

# Class10: Structural Bioinformatics 1

## AUTHOR

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## PDB statistics

The Protein Data Bank (PDB) is the main repository of biomolecular structures. Let's see what it contains:

Download a CSV file from the PDB site (accessible from "Analyze" > "PDB Statistics" > "by Experimental Method and Molecular Type"

```
stats <- read.csv("Data Export Summary.csv")
stats
```

	Molecular.Type	X.ray	EM	NMR	Integrative	Multiple.methods
1	Protein (only)	178,795	21,825	12,773	343	226
2	Protein/Oligosaccharide	10,363	3,564	34	8	11
3	Protein/NA	9,106	6,335	287	24	7
4	Nucleic acid (only)	3,132	221	1,566	3	15
5	Other	175	25	33	4	0
6	Oligosaccharide (only)	11	0	6	0	1
	Neutron Other Total					
1	84 32 214,078					
2	1 0 13,981					
3	0 0 15,759					
4	3 1 4,941					
5	0 0 237					
6	0 4 22					

Q1: What percentage of structures in the PDB are solved by X-Ray and Electron Microscopy.

```
#sum(stats$X.ray)
```

```
sum(stats$Neutron)
```

[1] 88

The comma in these numbers leads to the numbers here being read as characters

```
library(readr)
stats <- read_csv("Data Export Summary.csv")
```

Rows: 6 Columns: 9

— Column specification —

Delimiter: ","

```
chr (1): Molecular Type
dbl (4): Integrative, Multiple methods, Neutron, Other
num (4): X-ray, EM, NMR, Total
```

**i** Use `spec()` to retrieve the full column specification for this data.  
**i** Specify the column types or set `show\_col\_types = FALSE` to quiet this message.

stats

```
# A tibble: 6 × 9
`Molecular Type` `X-ray`    EM   NMR Integrative `Multiple methods` Neutron
<chr>           <dbl>    <dbl> <dbl>      <dbl>           <dbl>    <dbl>
1 Protein (only) 178795  21825 12773     343        226     84
2 Protein/Oligosacch... 10363   3564   34       8        11      1
3 Protein/NA       9106    6335   287      24        7      0
4 Nucleic acid (only) 3132    221   1566      3        15      3
5 Other            175     25    33       4        0      0
6 Oligosaccharide (o... 11      0     6       0        1      0
# i 2 more variables: Other <dbl>, Total <dbl>
```

Q1: What percentage of structures in the PDB are solved by X-Ray and Electron Microscopy.

```
(sum(stats$`X-ray`) + sum(stats$EM))/sum(stats$Total)
```

[1] 0.937892

The structures of X-ray and Electron Microscopy make up 93.78% of the data.

Q2: What proportion of structures in the PDB are protein?

```
(stats[1,9])/sum(stats$Total)
```

```
Total
1 0.8596889
```

