Energy-based Advection Modelling Using Bond Graphs: Additional Material

Peter Gawthrop. peter.gawthrop@unimelb.edu.au August 25, 2022

Contents

1	Introduction									
	1.1 Import some python code	1								
2	Pipe									
	2.1 Chain of compartments	2								
	2.2 Transfer-function analysis	9								
3	Coupled Advection and Transformation									
	3.1 Orifice connection	11								
	3.2 Pipe connection	14								
4	Enzyme-catalysed reaction									
	4.1 Split enzyme-catalysed reaction	20								
5	Circular advection									
	5.1 Orifice (Symmetric case): Bond graph model	21								
	5.1.1 Simulate									
	5.2 Circular advection: pipe connection									
6	Variable flow rate	32								
7	Pharmacokinetics									
	7.1 Parameters: Map78 Table I	40								
	7.2 Simulation									

1 Introduction

This notebook generates the figures for the paper: *Energy-based Advection Modelling Using Bond Graphs*. This is a tidied and extended version of the 2021 version.

1.1 Import some python code

The bond graph analysis uses a number of Python modules:

```
[1]: import numpy as np
import matplotlib.pyplot as plt
import IPython.display as disp

## Control toolbox
```

```
import control as con
## Bond graph tools
import BondGraphTools as bgt
print('BondGraphTools version', bgt.version)
## Stoichiometric analysis
import stoich as st
## SVG bg representation conversion
import svgBondGraph as sbg
## Modular bond graphs
import modularBondGraph as mbg
## Stoichiometric conversion
import stoichBondGraph as stbg
## Allow reloading of modules
import importlib as imp
## CSV reader etc
import csv
from os.path import exists
## Set quiet=False for verbose output
quiet = True
## Plot parameters
lw = 5
## Optional plotting
Plotting = True
def Savefig(plotname):
    if Plotting:
        plt.savefig(plotname)
```

BondGraphTools version 0.3.7

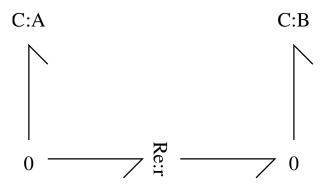
2 Pipe

2.1 Chain of compartments

This is the lumped model of advection considered in Section 3.1. of the paper. Using the modular extension to BGT, a chain of simple A \implies B reactions of length N is created.

```
[2]: ## Simple A <==> B reaction
sbg.model('AB_abg.svg')
import AB_abg
disp.SVG('AB_abg.svg')
```

[2]:



[3]:

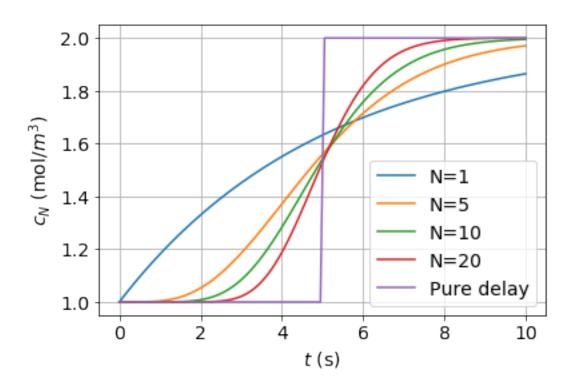
[4]:

$$A \Leftrightarrow B$$
 (1)

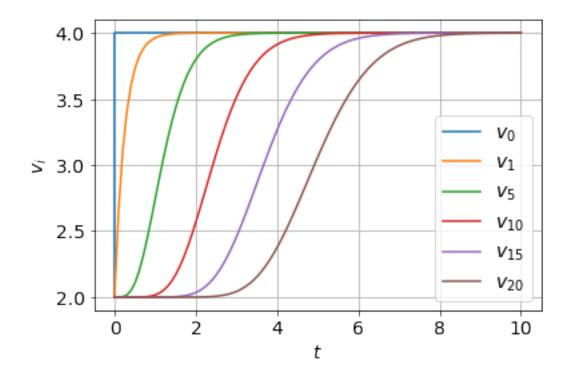
Exposing A and B
Creating ABChain
Creating ABO from AB within ABChain
Creating A from A within ABChain
Creating jA from 013 within ABChain
Creating B from A within ABChain
Creating jB from 013 within ABChain

 $A \Leftrightarrow B$ (2)

```
Pipe = mbg.chain(AB_abg.model(),inport = 'A',outport = 'B',
 →N=N_lump+1,rename_components=True,Comps=['R'],quiet=quiet)
   V_i = V_p/N_lump # Volume of lump
    parameter = {}
    UniDir = []
    for i in range(N_lump+1):
        reac = f'r_{i}'
        UniDir.append(reac)
        kap = f'kappa_{reac}'
        parameter[kap] = Q0
    s = st.stoich(Pipe,UniDir=UniDir, quiet=quiet)
    sc = st.statify(s,chemostats=['A','B'])
    species = s['species']
    disp.Latex(st.sprintrl(s))
    X0 = np.ones(s['n_X'])
    X0[species.index('A')] = 2
    for i in range(N_lump):
        spec = f'IS{i+1}'
        K = f'K_{spec}'
        parameter[K] = 1/V_i
        X0[species.index(spec)] = V_i
    dat = st.sim(s,sc=sc,t=t,parameter=parameter,X0=X0,quiet=quiet)
    r_N = f'r_{N_lump}'
    i_N = species.index(f'IS{N_lump}')
    x_N = dat['X'][:,i_N]
    plt.plot(t,x_N/V_i,label=f'N={N_lump}')
## Pure delay
y_del = 1+np.heaviside(t-delay,1)
plt.plot(t,y_del,label='Pure delay')
plt.grid()
plt.legend()
plt.xlabel('$t$ (s)')
plt.ylabel('$c_N$ (mol/$m^3$)')
Savefig('Figs/Pipe.pdf')
```



```
[6]: V = dat['V']
     reaction = s['reaction']
     I = [0,1,5,10,15,20]
     for i,reac in enumerate(reaction):
         if i in I:
              v = V[:,i]
              if i == 0:
                  vv = np.hstack((2,v))
                  tt = np.hstack((0,t))
              else:
                  tt = t
                  \Delta\Delta = \Delta
              vi = 'v_{{i}} + f'{{i}}' + '}'
              plt.plot(tt,vv,label=f'${vi}$')
     plt.legend()
     plt.grid()
     plt.xlabel('$t$')
     plt.ylabel('$v_i$')
     plt.show()
     Savefig('Figs/Pipe_v.pdf')
```



<Figure size 432x288 with 0 Axes>

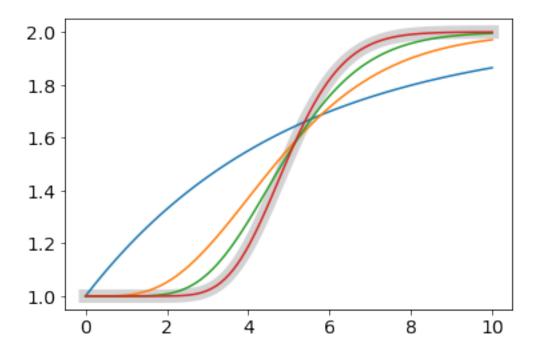
```
[7]: ## Compare with a constructed transfer function 1/(s+sT/N)^N
plt.plot(t,x_N/V_i,lw=10,color='lightgrey')
for N_lump in N_lumps:
    lump = con.tf(1,[delay/N_lump,1])
    lumps = con.tf(1,1)
    for i in range(N_lump):
        lumps = con.series(lumps,lump)
# print(lumps)
t,y = con.step_response(lumps,T=t)
    plt.plot(t,y+1)
```

```
u
------
```

RuntimeError Traceback (most recent call⊔ →last)

⊔ →-----

⇔last)	RuntimeError					Tracebad	ck (1	most	recen	nt c	all⊔
of nu ⊶	RuntimeError: umpy is 0xd	module	compiled	against	API	version	0xe	but	this	ver	sion _L
	ш										
⇔last)	RuntimeError					Tracebad	ck (1	most	recen	nt c	all <mark>u</mark>
⊶of nı	RuntimeError: umpy is 0xd	module	compiled	against	API	version	0xe	but	this	ver	sion _L
	ш										
⇔last)	RuntimeError					Tracebad	ck (1	most	recer	ıt c	all <mark>u</mark>
⇔of nu	RuntimeError: umpy is 0xd	module	compiled	against	API	version	0xe	but	this	ver	sion _L
package matrix	eterg/.local/l: s/scipy/sparse, (rcond=9.31351) n solve(Q, P)	/linalg/	$^\prime$ matfuncs	.py:710:		•	ng: [I11-0	condit	ion	ed



