

# Appendix - Implement LCA Analyses with MplusAutomation

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## Motivation for MplusAutomation workflow:

This R tutorial is intended to provide a template for estimating latent class analysis (LCA), integrating the unconditional LCA with a larger SEM model, and producing publication ready figures in a systematic manner. Using this approach all code for data pre-processing, model estimation, tabulation, and figure processing can be contained within a single script providing clear documentation. It is the authors' belief that this dramatically reduces risk of user-error, is conducive to open-science philosophy, and scientific transparency. All models are estimated using **Mplus** (Muthén & Muthén, 1998 - 2017) using the wrapping package **MplusAutomation** (Hallquist & Wiley, 2018). This method requires the user to have the proprietary software **Mplus** installed on their computer.

This approach also relies on the utility of **R-Projects**. This provides a structured framework for organizing all associated data files, Mplus input/output files, scripts, and figures. Given the large number of Mplus output files inherent to fitting mixture models, creating a system of project sub-folders greatly improves organization (i.e., folders; 'data', 'mplus\_files' 'figures', etc.) Additionally, the communication between R and Mplus requires the specification of file-paths, a procedure which is streamlined by use of **R-projects**. Due to the reliance on file-paths, the **here** package ensures that the syntax for file-paths is uniform across operating systems (e.g., Windows or Mac) enabling reproducibility.

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## Preparation

### Download the R-Project

Link to Github repository here: <https://github.com/garberadamc/LCA-COPING-BD>

For readers **unfamiliar with Github** and version controlled R-projects:

1. On the repository page, click the green **Code** button and in the menu choose option **Download ZIP**
2. Place the un-zipped downloaded folder on your desktop
3. Within this folder open the file with the blue cube icon that is file named **LCA-COPING-BD.Rproj**
4. Next open the file containing all analysis code named **rcode-lca-coping-bd.R**.

**Note:** If preferred users may follow analyses using the Rmarkdown script (**.Rmd**).

### Project folder organization

The following sub-folders will be used to contain files:

1. “data”; 2. “enum\_mplus”; 3. “3step\_mplus”; 4. “figures”

**Note regarding project location:** If the main project folder is located within too many nested folders it may result in a file-path error when estimating models with `MplusAutomation`.

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To install package {rhdf5}

```
if (!requireNamespace("BiocManager", quietly = TRUE))
  install.packages("BiocManager")

BiocManager::install("rhdf5")
```

Load packages

```
library(MplusAutomation)
library(tidyverse)
library(rhdf5)
library(here)
library(glue)
library(gt)
library(reshape2)
library(cowplot)
library(patchwork)
```

Read in data. File-paths are set using R-Projects & the {here} package

```
ies_data <- read_csv(here("data", "ies_bd_data.csv"),
  na=c("", ".", "999"))
```

---

## 1.1 Enumeration:

Estimate  $K$ -class models with 1 through 6 classes.

---

**Specification details that are specific to applied coping example:**

- Within the `lapply()` function 1:6 indicates the number of  $K$ -class models to estimate.
- The measurement model item names for the coping strategy example are listed after the `categorical = ...; & usevar = ...;` statements.
- Note, that for Latent Profile Analysis (LPA) the `categorical = ...;` statement would be removed so that items are estimated as continuous variables.

```

lca_k1_6 <- lapply(1:6, function(k) {
  lca_enum <- mplusObject(

    TITLE = glue("Class-{k} LCA Enumeration - Youth Coping Strategies"),

    VARIABLE = glue(
      "categorical = do1 do2 do3 do5 do6; !!! Coping strategy items for measurement model !!!
      usevar = do1 do2 do3 do5 do6;

      classes = c({k});"),

    ANALYSIS =
      "estimator = mlr;
      type = mixture;
      processors=10;",

    PLOT =
      "type = plot3;
      series = do1 do2 do3 do5 do6(*);",

    OUTPUT = "sampstat tech11 tech14;",

    usevariables = colnames(ies_data),
    rdata = ies_data)

  lca_enum_fit <- mplusModeler(lca_enum,
    dataout=glue(here("enum_mplus", "c_lca_enum_bd.dat")),
    modelout=glue(here("enum_mplus", "c{k}_lca_enum_bd.inp")) ,
    check=TRUE, run = TRUE, hashfilename = FALSE)
})

```

---

## 1.2 Generate Model Fit Summary Table

- This syntax can be used to compare model fit from the series of LCA models generated during enumeration ( *Table 4* in manuscript).
  - The code produces a table that is approximately in APA format.
- 

```

output_enum <- readModels(here("enum_mplus"), quiet = TRUE)

enum_extract <- LatexSummaryTable(output_enum,
  keepCols=c("Title", "Parameters", "LL", "BIC", "aBIC",
    "BLRT_PValue", "T11_VLMR_PValue", "Observations"))

```

Calculate relevant fit indices for summary table

```
allFit <- enum_extract %>%
  mutate(aBIC = -2*LL+Parameters*log((Observations+2)/24)) %>%
  mutate(CIAC = -2*LL+Parameters*(log(Observations)+1)) %>%
  mutate(AWE = -2*LL+2*Parameters*(log(Observations)+1.5)) %>%
  mutate(SIC = -.5*BIC) %>%
  mutate(expSIC = exp(SIC - max(SIC))) %>%
  mutate(BF = exp(SIC-lead(SIC))) %>%
  mutate(cmPk = expSIC/sum(expSIC)) %>%
  select(1:5,9:10,6:7,13,14) %>%
  arrange(Parameters)
```

