

# gratia: An R package for working with generalized additive models

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DOI: [DOI unavailable](#)

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Published: N/A

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## Summary

Generalized additive models (GAMs, Hastie & Tibshirani, 1990; Wood, 2017) are an extension of the generalized linear model that allows the effects of covariates to be modelled as smooth functions. GAMs are increasingly used in many areas of science (e.g. Pedersen et al., 2019; Simpson, 2018) because the smooth functions allow nonlinear relationships between covariates and the response to be learned from the data through the use of penalized splines. Within the R (R Core Team, 2024) ecosystem, Simon Wood's *mgcv* package (Wood, 2017) is widely used to fit GAMs and is a *Recommended* package that ships with R as part of the default install. A growing number of other R packages build upon *mgcv*, for example as an engine to fit specialised models not handled by *mgcv* itself (e.g. *GJMR*, Marra & Radice, 2023), or to make use of the wide range of splines available in *mgcv* (e.g. *brms*, Bürkner, 2017).

The *gratia* package builds upon *mgcv* by providing functions that make working with GAMs easier. *gratia* takes a *tidy* approach (Wickham, 2014) providing *ggplot2* (Wickham, 2016) replacements for *mgcv*'s base graphics-based plots, functions for model diagnostics and exploration of fitted models, and a family of functions for drawing samples from the posterior distribution of a fitted GAM. Additional functionality is provided to facilitate the teaching and understanding of GAMs.

## Generalized additive models

A GAM has the form

$$y_i \sim \mathcal{D}(\mu_i, \phi)$$

$$g(\mu_i) = \mathbf{A}_i \boldsymbol{\gamma} + \sum_{j=1}^J f_j(x_{ji})$$

where observations  $y_i$  are assumed to be conditionally distributed  $\mathcal{D}$  with expectation  $\mathbb{E}(y_i) = \mu_i$  and dispersion  $\phi$ . The expectation of  $y_i$  is given by a linear predictor of strictly parametric terms, whose model matrix is  $\mathbf{A}_i$  with parameters  $\boldsymbol{\gamma}$ , plus a sum of smooth functions of  $j = 1, \dots, J$  covariates  $f_j()$ .  $g()$  is a link function mapping values on the linear predictor to the scale of the response.

The smooth functions  $f_j$  are represented in the GAM using penalised splines, which are themselves formed as weighted sums of basis functions,  $b_k()$ , (De Boor, 2001) e.g.

$$f_j(x_{ij}) = \sum_{k=1}^K \beta_{jk} b_{jk}(x_{ij})$$

for a univariate spline. The weights,  $\beta_k$ , are model coefficients to be estimated alongside  $\boldsymbol{\gamma}$ . To avoid overfitting, estimates  $\hat{\beta}_{jk}$  and  $\hat{\boldsymbol{\gamma}}$  are sought to minimise the penalised log-likelihood of the model

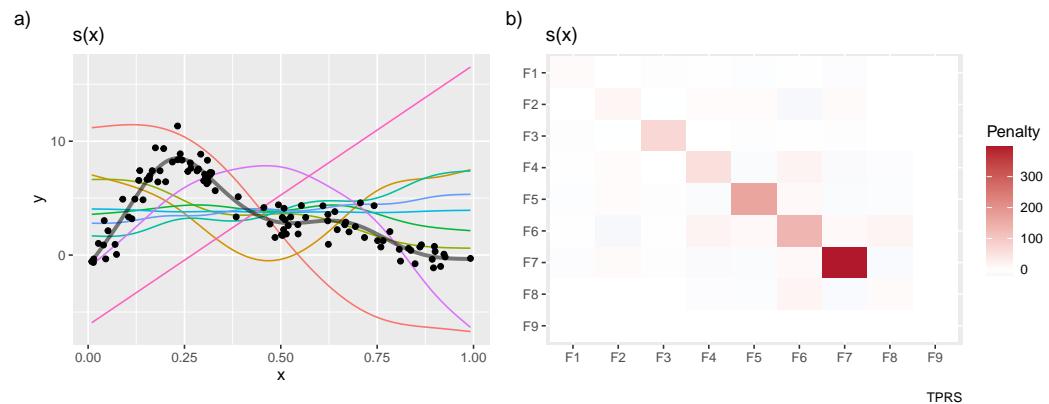
$$\mathcal{L}(\boldsymbol{\beta}) = \ell(\boldsymbol{\beta}) - \frac{1}{2\phi} \sum_j \lambda_j \boldsymbol{\beta}_j^\top \mathbf{S}_j \boldsymbol{\beta}_j$$

