1	Advances	in	Metagenomic	Binning	for	the	Reconstruction	of
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Microbial Species

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$_{\scriptscriptstyle ext{\tiny 1}}$ Glossary

- 22 CPU Central Processing Unit.
- 23 HMP Human Microbiome Project.
- ²⁴ MAG Metagenome-Assembled Genome.
- ²⁵ MSP Metagenomic Species Pan-genomes.
- VAE Variational Auto Encoders.

1 Abstract

- 28 New generations of sequencing platforms coupled with numerous bioinformatics tools have led to
- 29 rapid technological progress in metagenomics to investigate complex microorganism communities.
- 30 Nevertheless, a combination of different bioinformatic tools remains necessary to draw conclusions
- out of microbiota studies. As sequencing costs have dropped at a rate above 'Moore's law', a
- 32 greater number of large data sets are being produced than ever before. Newer algorithms that
- take advantage of the size of these datasets are continually being developed. Binning algorithms
- 34 are defined as the grouping of assembled metagenomic contigs by their genome of origin (Figure 1).
- 35 Selecting the most appropriate binning algorithm can be a daunting task and is influenced by many
- ₃₆ factors. This review details the current advances in the field of metagenomic binning.

2 Background

- The explosion in popularity and success in the field of metagenomics over the last 25 years can be
- ³⁹ largely attributed to the advances in computing technologies. An example of the outcomes of this
- 40 can be found in the Human Microbiome Project (HMP); a project that has been greatly improved
- 41 the understanding of the microbiome flora involved in human health and disease (Turnbaugh
- 42 et al., 2007). These advances have brought with them greater demands for storage, Central
- 43 Processing Unit (CPU) time, and consequently more efficient algorithms. The main function of
- binning tools is to reconstruct species/biological entities from metagenomic samples. Compared
- to amplicon, shotgun metagenome can provide functional gene profiles directly and reach a much
- 46 higher resolution of taxonomic annotation. However, due to the high demands on computational

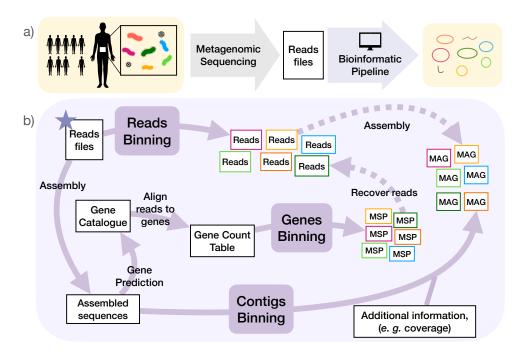


Figure 1: Overview of the existing binning strategies. a) Microbiome samples are collected and sequenced, the resulting sequences are processed, employing the existing binning strategies, which try to reconstruct the genome sequences from the original organisms. b) The scheme present a simplified workflow of the existing binning strategies, named contig-binning, reads-binning and genes-binning, represented as rounded rectangles in the middle of arrows. The star in the upper left indicates the entry point, simple arrows represent intermediate processing steps, black edge squares represent intermediate files, the color squares represent endpoint files. The binning strategies are not independent and can complement each other as shown by the dotted arrows.

resources, cost, and expertise necessary to perform this analysis, researchers have historically been limited in their capacity to collect and analyse sequencing data. As the cost of sequencing is rapidly falling, this burden has been significantly lessened. Accurate, robust, and suitable binning methods are crucial for the proper interpretation of any metagenomic dataset. Here we will briefly recapitulate recent binning algorithms and highlight some of the developments in the field, among them, the use of new algorithms and strategies employed to achieve the goal of identifying the organisms composing microbiome communities.