Create table using {gt}

```
allFit %>%
  mutate(Title = str_remove(Title, " LCA Enumeration - Youth Coping Strategies")) %>%
  gt() %>%
  tab_header(
    title = md("**Model Fit Summary Table**"), subtitle = md("&nbsp;") %>%
  cols_label(
    Title = "Classes",
    Parameters = md("Par"),
    LL = md("*LL*"),
    T11_VLMR_PValue = "VLMR",
    BLRT_PValue = "BLRT",
    BF = md("BF"),
    cmPk = md("*cmP_k*")) %>%
  tab_footnote(
    footnote = md(
      "*Note.* Par = parameters; *LL* = log likelihood;
      BIC = bayesian information criterion;
      aBIC = sample size adjusted BIC; CAIC = consistent Akaike information criterion;
      AWE = approximate weight of evidence criterion;
      BLRT = bootstrapped likelihood ratio test p-value;
      VLMR = Vuong-Lo-Mendell-Rubin adjusted likelihood ratio test p-value;
      cmPk = approximate correct model probability."),
    locations = cells_title()) %>%
  tab_options(column_labels.font.weight = "bold") %>%
  fmt_number(10,decimals = 2,
    drop_trailing_zeros=TRUE,
    suffixing = TRUE) %>%
  fmt_number(c(3:9,11), decimals = 0) %>%
  fmt_missing(1:11, missing_text = "--") %>%
  fmt(c(8:9,11), fns = function(x)
    ifelse(x<0.001, "<.001",
      scales::number(x, accuracy = 0.01))) %>%
  fmt(10, fns = function(x)
    ifelse(x>100, ">100",
      scales::number(x, accuracy = .1)))
```

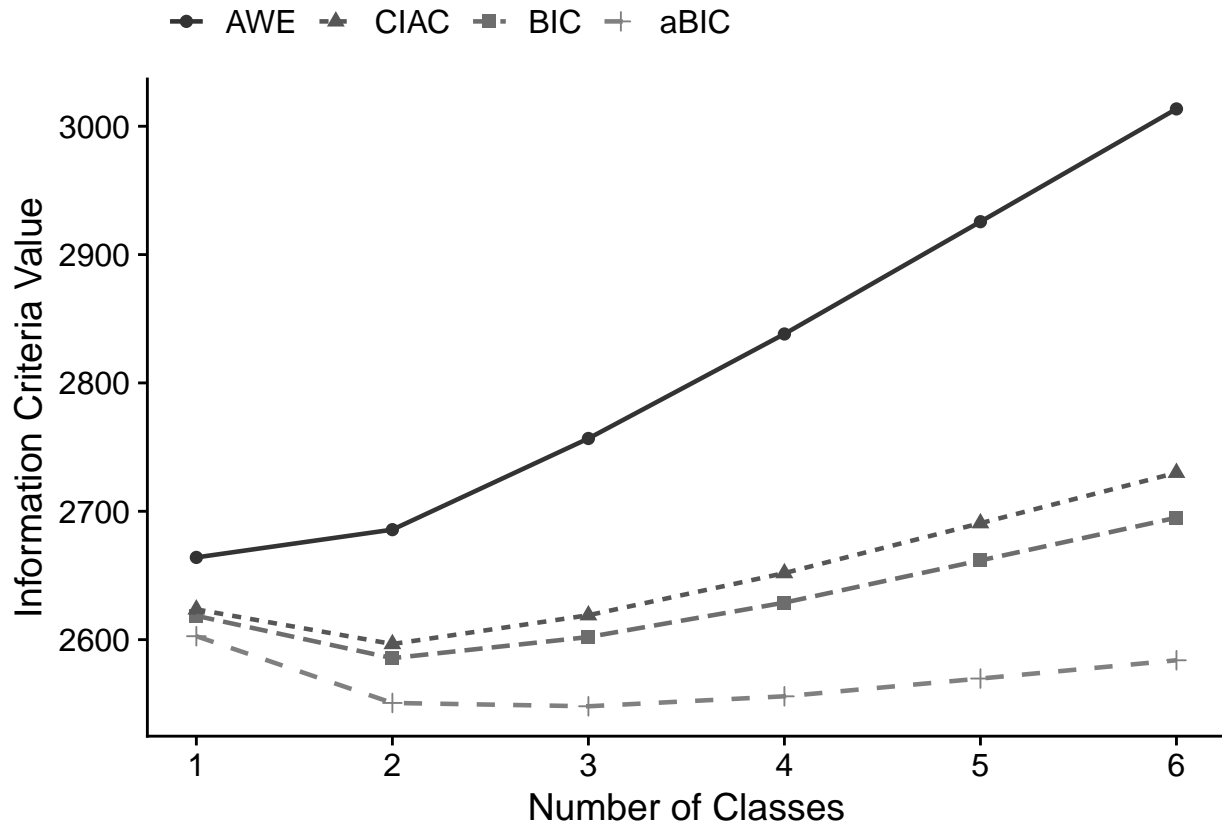
Model Fit Summary Table<sup>1</sup>

Classes	Par	LL	BIC	aBIC	CIAC	AWE	BLRT	VLMR	BF	cmP <sub>k</sub>
Class-1	5	-1,294	2,619	2,603	2,624	2,664	–	–	>100	<.001
Class-2	11	-1,259	2,586	2,551	2,597	2,686	<.001	<.001	>100	1.00
Class-3	17	-1,249	2,602	2,548	2,619	2,757	<.001	0.12	>100	<.001
Class-4	23	-1,244	2,629	2,556	2,652	2,838	0.27	0.01	>100	<.001
Class-5	29	-1,242	2,662	2,570	2,691	2,926	0.67	0.46	>100	<.001
Class-6	35	-1,241	2,695	2,584	2,730	3,014	0.67	0.49	–	<.001

<sup>1</sup>Note. Par = parameters; LL = log likelihood; BIC = bayesian information criterion; aBIC = sample size adjusted BIC; CAIC = consistent Akaike information criterion; AWE = approximate weight of evidence criterion; BLRT = bootstrapped likelihood ratio test p-value; VLMR = Vuong-Lo-Mendell-Rubin adjusted likelihood ratio test p-value; cmPk = approximate correct model probability.