The proportion is 85%

Q3: SKIP... Looking up HIV structures including 1HSG

## Visualizing the HIV-1 protease structure

We can use the Molstar viewer online: <https://molstar.org/viewer/>

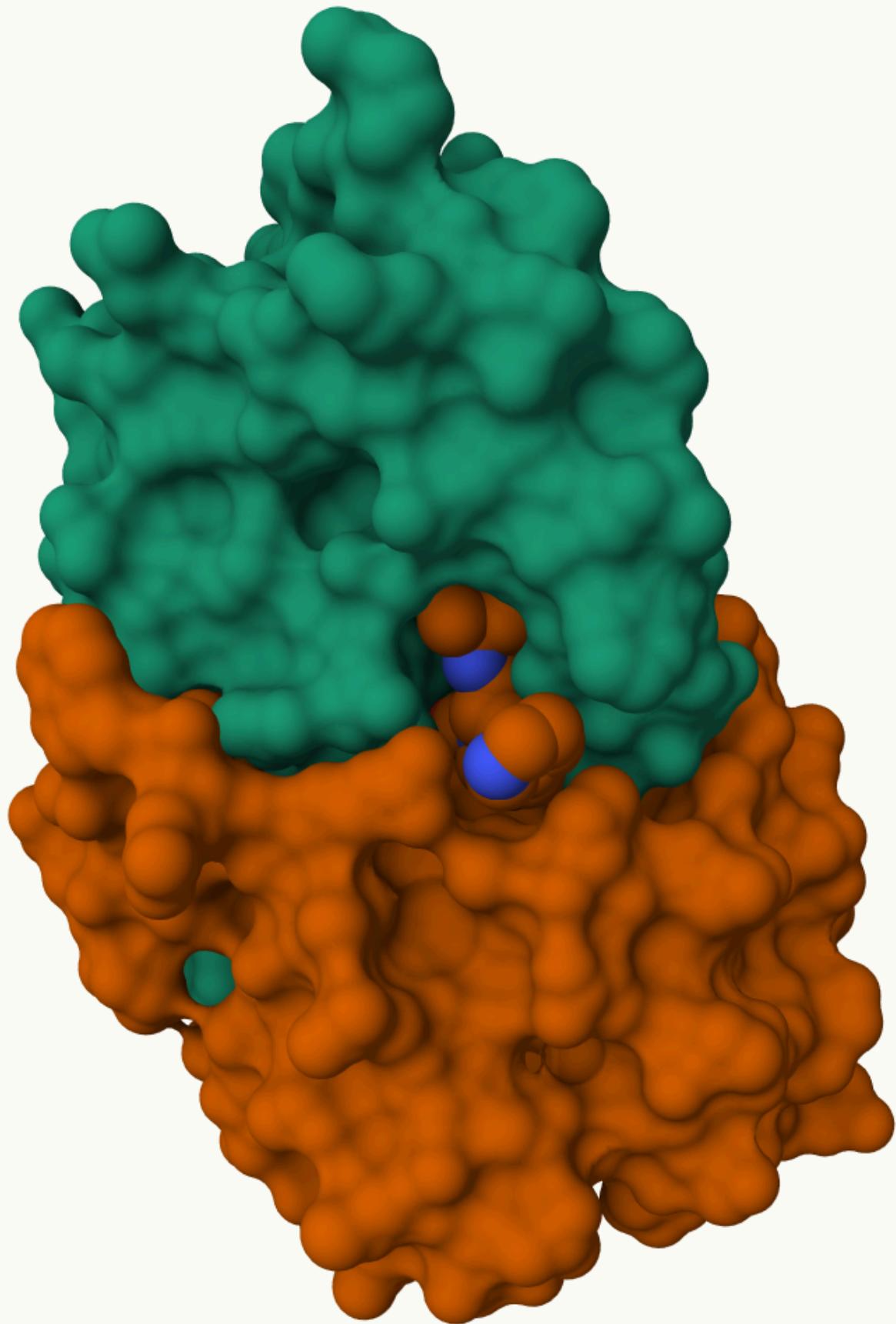


Figure 1: HIV Protease

Q4: Water molecules normally have 3 atoms. Why do we see just one atom per water molecule in this structure?

This is because the molecular model is only showing the central oxygen atom of the water, not the two additional hydrogens

Q5: There is a critical “conserved” water molecule in the binding site. Can you identify this water molecule? What residue number does this water molecule have

This water molecule has a residue number of HOH 308

Q6: Generate and save a figure clearly showing the two distinct chains of HIV-protease along with the ligand. You might also consider showing the catalytic residues ASP 25 in each chain and the critical water (we recommend “Ball & Stick” for these side-chains). Add this figure to your Quarto document.

