADLEKILG-
[Truncated_Name:12]1E4Y_A.pdb	T--KYAKVDGTPVAEVRADLEKILG-
[Truncated_Name:13]3X2S_A.pdb	T--KYAKVDGTPVAEVRADLEKILG-
[Truncated_Name:14]6HAP_A.pdb	T--KYAKVDGTPVCEVRADLEKILG-
[Truncated_Name:15]6HAM_A.pdb	T--KYAKVDGTPVCEVRADLEKILG-
[Truncated_Name:16]4K46_A.pdb	T--QYLKFDGTKAVAEVSAELEKALA-
[Truncated_Name:17]4NP6_A.pdb	T--QYLKFDGTKQVSEVSADIAKALA-
[Truncated_Name:18]3GMT_A.pdb	E-----NGLKAPA-----YRKISG-
[Truncated_Name:19]4PZL_A.pdb	KIPKYIKINGDQAVEKVSQDIFDQLNK

*

201 . . 227

Call:

```
pdbaln(files = files, fit = TRUE, exefile = "msa")
```

Class:

```
pdbs, fasta
```

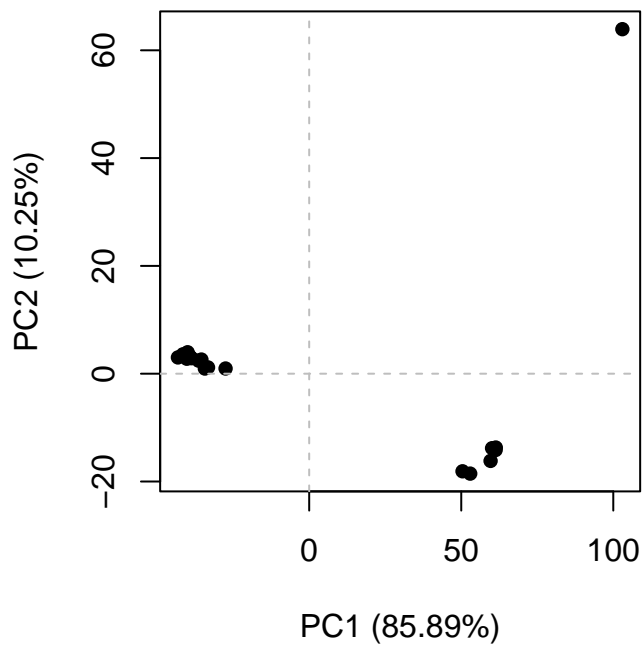
Alignment dimensions:

```
19 sequence rows; 227 position columns (199 non-gap, 28 gap)
```

```
+ attr: xyz, resno, b, chain, id, ali, resid, sse, call
```

Principal Component Analysis

```
# Perform PCA
pc.xray <- pca(pdbs)
plot(pc.xray, pc.axes = c(1,2))
```

To visualize the major structural variations in the ensemble the function `mktrj()` can be used to generate a trajectory PDB file by interpolating along a give PC (eigenvector):

```
# Visualize first principal component  
pc1 <- mktrj(pc.xray, pc=1, file="pc_1.pdb")
```

```
uniprot <- 248838887  
pdb <- 195610  
  
pdb/uniprot * 100
```

```
[1] 0.0786091
```