[8]:

$$A \stackrel{r_0}{\rightleftharpoons} IS_1 \qquad (3)$$

$$IS_1 \stackrel{r_1}{\rightleftharpoons} IS_2 \qquad (4)$$

$$IS_2 \stackrel{r_2}{\rightleftharpoons} IS_3 \qquad (5)$$

$$IS_3 \stackrel{r_3}{\longleftrightarrow} IS_4$$
 (6)

$$IS_4 \stackrel{r_4}{\longleftrightarrow} IS_5 \tag{7}$$

$$IS_5 \stackrel{r_5}{\Longleftrightarrow} IS_6$$
 (8)

$$IS_6 \stackrel{r_6}{\Longleftrightarrow} IS_7$$
 (9)

$$IS_7 \stackrel{r_7}{\Longleftrightarrow} IS_8$$
 (10)

$$IS_8 \stackrel{r_8}{\longleftarrow} IS_9 \tag{11}$$

$$IS_9 \stackrel{r_9}{\Longleftrightarrow} IS_{10} \tag{12}$$

$$IS_{10} \stackrel{r_{10}}{\longleftarrow} B \tag{13}$$

2.2 Transfer-function analysis

This chain of reactions has liner dynamics which are extracted to give a model in the Python Control Toolbox format. The effective timeconstant is computed from:

$$\tau = \sum_{i=1}^{N} 1/p_i \tag{14}$$

where p_i is the *i*th pole of the tranfer function relating the input and output of the chain.

```
[9]: imp.reload(st)
UniDir=[]
for i in range(N_lump+1):
        UniDir.append(f'r_{i}')
print(UniDir)
s = st.stoich(Pipe,UniDir=UniDir, quiet=quiet)
disp.Latex(st.sprintvl(s))
```

['r_0', 'r_1', 'r_2', 'r_3', 'r_4', 'r_5', 'r_6', 'r_7', 'r_8', 'r_9', 'r_10']
[9]:

$$v_{r0} = K_A \kappa_{r0} x_A \tag{15}$$

$$v_{r1} = K_{IS1} \kappa_{r1} x_{IS1} \tag{16}$$

$$v_{r2} = K_{IS2}\kappa_{r2}\chi_{IS2} \tag{17}$$

$$v_{r3} = K_{IS3}\kappa_{r3}\chi_{IS3} \tag{18}$$

$$v_{r4} = K_{IS4}\kappa_{r4}x_{IS4} \tag{19}$$

$$v_{r5} = K_{IS5}\kappa_{r5}x_{IS5} \tag{20}$$

$$v_{r6} = K_{IS6} \kappa_{r6} \chi_{IS6} \tag{21}$$

$$v_{r7} = K_{IS7}\kappa_{r7}\chi_{IS7} \tag{22}$$

$$v_{r8} = K_{IS8} \kappa_{r8} x_{IS8} \tag{23}$$

$$v_{r9} = K_{IS9}\kappa_{r9}\chi_{IS9} \tag{24}$$

$$v_{r10} = K_{IS10} \kappa_{r10} x_{IS10} \tag{25}$$

```
[10]: sc = st.statify(s,chemostats=['A','B'])
    sys = st.lin(s,sc)
# print(sys)
A,B,C,D = con.ssdata(sys)
    sys12 = con.ss(A,B[:,0],C[N_lump,:],0)
    print(con.tf(sys12))
    print(con.pole(sys12))
    tc = np.sum(-1/con.pole(sys12))
    gain = con.dcgain(sys12)
    print('Time constant =',tc)
    t_step,y_step = con.step_response(sys12,T=np.linspace(0,2*tc))
    plt.plot(t_step/tc,y_step.T/gain)
```

```
plt.vlines(1,0,max(y_step)/gain)
plt.grid()
plt.xlabel('$t$ (normalised)')
plt.ylabel('$y_{step}$ (normalised)')
```

⊔ →-----

RuntimeError

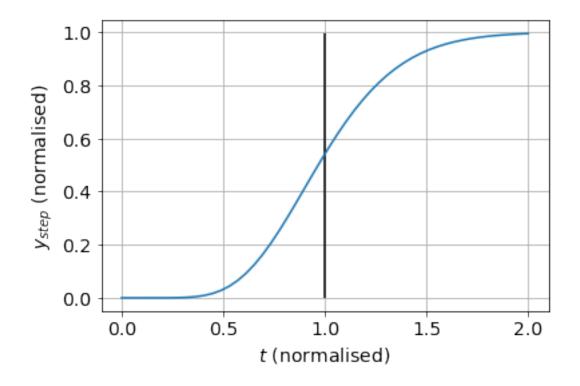
Traceback (most recent call⊔

→last)

 $-3.553e-15 s^9 - 7.105e-15 s^8 - 9.948e-14 s^7 - 2.842e-13 s^6 - 3.979e-13 s^5 - 4.547e-13 s^4 - 2.7e-13 s^3 - 1.137e-13 s^2 - 2.132e-14 s + 1$

[-1. -1. -1. -1. -1. -1. -1. -1. -1. -1.]Time constant = 10.0

[10]: Text(0,0.5,'\$y_{step}\$ (normalised)')



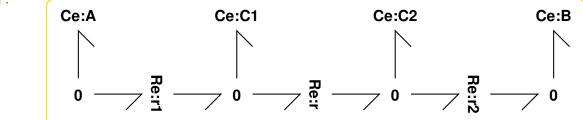
3 Coupled Advection and Transformation

This corresponds to Section 4 of the paper.

3.1 Orifice connection

```
[11]: ##
sbg.model('splitReos_abg.svg')
import splitReos_abg
disp.SVG('splitReos_abg.svg')
```

[11]:



```
[12]: ##Stoichiometry
s = st.stoich(splitReos_abg.model(),quiet=quiet)
chemostats = ['A','B']
sc = st.statify(s,chemostats=chemostats)
disp.Latex(st.sprintrl(s,chemformula=True))
```

[12]:

$$C_1 \stackrel{r}{\Longleftrightarrow} C_2$$
 (26)

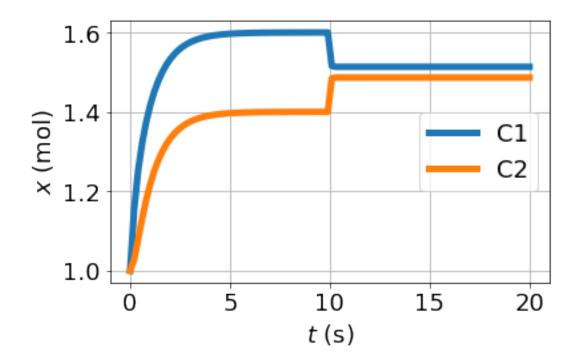
$$A \stackrel{r_1}{\longleftrightarrow} C_1 \tag{27}$$

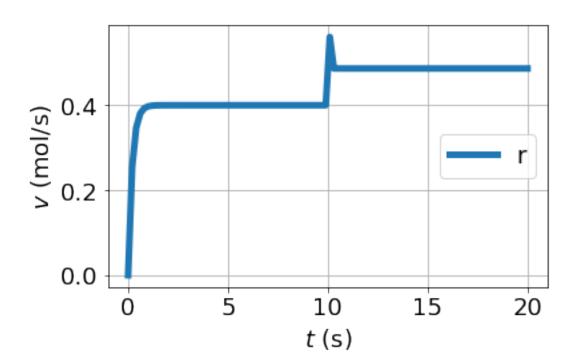
$$C_2 \stackrel{r_2}{\longleftrightarrow} B$$
 (28)

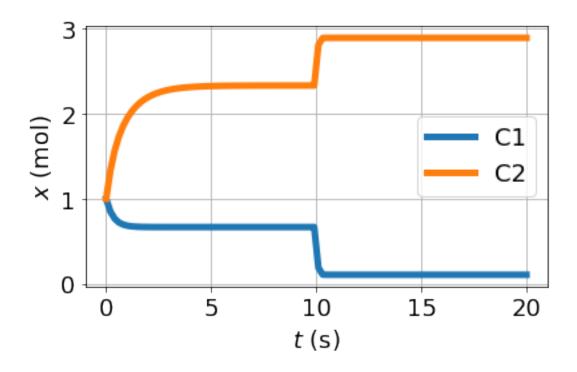
```
[13]: Q0 = 2
      t1 = 10
      f1 = 10-Q0
      t = np.linspace(0,20,100)
      Flow = f'(Q0)*(1+{f1}*np.heaviside(t-{t1},0.5))'
      #Flow = f'{Q0}'
      print(Flow)
      ## Simulation
      species = s['species']
      reaction = s['reaction']
      X0 = np.ones(s['n_X'])
      X0[species.index('A')] = 2
      parameter = {}
      Kappa = \{\}
      for unidir in [False,True]:
          if unidir:
```

