Lab 4 - Random splits, Iterators, & Handling large data

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Lab 4 outline

- 1. Randomly split data into 2 equal parts (calibration & validation samples)
- 2. Introduction to MplusAutomation with iterators
- 3. Dealing with large data

Getting started - following the routine...

- 1. Create an R-Project
- 2. Install packages (ONLY IF NEEDED)
- 3. Load packages

R-Project instructions:

- 1. click "NEW PROJECT" (upper right corner of window)
- 2. choose option "NEW DIRECTORY"
- 3. choose location of project (on desktop OR in a designated class folder)

Within R-studio under the files pane (bottom right):

- 1. click "New Folder" and name folder "data"
- 2. click "New Folder" and name folder "efa_mplus"
- 3. click "New Folder" and name folder "figures"

SKIP (return here if you recieve an error when loading packages)

- 1. First check if the packages load (run lines 32-39)
- 2. If an error is returned for package(s)
- 3. then run the following lines of code
- 4. AFTER ANY INSTALLATION... MUST RE-LOAD PACKAGES!
- ** NOTE: the following code ONLY needs to be run once per computer!

IF package rhdf5 does not load then run:

```
if (!requireNamespace("BiocManager", quietly = TRUE))
  install.packages("BiocManager")

BiocManager::install("rhdf5")
```

Lab 4 - Begin

loading packages...

```
library(janitor)
library(tidyverse)
library(haven)
library(MplusAutomation)
library(rhdf5)
```

```
library(here)
library(corrplot)
```

read in the raw dataset

```
lab_data <- read_spss(here("data", "lab_efa_cfa_data.sav"))</pre>
```

create a subset of the dataset called "school_trouble"

```
school_trouble <- lab_data %>%
select(41:55)
```

make a new codebook from the "school_trouble" subset

```
sjPlot::view_df(school_trouble)
```

write a CSV datafile

```
write_csv(school_trouble, here("data", "school_trouble_data.csv"))
```

read the unlabeled data back into R

```
trouble_data <- read_csv(here("data", "school_trouble_data.csv"))</pre>
```

check items to see if reverse coding is needed

Randomly split a sample into 2 equal parts

find the size of half of original sample.

The "floor()" function helps with rounding

```
smp_size <- floor(0.50 * nrow(trouble_data))</pre>
```

set the seed to make your partition reproducible

```
set.seed(123)
```

the function "sample()" will pick at random the values of the specified number

```
calibrate_smp <- sample(seq_len(nrow(trouble_data)), size = smp_size)</pre>
```

create two samples called "calibrate" & "validate"

```
calibrate <- trouble_data[calibrate_smp, ]
validate <- trouble_data[-calibrate_smp, ]</pre>
```

Let's run an EFA with the "calibrate" sample

```
m_efa_1 <- mplusObject(
  TITLE = "School Trouble EFA - LAB 4 DEMO",
  VARIABLE =
    "usevar = BYS22A-BYS24G;",

ANALYSIS =
    "type = efa 1 5;
    estimator = mlr;
    parallel=50; ! run parallel analysis",</pre>
```

Plot Parallel Analysis & Eigenvalues

read into R an Mplus output file

```
efa_summary <- readModels(here("efa_mplus", "lab4_efa1_trouble.out"))</pre>
```

extract relavent data & prepare dataframe for plot

pivot the dataframe to "long" format

plot using ggplot

save figure to the designated folder

```
ggsave(here("figures", "eigenvalue_elbow_rplot.png"), dpi=300, height=5, width=7, units="in")
```

Introduction to MplusAutomation with iterators

Alternate way to run an EFA with the "calibrate" sample

```
m_efa <- lapply(1:5, function(k) {</pre>
  m_efa2 <- mplusObject(</pre>
    TITLE = "School Trouble EFA - LAB 4 DEMO",
    VARIABLE =
      "usevar = BYS22A-BYS24G;",
    ANALYSIS =
      paste("type=efa", k, k),
    MODEL = "",
    PLOT = "type = plot3;",
    OUTPUT = "sampstat;",
    usevariables = colnames(calibrate),
    rdata = calibrate)
  m_efa_2_fit <- mplusModeler(m_efa2,</pre>
                               dataout=sprintf(here("efa_mplus2", "efa_trouble.dat"), k),
                               modelout=sprintf(here("efa_mplus2", "efa_%d_trouble.inp"), k),
                               check=TRUE, run = TRUE, hashfilename = FALSE)
})
```

Cleaning & subsetting large datasets

reading SPSS files is a lot slower than reading CSV formatted files

```
hsls_raw <- read_spss(here("data", "hsls_16_student_sub_v1.sav"))
```

make all column names "lower_snake_case" style

```
hsls_tidy <- hsls_raw %>% clean_names()
```

select usig the starts_with() function

```
hsls_x1 <- hsls_tidy %>%
select(starts_with("x1")) # columns with first 2 characters "x1"
```

select using the end_with() function

```
hsls_not_sex <- hsls_tidy %>%
select(!ends_with("sex")) # columns that do NOT end with "sex"
```

select using the end_with() function

```
hsls_science <- hsls_tidy %>%
  select(contains("sci")) # columns that contain characters "sci"

hsls_math <- hsls_tidy %>%
  select(contains(c("mth" , "math"))) # columns that contain "mth" or "math"
```

combine different select() arguements

```
hsls_math_sci <- hsls_tidy %>%
select(contains(c("mth" , "math", "sci"))) %>%
select(!starts_with("x1")) %>%
select(!ends_with("sex"))
```

remove characters from the variable names that are greater than 8 characters

```
names(hsls_math_sci) = str_sub(names(hsls_math_sci), 1, 8)
```

check if culumn names are unique

```
test.unique <- function(df) { ## function to identify unique columns

length1 <- length(colnames(df))
length2 <- length(unique(colnames(df)))
if (length1 - length2 > 0 ) {

print(paste("There are", length1 - length2, " duplicates", sep=" "))
}

test.unique(hsls_math_sci)
```

locate duplicates (this will find the column of the first duplicate)

```
anyDuplicated(colnames(hsls_math_sci))
names(hsls_math_sci)
```

Other functions to consider from the "stringr" package (part of tidyverse):

```
str_remove()
str_replace() # replace one string pattern with another
str_match()
str_pad() # to remove spaces
str_count()
str_detect()
str_dup()
str_extract_all()
```

End of lab 4 exercise

References:

Hallquist, M. N., & Wiley, J. F. (2018). MplusAutomation: An R Package for Facilitating Large-Scale Latent Variable Analyses in Mplus. Structural equation modeling: a multidisciplinary journal, 25(4), 621-638.