### 1.3 Plot Information Criteria

```
allFit %>% select(2:7) %>%
  rowid_to_column() %>%
  pivot_longer(`BIC`:`AWE`,
    names_to = "Index",
    values_to = "ic_value") %>%
  mutate(Index = factor(Index,
    levels = c("AWE", "CIAC", "BIC", "aBIC"))) %>%
  ggplot(aes(x = rowid, y = ic_value,
    color = Index, shape = Index,
    group = Index, lty = Index)) +
  geom_point(size = 2.0) + geom_line(size = .8) +
  scale_x_continuous(breaks = 1:6) +
  scale_colour_grey(end = .5) +
  theme_cowplot() +
  labs(x = "Number of Classes", y = "Information Criteria Value") +
  theme(legend.title = element_blank(),
    legend.position = "top")
```



Save plot in the figures folder.

```
ggsave(here("figures", "Fig2_IC_plot.png"), dpi=300, height=5, width=7, units="in")
```

## 1.4 Compare Conditional Item Probability Plots

This syntax produces a plot containing a series of LCA probability plot facets. This can be used to compare solutions across  $K$ -class models.

```
model_results <- data.frame()

for (i in 1:length(output_enum)) {
  temp <- output_enum[[i]]$parameters$unstandardized
  temp <- data.frame(unclass(temp)) %>%
    mutate(model = paste0(i, "-Class Model"))
  model_results <- rbind(model_results, temp)
}

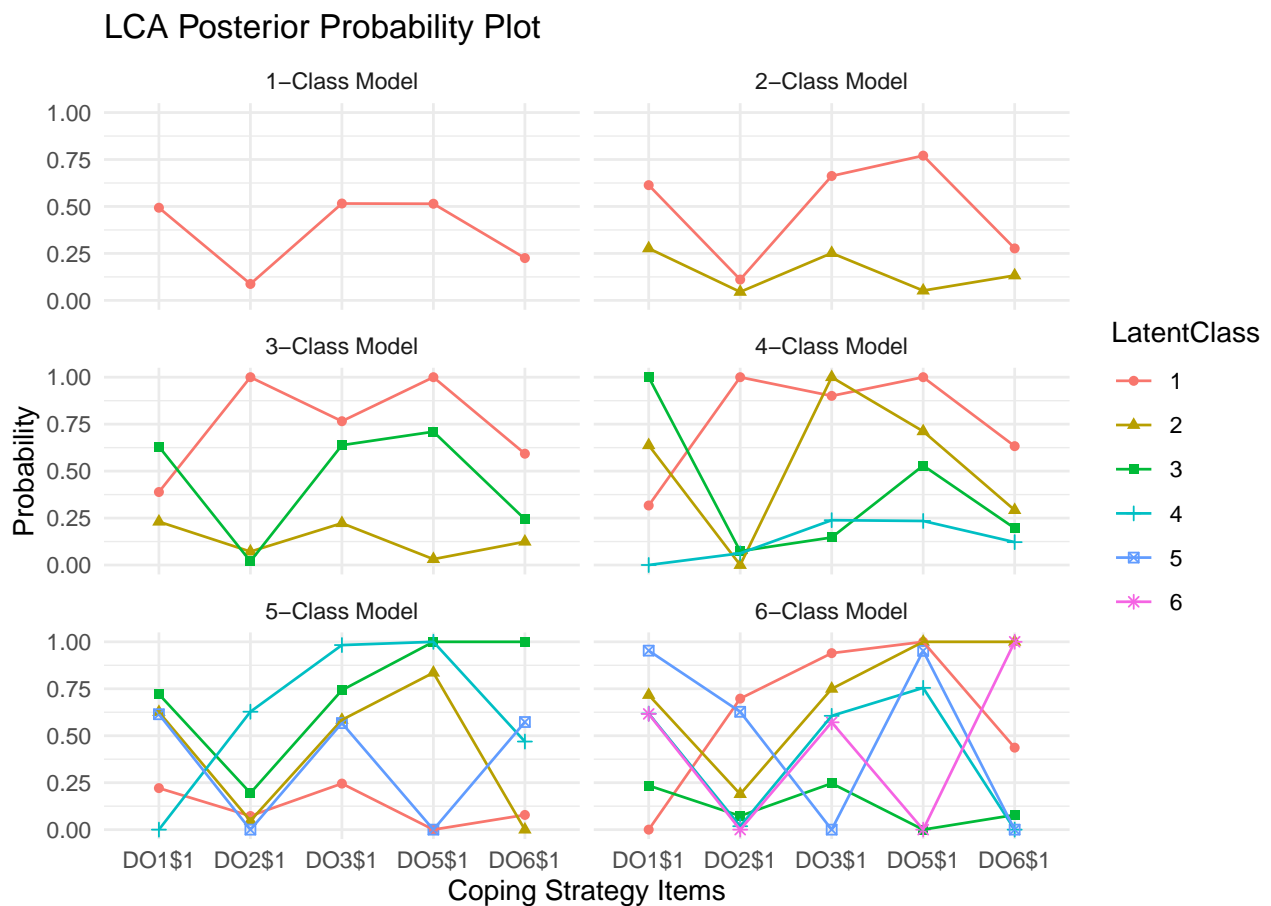
model_results <- model_results %>%
```

```

filter(paramHeader == "Thresholds") %>%
select(est, model, LatentClass, param) %>%
mutate(prob = (1 / (1 + exp(est))))

ggplot(model_results,
  aes(x = param, y = prob,
      color = LatentClass,
      shape = LatentClass,
      group = LatentClass)) +
geom_point() + geom_line() +
facet_wrap(~ model, ncol = 2) +
labs(title = "LCA Posterior Probability Plot",
      x = "Coping Strategy Items", y = "Probability") +
theme_minimal()

```



```

ggsave(here("figures", "FigA1_compare_kclass_LCAs.png"), dpi=300, height=4, width=6, units="in")

```

## 1.5 Plot Final Model - Conditional Item Probability Plot

This code generates the figure as presented in the manuscript for the final unconditional LCA model.