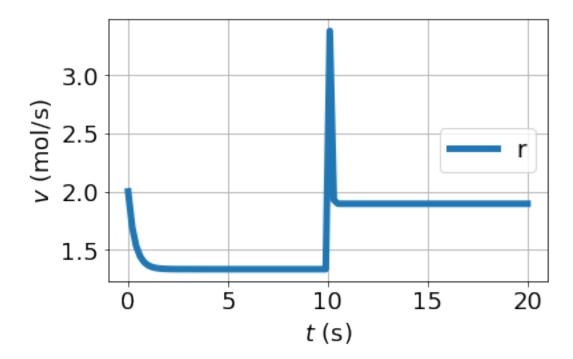
```
UniDir=['r']
    else:
        UniDir=[]
    parameter['kappa_r1'] = parameter['kappa_r2'] = 1
    parameter['kappa_r'] = Q0
    Kappa['r'] = Flow
    s = st.stoich(splitReos_abg.model(),UniDir=UniDir, quiet=quiet)
    sc = st.statify(s,chemostats=chemostats)
 →sim(s,sc=sc,t=t,parameter=parameter,Kappa=Kappa,X0=X0,quiet=quiet)
    X_old = {}
    specPlot = ['C1','C2']
    plt.rcParams.update({'font.size': 18})
   for spec in specPlot:
        x = dat['X'][:,species.index(spec)]
        plt.plot(t,x,label=spec,lw=lw)
        X_{old[spec]} = x
    plt.grid()
    plt.legend(loc='center right')
    plt.xlabel('$t$ (s)')
    plt.ylabel('$x$ (mol)')
    plt.tight_layout()
    name = f'Figs/splitReos_{unidir}'
    Savefig(name+'.pdf')
    plt.show()
    V_old = {}
    reacPlot = ['r']
    for reac in reacPlot:
        v = dat['V'][:,reaction.index(reac)]
        plt.plot(t,v,label=reac,lw=lw)
        V_old[reac] = v
    plt.grid()
    plt.legend(loc='center right')
    plt.xlabel('$t$ (s)')
    plt.ylabel('$v$ (mol/s)')
   plt.tight_layout()
    name = f'Figs/splitReos_{unidir}_v'
    Savefig(name+'.pdf')
    plt.show()
x_B_0 = dat['X'][:,species.index('B')]
     st.plot(s, dat, species=['A', 'B', 'C1', 'C2'])
      st.plot(s,dat,species=[],filename=f'{name}_v.pdf')
```

2*(1+8*np.heaviside(t-10,0.5))









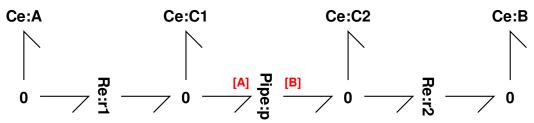
3.2 Pipe connection

```
[14]: ## Create Pipe component
N_lump = 5
V_p = 10
```

```
[15]: ## Split enzyme catalysed Reaction
    sbg.model('splitRep_abg.svg')
    import splitRep_abg
    disp.SVG('splitRep_abg.svg')
```

Creating subsystem: Pipe:p

[15]:



```
[16]: ##Stoichiometry+parameters
      parameter = {}
      Kappa = \{\}
      UniDir = []
      for i in range(N_lump+1):
          reac = f'p_r_{i}'
          UniDir.append(reac)
          kap = f'kappa_{reac}'
          parameter[kap] = Q0
          Kappa[reac] = Flow
      s = st.stoich(splitRep_abg.model(),UniDir=UniDir,quiet=quiet)
      species = s['species']
      sc = st.statify(s,chemostats=chemostats)
      ## Initial condition
      X0 = np.ones(s['n_X'])
      X0[species.index('A')] = 2
      ## Pipe parameters
      small = 1e-6
      if N_lump>0:
```

```
V_i = V_p/N_lump # Volume of lump
    for i in range(N_lump):
        spec = f'p_IS{i+1}'
        K = f'K_{spec}'
        parameter[K] = 1/V_i
        X0[species.index(spec)] = small
# print(parameter)
# print(X0)
# print(parameter)
# print(UniDir)
print(Kappa)
disp.Latex(st.sprintrl(s,chemformula=True))
```

 ${'p_r_0': '2*(1+8*np.heaviside(t-10,0.5))', 'p_r_1': }$ 2*(1+8*np.heaviside(t-10,0.5))', p_r_2' : 2*(1+8*np.heaviside(t-10,0.5))', $'p_r_3': '2*(1+8*np.heaviside(t-10,0.5))', 'p_r_4':$ $2*(1+8*np.heaviside(t-10,0.5))', 'p_r_5': '2*(1+8*np.heaviside(t-10,0.5))'$

[16]:

$$C_1 \xrightarrow{p_{r0}} p_I S_1 \tag{29}$$

$$p_{I}S_{1} \xrightarrow{p_{r1}} p_{I}S_{2} \tag{30}$$

$$p_{I}S_{2} \xrightarrow{p_{r2}} p_{I}S_{3} \tag{31}$$

$$p_{I}S_{3} \xrightarrow{p_{r3}} p_{I}S_{4}$$

$$(32)$$

$$p_{1}S_{4} \xrightarrow{p_{r4}} p_{1}S_{5}$$

$$p_{1}S_{5} \xrightarrow{p_{r5}} C_{2}$$

$$A \xrightarrow{r_{1}} C_{1}$$

$$(32)$$

$$(33)$$

$$(34)$$

$$p_{r}S_{5} \xrightarrow{p_{r5}} C_{2} \tag{34}$$

$$A \stackrel{r_1}{\longleftarrow} C_1 \tag{35}$$

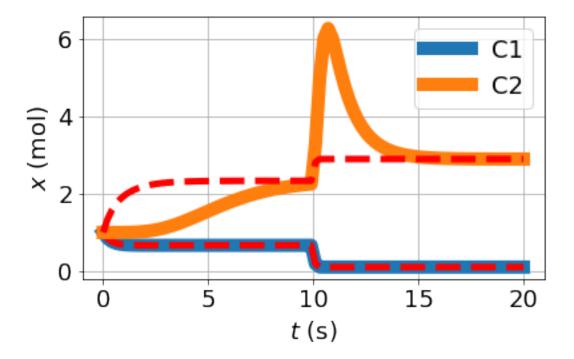
$$C_2 \stackrel{r_2}{\longleftrightarrow} B$$
 (36)

