



# Animal models of SARS-CoV-2 transmission

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SARS-CoV-2 emerged in China as a zoonotic virus in December 2019. The virus proved to be human-to-human transmissible and its global spread resulted in the ongoing COVID-19 pandemic, associated with high morbidity and mortality. Vaccines were developed at an unprecedented speed and proved to be efficacious in preventing disease, but it remains to be determined if vaccines are able to interrupt transmission. Moreover, virus variants of concern continue to emerge that appear more transmissible and/or less sensitive to virus-specific immune responses. Here, we briefly review the role of animal models in assessing prophylactic and therapeutic options to interrupt SARS-CoV-2 transmission.

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Current Opinion in Virology 2021, 50:8–16

This review comes from a themed issue on **Anti-viral strategies**

Edited by **Richard K Plemper**

For complete overview about the section, refer “[Engineering for viral resistance](#)”

Available online 29th June 2021

<https://doi.org/10.1016/j.coviro.2021.06.007>

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## Introduction

In December 2019 a cluster of patients with severe respiratory tract disease was notified in Wuhan, China. A novel coronavirus was rapidly identified as the causative agent and due to the close relationship to severe acute respiratory syndrome (SARS)-coronavirus the virus was named SARS-CoV-2 [1]. Efficient transmission, even by pre-symptomatic and asymptomatic individuals, combined with domestic and international travel, resulted in a pandemic of coronavirus disease-2019 (COVID-19). Phylogenetic comparison with previously identified coronaviruses suggested a zoonotic origin of SARS-CoV-2 [2–4]. A wide diversity of closely related betacoronaviruses was detected in bats and pangolins across Asia [5,6,7<sup>••</sup>], but the exact source species and potential intermediate host remain elusive.

## Human-to-human transmission

Since the majority of COVID-19 outbreaks occurred in household settings or events involving clusters of people in close contact, it is thought that SARS-CoV-2 is primarily transmitted via direct, indirect or close contact with infected individuals through contaminated secretions like saliva, respiratory droplets and aerosols. These droplets and aerosols can be expelled by coughing, sneezing, talking or singing and infect the subsequent host by reaching the respiratory tract or eyes [8–14]. Evidence for long-distance airborne transmission between humans is limited [15]. Despite consistent evidence that SARS-CoV-2 can survive on specific surfaces [16–18,19<sup>•</sup>], evidence of fomite transmission between humans is also limited. In addition to respiratory transmission, viable SARS-CoV-2 has been demonstrated in urine [20] and stool specimens [21,22] obtained from infected humans, but transmission via the fecal-oral route is thought to be of limited relevance in the spread of the virus among humans.

SARS-CoV-2 RNA can be detected by RT-PCR on nasopharyngeal swab material 1–3 days before symptom onset, with peak values around symptom onset, followed by a gradual decline over time [23–27]. Persistence of RNA for several weeks has been reported, especially in individuals with severe COVID-19 [23,24,28,29]. However, detection of infectious virus is a better measure for transmissibility [30<sup>••</sup>]. Shedding of infectious virus was initially not thought to occur for more than 8 days after symptom onset [24,31,32], but a recent study showed longer shedding of infectious virus in a selection of patients, which positively correlated with disease severity and viral load. In that study, infectious virus shedding became undetectable with the appearance of neutralizing antibodies in serum [30<sup>••</sup>]. Comparative studies have shown that the relationship between detection of viral RNA and infectiousness differs between persons with mild and severe disease [33]. The level of shedding of viral RNA differs greatly among individuals, in part explaining the observed highly skewed patterns of transmission towards a limited proportion of human-to-human contacts [34].

Transmission by individuals without symptoms was already suspected early in the pandemic. Two transmission ‘types’ can be distinguished: transmission by asymptomatic (infected people who never develop symptoms) or by pre-symptomatic (infected people who have not yet developed symptoms) individuals. The true extent of

asymptomatic infections is still unclear, but a recent systematic review and meta-analysis (that included several studies with limitations) estimated the proportion of truly asymptomatic cases to be 1 in 6 infections [35]. A study performed in close contacts of confirmed COVID-19 cases estimated the proportion to be 23% [36]. Both studies demonstrated transmission by asymptomatic individuals, albeit it to a lower extent compared to transmission by symptomatic patients. Pre-symptomatic transmission has often been demonstrated, and is in line with isolation of infectious virus as discussed above. Although frequently demonstrated, the estimated rates of pre-symptomatic transmission vary considerably: from 6.4% up to 44% [25,37,38]. Clearly such parameters are not fixed and may differ for emerging variants with increased transmissibility and/or disease severity.