where  $\ell$  is the log likelihood of the data at the parameter estimates,  $S_j$  are penalty matrices and  $\lambda_j$  are smoothing parameters associated with each smooth. Note that  $\beta$  now contains the coefficients  $\gamma$  and  $\beta_{jk}$ .  $\beta_j^T S_j \beta_j$  measures the wiggliness of  $f_j$ , which, with the default penalty, is the integrated squared second derivative of  $f_j$ . The smoothing parameters,  $\lambda$ , control the trade-off between fit to the data and the complexity of the estimated functions.

The default spline created by *mgcv*'s `s()` is a low rank, thin plate regression spline (Wood, 2003). Figure 1, shows the basis functions for such a spline fitted to data simulated from the function

$$f = 0.2x^{11}\{10(1-x)\}^6 + 10(10x)^3(1-x)^{10}$$

with additive Gaussian noise ( $\mu = 0, \sigma = 1$ ), and the associated penalty matrix, prepared using functions from *gratia*.



**Figure 1:** Basis functions (a) and associated penalty matrix (b) for a penalised, low rank, thin plate regression spline. a) shows the individual basis functions (thin coloured lines), as well as the data (black points) to which the GAM was fitted. The estimated smooth is shown as the thick grey line. b) shows the penalty matrix for the basis shown in a). Note the 9th basis function (labelled 'F9,' which is the linear function at the lower left to upper right in a), is not affected by the penalty as it has 0 second derivative everywhere, and hence the resulting penalty for this function is 0.

## Statement of need

*mgcv* is state-of-the-art software for fitting GAMs and their extensions to data sets on the order of millions of observations (e.g. Li & Wood, 2020; Wood, 2011; Wood et al., 2016). *mgcv* provides functions for plotting estimated smooth functions, as well as for producing model diagnostic plots. These functions produce plots using base graphics, the original plotting system for R. Additionally, *mgcv* returns fitted GAMs as complex list objects (see `?mgcv::gamObject`), the contents of which are not easily used for downstream analysis without careful study of *mgcv* and its help pages, plus a good understanding of GAMs themselves. One of the motivations driving the development of *gratia* was to provide equivalent plotting capabilities using the *ggplot2* package (Wickham, 2016). To facilitate this, *gratia* provides functions for representing the model components as objects using *tidy* principles, which are suitable for plotting with *ggplot2* or manipulation by packages in the *tidyverse* (e.g. Wickham et al., 2023). This functionality allows for high-level plotting using the `draw()` method, as well as easily customisable plot creation using lower-level functionality.

Taking a Bayesian approach to smoothing with penalized splines (Kimeldorf & Wahba, 1970; Silverman, 1985; Wahba, 1983, 1985; see Miller, 2019 for a summary), it can be shown that GAMs fitted by *mgcv* are an empirical Bayesian model with an improper multivariate normal prior on the basis function coefficients. Samples from the posterior distribution of these models can be used to estimate the uncertainty in quantities derived from a GAM. This can be invaluable in applied research, where, for example, a quantity of interest may arise as

an operation on predictions from the model. *gratia* provides functions for sampling from the posterior distribution of estimated smooths and from the model as a whole, where sampling can include the uncertainty in the estimated coefficients (`fitted_samples()`), the sampling uncertainty of the response (`predicted_samples()`), or both (`posterior_samples()`). By default, a Gaussian approximation to the posterior distribution is used, but a simple Metropolis Hasting sampler can be substituted (using `mgcv:::gam.mh()`), which has better performance when the posterior is not well approximated by a Gaussian approximation.

The teaching of GAMs can benefit from visualisation of the spline basis functions and associated penalty matrices. *gratia* provides this functionality via `basis()` and `penalty()`, which can be applied either to a smooth specification (e.g. `s(x, z, bs = "ds")`) or to a fitted GAM (see Figure 1). These functions expose functionality already available in *mgcv*, but supply outputs in a tidy format, which makes access to these features more intuitive than the original implementations in *mgcv*. Additional utility functions are provided, for example: `model_constant()`, `edf()`, `model_edf()`, `overview()`, and `inv_link()`, which extract the model intercept term (or terms), the effective degrees of freedom of individual smooths and the overall model, shows a summary of the fitted GAM, and extracts the inverse of the link function(s) used, respectively.