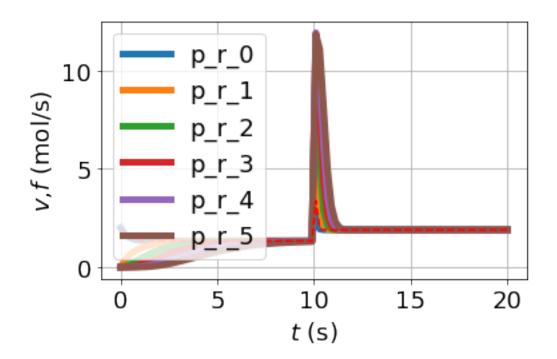
```
[17]: | ## Simulate
      dat = st.sim(s,sc=sc,t=t,parameter=parameter,X0=X0,Kappa=Kappa,quiet=quiet)
      plt.rcParams.update({'font.size': 18})
      for spec in specPlot:
          x = dat['X'][:,species.index(spec)]
          plt.plot(t,x,label=spec,lw=10)
          plt.plot(t,X_old[spec],color='red',ls='dashed',lw=5)
      plt.grid()
      plt.legend()
      plt.xlabel('$t$ (s)')
      plt.ylabel('$x$ (mol)')
      plt.tight_layout()
      Savefig('Figs/splitRep.pdf')
      plt.show()
      reaction = s['reaction']
```

```
for i in range(N_lump+1):
    reac = f'p_r_{i}'
    v = dat['V'][:,reaction.index(reac)]
    plt.plot(t,v,label=reac,lw=lw)

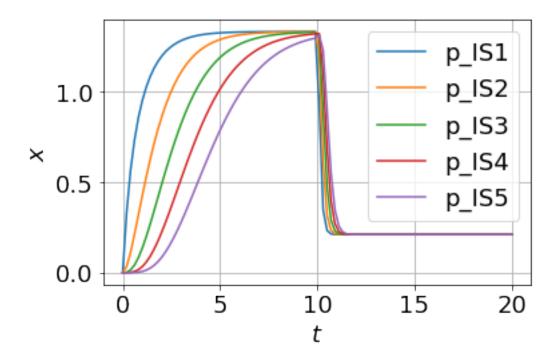
plt.plot(t,V_old['r'],color='red',ls='dashed')

plt.grid()
plt.legend()
plt.xlabel('$t$ (s)')
plt.ylabel('$v$,$f$ (mol/s)')
plt.tight_layout()
name = f'Figs/splitRep_v'
Savefig(name+'.pdf')
plt.show()
```





```
[18]: # st.plot(s,dat,reaction=[],species=['A','B','C1','C2'])
[19]: # st.plot(s,dat,species=[])
[20]: # st.plot(s,dat,reaction=[],species=['p_IS1'])
[21]: for i,spec in enumerate(species):
    if 'I' in spec:
        x = dat['X'][:,i]
        plt.plot(t,x,label=spec)
    plt.grid()
    plt.legend()
    plt.ylabel('$t$')
    plt.ylabel('$t$')
    plt.tight_layout()
    Savefig('Figs/splitRep_pipe.pdf')
```

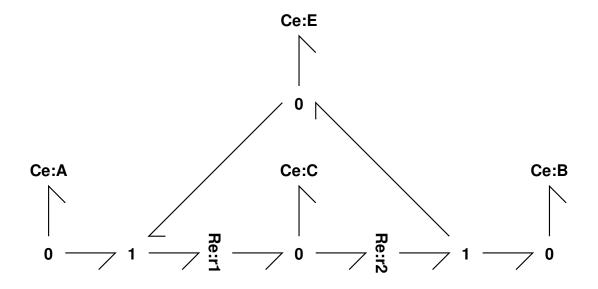


4 Enzyme-catalysed reaction

This section reprises the standard bond graph model of an enzyme-catalysed reaction - see ECR. In this case: - Ce_A represents the unbound oxygen entering the blood - Ce_B represents the unbound oxygen leaving the blood - Ce_E represents the unbound haemoglobin - Ce_C represents the haemoglobin bound to oxygen

This corresponds to a simple model of haemoglobin with one binding site (like myoglobin). A more detailed model wouPipe incorporate four cooperative binding sites - see Cooperative.

```
[22]: ## Enzyme catalysed reaction
    sbg.model('RE_abg.svg')
    import RE_abg
    disp.SVG('RE_abg.svg')
[22]:
```



```
[23]: ##Stoichiometry
s = st.stoich(RE_abg.model(),quiet=quiet)
chemostats = ['A','B']
sc = st.statify(s,chemostats=chemostats)
disp.Latex(st.sprintrl(s,chemformula=True))
```

[23]:

$$A + E \stackrel{r_1}{\rightleftharpoons} C \tag{37}$$

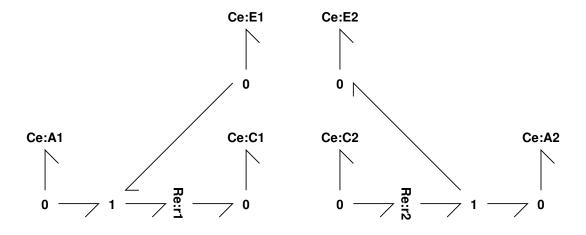
$$C \stackrel{\mathbf{r}_2}{\Longleftrightarrow} B + E \tag{38}$$

4.1 Split enzyme-catalysed reaction

In this bond graph, the components Ce_E and Ce_E are split: advection carries the coreponding molecules from one side to the other. An explicit bond graph for this coupling is provided below for tghe case were the advection time-delay is zero; the bond graph for the general case has not yet been determined.

```
[24]: ## Split enzyme catalysed reaction
sbg.model('splitRE_abg.svg')
import splitRE_abg
disp.SVG('splitRE_abg.svg')
```

[24]:



```
[25]: ##Stoichiometry
ss = st.stoich(splitRE_abg.model(),quiet=quiet)
chemostats = ['A1','A2']
ssc = st.statify(ss,chemostats=chemostats)
disp.Latex(st.sprintrl(ss,chemformula=True))
```

[25]:

$$A_1 + E_1 \xrightarrow{r_1} C_1$$

$$C_2 \xrightarrow{r_2} A_2 + E_2$$

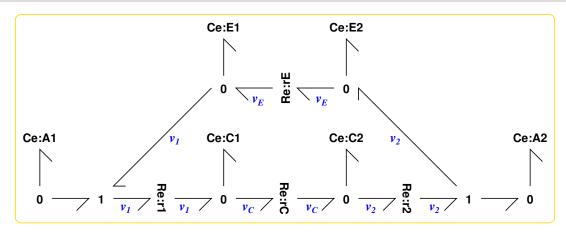
$$(39)$$

5 Circular advection

5.1 Orifice (Symmetric case): Bond graph model

```
[26]: ## Split enzyme catalysed reaction with symmetric connection
    sbg.model('splitREos_abg.svg')
    import splitREos_abg
    disp.SVG('splitREos_abg.svg')
```

[26]:



```
[27]: ##Stoichiometry
snd = st.stoich(splitREos_abg.model(),quiet=quiet)
chemostats = ['A1','A2']
sndc = st.statify(snd,chemostats=chemostats)
print(snd['species'])
print(snd['reaction'])
disp.Latex(st.sprintrl(snd,chemformula=True))
['A1', 'A2', 'C1', 'C2', 'E1', 'E2']
['r1', 'r2', 'rC', 'rE']
```

[27]:

$$A_1 + E_1 \stackrel{r_1}{\longleftarrow} C_1 \tag{41}$$

$$C_2 \stackrel{\mathbf{r}_2}{\longleftrightarrow} A_2 + E_2 \tag{42}$$

$$C_1 \stackrel{rC}{\longleftrightarrow} C_2 \tag{43}$$

$$E_2 \stackrel{rE}{\rightleftharpoons} E_1 \tag{44}$$

5.1.1 Simulate

Note that – the final steady state of the two cases d=0 and d>0 are the same – if the simulation is repeated with $f_0=100$, the $x_{C1}\approx x_{C2}$ and $x_{E1}\approx x_{E2}$ as predicted.

The two chemostats corresponding to A1 and A2 are:

$$x_{A1} = \begin{cases} 1 & \text{if } t < 1\\ 2 & \text{if } t \ge 1 \end{cases} \tag{45}$$

$$x_{A2} = 1 \tag{46}$$

```
[28]: ## Simulate split ECR with no delay
      i_{chemo} = []
      for chemo in chemostats:
          i_chemo.append(snd['species'].index(chemo))
      # print(i_chemo)
      ## Time
      tt = np.linspace(0, 20,10000)
      ## Parameters
      parameter = {}
      parameter['kappa_rC'] = Q0
      parameter['kappa_rE'] = Q0
      \# parameter['K_E'] = 0.5
      # parameter['K_C'] = 0.5
      ## Chemostats
      t_0 = 0
      X_{\text{chemo}} = \{ 'A1':f'1 + np.heaviside(t-\{t_0\},1)', 'A2':'1' \}
      ## Simulate
```

```
dat = st.sim(snd,sc=sndc,t=tt,parameter=parameter,X_chemo=X_chemo,quiet=False)

XXO = dat['X'][:,2:6]

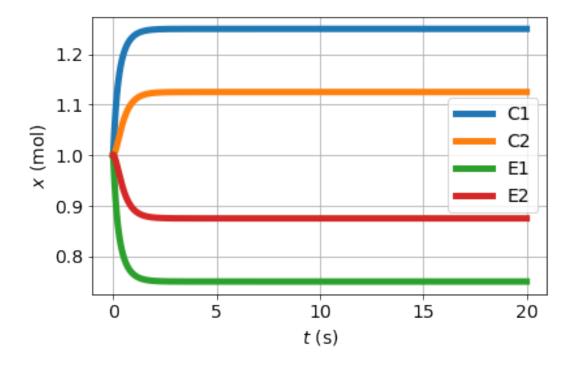
Legend = snd['species'][2:6]

VO = dat['V'][:,0:2]
dXO = dat['dX'][:,i_chemo]
```

