Coping Behavior Example - Accompanying R Code

Motivation for MplusAutomation workflow:

This R tutorial is intended to provide a template for estimating latent class analysis (LCA), integrating the unconditional LCA with a larger SEM model, and producing publication ready figures in a systematic manner. Using this approach all code for data pre-processing, model estimation, tabulation, and figure processing can be contained within a single script providing clear documentation. It is the authors' belief that this dramatically reduces risk of user-error, is conducive to open-science philosophy, and scientific transparency. All models are estimated using Mplus (Version 8.6; Muthén & Muthén, 1998 - 2017) using the wrapping package MplusAutomation (Hallquist & Wiley, 2018). This method requires the user to have the proprietary software Mplus installed on their computer.

This approach also relies on the utility of R-Projects. This provides a structured framework for organizing all associated data files, Mplus input/output files, scripts, and figures. Given the large number of Mplus output files inherent to fitting mixture models, creating a system of project sub-folders greatly improves organization (i.e., folders; 'data', 'mplus_files' 'figures', etc.) Additionally, the communication between R and Mplus requires the specification of file-paths, a procedure which is streamlined by use of R-projects. Due to the reliance on file-paths, the here package ensures that the syntax for file-paths is uniform across operating systems (e.g., Windows or Mac) enabling reproducibility.

Preparation

Download the R-Project

Link to Github repository here: LCA-COPING-BD

For readers new to Github & version controlled R-projects:

- 1. On the repository page, click the green Code button and in the menu choose option Download ZIP
- 2. Place the un-zipped downloaded folder on your desktop
- 3. Within this folder open the file with the blue cube icon that is file named LCA-COPING-BD. Rproj
- 4. Next open the file containing all analysis code named rcode-lca-coping-bd.R.

Note: If preferred users may follow analyses using the Rmarkdown script (.Rmd).

Project folder organization

The following sub-folders will be used to contain files within the project directory:

```
1. "data"; 2. "enum mplus"; 3. "3step mplus"; 4. "figures"
```

Note regarding location of project directory: If the main project folder is located within too many nested folders it may result in a file-path error when estimating models with MplusAutomation.

Load packages

```
library(MplusAutomation)  # A conduit between R & Mplus
library(here)  # To locate or send files within the Rproject folder
library(glue)  # To insert R code within strings
library(gt)  # For pretty tables
library(reshape2)  # For manipulating plot data
library(cowplot)  # For pretty plots
library(patchwork)  # To effortlessly combine plots
library(tidyverse)  # For everything else...
```

Read in data. File-paths are set using R-Projects & the {here} package

1.1 Enumeration:

Estimate *K*-class models with 1 through 6 classes.

Specification details that are specific to applied coping example:

- Within the lapply () function 1:6 indicates the number of K-class models to estimate.
- The measurement model item names for the coping strategy example are listed after the categorical = ...; & usevar = ...; statements.
- Note, that for Latent Profile Analysis (LPA) the categorical = ...; statement would be removed so that items are estimated as continuous variables.

```
lca_k1_6 <- lapply(1:6, function(k) {
  lca_enum <- mplusObject(

TITLE = glue("Class-{k} LCA Enumeration - Youth Coping Strategies"),

VARIABLE = glue(
  "categorical = do1 do2 do3 do5 do6; !!! Coping strategy items for measurement model !!!
  usevar = do1 do2 do3 do5 do6;</pre>
```

```
classes = c(\{k\});"),
  ANALYSIS =
    "estimator = mlr;
     ! optseed = 458733; !!! FOR REPLICATING PLOTS !!!
     type = mixture;
     starts = 500 100;
     processors=10;",
  PLOT =
    "type = plot3;
     series = do1 do2 do3 do5 do6(*);",
  OUTPUT = "tech11 tech14;",
  usevariables = colnames(ies_data),
  rdata = ies_data)
lca_enum_fit <- mplusModeler(lca_enum,</pre>
                  dataout=glue(here("enum_mplus", "c_lca_enum_bd.dat")),
                  modelout=glue(here("enum_mplus", "c{k}_lca_enum_bd.inp")) ,
                  check=TRUE, run = TRUE, hashfilename = FALSE)
})
```

1.2 Generate Model Fit Summary Table

- This syntax can be used to compare model fit from the series of LCA models generated during enumeration (*Table 2* in manuscript).
- The code produces a table that is approximately in APA format.