The overall aim of *gratia* is to abstract away some of the complexity of working with GAMs fitted using *mgcv* to allow researchers to focus on using and interrogating their model rather than the technical R programming needed to achieve this. As a result, *gratia* is increasingly being used by researchers in many fields, and has, at the time of writing, been cited over 200 times (data from Google Scholar).

## Example usage

In this short example, I illustrate a few of the features of *gratia* using a data set of sea surface chlorophyll *a* measurements at a number of locations in the Atlantic Ocean, whose spatial locations are given as geographical coordinates (`lat` and `lon`), plus two additional covariates; `bathy`, the depth of the ocean, in metres, at the sampling location, and `jul.day`, the day of the year in which the observation was made. These data are in the `chl` dataset provided by the `gamair` package accompanying Wood (2017).

The packages required for this example are loaded, as is the data set `chl` with

```
pkgs <- c("mgcv", "gamair", "gratia", "ggplot2", "dplyr", "ggdist")
loaded <- vapply(pkgs, library, logical(1L), logical.return = TRUE,
                 character.only = TRUE)
data(chl, package = "gamair")
```

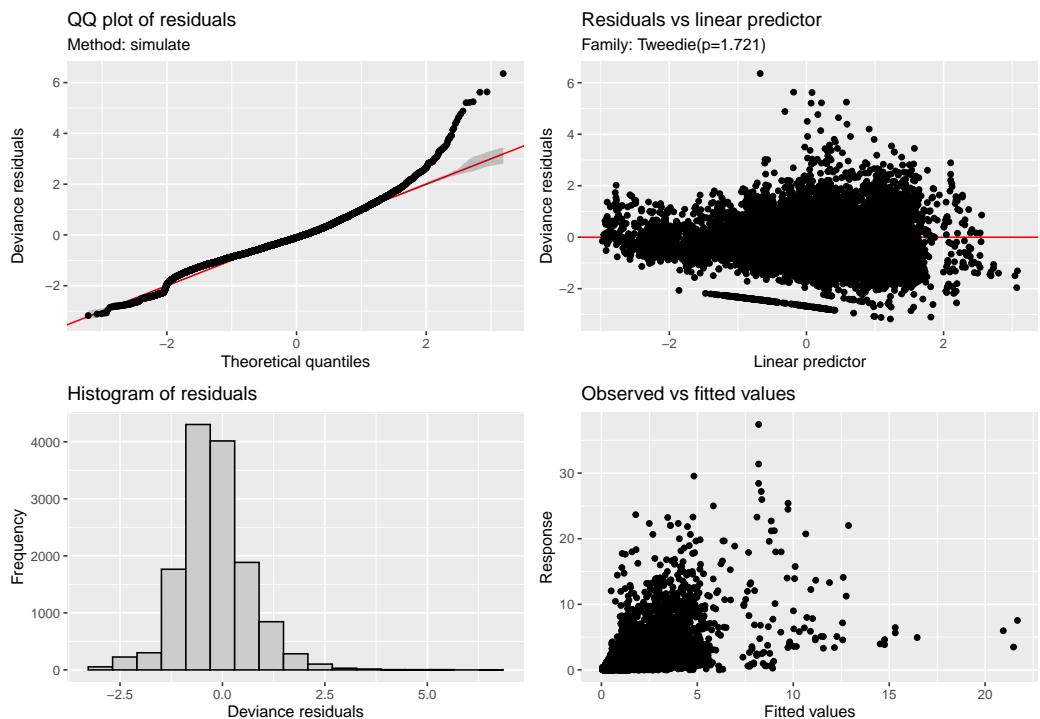
A simple GAM for these data is to model the response (`chl`) with a spatial smooth of latitude (`lat`) and longitude (`lon`) as covariates. Here, I use a spline on the sphere (SOS) smoother built using a Duchon spline with second order derivative penalty (Duchon, 1977). Additional terms included in the linear predictor are a smooth of the day of year of sample collection (`jul.day`) and a smooth of ocean depth (`bath`). The response is assumed to be conditionally distributed Tweedie, with the power parameter (`p`) of the distribution estimated during fitting. Model coefficients and smoothing parameters are estimated using restricted maximum likelihood (Wood, 2011)

```
ctrl <- gam.control(nthreads = 10)
m1 <- gam(
  chl ~ s(lat, lon, bs = "sos", m = -1, k = 150) +
    s(jul.day, bs = "cr", k = 20) +
    s(bath, k = 10),
  data = chl, method = "REML", control = ctrl, family = tw()
)
```