₅₄ 3 Overview of recent methods for metagenomic binning

55 3.1 Progress in recent binning strategies

A metagenomic sample is comprised of many organisms and the goal of binning is to reconstruct the sequences from each organism present in the original sample. The majority of binning tools are oriented toward clustering contigs (contig-binning) into bins, which may represent the genome from a single biological entity/organism. A Metagenome-Assembled Genome (MAG) is a singletaxon assembly based on one or more binned metagenomes that has been asserted to be a close representation to an individual genome that could match an already existing isolate or represent a novel isolate. Current contig-binning tools are commonly reference free (i.e. they do not depend on reference sequences to perform clustering) and rely on coverage information and sequence composition. Progress in contig-binning algorithms can be seen in the proposals to integrate new sources of information (for example, from scaffold-graphs (Binnacle), paired-end reads (CO-CACOLA), or 3D contact information (MetaTOR)) and state of the art algorithms in machine learning (CoCoNet, VAMB). We also notice the development of Bin refinement tools (DAS-tool, Binning Refiner) that rely on the outputs from multiple contig-binning algorithms and combine them to produce better results (Sieber et al., 2018). Binning of contigs have played a central role in software development in the field, a review on the benchmarking of binning algorithms was done by Yue et al., 2020. Beside contig-binning tools we can also distinguish read-binning tools and 71 co-abundant-gene-binning tools. The main purpose of read-binning tools is to pre-process reads into clusters for a posterior targeted assembly. Here we find reference-free and non-referencefree tools, and tools designed for short-read or long-read sequencing technologies. Among the binning tools developed in recent years, a subset of them are dedicated to cluster reads (readbinning) (MetaBBC-LR, BioBloom Tools, CLAME, LVQ-KNN, Meta VW, HirBin, MEGAN-LR)

(Wickramarachchi, Mallawaarachchi, Rajan, & Lin, 2020; Chu et al., 2014; Benavides, Isaza,
 Niño-García, Alzate, & Cabarcas, 2018; Belka, Fischer, Pohlmann, Beer, & Höper, 2018; Vervier,
 Mahé, Tournoud, Veyrieras, & Vert, 2016; Österlund, Jonsson, & Kristiansson, 2017; Huson et al., 2018).

81 3.1.1 Binning co-abundant genes

Binning of co-abundant genes represents an alternative proposal to reconstruct species/biological entities from a set of metagenomic samples. Co-abundant gene binning methods assumes that each gene coming from a shared chromosome will display proportional abundances across samples. Therefore, if there are enough samples from a similar environment you can identify the sets of genes from a common organism of origin (MLGs Chameleon-clust 2012, CAGs and MGSs Canopy 2014, Markovclust-MGCs Karlsson 2013, Metagenomic Species Pan-genomess (MSPs) MSPminner 2018) (Karypis, Han, & Kumar, 1999; Plaza Oñate et al., 2019). The MSPminer software was developed to exploit this approach. MSPminer introduced a robust proportionality measure to detect co-abundance but no necessarily co-occurrance. This tool groups co-abundant genes into MSPs and classifies genes within an MSP as core, accessory and shared. Core genes are present in all strains, accessory are present only in some, and the shared category applies for those genes which may be present in more than one MSP due to horizontal transfer (Tettelin et al., 2005). Factors that impact directly on MSP quality include sample composition, sequencing depth, and previous bioinforamtic steps to build the reference gene dataset and map the reads. MSPs can be used for taxonomic profiling of new samples from similar ecosystems at the species level, and also to compare strains between samples by building a presence/absence table of accessory genes and for biomarker discovery. By binning contigs carrying genes from the same MSP it is also possible to build a MAG.

3.1.2 Binning microbial genomes with deep learning

The integration of deep learning techniques has revolutionised the field of metagenomics. Deep learning approaches have benefitted from the rapid acceleration in GPU efficiency over the past few years. The Software VAMB and CoCoNet constitute two such examples that employ deep learning for binning (Nissen et al., n.d.; Arisdakessian, Nigro, Steward, Poisson, & Belcaid, 2021).