Calculate relevant fit indices for summary table

```
allFit <- enum_extract %>%
  mutate(aBIC = -2*LL+Parameters*log((Observations+2)/24)) %>%
  mutate(CIAC = -2*LL+Parameters*(log(Observations)+1)) %>%
  mutate(AWE = -2*LL+2*Parameters*(log(Observations)+1.5)) %>%
  mutate(SIC = -.5*BIC) %>%
  mutate(expSIC = exp(SIC - max(SIC))) %>%
  mutate(expSIC = exp(SIC - max(SIC))) %>%
  mutate(BF = exp(SIC-lead(SIC))) %>%
  mutate(cmPk = expSIC/sum(expSIC)) %>%
  select(1:5,9:10,6:7,13,14) %>%
  arrange(Parameters)

allFit %>%
  mutate(Title = str_remove(Title, " LCA Enumeration - Youth Coping Strategies")) %>%
  gt() %>%
```

```
tab_header(
 title = md("**Model Fit Summary Table**"), subtitle = md(" ")) %>%
cols label(
 Title = "Classes",
 Parameters = md("Par"),
 LL = md("*LL*"),
 T11_VLMR_PValue = "VLMR",
 BLRT PValue = "BLRT",
 BF = md("BF"),
 cmPk = md("*cmPk*")) %>%
tab_footnote(
  footnote = md(
  "*Note.* Par = parameters; *LL* = log likelihood;
   BIC = bayesian information criterion;
   aBIC = sample size adjusted BIC; CAIC = consistent Akaike information criterion;
   AWE = approximate weight of evidence criterion;
   BLRT = bootstrapped likelihood ratio test p-value;
   VLMR = Vuong-Lo-Mendell-Rubin adjusted likelihood ratio test p-value;
    cmPk = approximate correct model probability."),
 locations = cells_title()) %>%
tab options(column labels.font.weight = "bold") %>%
fmt_number(10,decimals = 2,
           drop_trailing_zeros=TRUE,
           suffixing = TRUE) %>%
fmt number(c(3:9,11),
           decimals = 0) %>%
fmt_missing(1:11,
            missing_text = "--") %>%
fmt(c(8:9,11),
  fns = function(x)
  ifelse(x<0.001, "<.001",
         scales::number(x, accuracy = 0.01))) %>%
fmt(10, fns = function(x))
  ifelse(x>100, ">100",
         scales::number(x, accuracy = .1)))
```

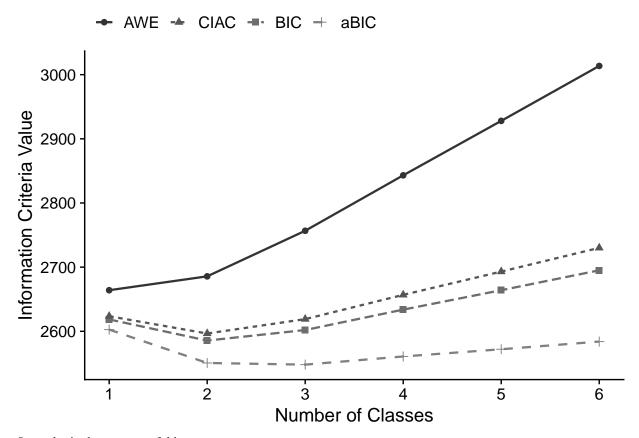
Model Fit Summary Table¹

Classes	Par	LL	BIC	aBIC	CIAC	AWE	BLRT	VLMR	BF	cmPk
Class-1	5	-1,294	2,619	2,603	2,624	2,664	_	_	>100	<.001
Class-2	11	-1,259	2,586	2,551	2,597	2,686	<.001	<.001	>100	1.00
Class-3	17	-1,249	2,602	2,548	2,619	2,757	<.001	0.12	>100	<.001
Class-4	23	-1,247	2,634	2,561	2,657	2,843	1.00	0.63	>100	<.001
Class-5	29	-1,244	2,664	2,572	2,693	2,928	1.00	0.70	>100	<.001
Class-6	35	-1,241	2,695	2,584	2,730	3,014	0.67	0.16	_	<.001

¹Note. Par = parameters; LL = log likelihood; BIC = bayesian information criterion; aBIC = sample size adjusted BIC; CAIC = consistent Akaike information criterion; AWE = approximate weight of evidence criterion; BLRT = bootstrapped likelihood ratio test p-value; VLMR = Vuong-Lo-Mendell-Rubin adjusted likelihood ratio test p-value; cmPk = approximate correct model probability.