---

This syntax creates a function called `plot_lca_function` that requires 7 arguments (inputs):

- `model_name`: name of Mplus model object (e.g., `model_step1`)
- `item_num`: the number of items in LCA measurement model (e.g., 5)
- `class_num`: the number of classes ( $k$ ) in LCA model (e.g., 3)
- `item_labels`: the item labels for x-axis (e.g., `c("Enjoy", "Useful", "Logical", "Job", "Adult")`)
- `class_labels`: the class label names (e.g., `c("Adaptive Coping", "Externalizing Behavior", "No Coping")`)
- `class_legend_order` = change the order that class names are listed in the plot legend (e.g., `c(2,1,3)`)
- `plot_title`: include the title of the plot here (e.g., "LCA Posterior Probability Plot")

Read in plot data from Mplus output file `Step1_3step_BD.out`

```
model_step1 <- readModels(here("3step_mplus", "Step1_3step_BD.out"), quiet = TRUE)

plot_lca_function <- function(model_name,item_num,class_num,item_labels,
                              class_labels,class_legend_order,plot_title){

mplus_model <- as.data.frame(model_name$gh5$means_and_variances_data$estimated_probs$values)
plot_data <- mplus_model[seq(2, 2*item_num, 2),]

c_size <- as.data.frame(model_name$class_counts$modelEstimated$proportion)
colnames(c_size) <- paste0("cs")
c_size <- c_size %>% mutate(cs = round(cs*100, 2))
colnames(plot_data) <- paste0(class_labels, glue(" ({c_size[1:class_num,]}%))")
plot_data <- plot_data %>% relocate(class_legend_order)

plot_data <- cbind(Var = paste0("U", 1:item_num), plot_data)
plot_data$Var <- factor(plot_data$Var,
                      labels = item_labels)
plot_data$Var <- fct_inorder(plot_data$Var)

pd_long_data <- melt(plot_data, id.vars = "Var")

# This syntax uses the data-frame created above to produce the plot with `ggplot()`

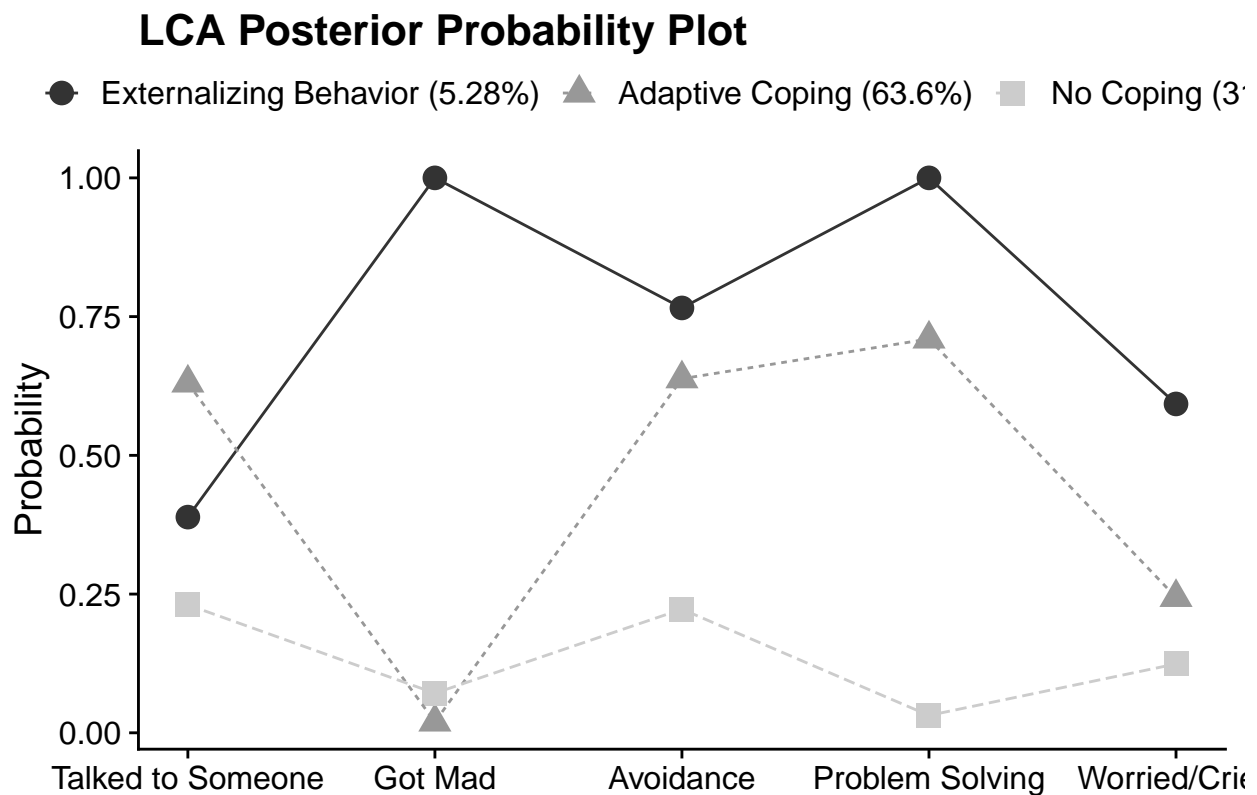
p <- pd_long_data %>%
  ggplot(aes(x = as.integer(Var), y = value,
             shape = variable, colour = variable, lty = variable)) +
  geom_point(size = 4) + geom_line() +
  scale_x_continuous("", breaks = 1:5, labels = plot_data$Var) +
  scale_colour_grey() +
  labs(title = plot_title, y = "Probability") +
  theme_cowplot() +
  theme(legend.title = element_blank(),
        legend.position = "top")

p
return(p)
}
```



Run the `plot_lca_function` by specifying each input (*Figure 1*)

```
plot_lca_function(  
  model_name = model_step1,  
  item_num = 5,  
  class_num = 3,  
  item_labels = c("Talked to Someone", "Got Mad", "Avoidance", "Problem Solving", "Worried/Cried"),  
  class_labels = c("Adaptive Coping", "Externalizing Behavior", "No Coping"),  
  class_legend_order = c(2,1,3),  
  plot_title = "LCA Posterior Probability Plot"  
)
```



```
ggsave(here("figures", "Fig1_LCA_3-Class.png"), dpi=300, height=5, width=7, units="in")
```