Setting kappa_rC to 2 Setting kappa_rE to 2

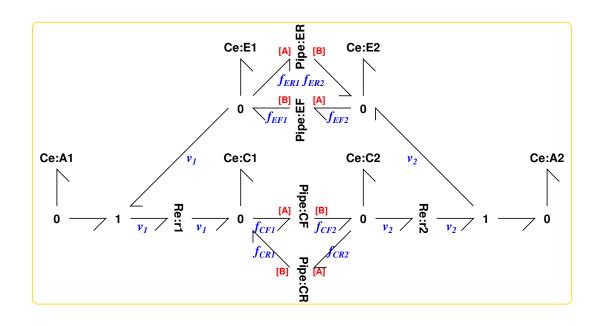
```
[29]: ## Plot
plt.rcParams.update({'font.size': 14})
## Orifice
plt.plot(tt,XXO,lw=lw)

plt.legend(Legend)
plt.grid()
plt.xlabel('$t$ (s)')
plt.ylabel('$x$ (mol)')
plt.tight_layout()
Savefig('Figs/OrificeSim.pdf')
```



5.2 Circular advection: pipe connection

```
[30]: disp.SVG('splitREp_abg.svg')
[30]:
```



```
[31]: def CircularAdvection(N_lump=5,Flow=f'{Q0}', quiet=True):
          ## Create Pipe component
          ## Note: N_lump = 0 corresponds to orifice case.
          Pipe = mbg.chain(AB_abg.model(),inport = 'A',outport = 'B',
       →N=N_lump+1,rename_components=True,Comps=['R'],quiet=quiet)
          s = st.stoich(Pipe,UniDir=None, quiet=quiet)
          s['name'] = 'Pipe_abg'
          stbg.model(s)
          ## Set uni directional Re components
          UniDir = []
          for i in range(N_lump+1):
              for comp in ['EF','ER','CF','CR']:
                  ud = f'\{comp\}_r_{i}'
                  UniDir.append(ud)
          # print(UniDir)
          ## Create stoichiometry
          sbg.model('splitREp_abg.svg')
          import Pipe_abg
          imp.reload(Pipe_abg)
          import splitREp_abg
          imp.reload(splitREp_abg)
          sPipe = st.stoich(splitREp_abg.model(),UniDir=UniDir,quiet=quiet)
          if not quiet:
              disp.Latex(st.sprintrl(sPipe,chemformula=True))
          ## Setup parameters
          parameter = {}
```

```
Kappa = \{\}
    ## Assume K=KK is same for all species
    ## Note that in the simulation code K corresponds to KK/V_i
    KK = 1
    for i in range(N_lump+1):
        for comp in ['EF', 'ER', 'CF', 'CR']:
            kap = f'\{comp\}_r_{i}'
            Kappa[kap] = f'({Flow})/{KK}'
            \#parameter[kap] = Q0
            if i>0:
                V_i = V_p/N_lump
                KO = KK/V_i
                KOF = KO
                KOR = KO
                K = f'K_{comp}_{IS}{i}'
                if 'F' in comp:
                    parameter[K] = KOF
                else:
                    parameter[K] = KOR
    ## Setup initial states
    XO = []
    small = 1e-6
    for spec in sPipe['species']:
         print(spec)
        if 'IS' in spec:
            K = f'K_{spec}'
            Xi = 1/parameter[K]
              Xi = small
#
        else:
            Xi = 1
        X0.append(Xi)
    X0 = np.array(X0)
    print(Kappa)
     print(sPipe['reaction'])
    ## Simulate
    chemostats=['A1','A2']
    i_chemo = []
    for chemo in chemostats:
        i_chemo.append(sPipe['species'].index(chemo))
    # print(i_chemo)
    sPipec = st.statify(sPipe,chemostats=chemostats)
    dat = st.
 →sim(sPipe,sc=sPipec,t=tt,parameter=parameter,Kappa=Kappa,X0=X0,X_chemo=X_chemo,quiet=quiet
    return sPipe,dat,i_chemo
```

```
[32]: #Orifice (Pipe with no lumps)
N_lump = 0
sPipe0,dat0,i_chemo0 = CircularAdvection(N_lump=N_lump)

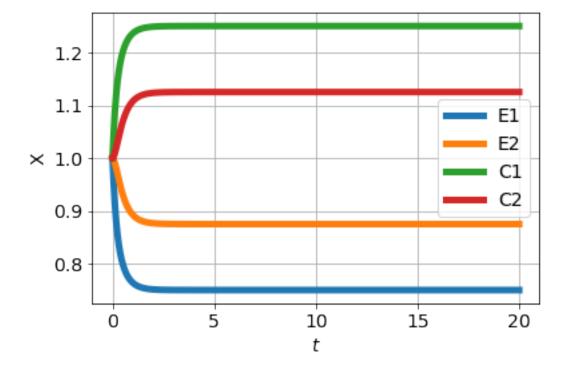
print('N_lump =', N_lump)

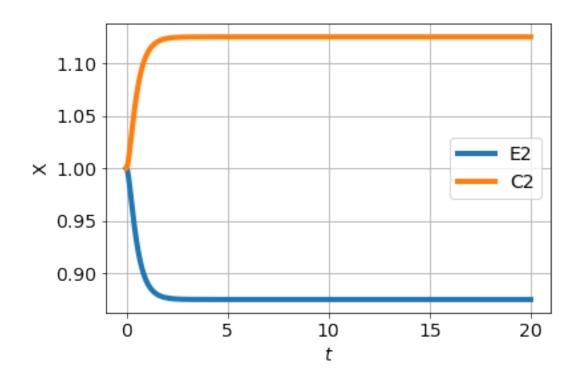
plt.rcParams.update({'font.size': 14})

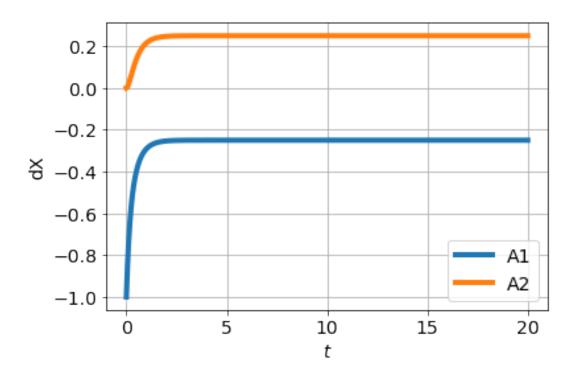
st.plot(sPipe0,dat0,species=['E1','E2','C1','C2'], reaction = [],lw=5)
#st.plot(sPipe,dat,species=[], reaction = ['PipeEF_r_0','PipeCF_r_0'])
st.plot(sPipe0,dat0,species=['E2','C2'], reaction = [])
st.plot(sPipe0,dat0,dX=True,species=['A1','A2'], reaction = [])
```

Creating subsystem: Pipe:CF Creating subsystem: Pipe:CR Creating subsystem: Pipe:EF Creating subsystem: Pipe:ER