### Emergence of other zoonotic coronaviruses

Two other zoonotic coronaviruses recently emerged to cause outbreaks in humans. In 2003 SARS-coronavirus emerged from bats [39,40], most likely via infection of palm civets as intermediate host [41]. SARS-CoV caused a large outbreak that was initiated by several superspreading events and subsequently amplified in hospitals [42]. The outbreak was associated with high case-fatality rates. However, in contrast to SARS-CoV-2, SARS-CoV proved to be only moderately transmissible among humans. This was probably due to the fact that viral excretion peaked relatively late, around 10 days post symptoms onset [43–45], leading to a predominant occurrence of transmission in the second week of illness. At this stage infectious patients were often hospitalized and most cases of SARS-CoV human-to-human transmission occurred in healthcare settings, potentially due to aerosol-generating procedures. Transmission by pre- or asymptomatic individuals (as described above for SARS-CoV-2) proved to be limited [46]. This strongly facilitated case-based surveillance, contact tracing and isolation measures. Although SARS-CoV was spread to multiple countries, the outbreak was fully contained by non-pharmaceutical interventions [47,48].

In 2012 another zoonotic coronavirus was identified as the causative agent of a cluster of severe respiratory tract disease patients in the Middle East and was named Middle East Respiratory Syndrome (MERS)-coronavirus [49]. Dromedary camels were identified as intermediate host [50,51] and it is thought that the virus likely originated from bats (similar to SARS-CoV and SARS-CoV-2) [49,52]. Although MERS-CoV rapidly spread globally [53], it is generally regarded less human-to-human transmissible than SARS-CoV or SARS-CoV-2, with an early estimate of an  $R_0$  below 1 in the general community and in households [54\*]. In healthcare settings human-to-human transmission has led to large outbreaks with higher  $R_0$ , providing a warning sign for pandemic potential [55–57]. Continuous zoonotic events are being reported. The

current consensus is that close and prolonged contact with an index case seems to be required for MERS-CoV transmission and adaptation would be required for efficient human-to-human transmission.

### COVID-19 outbreak containment

Strategies to contain the COVID-19 pandemic were initially limited to non-pharmaceutical interventions, including lockdowns and physical distancing measures. Within a year after onset of the pandemic, several vaccines were developed and shown to be safe and effective in large double-blind placebo-controlled clinical studies [58\*\*,59\*\*,60\*\*,61\*\*]. Large-scale implementation of these vaccines will reduce morbidity and mortality, alleviate the pressure on health care systems and relieve non-pharmaceutical interventions. However, it remains unclear how well vaccines protect from upper respiratory tract shedding and subsequent viral transmission. A recent study found the first evidence of reduced incidence of infection in household contacts of vaccinated healthcare workers, but the level of protection from shedding may differ for different variants, with age, and with time since vaccination [62]. In addition, it remains to be determined what vaccination coverage will be reached eventually, given vaccine hesitancy and disparities in access. Therefore, for the months and years ahead, it is crucial to continue to explore potential additional countermeasures that inhibit SARS-CoV-2 transmission.

### Animal models to study coronavirus transmission

Animal models have been instrumental to gain insight into SARS-CoV, MERS-CoV and SARS-CoV-2 replication kinetics, shedding, pathogenesis and medical countermeasures, and have been reviewed elsewhere [63\*,64–66]. These models include non-human primates (NHPs), cats, dromedary camels, ferrets, hamsters, rabbits and (transgenic) mice. Since coronaviruses emerge after zoonotic introduction, it is essential to study coronavirus transmission potential and evaluate transmission prevention by medical countermeasures. Animal models of transmission are therefore crucial. For SARS-CoV-2 specifically, this includes assessment of transmissibility of emerging variants, or transmission in the presence of vaccine-induced immunity. In this review, we provide a brief summary of the currently employed animal models to study different aspects of COVID-19, with a focus on transmission in combination with therapeutics that could interrupt transmission.