Model diagnostic plots can be produced using `appraise()`, which by default produces four plots: i) a QQ plot of model residuals, with theoretical quantiles and reference bands generated following Augustin et al. (2012), ii) a plot of residuals (deviance residuals are the default) against linear predictor values, iii) a histogram of residuals, and iv) a plot of observed versus fitted values. Model diagnostic plots for the model, with simulated residuals-based reference bands on the QQ plot, are produced with

```
appraise(m1, method = "simulate")
```

which show significant heteroscedasticity and departure from the conditional distribution of the response given the model (Figure 2).



**Figure 2:** Model diagnostic plots for the GAM fitted to the ocean chlorophyll  $a$  data produced by the `appraise()` function. The four plots produced are: i) a QQ plot of model residuals, with theoretical quantiles and reference bands generated following Augustin et al. (2012) (upper left), ii) a plot of residuals (deviance residuals are default) against linear predictor values (upper right), iii) a histogram of deviance residuals (lower left), and iv) a plot of observed versus fitted values (lower right)

The problems with the model apparent in the diagnostics plots are probably due to important controls on chlorophyll  $a$  missing from the covariates available in the example data. However, the original model assumed constant values for the scale,  $\varphi$ , and the power parameter  $p$ , which may be too inflexible given the absence of important effects in the model. A distributional GAM, where linear predictors for all distributional parameters, may improve the model diagnostics.

A distributional GAM for  $\mathcal{D}$  Tweedie, with linear predictors for  $\mu$ ,  $p$ , and  $\varphi$  is fitted below using `mgcv`'s `twlss()` family

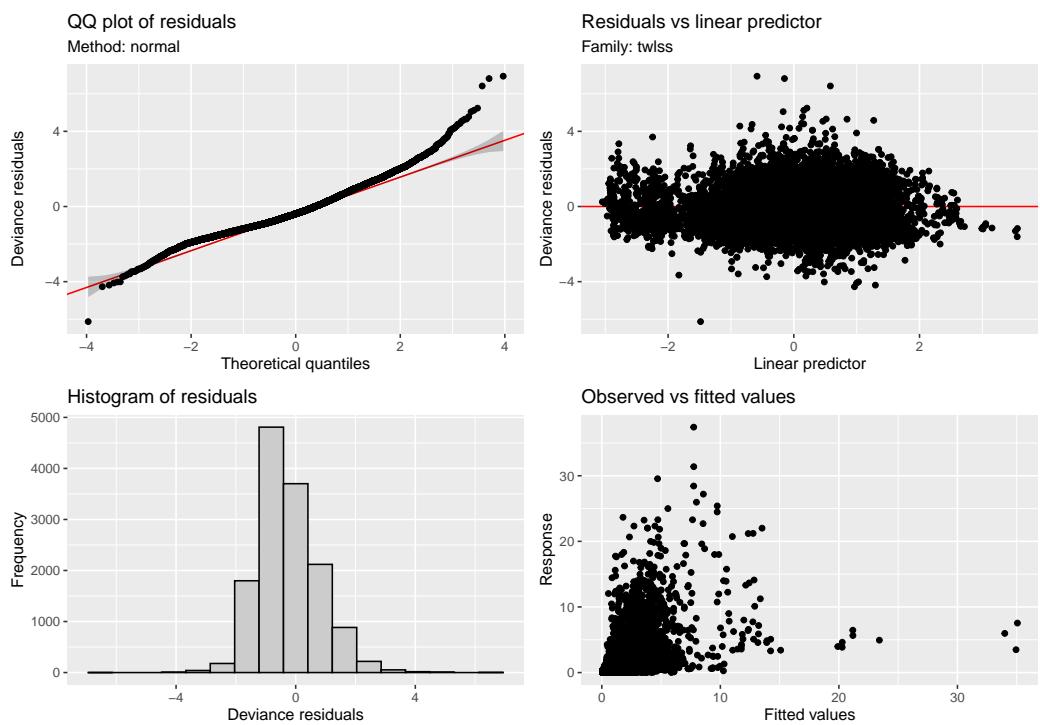
```
m2 <- gam(
  list(
    chl ~ s(lat, lon, bs = "sos", m = -1, k = 150) + # location
    s(jul.day, bs = "cr", k = 20) +
    s(bath, k = 10),
    ~ s(lat, lon, bs = "sos", m = -1, k = 100) +      # power
```

```

  s(jul.day, bs = "cr", k = 20) +
  s(bath, k = 10),
~ s(lat, lon, bs = "sos", m = -1, k = 100) +      # scale
  s(jul.day, bs = "cr", k = 20) +
  s(bath, k = 10)),
  data = chl, method = "REML", control = ctrl, family = twlss()
)