---

## 1.6 Manual “3-Step” ML Auxiliary Variable Integration Method

---

Step 1 - Estimate the unconditional model with all covariate & distal outcome variables mentioned in the auxiliary statement.

```
m_step1 <- mplusObject(
  TITLE = "Step1_3step_automation Behavioral Disorder",
  VARIABLE =
    "categorical = do1 do2 do3 do5 do6;

    usevar = do1 do2 do3 do5 do6;

    classes = c(3);

    !!! NOTE: All auxiliary variables to be considered in the final model should be listed here !!!
    auxiliary =
    FEMALE ETHN_CMP SOC_STRS
    BOTHR_U negmood1 posmood1;",

  ANALYSIS =
    "estimator = mlr;
    type = mixture;
    starts = 500 100;",

  SAVEDATA =
    "!!! NOTE: This saved dataset will contain class probabilities and modal assignment columns !!!
    File=3step_BD_savedata_012020.dat;
    Save=cprob;
    Missflag= 999;",

  MODEL = "",
  OUTPUT = "",

  PLOT =
    "type = plot3;
    series = do1 do2 do3 do5 do6(*);",

  usevariables = colnames(ies_data),
  rdata = ies_data)

m_step1_fit <- mplusModeler(m_step1,
  dataout=here("3step_mplus", "Step1_3step_BD.dat"),
  modelout=here("3step_mplus", "Step1_3step_BD.inp") ,
  check=TRUE, run = TRUE, hashfilename = FALSE)
```

---

## Step 2 - Extract logits & saved data from the step 1 unconditional model.

Extract logits for the classification probabilities for the most likely latent class

```
logit_cprobs <- as.data.frame(m_step1_fit[["results"]]  
                             [["class_counts"]]  
                             [["logitProbs.mostLikely"]])
```

Extract saved data from the step 1 model `mplusObject` named “m\_step1\_fit”

```
savedata <- as.data.frame(m_step1_fit[["results"]]  
                          [["savedata"]])
```

Rename the column in `savedata` for “C” and change to “N”

```
colnames(savedata)[colnames(savedata)=="C"] <- "N"
```

## Step 3 (part 1) - Estimate the unconditional model with logits from step 2.

This model is estimated to check that the class proportions are approximately the same as in step 1.

```
m_step2 <- mplusObject(  
  TITLE = "Step2_3step_automation Behavioral Disorder",  
  
  VARIABLE =  
    "nominal=N;  
    USEVAR = n;  
    missing are all (999);  
    classes = c(3); ",  
  
  ANALYSIS =  
    "estimator = mlr;  
    type = mixture;  
    starts = 0;",  
  
  MODEL =  
    glue(  
      "%C#1%  
      [n#1@{logit_cprobs[1,1]}};  
      [n#2@{logit_cprobs[1,2]}};  
  
      %C#2%  
      [n#1@{logit_cprobs[2,1]}};  
      [n#2@{logit_cprobs[2,2]}};  
  
      %C#3%  
      [n#1@{logit_cprobs[3,1]}};  
      [n#2@{logit_cprobs[3,2]}}];"),  
  
  OUTPUT = "!tech11  tech14 res;",
```

```

PLOT =
"!type = plot3;
!series = do1 do2 do3 do5 do6(*);",

usevariables = colnames(savedata),
rdata = savedata)

m_step2_fit <- mplusModeler(m_step2,
                           dataout=here("3step_mplus", "Step2_3step_BD.dat"),
                           modelout=here("3step_mplus", "Step2_3step_BD.inp"),
                           check=TRUE, run = TRUE, hashfilename = FALSE)

```

---

Step 3 (part 2) - Add covariates & distal outcomes to the model.

## 1.7 Moderation - Estimate the final SEM Model

---

### Specification details:

- This example contains two distal outcomes (POSMOOD1 & NEGMOOD1) and one binary covariate (SOC\_STRS).
- Under each class-specific statement (e.g., %C#1%) the distal outcomes are mentioned to estimate the intercept parameters.
- Moderation is specified by mentioning the "outcome ON covariate;" syntax under each of the class-specific statements.
- Note that the binary covariate is centered so that reported distal means (intercepts) are estimated at the weighted average of Social Stress.

```

m_step3 <- mplusObject(
  TITLE = "Step3_3step_automation Behavioral Disorder",

  VARIABLE =
  "nominal = N;
  usevar = n;
  missing are all (999);

  usevar = SOC_STRS POSMOOD1 NEGMOOD1;
  classes = c(3); ",

  DEFINE =
  "Center SOC_STRS (Grandmean);",

  ANALYSIS =
  "estimator = mlr;
  type = mixture;
  starts = 0;",

  MODEL =

```

```

glue(
"!DISTAL = POSMOOD1 NEGMOOD1
!MODERATOR = SOC_STRS

%OVERALL%
POSMOOD1 on SOC_STRS;
POSMOOD1;

NEGMOOD1 on SOC_STRS;
NEGMOOD1;

%C#1%
[n#1@{logit_cprobs[1,1]}};
[n#2@{logit_cprobs[1,2]}};

[NEGMOOD1] (m01);
NEGMOOD1;
NEGMOOD1 on SOC_STRS (s01);    !!! estimate conditional intercept !!!
                                !!! estimate conditional regression !!!

[POSMOOD1] (m1);
POSMOOD1;
POSMOOD1 on SOC_STRS (s1);

%C#2%
[n#1@{logit_cprobs[2,1]}};
[n#2@{logit_cprobs[2,2]}};

[NEGMOOD1] (m02);
NEGMOOD1;
NEGMOOD1 on SOC_STRS (s02);

[POSMOOD1] (m2);
POSMOOD1;
POSMOOD1 on SOC_STRS (s2);

%C#3%
[n#1@{logit_cprobs[3,1]}};
[n#2@{logit_cprobs[3,2]}};

[NEGMOOD1] (m03);
NEGMOOD1;
NEGMOOD1 on SOC_STRS (s03);

[POSMOOD1] (m3);
POSMOOD1;
POSMOOD1 on SOC_STRS (s3);"),

MODELCONSTRAINT =
"New (diff12 diff13
diff23 slope12 slope13
slope23 ndiff12 ndiff13
ndiff23 nslope12 nslope13
nslope23);