The main novelty of VAMB is the application of the Deep Learning technique known as Variational Auto Encoders (VAE). In this case, VAEs learn how to integrate two data types, coabun-

dance and k-mer composition. The resulting latent representation is able to cluster better than either of the inputs alone. In principle this technology is not limited by only two input data types. VAMB also applies a "mulitsplit" approach whereby each cluster should correspond to an organism representation across samples and each bin in a cluster to a per-sample representation of the genome of that organism.

The CoCoNet software uses deep learning and clustering to bin contigs into clusters representing species present in the samples. The algorithm consists of two phases. During the first phase, a neural network is trained to estimate the probability that two contigs arise from the same genome given their composition and coverage information. The second use a heuristic to bin the contigs using the probabilities inferred in the first stage An interesting feature in CoCoNet is it was trained on viral genomes. In the following section we discuss more about binning on viral genomes.

3.2 Binning of viral genomes

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Most binning algorithms are designed for prokariotic organisms leaving viruses out of the soft-120 ware scope. In recent years the virome and its importance in health and disease has recognised. CoCoNet uses deep leaning to model co-ocurrence of contigs from the same viral genome. The 122 network was optimized for diverse viral metagenomes, the network learns to model coverage variability within samples, a critical feature in viral metagenomes where DNA amplification methods 124 are needed to increase input genetic material. VirBin clusters contigs for genome reconstruction of 125 viral strains, different strains within viral species may show different biological properties such as transmissibility or virulence. Composition based features are usually are not enough to separate 127 haplotypes, VirBin receives contigs as inputs and outputs the estimated number of haplotypes via contig alignment and returns the contigs for each haplotype based on relative abundance distribu-129 tion, when the contigs are long enough VirBin produce better results. Newer strategies has been proposed and employed to reconstruct viral genomes from metagenomic samples, in a recent work 131 (Nayfach et al., 2021) a new compendium of 189680 DNA viruses from the human gut microbiome 132 was produced. In this work they use viral informative features including presence of viral protein 133 families (Paez-Espino et al., 2016), absence of non-viral families (El-Gebali et al., 2019), gene 134 strand switch rate (Roux, Beloin, & Ghigo, 2005), and the score produced from the VirFinder (Ren, Ahlgren, Lu, Fuhrman, & Sun, 2017) software.

3.3 Binning Pipelines

Other advances in binning can be found in the integration of existing tools and software into 138 bioinformatic pipelines. These innovations allow the automatic complete processing of read samples into bins or the addition of extra processing steps to address specific biological questions or 140 problems related to the sample of origin. MetaWRAP is a modular pipeline ready to perform common tasks in metagenomic analysis, starting from read quality checks up to bin creation, 142 refinement, reassembly quantification, taxonomic annotation and functional annotation. MAGO 143 pipeline integrates metagenome assembly, binning, bin improvement, bin quality check, bin func-144 tional annotation, and bin taxonomic annotation. SqueezeMeta also integrates external software 145 to perform the complete analysis of metagenomic samples from sequences reading to MAG construction and annotation (Tamames & Puente-Sánchez, 2019) nf-mag supports both short and 147 long reads, performs quality and adapter trimming, quality check, performs assembly, binning, checks bin quality and assigns taxonomy (Ewels et al., 2020). Autometa was developed to deal 149 with non-model Eukariotic host contamination and complex single metagenomes, the application integrate sequence homology, nucleotide composition, coverage and single-copy marker genes to 151 separate microbial genomes from non model host genomes (Miller et al., 2019). Sequex is a tool 152 written in R which separates endosymbionts from their host sequences (Chiodi et al., 2019). Their 153 approach uses specific features in endosymbiotic systems to better solve this problem. This tool 154 combines partial taxonomic annotations obtained trough homology searches and sequence compo-155 sition to predict the contig's organism of origin from host and its endosymbionts and helps the user 156 to understand how effective is the classification. Reproducibility, scalability, and ease of use from 157 people with little computational experience are attractive features that pipelines for metagenomic 158 analysis provide.