1.3 Plot Information Criteria

```
allFit %>% select(2:7) %>%
 rowid_to_column() %>%
 pivot longer(`BIC`: `AWE`,
   names_to = "Index",
   values_to = "ic_value") %>%
  mutate(Index = factor(Index,
   levels = c("AWE","CIAC","BIC","aBIC"))) %>%
  ggplot(aes(x = rowid, y = ic_value,
   color = Index, shape = Index,
   group = Index, lty = Index)) +
  geom_point(size = 2.0) + geom_line(size = .8) +
  scale_x_continuous(breaks = 1:6) +
  scale_colour_grey(end = .5) +
  theme_cowplot() +
  labs(x = "Number of Classes", y = "Information Criteria Value") +
  theme(legend.title = element_blank(),
       legend.position = "top")
```



Save plot in the figures folder.

```
ggsave(here("figures", "FigS1_IC_plot.png"), dpi=300, height=5, width=7, units="in")
```

1.4 Compare Conditional Item Probability Plots

This syntax produces a plot containing a series of LCA probability plot facets. This can be used to compare solutions across *K*-class models.

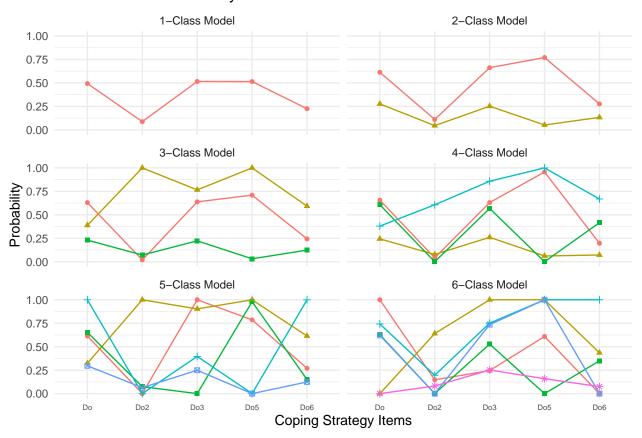
```
model_results <- data.frame()

for (i in 1:length(output_enum)) {
   temp <- data.frame(unclass(output_enum[[i]]$parameters$unstandardized)) %>%
      mutate(model = pasteO(i, "-Class Model"))
   model_results <- rbind(model_results, temp) }

pp_plots <- model_results %>%
   filter(paramHeader == "Thresholds") %>% select(est, model, LatentClass, param) %>%
   mutate(prob = (1 / (1 + exp(est))), param = str_to_title(str_remove_all(param, "[$1]")))
```

```
ggplot(pp_plots,
    aes(x = param, y = prob,
        color = LatentClass,
        shape = LatentClass,
        group = LatentClass)) +
geom_point() + geom_line() +
facet_wrap(~ model, ncol = 2) +
labs(title = "LCA Posterior Probability Plot",
        x = "Coping Strategy Items", y = "Probability") +
theme_minimal() + theme(legend.position = "none", axis.text.x = element_text(size = 6))
```

LCA Posterior Probability Plot



ggsave(here("figures","FigS1_compare_kclass_LCAs.png"), dpi=300, height=4, width=6, units="in")

1.5 Plot Final Model - Conditional Item Probability Plot

This syntax generates the figure as presented in the manuscript for the final unconditional LCA model.