```

```

diff12 = m1-m2;    ndiff12 = m01-m02;
diff13 = m1-m3;    ndiff13 = m01-m03;
diff23 = m2-m3;    ndiff23 = m02-m03;
slope12 = s1-s2;   nslope12 = s01-s02;
slope13 = s1-s3;   nslope13 = s01-s03;
slope23 = s2-s3;   nslope23 = s02-s03;

MODELTEST =
## NOTE: Only a single Wald test can be conducted per model run. Therefore,
## this example requires running separate models for each omnibus test (e.g.,
## 4 models; 2 outcomes and 2 slope coefficients). This can be done by
## commenting out all but one test and then making multiple input/output files.

"m1=m2;          !!! Distal outcome omnibus Wald test for `POSMOOD1` !!!
m2=m3;

!s1=s2;          !!! Slope difference omnibus Wald test `POSMOOD1 on SOC_STRS` !!!
!s2=s3;

!m01=m02;        !!! Distal outcome omnibus Wald test for `NEGMOOD1` !!!
!m02=m03;

!s01=s02;        !!! Slope difference omnibus Wald test for `POSMOOD1 on SOC_STRS` !!!
!s02=s03;",

usevariables = colnames(savedata),
rdata = savedata)

m_step3_fit <- mplusModeler(m_step3,
                           dataout=here("3step_mplus", "Step3_3step_BD.dat"),
                           modelout=here("3step_mplus", "Step3_3step_BD.inp"),
                           check=TRUE, run = TRUE, hashfilename = FALSE)

```

## End of 3-step procedure

---

**Estimate step 3 model with covariate un-centered for simple-slopes plots.** This puts the intercepts fixed at the first level of the covariate (SOC\_STRS). These values are needed to produce the simple slope plots.

**Note:** Here the `update()` function is used to take the previous model and remove the Mplus syntax within the `DEFINE` statement that was used to center the covariate `Social Stress`. Next, the updated model input syntax is used to estimate a new model. To learn more about the `update` function see the `MplusAutomation` tutorial article (<https://www.tandfonline.com/doi/pdf/10.1080/10705511.2017.1402334>).

```

m_uncen <- update(m_step3,
  DEFINE = ~" ") # This update removes the centering syntax from the model object `m_step3`

m_uncen_fit <- mplusModeler(m_uncen,
                           dataout=here("3step_mplus", "Step3_3step_BD.dat"),
                           modelout=here("3step_mplus", "Step3_uncentered_BD.inp"),
                           check=TRUE, run = TRUE, hashfilename = FALSE)UCSB_Navy_mark.png

```

---

## 1.8 Distal Outcome Plot

---

**Note:** The distal outcome means are estimated with the binary covariate (SOC\_STRS) at the weighted average. This is specified by centering social stress as shown in the **Step-3** model syntax.

This syntax reads in the **Step3** model & extract model parameter estimates.

```
model_step3 <- readModels(here("3step_mplus", "Step3_3step_BD.out"), quiet = TRUE)

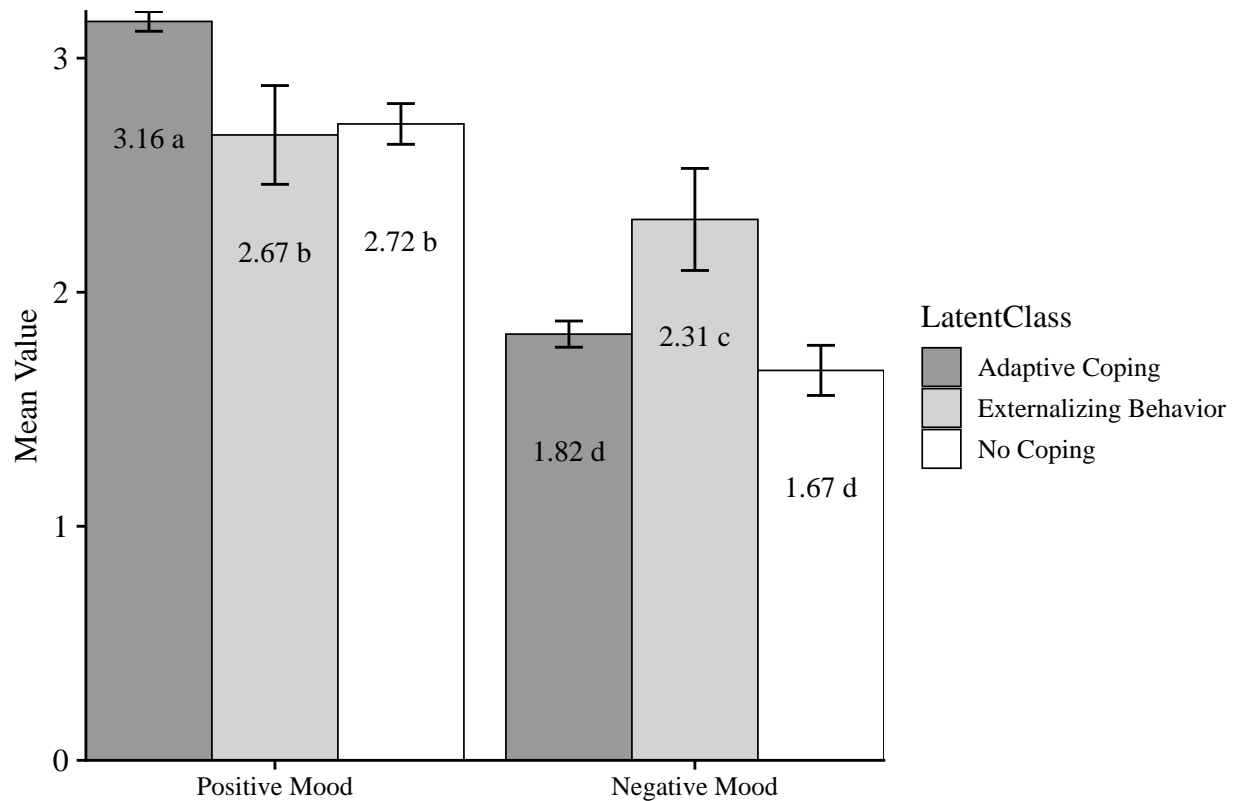
model_step3 <- data.frame(model_step3$parameters$unstandardized)
```

This syntax is used to create the data-frame that produces the distal outcome bar plot.

```
distal_data <- model_step3 %>%
  filter(paramHeader == "Intercepts") %>%
  mutate(param = case_when(
    param == "POSMOOD1" ~ "Positive Mood",
    param == "NEGMOOD1" ~ "Negative Mood")) %>%
  mutate(LatentClass = factor(LatentClass,
    labels = c("Adaptive Coping", "Externalizing Behavior", "No Coping"))) %>%
  mutate(value_labels = c("3.16 a", "1.82 d", "2.67 b", "2.31 c", "2.72 b", "1.67 d"))
```