4 Challenges in appropriate binning algorithm selection

A number of aspects should be considered when performing binning analysis on metagenomic samples. A list of binning algorithms, including some that we have not mentioned above, are listed
here Table 1. Computational resources available, sequencing technology, number of samples, and
the sample's source are important factors to consider. Some tools employ more resources than
others, and some perform better under specific circumstances (as reviewed by Yue et al., 2020).

If you are dealing with a large number of samples, a gene-binning strategy could be taken into

Table 1: Comparison of popular binning algorithms updated since 2017.

		T T T	1		
Software/Algorithm	Year	Description/purpose	Comment/Highlight	Doi	PubmedID
+oNoDoD	9091	Deer learning tool for Vinal Metagenome Binning	Reconstructe vival genomes	10 1003 /bioinformatics /btab913	33899801
Dinnell	2021	Tiene coeffolds to impress Meteoromic him and	Incompanted coeffeld information	10 9980 /fmich 9091 698561	99717099
Dinnacie	2021	Using scariolds to improve inclagenomic bin qua	incorporates scanoid information	10.3009/IMICD.2021.030301	0000011000
VAMB	707	Metagenome binning using deep variational auto	Autoencoder algorithm, fast processing	10.1038/841387-020-00777-4	33398103
phyloFlash	2020	ssrRNA profiling and MAG assembly	incorporates ssrRNA profiling info into MAG as	10.1128/mSystems.00920-20	33109753
MetaBCC-LR	2020	Metagenomic binning for Long-Reads	Suitable for Long Reads sequencing technology	10.1093/bioinformatics/btaa441	32657364
BioBloom Tools	2020	Reads binning for targeted assembly, alignment	Data preparation for targeted assembly, using	10.1073/pnas.1903436117	32641514
GraphBin	2020	Refined binning of metagenomic contigs using a	Incorporates assembly graphs information	10.1093/bioinformatics/btaa180	32167528
MetaSIPSim	2020	Simulating metagenomic stable isotope probing	Augment binning resolution with extra experime	10.1186/s12859-020-3372-6	32000676
MetaCon	2019	Unsupervised binning k-mers and coverage, focu	Focus different lengths contigs	10.1186/s12859-019-2904-4	31757198
VirBin	2019	Binning viral haplotypes from assembled contigs	Viral haplotypes MAGs	10.1186/s12859-019-3138-1	31684876
MAGO (*only tool pipeline)	2019	Framework for Production and analysis of MAGs	pipeline	10.1093/molbev/msz237	31633780
SeqDex	2019	Genome separation of Endosymbionts from mixed	Identification of endosymbiont	10.3389/fgene.2019.00853	31608107
MetaTOR	2019	High quality MAGs from mammalian guts using me	Incorporates 3D contact information	10.3389/fgene.2019.00753	31481973
MetaBAT (v2)	2019	Adatptative binning algorithm for genome recon	Eliminates manual parameter tuning from previo	10.7717/peerj.7359	31388474
MetaBMF	2019	Scalable binning algorithm for large scale met	Employs sample X contigs of mapped read counts	10.1093/bioinformatics/btz577	31347687
PolyCRACKER	2019	Method for partitioning polyploid sub genomes	Haplotypes for polyploid genomes	10.1186/s12864-019-5828-5	31299888
SolidBin	2019	Improving metagenome binning with semi-supervi	NaN	10.1093/bioinformatics/btz253	30977806