```

This model has much better model diagnostics although some large residuals remain (Figure 3). Note that the QQ plot uses theoretical quantiles from a standard normal distribution as the simulation-based values are not currently available in *mgcv* or *gratia* for some of the distributional families, including the *twlss()* family, and as such, the reference bands may not be appropriate.



**Figure 3:** Model diagnostic plots for the distributional GAM fitted to the ocean chlorophyll *a* data produced by the *appraise()* function. Refer to the caption for Figure 2 for a description of the plots shown.

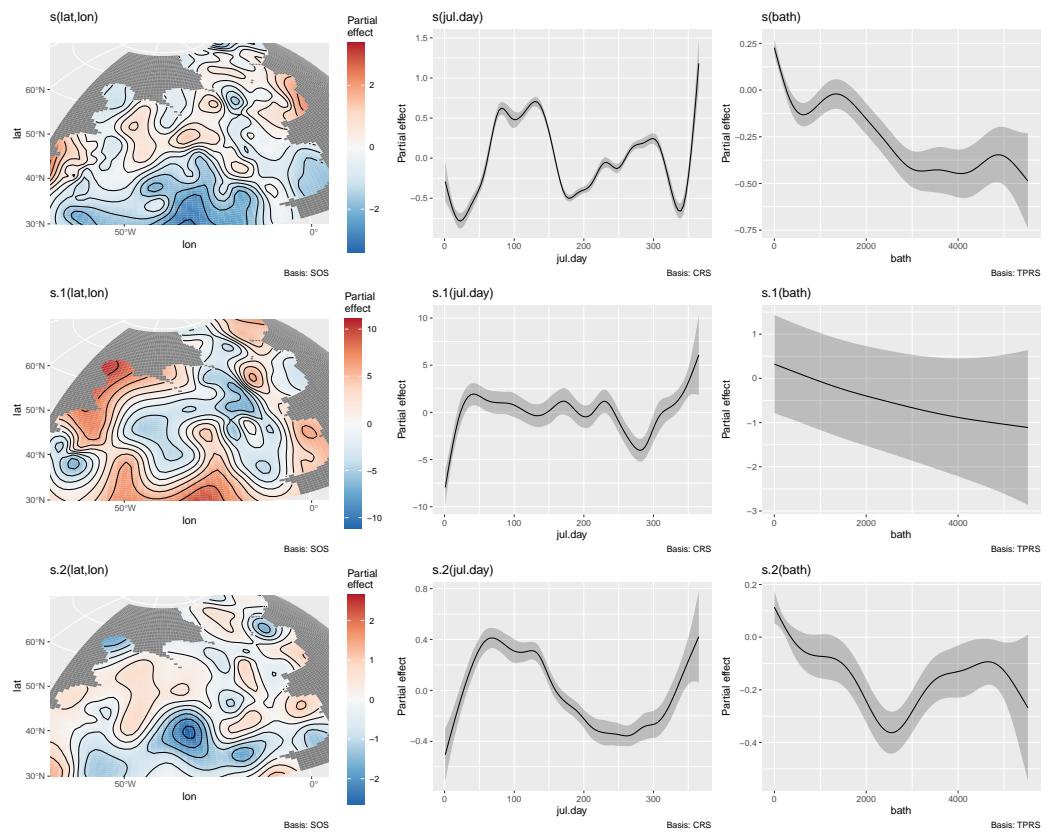
*gratia* can handle distributional GAMs fitted with *mgcv* and also *GJRM*'s *gamlss()*. Below, the estimated smooths from *m2* are plotted using *draw()*

```

crs <- "+proj=ortho +lat_0=20 +lon_0=-40"
draw(m2, crs = crs, default_crs = 4326, dist = 0.05, rug = FALSE)

