Metagenomic Binning Pipelines - the State of the Art

1 Abstract

- Decision tree graphical abstract for the choice of binning algorithm
- Features that distinguish binning algorithms
- Some guidelines for choosing the correct binning techniques appropriate for a given study

New generations of sequencing platforms coupled to numerous bioinformatics tools have led to rapid technological progress in metagenomics and metatranscriptomics to investigate complex microorganism communities. Nevertheless, a combination of different bioinformatic tools remains necessary to draw conclusions out of microbiota studies. As sequencing costs have dropped at a rate above 'Moore's law', bigger data sets are available, and proportional costs of analysis have risen as a consequence. Binning is the grouping of assembled metagenomic contigs by their genome of origin. Algorithms for binning are a rapidly evolving field. The number of these algorithms are growing over time. Selecting the most appropriate binning algorithm can be a daunting task and is influenced on computational resources available and experimental variables relating to the sequencing. This review serves as a roadmap to direct the researcher to the binning algorithm that best suits their needs.

2 Background

- General introduction to history of binning
- increase in popularity of the field of metagenomics
- Talk about reduced cost in sequencing and scheduling efficiency in pipelines
- Talk about HMP

Compared to amplicon, shotgun metagenome can provide functional gene profiles directly and reach a much higher resolution of taxonomic annotation. However, due to the high demands on computational resources, cost, and expertise necessary to perform this analysis, this method has been used The substantial decrease in cost of sequencing however has lessened this burden. At the time of writing, shotgun metagenomic sequeng costs on average three times as much as 16S sequencing comparitavely. A review on the benchmarking binning algorithms was done by Yue et al., 2020.

3 Methods for metagenomic binning

- Not sure if to focus on this or the appropriateness
- The increased impact of machine learning in analysis
- Short section just for past-present-future completeness

3.1 Metagenome Assembled Genomes

A Metagenome-Assembled Genome (MAG) is a single-taxon assembly based on one or more binned metagenomes that has been asserted to be a close representation to an actual individual genome (that could match an already existing isolate or represent a novel isolate).

3.2 Metagenomic Species Pan-genomes

Microbial pan-genomes are gene repertoires composed of core genes present in all strains and accessory genes present in only some of them (Medini et al., 2005). In a shotgun metagenomic sequencing context, we define as shared the genes detected in some samples where the species is not present.

A strain found in a sample is an instance of the species pan-genome: it is made of all the species (shared) core genes and of a subset of (shared) accessory genes. Core genes are suitable for taxonomic profiling at species-level while accessory genes can be used to compare strains across samples. Genes tagged as shared should be used carefully as they contain false positives counts or are subject to horizontal transfer.

We assumed that core genes of a microbial species should be consistently detected in samples where the species is present if sequencing depth allows (co-occurrence) and should yield directly proportional mapped reads counts across samples (co-abundance). Remarkably, a core and an accessory gene should have proportional counts only in the subset of samples carrying a strain with that accessory gene

3.3 Megagenome Assembled Genome

3.4 Binning microbial genomes with deep learning

The integration of deep learning techniques into the field of metagenomics has revolutionised the field of metagenomics. The VAMB pipeline was developed to take advantage of variational autoencoders; a generative machine learning model that uses a combination Improved metagenome binning and assembly using deep variational autoencoders Nature biotechnology - 4th Jan 2021 the VAMB pipeline (Nissen et al., n.d.)

4 Chosing the most appropriate binning algorithm

Resource management is an important factor in the choice of binning algorithm. The tradeoff between number of CPU's, memory, and time are important considerations. Pipeline vs standalone? Alignment based or alignment free An analysis pipeline is defined as a program that combines several programs in a defined order to complete a complex analysis. Improperly developed, validated, and/or monitored pipelines may generate inaccurate results.