Autometa	2019	extraction of microbial genomes from individua	Handles eukaryotic contamination	10.1093/nar/gkz148	30838416
MLBP MrGBP (Algorithm)	2019	Signal processing method for alignment free me	Alternative description of sequences designed	10.1038/s41598-018-38197-9	30770850
CLAME	2018	Aligment based algorithm allowed description o	Aligment based for reads	10.1186/s12864-018-5191-y	30537931
3D BH SNE (Algorithm)	2018	Fuzzy binning of metagenomic sequence fragments	Horizontal gene transfer and regions of uncert	10.1109/EMBC.2018.8512529	30440633
LVQ-KNN	2018	Composition based RNA or DNA binning of short	Classify into DNA or RNA sequence	10.1016/j.virusres.2018.10.002	30291874
MSPminer	2018	Abundance based reconstitution of microbial pa	Pan genome reconstitution	10.1093/bioinformatics/bty830	30252023
MetaWRAP*	2018	Flexible pipeline for genome resolved metageno	Hybrid bin extraction algorithm	10.1186/s40168-018-0541-1	30219103
MetaVW	2018	Large scale Machine Learning Sequence classifi	Machine learning for reads based on Khmer profile	10.1007/978-1-4939-8561-6_2	30030800
Opal (algorithm*)	2018	Metagenomic binning through low density binning	Improvement at higher taxonomic levels, discov	10.1093/bioinformatics/bty611	30010790
BMC3C	2018	Binning contigs using codon usage sequence com	Add codon usage information	10.1093/bioinformatics/bty519	29947757
AMBER tool	2018	Assessment of Metagenome Binners	NaN	10.1093/gigascience/giy069	29893851
DAS Tool	2018	Derreplication aggregation and scoring strategy	Combines several binning algorithm results	10.1038/s41564-018-0171-1	29807988
MEGAN-LR	2018	Long Read/ contigs taxonomic binning	Aligment of long reads against reference seque	10.1186/s13062-018-0208-7	29678199
CoMet	2018	Binning workflow using contain coverage and co	Single sample, include gc content and 4mer fr	10.1186/s12859-017-1967-3	29297295
MetaGen	2017	reference-free learning with multiple metageno	Requires multiple samples	10.1186/s13059-017-1323-y	28974263
d2sBin add onn	2017	Improved formula for calculate oligonucleotide	Math formula to calculate oligo sequence dissi	10.1186/s12859-017-1835-1	28931373
BusyBee Web	2017	Bootstrapped supervises binning and annotation	2d interactive scatterplots supervised binning	10.1093/nar/gkx348	28472498
ICoVer	2017	Interactive visualisation tool for verificatio	Interactive visualisation tool	10.1186/s12859-017-1653-5	28464793
HirBin*	2017	High resolution identification of differential	Supervised annotation, unsupervised clustering	10.1186/s12864-017-3686-6	28431529
BinSanity	2017	Unsupervised clustering using coverage and aff	Reduce bias for high/low abundance	10.7717/peerj.3035	28289564
Binning_refinner	2017	Improve genome bins through the combination of	Combination of different binning algorithms	10.1093/bioinformatics/btx086	28186226
IFCM add on	2016	Improved binning using Fuzzy C-Means Method	Add estimated distribution of real genome lengths	10.1109/TCBB.2016.2576452	27295684
COCACOLA	2016	binning contigs using composition, read covera	Adds paired end read and coaligment information	10.1093/bioinformatics/btw290	27256312
GroopM (v2)	2014	Tool for automatic recovery of population geno	Adds differential coverage to complement compo	10.7717/peerj.603	25289188