 $N_{lump} = 0$

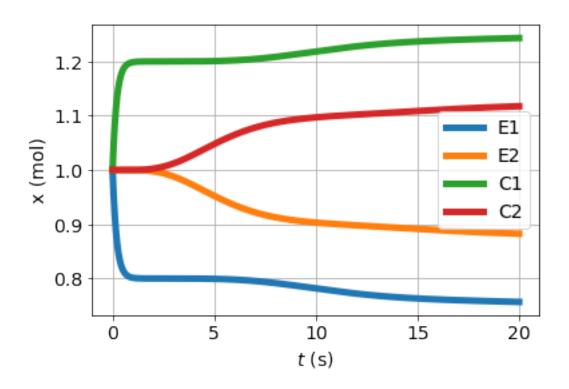




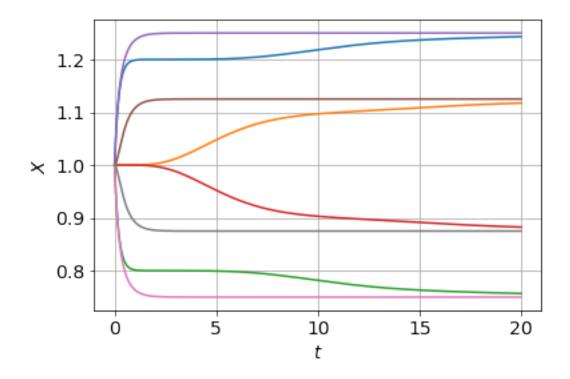


```
[33]: # ## Show final states
# final0 = dat0['X'][-1,:]
# for i,spec in enumerate(sPipe0['species']):
# print(f'Final value {spec} = {final0[i]:.2f}')
```

```
[34]: ## Show reactions
       disp.Latex(st.sprintrl(sPipe0))
[34]:
                                                     C1 \rightarrow C2
                                                                                                          (47)
                                                     C2 \rightarrow C1
                                                                                                          (48)
                                                     E2 \rightarrow E1
                                                                                                          (49)
                                                     E1 \rightarrow E2
                                                                                                          (50)
                                               A1 + E1 \Leftrightarrow C1
                                                                                                          (51)
                                                     C2 \Leftrightarrow A2 + E2
                                                                                                          (52)
[35]: ##Show flow equations
       disp.Latex(st.sprintvl(sPipe0))
[35]:
                                     v_{CFr0} = K_{C1} \kappa_{CFr0} x_{C1}
                                                                                                          (53)
                                     v_{CRr0} = K_{C2}\kappa_{CRr0}x_{C2}
                                                                                                          (54)
                                     v_{EFr0} = K_{E2} \kappa_{EFr0} x_{E2}
                                                                                                          (55)
                                     v_{ERr0} = K_{E1} \kappa_{ERr0} x_{E1}
                                                                                                          (56)
                                       v_{r1} = \kappa_{r1} (K_{A1}K_{E1}x_{A1}x_{E1} - K_{C1}x_{C1})
                                                                                                          (57)
                                       v_{r2} = \kappa_{r2} \left( -K_{A2}K_{E2}x_{A2}x_{E2} + K_{C2}x_{C2} \right)
                                                                                                          (58)
[36]: #Pipe
       N_{lump} = 5
       sPipe,dat,i_chemo = CircularAdvection(N_lump=N_lump)
       print('N_lump =', N_lump)
      Creating subsystem: Pipe:CF
      Creating subsystem: Pipe:CR
      Creating subsystem: Pipe:EF
      Creating subsystem: Pipe:ER
      N_{lump} = 5
[37]: #Pipe - plot
       imp.reload(st)
       st.plot(sPipe,dat,species=['E1','E2','C1','C2'], reaction = [],lw=lw,
                  xlabel='$t$ (s)',ylabel='x (mol)')
        #st.plot(sPipe,dat,species=[], reaction = ['PipeEF_r_0','PipeCF_r_0'])
        #st.plot(sPipe,dat,species=['E2','C2'], reaction = [])
        #st.plot(sPipe, dat, dX=True, species=['A1', 'A2'], reaction = [])
```



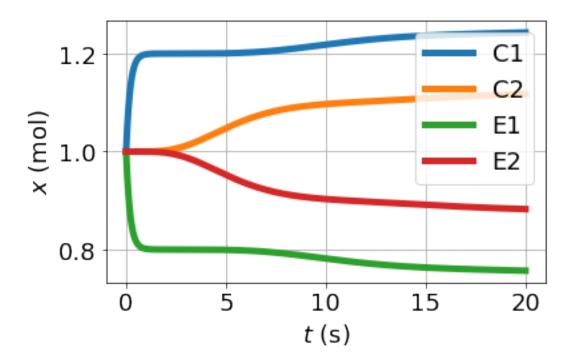
```
[38]:  # ## Show final states
      # final = dat['X'][-1,:]
      # for i,spec in enumerate(sPipe['species']):
            print(f'Final value {spec} = {final[i]:.3f}')
[39]: ## Extract data
      XXPipe = dat['X'][:,2:6]
      VPipe = dat['V'][:,[0,N_lump+1]]
      dXPipe = dat['dX'][:,i_chemo]
      dXPipe0 = dat0['dX'][:,i_chemo]
      ## Plot and compare
      ## Pipe
      plt.plot(tt,XXPipe)
      ## Orifice
      plt.plot(tt,XX0)
      plt.grid()
      plt.xlabel('$t$')
      plt.ylabel('$X$')
```



```
[40]: ## Plot

## Pipe
## Font
plt.rcParams.update({'font.size': 18})
plt.plot(tt,XXPipe,lw=lw)

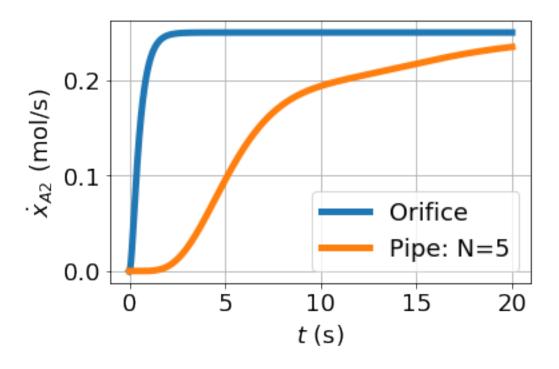
plt.legend(Legend)
plt.grid()
plt.xlabel('$t$ (s)')
plt.ylabel('$x$ (mol)')
plt.tight_layout()
Savefig('Figs/PipeSim.pdf')
```



```
[41]: ## Plot Chemostat flows
##Orifice
plt.plot(tt,dX0[:,1],label='Orifice',lw=lw)

## Pipe
#plt.plot(tt,dXPipe0[:,1],label='Pipe: N=0',linestyle='dashed',color='black')
plt.plot(tt,dXPipe[:,1],label=f'Pipe: N={N_lump}',lw=lw)

plt.grid()
plt.legend()
plt.xlabel('$t$ (s)')
plt.ylabel('$t$ (s)')
plt.ylabel('$\dot{x}_{A2}$ (mol/s)')
plt.tight_layout()
Savefig('Figs/PipeSim_flow.pdf')
```

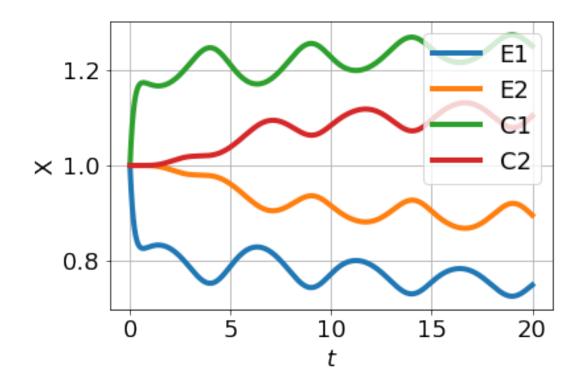


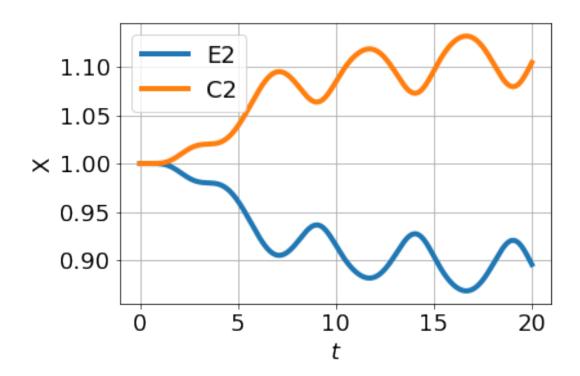
6 Variable flow rate

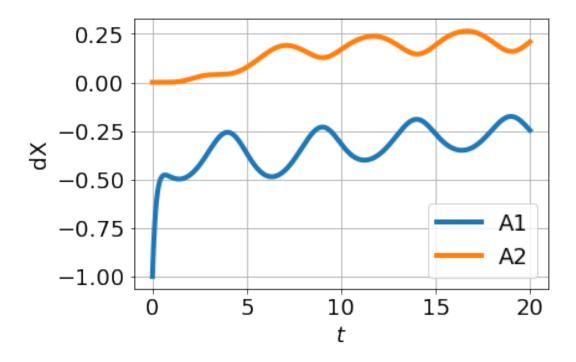
```
[42]: N_lump = 5
Period = 5
Flow = f'({Q0})*(1+0.5*np.sin(2*np.pi*t/{Period}))'
print('Flow:',Flow)
sPipe,dat,i_chemo = CircularAdvection(N_lump=N_lump,Flow=Flow)

print('N_lump =', N_lump)
st.plot(sPipe,dat,species=['E1','E2','C1','C2'], reaction = [])
#st.plot(sPipe,dat,species=[], reaction = ['PipeEF_r_0','PipeCF_r_0'])
st.plot(sPipe,dat,species=['E2','C2'], reaction = [])
st.plot(sPipe,dat,dX=True,species=['A1','A2'], reaction = [])
```