### Plot distal outcomes grouped by class (*Figure 3*)

```
ggplot(distal_data,
  aes(fill=LatentClass, y=est, x=fct_rev(param))) +
  geom_bar(position="dodge", stat="identity", color="black", size=.3) +
  geom_errorbar(aes(ymin=est-se, ymax=est+se),
    width=.2, position=position_dodge(.9)) +
  geom_text(aes(y = est -.5, label = value_labels),
    family="Times New Roman", size=4,
    position=position_dodge(.9)) +
  scale_fill_grey(start = 0.6, end = 1.0) +
  theme_cowplot() +
  xlab("") + ylab("Mean Value") +
  theme(text=element_text(family="Times New Roman", size=12),
    axis.text.x=element_text(size=10)) +
  coord_cartesian(expand = FALSE)
```



```
ggsave(here("figures", "Fig3_distal_barplot.png"), dpi=300, height=4, width=6, units="in")
```

## 1.9 Simple Slope Plots

**Note:** The un-centered distal intercepts represent the conditional means when the binary covariate is at its first level `SOC_STRS = 0` (i.e., no social stress). Therefore, the conditional mean for `SOC_STRS = 1` (i.e., social stress) can be calculated by adding the associated slope coefficient to the intercept.

Read in the un-centered model & extract relevant parameters

```
model_uncen <- readModels(here("3step_mplus", "Step3_uncentered_BD.out"), quiet = TRUE)

model_uncen <- data.frame(model_uncen$parameters$unstandardized)

slope_data <- model_uncen %>%
  filter(str_detect(paramHeader, 'ON|Inter')) %>%
  unite("param", paramHeader:param, remove = TRUE) %>%
  mutate(param = str_replace(param, "MOOD1.ON_SOC_STRS", "_COEF")) %>%
  mutate(param = str_remove_all(param, "Intercepts_|MOOD1")) %>%
  mutate(LatentClass = factor(LatentClass,
    labels = c("Adaptive Coping (63.6%)", "Externalizing Behavior (5.3%)", "No Coping (31.1%)")))
```



---

## Positive mood simple slope graph

---

Prepare data-frame for plotting

```
pos_data <- slope_data %>%
  filter(str_detect(param, 'POS'))

pos_wide <- pos_data %>%
  select(param, est, LatentClass) %>%
  pivot_wider(names_from = param, values_from = est) %>%
  rename("No.Social.Stress" = 'POS') %>%
  mutate(Social.Stress = No.Social.Stress + POS_COEF) %>% # calc. condit. means `SOC_STRS = 1`
  select(-POS_COEF)

plot_pos <- melt(pos_wide, id.vars = "LatentClass") %>%
  mutate(variable = factor(variable,
    levels = c("No.Social.Stress", "Social.Stress"),
    labels = c("No Social Stress", "Social Stress")))
```

Plot positive mood simple slope graph

```
p_plot <- ggplot(plot_pos,
  aes(y=value, x=variable,
    color=LatentClass,
    group=LatentClass,
    shape=LatentClass,
    lty=LatentClass)) +
  geom_point(size = 4) + geom_line() +
  xlab("") + ylab("Positive Mood") + ylim(2.5, 3.5) +
  scale_colour_grey() +
  theme_classic() +
  theme(text=element_text(family="Times New Roman", size=12),
    axis.text.x=element_text(size=12),
    legend.text = element_text(family="Times New Roman", size=10),
    legend.position = "top", legend.title = element_blank()) +
  annotate(geom = "text",
    x = 1.8, y = 2.77,
    label = "N.S.", color = "black") +
  annotate(geom = "text",
    x = 1.8, y = 2.60,
    label = "N.S.", color = "black")
```

---

## Negative mood simple slope graph

---

Prepare data-frame for plotting

```
neg_data <- slope_data %>%
  filter(str_detect(param, 'NEG'))

neg_wide <- neg_data %>%
  select(param, est, LatentClass) %>%
  pivot_wider(names_from = param, values_from = est) %>%
  rename("No.Social.Stress" = 'NEG') %>%
  mutate(Social.Stress = No.Social.Stress + NEG_COEF) %>% # calculate means for `SOC_STRS = 1`
  select(-NEG_COEF)

plot_neg <- melt(neg_wide, id.vars = "LatentClass") %>%
  mutate(variable = factor(variable,
    levels = c("No.Social.Stress", "Social.Stress"),
    labels = c("No Social Stress", "Social Stress")))
```