consideration. Tools such as CoMet were built around single sample binning (Herath, Tang, Tan-167 don, Ackland, & Halgamuge, 2017). Long read sequence technology is gaining momentum and some tools also integrate the characteristic features generated with this technology. The environ-169 ment under study also play an important role for binning. Sometimes there exists host organisms whose genome sequences would be removed before starting the analysis. The environment also 171 has a profound effect on the sample's diversity with samples that have greater diversity requiring 172 greater sequencing depth making binning more difficult. It is also difficult for binning tools to 173 discern between similar strains within the same sample. It is also worth mentioning that there 174 is no mutual exclusivity between the currently available tools and it is possible to benefit from 175 the relative advantages each has to offer and merge the results depending of the aim of the study. 176 Besides binning, other types of metagenomic analysis can be performed on microbiomes. Recent reviews provide an overview of the complete process and practical guides to apply available 178 software (Breitwieser, Lu, & Salzberg, 2019).

5 Conclusion

Popularity and successes of metagenomic binning have accelerated in the last ten years. Current 181 limitations that still remain include the difficulty in classifying similar strains within samples. They additionally do not perform well assigning 16S sequences to bins likely due to the high copy 183 number of these sequences within a genome. As binning has been focused mainly in prokariotic organisms, binning of organisms outside prokariotes need more development. Although there 185 have been significance advances in the characterisation of viral genomes as of late (Nayfach et al., 2021), the huge diversity in viral genomes still poses a challenge for current methodologies. The 187 continuously increasing number of sequences available require more efficient/faster algorithms and 188 new strategies to reconstruct single organisms from environmental samples. However, with the breakneck pace of technological advancements in computing resources, this requirement is sure to be met and will pave the way for greater insights into the microbial world. With the integration 191 of Machine learning algorithms into binning, we expect to see significant developments in the near 192 future. 193

94 References

Arisdakessian, C. G., Nigro, O. D., Steward, G. F., Poisson, G., & Belcaid, M. (2021). Coconet:

- an efficient deep learning tool for viral metagenome binning. *Bioinformatics*.
- Belka, A., Fischer, M., Pohlmann, A., Beer, M., & Höper, D. (2018). Lvq-knn: Composition-based dna/rna binning of short nucleotide sequences utilizing a prototype-based k-nearest neighbor approach. Virus research, 258, 55–63.
- Benavides, A., Isaza, J. P., Niño-García, J. P., Alzate, J. F., & Cabarcas, F. (2018). Clame: a new alignment-based binning algorithm allows the genomic description of a novel xanthomonadaceae from the colombian andes. *BMC genomics*, 19(8), 9–30.
- Breitwieser, F. P., Lu, J., & Salzberg, S. L. (2019). A review of methods and databases for metagenomic classification and assembly. *Briefings in bioinformatics*, 20(4), 1125–1136.
- Chiodi, A., Comandatore, F., Sassera, D., Petroni, G., Bandi, C., & Brilli, M. (2019). Seqdeχ: a
 sequence deconvolution tool for genome separation of endosymbionts from mixed sequencing
 samples. Frontiers in genetics, 10, 853.
- Chu, J., Sadeghi, S., Raymond, A., Jackman, S. D., Nip, K. M., Mar, R., ... Birol, I. (2014).

 Biobloom tools: fast, accurate and memory-efficient host species sequence screening using

 bloom filters. *Bioinformatics*, 30 (23), 3402–3404.
- El-Gebali, S., Mistry, J., Bateman, A., Eddy, S. R., Luciani, A., Potter, S. C., ... others (2019).

 The pfam protein families database in 2019. *Nucleic acids research*, 47(D1), D427–D432.
- Ewels, P. A., Peltzer, A., Fillinger, S., Patel, H., Alneberg, J., Wilm, A., ... Nahnsen, S. (2020).
- The nf-core framework for community-curated bioinformatics pipelines. Nature biotechnol- ogy, 38(3), 276-278.
- Herath, D., Tang, S.-L., Tandon, K., Ackland, D., & Halgamuge, S. K. (2017). Comet: a workflow using contig coverage and composition for binning a metagenomic sample with high precision.

 BMC bioinformatics, 18(16), 161–172.
- Huson, D. H., Albrecht, B., Bağcı, C., Bessarab, I., Gorska, A., Jolic, D., & Williams, R. B. (2018). Megan-lr: new algorithms allow accurate binning and easy interactive exploration of metagenomic long reads and contigs. *Biology direct*, 13(1), 1–17.
- Karypis, G., Han, E.-H., & Kumar, V. (1999). Chameleon: Hierarchical clustering using dynamic modeling. *Computer*, 32(8), 68–75.
- Miller, I. J., Rees, E. R., Ross, J., Miller, I., Baxa, J., Lopera, J., . . . Kwan, J. C. (2019). Autometa: automated extraction of microbial genomes from individual shotgun metagenomes.