Create a function called plot_lca_function that requires 7 arguments (inputs):

- model_name: name of Mplus model object (e.g., model_step1)
- item_num: the number of items in LCA measurement model (e.g., 5)
- class_num: the number of classes (k) in LCA model (e.g., 3)
- item_labels: the item labels for x-axis (e.g., c("Enjoy", "Useful", "Logical", "Job", "Adult"))
- class_labels: the class label names (e.g., c("Adaptive Coping", "Externalizing Behavior", "No Coping"))
- class_legend_order = change the order that class names are listed in the plot legend (e.g., c(2,1,3))
- plot title: include the title of the plot here (e.g., "LCA Posterior Probability Plot")

```
plot_lca_function <- function(model_name,item_num,class_num,item_labels,</pre>
                               class_labels,class_legend_order,plot_title){
mplus_model <- as.data.frame(model_name$gh5$means_and_variances_data$estimated_probs$values)
plot_data <- mplus_model[seq(2, 2*item_num, 2),]</pre>
c_size <- as.data.frame(model_name$class_counts$modelEstimated$proportion)</pre>
colnames(c size) <- paste0("cs")</pre>
c_size <- c_size %>% mutate(cs = round(cs*100, 2))
colnames(plot_data) <- paste0(class_labels, glue(" ({c_size[1:class_num,]}%)"))</pre>
plot_data <- plot_data %>% relocate(class_legend_order)
plot_data <- cbind(Var = paste0("U", 1:item_num), plot_data)</pre>
plot_data$Var <- factor(plot_data$Var,</pre>
               labels = item_labels)
plot_data$Var <- fct_inorder(plot_data$Var)</pre>
pd_long_data <- melt(plot_data, id.vars = "Var")</pre>
# This syntax uses the data.frame created above to produce the plot with `qqplot()`
p <- pd_long_data %>%
  ggplot(aes(x = as.integer(Var), y = value,
  shape = variable, colour = variable, lty = variable)) +
  geom_point(size = 4) + geom_line() +
  scale_x_continuous("", breaks = 1:5, labels = plot_data$Var) +
  scale colour grey() +
  labs(title = plot_title, y = "Probability") +
  theme_cowplot() +
  theme(legend.title = element_blank(),
        legend.position = "top") +
  coord_cartesian(xlim = c(.9, 5.4), ylim = c(-.05, 1.05), expand = FALSE)
```

```
p
return(p)
}
```

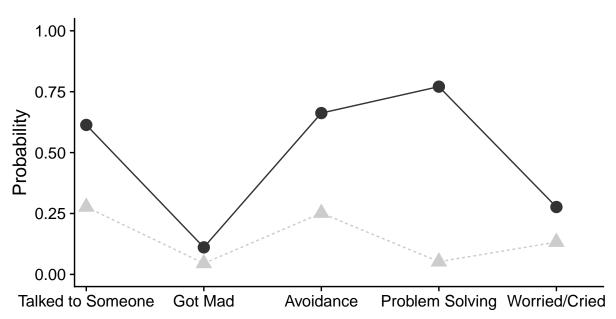
Run the plot_lca_function for the 3-class & 2-class models

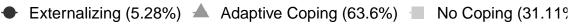
```
c3 <- plot_lca_function(</pre>
 model_name = output_enum[[3]],
 item_num = 5,
 class_num = 3,
  item_labels = c("Talked to Someone", "Got Mad", "Avoidance", "Problem Solving", "Worried/Cried"),
  class_labels = c("Adaptive Coping", "Externalizing", "No Coping"),
  class_legend_order = c(2,1,3),
  plot_title = ""
  )
c2 <- plot_lca_function(</pre>
model_name = output_enum[[2]],
item_num = 5,
class_num = 2,
item_labels = c("Talked to Someone", "Got Mad", "Avoidance", "Problem Solving", "Worried/Cried"),
class_labels = c("Class 1","Class 2"),
class_legend_order = c(1,2),
plot_title = ""
)
```

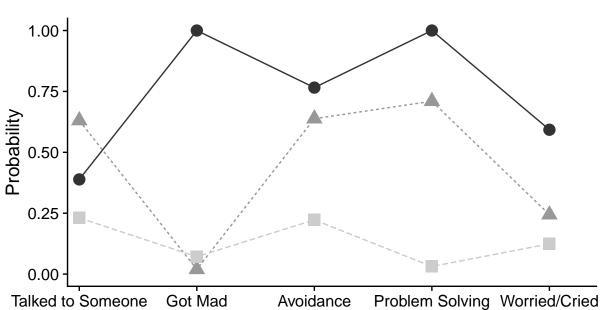
Combine the 3-class and 2-class LCA Posterior Probability Plots (Figure 1)