Plot negative mood simple slope graph

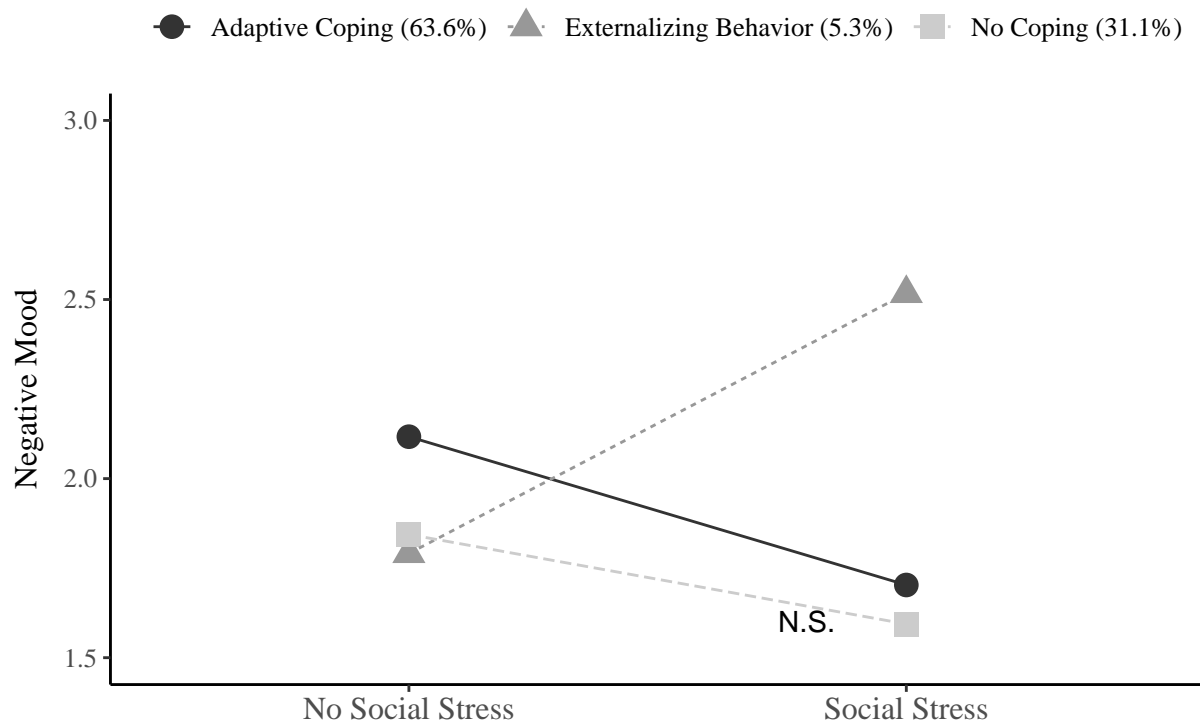
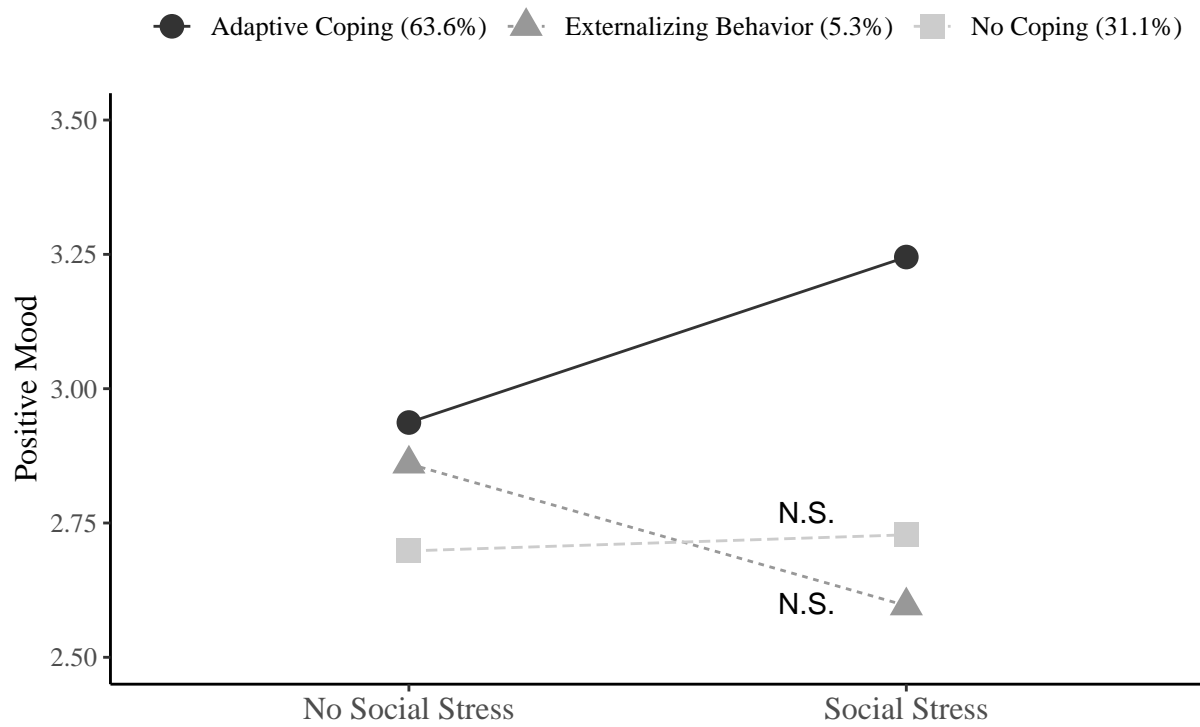
```
n_plot <- ggplot(plot_neg,
  aes(y=value, x=variable,
    color=LatentClass,
    group=LatentClass,
    shape=LatentClass,
    lty=LatentClass)) +
  geom_point(size=4) + geom_line() +
  xlab("") + ylab("Negative Mood") + ylim(1.5,3)+
  scale_colour_grey() + theme_classic() +
  theme(text=element_text(family="Times New Roman", color = "black", size=12),
    axis.text.x=element_text(size=12),
    legend.text = element_text(family="Times New Roman", size=10),
    legend.position = "top", legend.title = element_blank()) +
  annotate(geom = "text",
    x = 1.8, y = 1.6,
    label = "N.S.", color = "black")
```

---

Combine the two simple slopes graphs for distal outcomes positive & negative mood (*Figure 5*)

Note: This code combines 2 ggplot objects using the {patchwork} package.

p\_plot / n\_plot



ggsave(here("figures", "Fig5\_simple\_slopes.png"), dpi=300, height=8.5, width=6.5, units="in")

---

## 2.0 Response Pattern Table

To replicate the response pattern table produced in this study (*Table 3*) see the associated `R/MplusAutomation` code example here:

<https://garberadamc.github.io/project-site/Lab10.1-response-patterns>

Additional examples using `R/MplusAutomation` for a range of analyses can be found here:

<https://garberadamc.github.io/project-site>

---

### References:

- Hallquist, Michael N., and Joshua F. Wiley. 2018. “MplusAutomation: An R Package for Facilitating Large-Scale Latent Variable Analyses in Mplus.” *Structural Equation Modeling*, 1–18. <https://doi.org/10.1080/10705511.2017.1402334>.
- Miller, Jon D. Longitudinal Study of American Youth (LSAY), Seventh Grade Data, 1987-1988; 2015-2016. Ann Arbor, MI: Inter-university Consortium for Political and Social Research [distributor], 2019-04-23. <https://doi.org/10.3886/ICPSR37287.v1>
- Müller, Kirill. 2017. Here: A Simpler Way to Find Your Files. <https://CRAN.R-project.org/package=here>.
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