4.1 Binning for viral genomes

New insights from uncultivated genomes of the global human gut microbiome Nature - 13th March 2019 (Nayfach, Shi, Seshadri, Pollard, & Kyrpides, 2019)

5 Conclusion

- New and open areas of research in which the application of metagenomic pipelines are relevant
- HMP and other
- The increased impact of machine learning in analysis

Table 1: Comparison of binning algorithms

| | | Table 1. Companie | | on or binimis assorbinims | | |
|---|---------------|---|----------------|--|---|-------------------------|
| Software/Algorithm | Year | Description/purpose | Reference free | Comment/Highlight | Software category/ Topic | Input/Output data typ |
| 2 0 0 | 1000 | | 3.5 | | | : |
| CoCoNet | 2021 | Deep learning tool for Viral Metagenome Binning | res | Reconstructs viral genomes | MAG binning | Contigs, coverge (read |
| Binnacle | 2021 | Using scarfolds to improve Metagenomic bin quality | res | Incorporates scanoid information | MAG binning-remner. | Contigs, coverage, bins |
| VAMB | 2021 | Metagenome binning using deep variational autoe | Yes | Autoencoder algorithm, fast processing | MAG binning | Assembled contigs/ bit |
| phyloFlash | 2020 | ssrRNA profiling and MAG assembly | No | incorporates ssrRNA profiling info into MAG ass | SSU rRNA assembly/ connect SSUrRNA to MAG | Reads and reference da |
| MetaBCC-LR | 2020 | Metagenomic binning for Long-Reads | Yes | Suitable for Long Reads sequencing technology | LongReads binning | Long reads/ Bin identi |
| BioBloom Tools | 2020 | Reads binning for targeted assembly, alignment | No | Data preparation for targeted assembly, using s | Reads binning | Reads and reference se |
| 0 | 2020 | Binning unassembled short reads based on k-mer | Yes | Data preparation for targeted assembly, low abu | Reads binning | NaN |
| GranhBin | 2020 | Refined binning of metagenomic contigs using as | Z | Incorporates assembly graphs information | MAG binning-refiner* | Z |
| Month of the country | 000 | remined priming of medagenomic configs using as: | NI-NI | Morporace assembly graphs information | MAC SIMILING TOTAL | NI - NI |
| Metasirsim | 2020 | Simulating metagenomic stable isotope probing d | Nan | Augment binning resolution with extra experimen | Data Simulation/ DINA-SIF | Ivaiv |
| MetaCon | 2019 | Unsupervised binning k-mers and coverage, focus | NaN | Focus different lengths contigs | MAG binning | NaN |
| VirBin | 2019 | Binning viral haplotypes from assembled contigs | NaN | Viral haplotypes MAGs | Haplotype/MAG binning* | NaN |
| MAGO (*only tool pipeline) | 2019 | Framework for Production and analysis of MAGs | NaN | pipeline | MAG binning-refiner pipeline | NaN |
| SeaDex | 2019 | Genome separation of Endosymbionts from mixed s | ZeZ | Identification of endosymbiont | MAG binning* | ZeZ |
| MetaTOB | 2019 | High quality MAGs from mammalian guts using met | Z | Incorporates 3D contact information | MAG binning | Z |
| Mota BAT 9 | 0100 | Adatatative binning algorithm for genome recons | Z | Eliminates manual narameter tuning from pression | MAC binning | NeN |
| Trees of | 0100 | Additionally Diming algorithm for genome recons | NI NI | Eliminates manda parameter tuning nom previou | Mind Diming | NI NI |
| MetaBMF | 5018 | Scalable binning algorithm for large scale meta | NaN | Employs sample X contigs of mapped read counts | MAG binning | NaN |
| PolyCRACKER | 2019 | Method for partitioning polyploid sub genomes b | NaN | Haplotypes for polyploid genomes | Haplotype binning | NaN |
| SolidBin | 2019 | Improving metagenome binning with semi-supervis | NaN | NaN | MAG binning | NaN |
| Autometa | 2019 | extraction of microbial genomes from individual | NaN | Handles eukaryotic contamination | MAG binning / pipeline | NaN |
| MLBP MrGBP (Algorithm) | 2019 | Signal processing method for alignment free met | NaN | Alternative description of sequences designed f | MAG binning* | NaN |