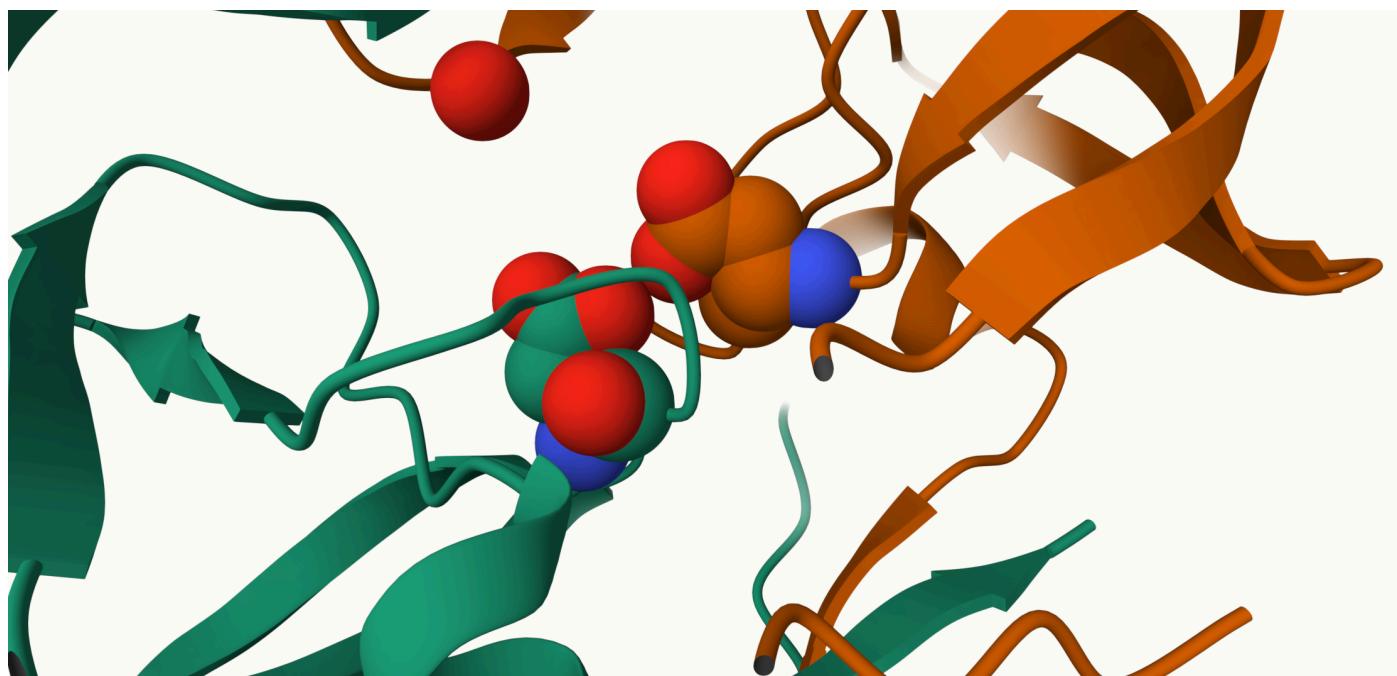


Figure 2: ASP 25 and the important active site water molecule

## Introduction to Bio3D in R

```
library(bio3d)
```

```
Warning: package 'bio3d' was built under R version 4.4.3
```

```
 pdb <- read.pdb("1hsg")
```

Note: Accessing on-line PDB file

`pdb`

Call: `read.pdb(file = "1hsg")`

Total Models#: 1

Total Atoms#: 1686, XYZs#: 5058 Chains#: 2 (values: A B)

Protein Atoms#: 1514 (residues/Calpha atoms#: 198)

Nucleic acid Atoms#: 0 (residues/phosphate atoms#: 0)

Non-protein/nucleic Atoms#: 172 (residues: 128)

Non-protein/nucleic resid values: [ HOH (127), MK1 (1) ]

Protein sequence:

```
PQITLWQRPLVTIKIGGQLKEALLDTGADDTVLEEMSLPGRWPKMIGGGIGGFIKVRQYD  
QILIEICGHKAIGTVLGPTPVNIIGRNLLTQIGCTLNFPQITLWQRPLVTIKIGGQLKE  
ALLDTGADDTVLEEMSLPGRWPKMIGGGIGGFIKVRQYDQILIEICGHKAIGTVLGPTP  
VNIIGRNLLTQIGCTLNF
```

+ attr: atom, xyz, seqres, helix, sheet,  
calpha, remark, call

Q7: How many amino acid residues are there in this pdb object?

There are 128 amino acids

Q8: Name one of the two non-protein residues?

There is both water molecules and MK1, merk 1

Q9: How many protein chains are in this structure?

There are 2 protein chains

```
head(pdb$atom)
```

	type	eleno	elety	alt	resid	chain	resno	insert	x	y	z	o	b
1	ATOM	1	N	<NA>	PRO	A	1	<NA>	29.361	39.686	5.862	1	38.10
2	ATOM	2	CA	<NA>	PRO	A	1	<NA>	30.307	38.663	5.319	1	40.62
3	ATOM	3	C	<NA>	PRO	A	1	<NA>	29.760	38.071	4.022	1	42.64
4	ATOM	4	O	<NA>	PRO	A	1	<NA>	28.600	38.302	3.676	1	43.40
5	ATOM	5	CB	<NA>	PRO	A	1	<NA>	30.508	37.541	6.342	1	37.87

```

6 ATOM      6   CG <NA>    PRO     A     1   <NA> 29.296 37.591 7.162 1 38.40
  segid elesy charge
1  <NA>      N   <NA>
2  <NA>      C   <NA>
3  <NA>      C   <NA>
4  <NA>      O   <NA>
5  <NA>      C   <NA>
6  <NA>      C   <NA>

```

## Predicting functional motions of a single structure

Read an ADK structure from the pdb database

```
adk <- read.pdb("6s36")
```

Note: Accessing on-line PDB file  
PDB has ALT records, taking A only, rm.alt=TRUE

```
adk
```

Call: read.pdb(file = "6s36")

Total Models#: 1  
Total Atoms#: 1898, XYZs#: 5694 Chains#: 1 (values: A)  
Protein Atoms#: 1654 (residues/Calpha atoms#: 214)  
Nucleic acid Atoms#: 0 (residues/phosphate atoms#: 0)  
Non-protein/nucleic Atoms#: 244 (residues: 244)  
Non-protein/nucleic resid values: [ CL (3), HOH (238), MG (2), NA (1) ]