Note: This code combines 2 ggplot objects using the {patchwork} package.

c2/c3







ggsave(here("figures", "Fig1_LCAs_Combined.png"), dpi=300, height=8.5, width=7, units="in")

Step 1 - Estimate the unconditional model with all covariate & distal outcome variables mentioned in the auxiliary statement.

```
m_step1 <- mplusObject(</pre>
 TITLE = "Step1_3step_automation Behavioral Disorder",
  VARIABLE =
   "categorical = do1 do2 do3 do5 do6;
   usevar = do1 do2 do3 do5 do6;
   classes = c(3);
   !!! NOTE: All auxiliary variables to be considered in the final model should be listed here !!!
   auxiliary =
   SOC_STRS negmood1 posmood1;",
  ANALYSIS =
   "estimator = mlr;
   type = mixture;
   starts = 500 100;",
  SAVEDATA =
   "!!! NOTE: This saved dataset will contain class probabilities and modal assignment columns !!!
   File=3step_BD_savedata_092021.dat;
   Save=cprob;
   Missflag= 999;",
 PLOT =
  "type = plot3;
   series = do1 do2 do3 do5 do6(*);",
 usevariables = colnames(ies_data),
 rdata = ies_data)
m_step1_fit <- mplusModeler(m_step1,</pre>
                 dataout=here("3step_mplus", "Step1_3step_BD.dat"),
                 modelout=here("3step_mplus", "Step1_3step_BD.inp") ,
                 check=TRUE, run = TRUE, hashfilename = FALSE)
```

Step 2 - Extract logits & saved data from the step 1 unconditional model.

Extract logits for the classification probabilities for the most likely latent class

Extract saved data from the step 1 model mplusObject named "m_step1_fit"

Rename the column in savedata for "C" and change to "N"

```
colnames(savedata)[colnames(savedata)=="C"] <- "N"
```

Step 3 (part 1) - Estimate the unconditional model with logits from step 2.

This model is estimated to check that the class proportions are approximately the same as in step 1.

```
m_step2 <- mplusObject(</pre>
 TITLE = "Step2_3step_automation Behavioral Disorder",
 VARIABLE =
 "nominal=N;
 USEVAR = n;
 missing are all (999);
  classes = c(3); ",
 ANALYSIS =
 "estimator = mlr;
 type = mixture;
  starts = 0;",
 MODEL =
    glue(
 "%C#1%
  [n#1@{logit_cprobs[1,1]}];
  [n#20{logit_cprobs[1,2]}];
  %C#2%
  [n#1@{logit_cprobs[2,1]}];
  [n#20{logit_cprobs[2,2]}];
 %C#3%
  [n#1@{logit_cprobs[3,1]}];
  [n#20{logit_cprobs[3,2]}];"),
 PLOT =
 "!type = plot3;
 !series = do1 do2 do3 do5 do6(*);",
 usevariables = colnames(savedata),
 rdata = savedata)
```

Step 3 (part 2) - Add covariates & distal outcomes to the model.

1.7 Moderation - Estimate the final SEM Model

Specification details:

- This example contains two distal outcomes (POSMOOD1 & NEGMOOD1) and one binary covariate (SOC STRS).
- Under each class-specific statement (e.g., %C#1%) the distal outcomes are mentioned to estimate the intercept parameters.
- Moderation is specified by mentioning the statement "OUTCOME on COVARIATE;" under each of the class-specific statements (e.g., %C#1%).
- Class-specific distal means are specified by mentioning the outcome variable inside square brackets (e.g., [POSMOOD1];) repeated under each class-specific model statement. Variances are specified by mentioning the outcome variable without brackets (e.g., POSMOOD1;)
- Parameters are labeled by adding parentheses following parameter name (e.g., [NEGMOOD1] (m01)) this label can be then used to conduct additional tests within the MODELCONSTRAINT = section of the input file.
- Note that the binary covariate (SOC_STRS) is centered so that reported distal means (intercepts) are estimated at the weighted average of Social Stress.

```
m_step3 <- mplusObject(</pre>
  TITLE = "Step3 3step automation Behavioral Disorder",
  VARIABLE =
 "nominal = N;
  usevar = n;
  missing are all (999);
  usevar = SOC_STRS POSMOOD1 NEGMOOD1;
  classes = c(3); ",
  DEFINE =
 "Center SOC_STRS (Grandmean);",
  ANALYSIS =
 "estimator = mlr;
  type = mixture;
  starts = 0;",
 MODEL =
  glue(
 "!!! DISTAL = POSMOOD1 NEGMOOD1 !!!
```

```
!!! MODERATOR = SOC_STRS
  %OVERALL%
  POSMOOD1 on SOC_STRS;
  POSMOOD1;
  NEGMOOD1 on SOC_STRS;
  NEGMOOD1;
  %C#1%
  [n#1@{logit_cprobs[1,1]}];
  [n#20{logit_cprobs[1,2]}];
  [NEGMOOD1] (mO1);
                               !!! estimate conditional intercept !!!
  NEGMOOD1;
                                 !!! estimate conditional variance !!!
  NEGMOOD1 on SOC_STRS (s01); !!! estimate conditional regression !!!
  [POSMOOD1] (m1);
  POSMOOD1;
  POSMOOD1 on SOC_STRS (s1);
  %C#2%
  [n#10{logit_cprobs[2,1]}];
  [n#2@{logit_cprobs[2,2]}];
  [NEGMOOD1] (m02);
  NEGMOOD1;
  NEGMOOD1 on SOC_STRS (s02);
  [POSMOOD1] (m2);
  POSMOOD1;
  POSMOOD1 on SOC_STRS (s2);
  %C#3%
  [n#1@{logit_cprobs[3,1]}];
  [n#2@{logit_cprobs[3,2]}];
  [NEGMOOD1] (mO3);
  NEGMOOD1;
  NEGMOOD1 on SOC_STRS (s03);
  [POSMOOD1] (m3);
  POSMOOD1;
  POSMOOD1 on SOC STRS (s3);"),
  MODELCONSTRAINT =
 "New (diff12 diff13
                                 !!! Testing equivalenve of the conditional intercepts and slope !!!
  diff23 slope12 slope13
  slope23 ndiff12 ndiff13
  ndiff23 nslope12 nslope13
  nslope23);
  diff12 = m1-m2; ndiff12 = m01-m02;
```

```
diff13 = m1-m3; ndiff13 = m01-m03;
  diff23 = m2-m3;    ndiff23 = m02-m03;
  slope12 = s1-s2; nslope12 = s01-s02;
  slope13 = s1-s3; nslope13 = s01-s03;
  slope23 = s2-s3; nslope23 = s02-s03;",
  MODELTEST =
  ## NOTE: Only a single Wald test can be conducted per model run. Therefore,
  ## this example requires running separate models for each omnibus test (e.g.,
  ## 4 models; 2 outcomes and 2 slope coefficients). This can be done by
  ## commenting out all but one test and then making multiple input/output files.
               !!! Distal outcome omnibus Wald test for `POSMOOD1` !!!
 "m1=m2;
  m2=m3;
  !s1=s2;
               !!! Slope difference omnibus Wald test `POSMOOD1 on SOC_STRS` !!!
  !s2=s3;
               !!! Distal outcome omnibus Wald test for `NEGMOOD1` !!!
  !m01=m02;
  !m02=m03;
  !s01=s02;
               !!! Slope difference omnibus Wald test for `POSMOOD1 on SOC_STRS` !!!
  !s02=s03;",
  usevariables = colnames(savedata),
  rdata = savedata)
m_step3_fit <- mplusModeler(m_step3,</pre>
                 dataout=here("3step_mplus", "Step3_3step_BD.dat"),
                 modelout=here("3step_mplus", "Step3_3step_BD.inp"),
                 check=TRUE, run = TRUE, hashfilename = FALSE)
```

End of 3-step procedure

Estimate step 3 moderation model with covariate un-centered to produce simple-slopes plots

• Intercepts is fixed at the first level of the covariate SOC_STRS

Note: Here the update() function is used to take the previous model and remove the Mplus syntax within the DEFINE statement that was used to center the covariate Social Stress. Next, the updated model input syntax is used to estimate a new model. To learn more about the update function see the MplusAutomation tutorial article (https://www.tandfonline.com/doi/pdf/10.1080/10705511.2017.1402334).