| CLAME | 2018 | Aligment based algorithm allowed description of | No | Aligment based for reads | Reads binning* | NaN |
| 3D BH SNE (Algorithm) | 2018 | Fuzzy binning of metagenomic sequence fragments | Z | Horizontal gene transfer and regions of uncerta | MAG binning /Fuzzy binning | NeX |
| LVO-KNN | 2018 | Composition based RNA or DNA binning of short s | Z | Classify into DNA or RNA securence | Reads binning | NoN |
| MOD | 0 100 | Attendance been been distriction of minutes of some | NaM | Dec account mod Division and demon | Mod Lining | NEW |
| Mor miner | 2010 | Abundance based reconstitution of microbial pan | N'S | ran genome reconstitution | Mar bining | Nain 1 |
| Meta WRAP* | 2018 | Flexible pipeline for genome resolved metagenom | NaN | Hybrid bin extraction algorithm | MAG binning-refiner pipeline | NaN |
| MetaVW | 2018 | Large scale Machine Learning Sequence classific | NaN | Machine learning for reads based on Khmer profile | Reads binning | NaN |
| Opal (algorithm*) | 2018 | Metagenomic binning through low density binning | NaN | Improvement at higher taxonomic levels, discove | MAG binning | NaN |
| BMC3C | 2018 | Binning contigs using codon usage sequence comp | NaN | Add codon usage information | MAG binning | NaN |
| AMBER tool | 2018 | Assessment of Metagenome Binners | NaN | NaN | Benchmark tool for binning software | NaN |
| DAS Tool | 2018 | Derreplication aggregation and scoring strategy | NaN | Combines several binning algorithm results | MAG binning-refiner | NaN |
| MEGAN-LR | 2018 | Long Read/ contigs taxonomic binning | No | Aligment of long reads against reference sequences | NaN | NaN |
| CoMet | 2018 | Binning workflow using contain coverage and com | NaN | Single sample, include gc content and 4mer fre | MAG binning | NaN |
| ė. | 2017 | Metagenomic binning and association of plasmids | NaN | Plasmid banning at strain level using methylati | MAG binning | NaN |
| MetaGen | 2017 | reference-free learning with multiple metagenom | NaN | Requires multiple samples | MAG binning | NaN |
| d2sBin add onn | 2017 | Improved formula for calculate oligonucleotide | ZaZ | Math formula to calculate oligo sequence dissim | NA, only formula for dissimilarity | ZaZ |
| BusyBee Web | 2017 | Bootstrapped supervises binning and annotation | NaN | 2d interactive scatterplots supervised binning | refiner | ZeZ |
| ICoVer | 2017 | Interactive visualisation tool for verification | NaN | Interactive visualisation tool | MAG refiner | NaN |
| HirBin* | 2017 | High resolution identification of differentiall | NaN | Supervised annotation, unsupervised clustering | Reads binning | NaN |
| BinSanity | 2017 | Unsupervised clustering using coverage and affi | ZeZ | Reduce bias for high/low abundance | MAG binning | NaN |
| Binning refinner | 2017 | Improve genome bins through the combination of | Z | Combination of different binning algorithms | MAG hinning-refiner | NeN |
| IFCM add on | 2016 | Improved binning using Fuzzy C-Means Method | Z | Add estimated distribution of real genome lengths | MAG binning | Z |
| A TOO A COO | 2016 | hinning conting using composition need covered | Z | Adde naired and read and coaligment information | MAG binning | Paired and contine |
| COMOCON Company (9) | 2010 | The few enterestic measure of meaning manner | New | Adda differential commons to complement common | MAG binning | No.N |
| Groopini (2) | \$10 7 | tool for automatic recovery of population genom | NEW | Adds differential coverage to complement compos | MAG DIMING | NTRNI |

- Short section just for past-present-future completeness
- Future developments for metagenomic analysis

References

- Nayfach, S., Shi, Z. J., Seshadri, R., Pollard, K. S., & Kyrpides, N. C. (2019). New insights from uncultivated genomes of the global human gut microbiome. *Nature*, 568 (7753), 505–510.
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