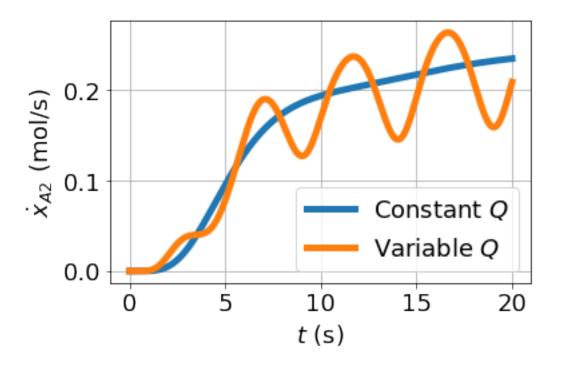
Flow: (2)*(1+0.5*np.sin(2*np.pi*t/5))
Creating subsystem: Pipe:CF
Creating subsystem: Pipe:CR
Creating subsystem: Pipe:EF
Creating subsystem: Pipe:ER
N_lump = 5





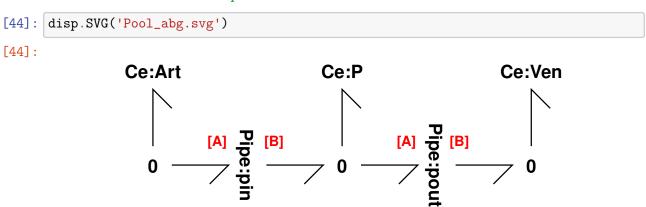


```
[43]: ## Extract data
      XXPipev = dat['X'][:,2:6]
      VPipev = dat['V'][:,[0,N_lump+1]]
      dXPipev = dat['dX'][:,i_chemo]
      ## Plot Chemostat flows
      ## Pipe
      ## Font
      plt.rcParams.update({'font.size': 18})
      plt.plot(tt,dXPipe[:,1],label='Constant $Q$',lw=lw)
      plt.plot(tt,dXPipev[:,1],label='Variable $Q$',lw=lw)
      \#plt.plot(tt,Q/4,ls='dashed',label='\$Q/4\$')
      plt.grid()
      plt.legend()
      plt.xlabel('$t$ (s)')
      plt.ylabel('\$\dot\{x\}_{A2}\$ (mol/s)')
      plt.tight_layout()
      Savefig('Figs/PipeSim_flow_vary.pdf')
```

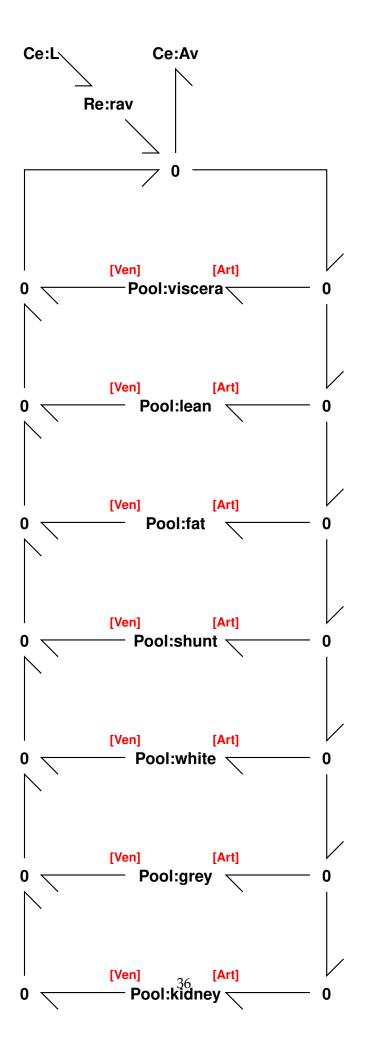


7 Pharmacokinetics

The data is taken from Mapleson (1973).



```
[45]: disp.SVG('PK_abg.svg')
[45]:
```



```
[46]: def Pool(N_lump=0, quiet=True):
          ## Create Pipe component
          ## Note: N_lump = 0 corresponds to orifice case.
          Pipe = mbg.chain(AB_abg.model(),inport = 'A',outport = 'B',
       →N=N_lump+1, rename_components=True, Comps=['R'], quiet=quiet)
          s = st.stoich(Pipe,UniDir=None, quiet=quiet)
          s['name'] = 'Pipe_abg'
          stbg.model(s)
          ## Set unidirectional Re components
          UniDir = []
          for i in range(N_lump+1):
              for comp in ['pin','pout']:
                  ud = f'\{comp\}_r_{i}'
                  UniDir.append(ud)
          # print(UniDir)
          ## Create stoichiometry
          sbg.model('Pool_abg.svg')
          import Pipe_abg
          imp.reload(Pipe_abg)
          import Pool_abg
          imp.reload(Pool_abg)
          sPool = st.stoich(Pool_abg.model(),UniDir=UniDir,quiet=quiet)
          if not quiet:
              disp.Latex(st.sprintrl(sPool,chemformula=True))
          stbg.model(sPool,filename='Pool_abg')
          return UniDir
      def setPool(N_lump=0,Flow=f'{Q0}', quiet=True):
          ## Setup parameters
          parameter = {}
          Kappa = \{\}
          ## Assume K=KK is same for all species
          ## Note that in the simulation code K corresponds to KK/V_i
          KK = 1
          for i in range(N_lump+1):
              for comp in ['pin','pout']:
                  kap = f'\{comp\}_r_{i}'
```

```
Kappa[kap] = f'({Flow})/{KK}'
                  \#parameter[kap] = Q0
                  if i>0:
                      V_i = V_p/N_lump
                      KO = KK/V_i
                      KOF = KO
                      KOR = KO
                      K = f'K_{comp}_{IS\{i\}'}
                      if 'f' in comp:
                          parameter[K] = KOF
                      else:
                          parameter[K] = KOR
          ## Setup initial states
          XO = []
          for spec in sPool['species']:
                print(spec)
              if 'IS' in spec:
                  K = f'K_{spec}'
                  Xi = 1/parameter[K]
              else:
              X0.append(Xi)
          X0 = np.array(X0)
          return parameter, XO
[47]: N_{lump} = 5
      UniDir = Pool(N_lump=N_lump)
      import Pool_abg
      imp.reload(Pool_abg)
      sPool = st.stoich(Pool_abg.model(),UniDir=UniDir,quiet=quiet)
      disp.Latex(st.sprintrl(sPool,chemformula=True))
     Creating subsystem: Pipe:pin
     Creating subsystem: Pipe:pout
[47]:
```

$$Art \xrightarrow{pin_{r0}} pin_{r}S_{1}$$
 (59)

$$pin_{I}S_{1} \xrightarrow{pin_{r1}} pin_{I}S_{2}$$
 (60)

$$pin_{I}S_{2} \xrightarrow{pin_{r2}} pin_{I}S_{3}$$
 (61)

$$pin_{I}S_{3} \xrightarrow{pin_{r3}} pin_{I}S_{4}$$
 (62)

$$pin_{I}S_{4} \xrightarrow{pin_{r4}} pin_{I}S_{5}$$
 (63)

$$pin_{I}S_{5} \xrightarrow{pin_{r5}} P$$
 (64)

$$P \xrightarrow{pout_{r0}} pout_{I}S_{1}$$
 (65)

$$pout_{I}S_{1} \xrightarrow{pout_{r1}} pout_{I}S_{2}$$
(66)

$$pout_{I}S_{2} \xrightarrow{pout_{r_{2}}} pout_{I}S_{3}$$

$$pout_{r_{3}}$$

$$pout_{r_{3}}$$

$$pout_{r_{3}}$$

$$pout_{r_{3}}$$

$$pout_{r_{3}}$$

$$pout_{r_{3}}$$

$$pout_{r_{3}}$$

$$pout_{r_{3}}$$

$$pout_{I}S_{3} \xrightarrow{pout_{r3}} pout_{I}S_{4}$$
 (68)

$$pout_{I}S_{4} \xrightarrow{pout_{r_{4}}} pout_{I}S_{5}$$

$$pout_{r_{5}} \qquad (69)$$

$$pout_{I}S_{5} \xrightarrow{pout_{r5}} Ven$$
 (70)

```
[48]: import Pool_abg
      imp.reload(Pool_abg)
      sbg.model('PK_abg.svg')
      import PK_abg
      imp.reload(PK_abg)
      pools = ['viscera','lean','fat','shunt','white','grey','kidney']
      PK_UniDir = []
      for pool in pools:
          for ud in UniDir:
              PKud = f'{pool}_{ud}'
              PK_UniDir.append(PKud)
      # print(PK_UniDir)
      sPK = st.stoich(PK_abg.model(),UniDir=PK_UniDir, quiet=quiet)
```