1.8 Distal Outcome Plot

Note: The distal outcome means are estimated with the binary covariate (SOC_STRS) at the weighted average. This is specified by centering social stress as shown in the Step-3 model syntax.

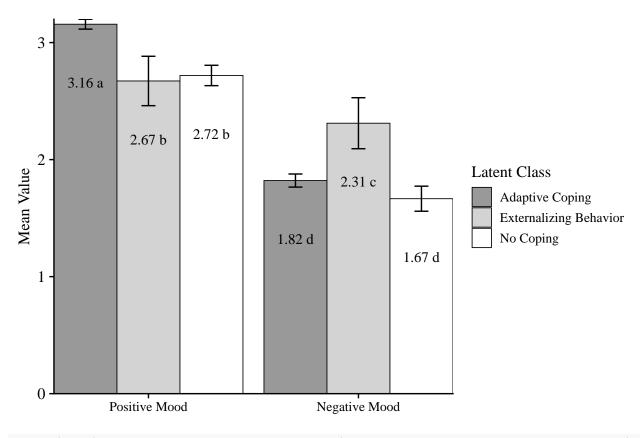
This syntax reads in the Step3 model & extract model parameter estimates.

```
model_step3 <- readModels(here("3step_mplus", "Step3_3step_BD.out"), quiet = TRUE)
model_step3 <- data.frame(model_step3$parameters$unstandardized)</pre>
```

This syntax is used to create the data-frame that produces the distal outcome bar plot.

```
distal_data <- model_step3 %>%
  filter(paramHeader == "Intercepts") %>%
  mutate(param = case_when(
    param == "POSMOOD1" ~ "Positive Mood",
    param == "NEGMOOD1" ~ "Negative Mood")) %>%
  mutate(LatentClass = factor(LatentClass,
    labels = c("Adaptive Coping", "Externalizing Behavior", "No Coping"))) %>%
  mutate(value_labels = c("3.16 a", "1.82 d", "2.67 b", "2.31 c", "2.72 b", "1.67 d"))
```

Plot distal outcomes grouped by class (Figure 3)



ggsave(here("figures","Fig3_distal_barplot.png"), dpi=300, height=4, width=6, units="in")

1.9 Simple Slope Plots

Note: The un-centered distal intercepts represent the conditional means when the binary covariate is at its first level SOC_STRS = 0 (i.e., no social stress). Therefore, the conditional mean for SOC_STRS = 1 (i.e., social stress) can be calculated by adding the associated slope coefficient to the intercept.

Read in the un-centered model & extract relevant parameters

```
model_uncen <- readModels(here("3step_mplus", "Step3_uncentered_BD.out"), quiet = TRUE)

model_uncen <- data.frame(model_uncen$parameters$unstandardized)

slope_data <- model_uncen %>%
   filter(str_detect(paramHeader, 'ON|Inter')) %>%
   unite("param", paramHeader:param, remove = TRUE) %>%
   mutate(param = str_replace(param, "MOOD1.ON_SOC_STRS", "_COEF")) %>%
   mutate(param = str_remove_all(param, "Intercepts_|MOOD1")) %>%
```

```
mutate(LatentClass = factor(LatentClass,
    labels = c("Adaptive Coping (63.6%)", "Externalizing Behavior (5.3%)", "No Coping (31.1%)")))
```

Positive mood simple slope graph

Prepare data-frame for plotting

Plot positive mood simple slope graph

```
p_plot <- ggplot(plot_pos,</pre>
            aes(y=value, x=variable,
                color=LatentClass,
                group=LatentClass,
                shape=LatentClass,
                lty=LatentClass)) +
  geom_point(size = 4) + geom_line() +
  xlab("") + ylab("Positive Mood") + ylim(2.5,3.5)+
  scale_colour_grey() +
  theme_classic() +
  theme(text=element_text(family="Times New Roman", size=12),
        axis.text.x=element_text(size=12),
        legend.text = element_text(family="Times New Roman", size=10),
        legend.position = "top", legend.title = element blank()) +
      annotate(geom = "text",
           x = 1.8, y = 2.77,
           label = "N.S.", color = "black") +
      annotate(geom = "text",
           x = 1.8, y = 2.60,
           label = "N.S.", color = "black")
```