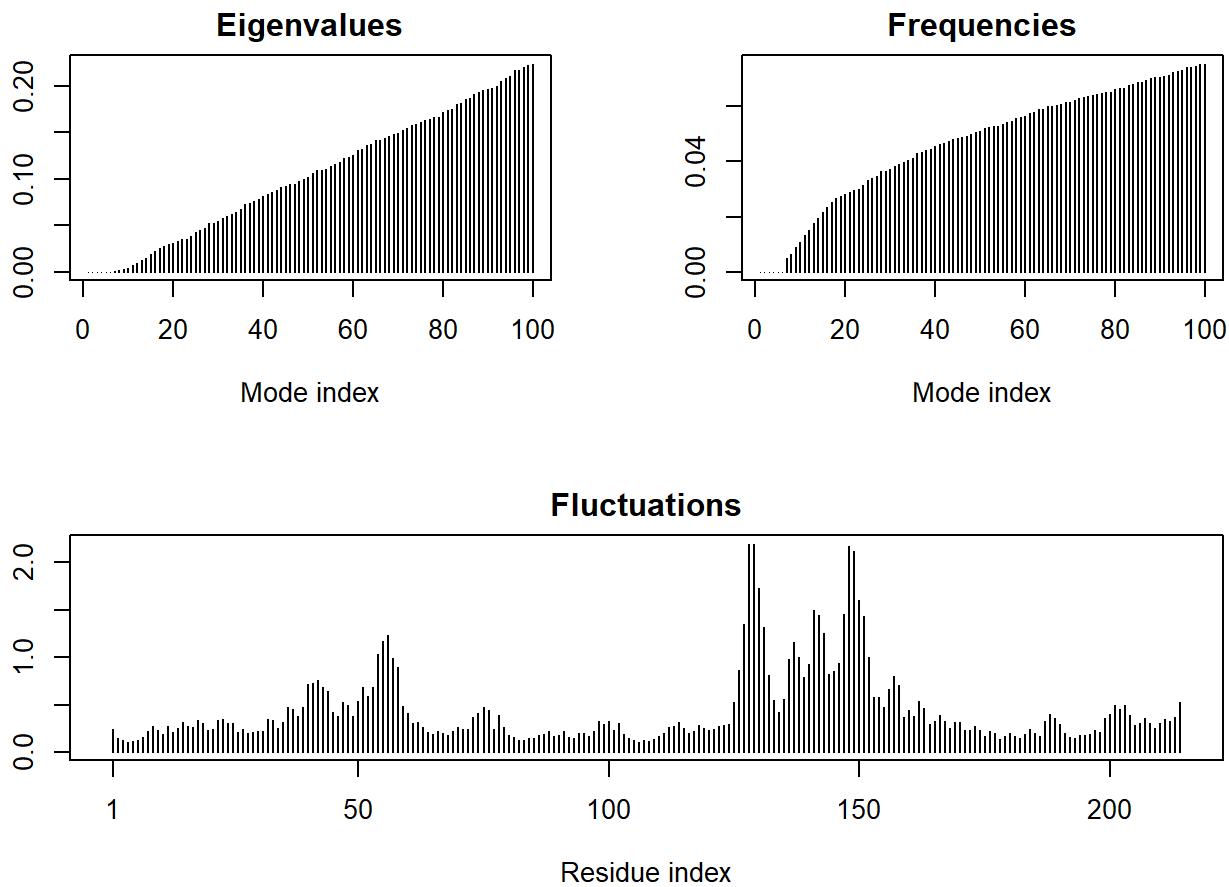
Protein sequence:  
MRIILLGAPGAGKGQTQAFIMEKYGIPQISTGDMRLRAAVKSGSELGKQAKDIMDAGKLVT  
DELVIALVKERIAQEDCRNGFLLDGFPRTIPQADAMKEAGINVDYVLEFDVPDELIVDKI  
VGRRVHAPSGRVYHVKFNPVKEGKDDVTGEELTTRKDDQEETRKRLVEYHQMTAPLIG  
YYSKAEAGNTKYAKVDTKPVAEVRADLEKILG

+ attr: atom, xyz, seqres, helix, sheet,  
calpha, remark, call

```
m <- nma(adk)
```

Building Hessian... Done in 0.02 seconds.  
Diagonalizing Hessian... Done in 0.34 seconds.

```
plot(m)
```



Write our results as a new trajectory/movie of predicted motions using `mktrj`

```
mktrj(m, file="adk_m7.pdb")
```