 Nucleic acids research, 47(10), e57–e57.
- Nayfach, S., Páez-Espino, D., Call, L., Low, S. J., Sberro, H., Ivanova, N. N., ... others (2021).

- Metagenomic compendium of 189,680 dna viruses from the human gut microbiome. *Nature*Microbiology, 1–11.
- Nissen, J. N., Johansen, J., Allesøe, R. L., Sønderby, C. K., Armenteros, J. J. A., Grønbech,
 C. H., ... others (n.d.). Improved metagenome binning and assembly using deep variational
 autoencoders. *Nature Biotechnology*, 1–6.
- Osterlund, T., Jonsson, V., & Kristiansson, E. (2017). Hirbin: high-resolution identification of differentially abundant functions in metagenomes. *BMC genomics*, 18(1), 1–11.
- Paez-Espino, D., Eloe-Fadrosh, E. A., Pavlopoulos, G. A., Thomas, A. D., Huntemann, M.,
 Mikhailova, N., ... Kyrpides, N. C. (2016). Uncovering earth's virome. Nature, 536 (7617),
 425–430.
- Plaza Oñate, F., Le Chatelier, E., Almeida, M., Cervino, A. C., Gauthier, F., Magoulès, F., ...

 Pichaud, M. (2019). Mspminer: abundance-based reconstitution of microbial pan-genomes
 from shotgun metagenomic data. *Bioinformatics*, 35(9), 1544–1552.
- Ren, J., Ahlgren, N. A., Lu, Y. Y., Fuhrman, J. A., & Sun, F. (2017). Virfinder: a novel k-mer based tool for identifying viral sequences from assembled metagenomic data. *Microbiome*, 5(1), 1–20.
- Roux, A., Beloin, C., & Ghigo, J.-M. (2005). Combined inactivation and expression strategy
 to study gene function under physiological conditions: application to identification of new
 escherichia coli adhesins. *Journal of bacteriology*, 187(3), 1001–1013.
- Sieber, C. M., Probst, A. J., Sharrar, A., Thomas, B. C., Hess, M., Tringe, S. G., & Banfield,

 J. F. (2018). Recovery of genomes from metagenomes via a dereplication, aggregation and

 scoring strategy. *Nature microbiology*, 3(7), 836–843.
- Tamames, J., & Puente-Sánchez, F. (2019). Squeezemeta, a highly portable, fully automatic metagenomic analysis pipeline. Frontiers in microbiology, 9, 3349.
- Tettelin, H., Masignani, V., Cieslewicz, M. J., Donati, C., Medini, D., Ward, N. L., . . . others

 (2005). Genome analysis of multiple pathogenic isolates of streptococcus agalactiae: implications for the microbial "pan-genome". *Proceedings of the National Academy of Sciences*,

 102(39), 13950–13955.
- Turnbaugh, P. J., Ley, R. E., Hamady, M., Fraser-Liggett, C. M., Knight, R., & Gordon, J. I.
 (2007). The human microbiome project. *Nature*, 449 (7164), 804–810.
- Vervier, K., Mahé, P., Tournoud, M., Veyrieras, J.-B., & Vert, J.-P. (2016). Large-scale machine learning for metagenomics sequence classification. *Bioinformatics*, 32(7), 1023–1032.

- Wickramarachchi, A., Mallawaarachchi, V., Rajan, V., & Lin, Y. (2020). Metabcc-lr: meta
 genomics b inning by c overage and c omposition for l ong r eads. *Bioinformatics*,
 36 (Supplement_1), i3-i11.

 Yue, Y., Huang, H., Qi, Z., Dou, H.-M., Liu, X.-Y., Han, T.-F., ..., Tu, J. (2020). Evaluating
- Yue, Y., Huang, H., Qi, Z., Dou, H.-M., Liu, X.-Y., Han, T.-F., ... Tu, J. (2020). Evaluating metagenomics tools for genome binning with real metagenomic datasets and cami datasets.

 BMC bioinformatics, 21(1), 1–15.

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