Transmission studies in animal models are relatively scarce. Natural SARS-CoV-2 transmission in animals has been described in companion animals (e.g. cats and dogs) and on mink farms. Experimental animal transmission models remain limited to the cat, ferret and hamster model. Although it has been shown that rabbits are susceptible to experimental SARS-CoV-2 infection, given

**Table 1****Overview of transmission models**

	Mouse	Cat	Hamster	Ferret	Fruit bat	NHP <sup>d</sup>
Animal handling BSL3	Easy	Difficult	Easy	Intermediate	Difficult	Difficult
Species relevance	Limited	Limited	Limited	Sialic acids RT <sup>a</sup> resemble human	Close to original host	Close to human
Naturally susceptible	Only variants <sup>b</sup>	Yes	Yes	Yes	Not fully	Yes
Clinical presentation	Variable	Limited disease	Severe disease	Limited disease	Limited disease	Limited disease
Transmission efficacy <sup>c</sup>	Unknown	DC: efficient Aer: not efficient	DC: efficient Aer: efficient	DC: efficient Aer: possible	Limited	Unknown
Reagents availability	Widespread	Limited	Limited	Limited	Limited	Widespread
Transmission literature	Not available	Limited	Multiple publications	Multiple publications	Limited	Not available
References	N/A	[74,76,77]	[97**,104,108]	[87,89,91,94**]	[89]	N/A

<sup>a</sup> RT = respiratory tract.<sup>b</sup> mouse susceptibility differs per model, dependent on the use of mouse strain / transgenic mice. Some models are susceptible to variants of concern [63\*,117].<sup>c</sup> DC = direct contact, aer = aerosol.<sup>d</sup> NHP = non-human primates.

the high dose needed to establish productive infection, transmission of the virus from rabbit-to-rabbit is unlikely [67]. Ferrets thus far appear an ideal model to study transmission in the absence of clinical signs, whereas hamsters are more appropriate to study transmission in the context of pathology and severe disease. NHPs are rarely used for transmission studies, in part due to practical limitations in high-containment facilities. An overview of the characteristics of different animal models and their usability in transmission studies is shown in Table 1.

### Human-to-animal transmission: companion animals

Naturally occurring human-to-animal transmission was already reported during the SARS-CoV outbreak in 2002/2003. Viral RNA was detected in oropharyngeal swabs obtained from cats after contact with infected individuals, and infections were confirmed serologically [68]. Susceptibility of cats was subsequently confirmed experimentally [69]. Because of these reports, several studies performed in Hong Kong describe swabbing of companion animals in close contact with confirmed COVID-19 cases. Here, household transmission to dogs [70] and cats [71] was observed, but in most cases infected animals remained asymptomatic [72]. This remains to be determined for emerging variants [73]. Experimental infections confirmed susceptibility of cats, but dogs proved poorly susceptible [74,75].

Cats were also evaluated as an animal model for SARS-CoV-2 transmission. A first study assessed airborne transmission among cats housed in adjacent cages (also allowing for respiratory droplet transmission) and

demonstrated that experimental transmission is feasible, although this remained limited to infection of 1 out of 3 sentinel animals [75]. Direct contact transmission in cats was more robust, showing infection of all sentinel animals in several separate small studies [74,76,77].

### Human-to-animal transmission: outbreaks on mink farms

Large outbreaks of SARS-CoV-2 infection have been reported on mink (*Neovison vison*) farms in the Netherlands, Spain, Italy, Sweden, Greece, the US and Denmark, initiated by a human-to-mink transmission [78]. Circulation and spread initially went unnoticed as mustelids usually develop asymptomatic or mild disease, and transmission on some farms was only confirmed after serological surveys. However, upper and lower respiratory tract involvement has been described [79,80]. Transmission on mink farms was rapid and widespread after initial introduction. Once introduced, it has proven difficult to stop transmission on a farm, and ongoing transmission between farms was described [81\*]. Large-scale culling of mink was initiated when it was observed that ongoing SARS-CoV-2 transmission led to an accumulation of mutations in the S protein, and that these viruses could spill back from mink farms into the community in Denmark and possibly the Netherlands [81\*,82–84].

Experimental infection or transmission studies in mink have not been performed to date. Mink belong to the family of mustelids (*Mustelidae*), together with ferrets. Ferrets were already shown to be highly susceptible to SARS-CoV in 2003 [69], and are now in widespread use to study SARS-CoV-2 transmission and pathogenesis.