Negative mood simple slope graph

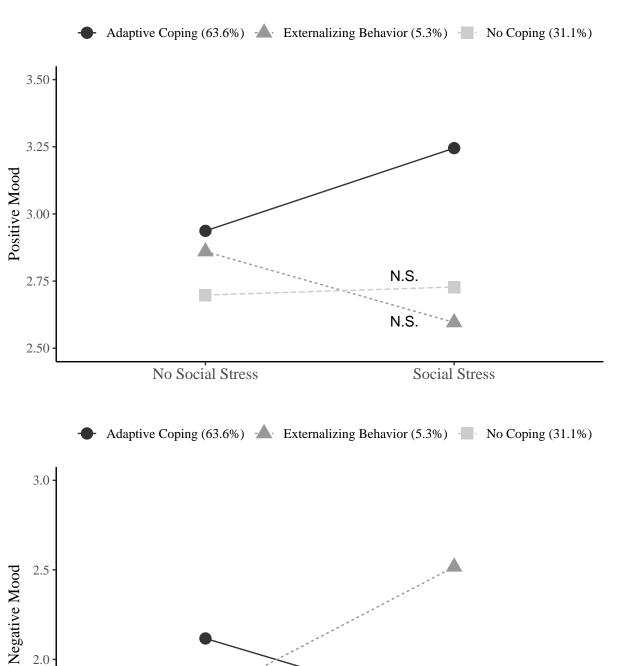
Prepare data-frame for plotting

Plot negative mood simple slope graph

```
n_plot <- ggplot(plot_neg,</pre>
            aes(y=value, x=variable,
            color=LatentClass,
            group=LatentClass,
            shape=LatentClass,
            lty=LatentClass)) +
  geom_point(size=4) + geom_line() +
  xlab("") + ylab("Negative Mood") + ylim(1.5,3)+
  scale_colour_grey() + theme_classic() +
  theme(text=element_text(family="Times New Roman", color = "black", size=12),
        axis.text.x=element_text(size=12),
        legend.text = element_text(family="Times New Roman", size=10),
       legend.position = "top", legend.title = element_blank()) +
    annotate(geom = "text",
       x = 1.8, y = 1.6,
       label = "N.S.", color = "black")
```

Combine the two simple slopes graphs for distal outcomes positive & negative mood (Figure 4)

```
p_plot / n_plot # combines plots using the {patchwork} package
```



No Social Stress

ggsave(here("figures", "Fig4_simple_slopes.png"), dpi=300, height=8.5, width=6.5, units="in")

2.0 Repsonse Pattern Table

To replicate the response pattern table produced in this study (*Table 3*) see the associated R/MplusAutomation code example here:

Lab10.1-Response-Patterns

Additional R/MplusAutomation Examples

Tidy-Workflow-with-MplusAutomation

References:

Hallquist, Michael N., and Joshua F. Wiley. 2018. "MplusAutomation: An R Package for FacilitatingLarge-Scale Latent Variable Analyses in Mplus." Structural Equation Modeling, 1–18. https://doi.org/10.1080/10705511.2017. 1402334.

Miller, Jon D. Longitudinal Study of American Youth (LSAY), Seventh Grade Data, 1987-1988; 2015-2016. Ann Arbor, MI: Inter-university Consortium for Political and Social Research [distributor], 2019-04-23. https://doi.org/10.3886/ICPSR37287.v1

Müller, Kirill. 2017.Here: A Simpler Way to Find Your Files. https://CRAN.R-project.org/package=here.

Muthen L.K., & Muthen B.O. (1998-2017) Mplus User's Guide. Eight Edition. Los Angelos, CA: Muthen & Muthen.

R Core Team. 2019.R: A Language and Environment for Statistical Computing. Vienna, Austria: R Foundation for Statistical Computing. https://www.R-project.org/.

Wickham H, et al. (2019). "Welcome to the tidyverse." Journal of Open Source Software, 4(43), 1686. doi: 10.21105/joss.01686.