```

Here, we see a specialised plot drawn for spline-on-the-sphere smooths  $f(\text{lat}_i, \text{lon}_i)$  (Figure 4), which uses *ggplot2::coord\_sf()* and functionality from the *sf* package (Pebesma, 2018; Pebesma & Bivand, 2023) to visualise the smooth via an orthographic projection.



**Figure 4:** Estimated smooth functions for the distributional GAM, `m2`, fitted to the ocean chlorophyll *a* data. The first row of plots is for the linear predictor of the conditional mean chlorophyll *a*, while the second and third rows are for the conditional power parameter and conditional scale, respectively. The shaded ribbons are 95% Bayesian credible intervals.

If the provided plots are insufficient for users' needs, lower-level functionality is provided by *gratia* to facilitate bespoke plotting with `ggplot2`. For example, to evaluate the SOS smooth at a grid (50x50) of values over the range of the covariates, we use `smooth_estimates()` and add a Bayesian credible interval with `add_confint()`:

```
smooth_estimates(m2, select = "s(lat,lon)", n = 50) |>
  add_confint()
```

This returns a data frame of the requested values, which is easily plotted using `ggplot()`.

## Posterior sampling

Perhaps we are interested in the average expected chlorophyll *a* between 40–50 degrees N and 40–50 degrees W. It would be quite a simple matter to calculate this value from the fitted model: we first create a slice through the data for the spatial locations were are interested in using the `data_slice()` function, which ensures that `ds` contains everything we need to predict from the fitted model

```
ds <- data_slice(m2,
  lat = evenly(lat, lower = 40, upper = 50, by = 0.5),
  lon = evenly(lon, lower = -50, upper = -40, by = 0.5))
```

Next, `fitted_values()` returns the predicted values at the specified locations. I only include the spatial effects, excluding the effects of ocean depth and day of year:

```
use <- c("(Intercept)", "s(lat,lon)" # , "s.1(lat,lon)", "s.2(lat,lon)")
fv <- fitted_values(m2, data = ds, terms = use) # predict
```

Finally, I summarise the predictions for the location parameter to yield the average of the predicted values

```
fv |>
  filter(.parameter == "location") |>
  summarise(chl_a = mean(.fitted))

## # A tibble: 1 x 1
##   chl_a
##   <dbl>
## 1  1.07
```

While this is an acceptable answer to the question, it lacks an uncertainty estimate. This is where posterior sampling is useful. With a small modification of the above code and a little data wrangling, we can produce an uncertainty estimate ,using `fitted_samples()` to generate posterior draws of the expected chlorophyll *a*:

```
fs <- fitted_samples(m2,               # model
                     data = ds,          # values of covariates to predict at
                     terms = use,         # which terms to include in predictions
                     n = 10000,           # number of posterior draws
                     method = "gaussian", # Gaussian approximation to the posterior
                     unconditional = TRUE, # incl uncertainty for estimating lambda
                     n_cores = 4,          # how many CPU cores to compute MVN samples
                     seed = 342)           # set the random number seed, used internally
```

The posterior draws can then be summarised as before, except now the average chlorophyll *a* is calculated separately for each posterior draw (`.draw`)

```
fs |>                                # take the posterior draws
  group_by(.draw) |>                  # group them by `.`.draw` 
  summarise(chl_a = mean(.fitted)) |> # compute mean of fitted chl a
  ggdist::median_qi()                 # summarise posterior

## # A tibble: 1 x 6
##   chl_a .lower .upper .width .point .interval
##   <dbl>  <dbl>  <dbl>  <dbl> <chr>  <chr>
## 1  1.07   0.870  1.33   0.95 median qi
```

The posterior distribution of average chlorophyll *a* is summarised using `median_qi()` from the `ggdist` package (Kay, 2024a, 2024b). While it would be a simple matter to compute the interval with base R commands, the use of `median_qi()` illustrates how *gratia* tries to interact with other packages.

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