Creating subsystem: Pool:fat Creating subsystem: Pool:grey Creating subsystem: Pool:kidney Creating subsystem: Pool:lean Creating subsystem: Pool:shunt Creating subsystem: Pool:viscera Creating subsystem: Pool:white

```
[49]: # disp.Latex(st.sprintrl(sPK,chemformula=True))
```

[50]: # disp.Latex(st.sprintvl(sPK))

7.1 Parameters: Map78 Table I

```
[51]: ## Parameters: Volume - litre
      Volume={}
      Volume['viscera'] = 6.2
      Volume['lean'] = 39.2
      Volume['fat'] = 12.2
      Volume['shunt'] = 1e-6
      Volume['white'] = 0.0007
      Volume['grey'] = 0.0007
      Volume['kidney'] = 0.0007
      Volume['arterial'] = 1.4
      Volume['venous'] = 4.0
[52]: # ## Parameters: Flow (OLD)
      # FlowFraction={}
      # FlowFraction['viscera'] = 0.63
      # FlowFraction['lean'] = 0.131
      # FlowFraction['fat'] = 0.04
      # FlowFraction['shunt'] = 0.199
      # FlowFraction['white'] = 0.000022
      # FlowFraction['grey'] = 0.000086
      # FlowFraction['kidney'] = 0.000432
[53]: ## Parameters: Flow (New)
      FlowFraction={}
      FlowFraction['viscera'] = 0.399
      FlowFraction['lean'] = 0.364
      FlowFraction['fat'] = 0.111
      FlowFraction['shunt'] = 0.126
      FlowFraction['white'] = 0.000014
      FlowFraction['grey'] = 0.000055
      FlowFraction['kidney'] = 0.000274
[54]: ## Parameters: Partition coefficient
      PartitionCoefficient = {}
      for pool in pools:
          if pool in ['fat']:
              PartitionCoefficient[pool] = 1.40
          else:
              PartitionCoefficient[pool] = 0.46
      print(PartitionCoefficient)
     {'viscera': 0.46, 'lean': 0.46, 'fat': 1.4, 'shunt': 0.46, 'white': 0.46,
     'grey': 0.46, 'kidney': 0.46}
[55]: Q = 6.48/60 \# lit/sec Table II
      Delay = \{\}
```

```
Delay_min = {}
for blood in ['arterial', 'venous']:
    Delay[blood] = Volume[blood]/Q
    Delay_min[blood] = Delay[blood]/60
    print(f'Delay ({blood}) = {Delay[blood]:.2f} sec = {Delay_min[blood]:0.
 \rightarrow 2f} min')
parameter = {}
## Lung diffusion parameters
parameter['kappa_rav'] = 0.5
parameter['K_L'] = 1
parameter['K_Av'] = 0.05
parameter['kappa_rav'] = 0.5
parameter['K_L'] = 1
parameter['K_Av'] = 0.05
## Pool parameters
for pool in pools:
    ## Pool Ce:P
    K = f'K_{pool}'
    par = 1/(Volume[pool]*PartitionCoefficient[pool])
     frac = VolumeFraction[pool]
      frac = 0.5 # FIXME
    parameter[f'{K}_P'] = par
    ## Pool pipes RA components
    for i in range(N_lump+1):
        for inout in ['in','out']:
            kappa = f'kappa_{pool}_p\{inout\}_r_{i}'
            parameter[kappa] = Q*FlowFraction[pool]
    ## Pool pipes C components
    if N_lump>0:
        for i in range(N_lump):
            for inout in ['in','out']:
                K = f'K_{pool}_p\{inout\}_IS\{i+1\}'
                if inout in ['in']:
                    V_i = FlowFraction[pool]*Volume['arterial']/N_lump
                    V_i = FlowFraction[pool]*Volume['venous']/N_lump
                parameter[K] = (1/V_i)
#print(parameter)
```

Delay (arterial) = 12.96 sec = 0.22 min

```
Delay (venous) = 37.04 \text{ sec} = 0.62 \text{ min}
```

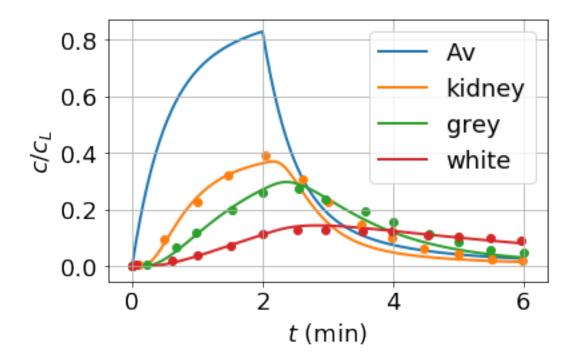
7.2 Simulation

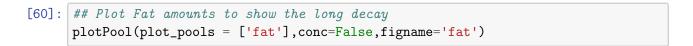
```
[56]: ## Simulation
      t = np.linspace(0,360,1000)
      t = np.linspace(0,360,100)
      t_ss = np.linspace(0,10000,1000)
      t_1 = 120
      ## Initial conditions
      x_min = 1e-6
      X0 = x_{\min} \cdot p.ones(sPK['n_X'])
      ## Chemostats
      chemostats = ['L']
      scPK = st.statify(sPK,chemostats=chemostats)
      X_{\text{chemo}} = \{'L': f'\{x_{\min}\} + 1*(t<\{t_{1}\})'\}
      # print(X_chemo)
      ## Simulate - steady state
      dat_ss = st.sim(sPK,sc=scPK,t=t_ss,X0=X0,parameter=parameter)
      X_ss = dat_ss['X'][-1,:]
      #print(X_ss)
      ## Simulate
      dat = st.sim(sPK,sc=scPK,t=t,X0=X_ss,parameter=parameter,X_chemo=X_chemo)
[57]: | ##Extract Mapleson data
      def getCSV(name):
          \mathbf{x} = [0]
          y = [0]
          filename = name+'.csv'
          if exists(filename):
               with open(filename) as csvfile:
                   csvReader = csv.reader(csvfile, delimiter=',')
                   for row in csvReader:
                       x += [float(row[0])]
                       y += [float(row[1])]
          return np.array(x),np.array(y)
      \# X = \{\}
      \# Y = \{\}
      # x,y = getCSV('kidney')
      # plt.plot(x,y)
[58]: ## Function to plot pool data
      def
```

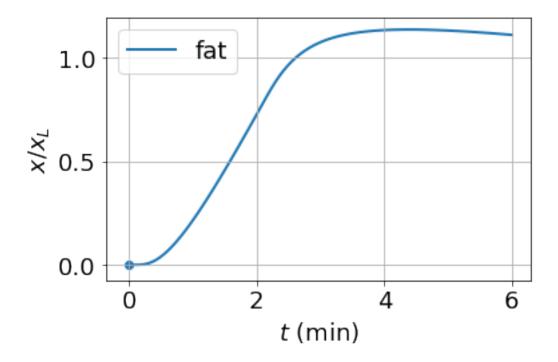
→plotPool(plot_pools=['Av','kidney','grey','white'],figname='sim',conc=True):

```
## Extract state
  species = sPK['species']
  X = dat['X']
   ## Font
  plt.rcParams.update({'font.size': 18})
   ## Compute concentration, and plot
  for pool in plot_pools:
       if pool in ['L','Av']:
           name = f'{pool}'
           par = f'K_{pool}'
           vol = 1/parameter[par]
       else:
           name = f'{pool}_P'
           vol = Volume[pool]
         print(name)
       i_pool = species.index(name)
       if conc:
           y = X[:,i_pool]/vol
       else:
           y = X[:,i_pool]
       plt.plot(t/60,y,label=pool,lw=2,zorder=0)
  for pool in plot_pools:
       t_map,y_map = getCSV(pool)
       ## Normalisation factor (units of Tension) to make data
       ## commensurate with simulation data which is normalised _{\!\!\! \sqcup}
\rightarrow concentration.
       T_L = 1000 \#mmHg
       ## Plot as dots on top of simulation data - same colours
       plt.scatter(t_map,y_map/T_L,zorder=1)
  plt.legend()
  plt.grid()
  plt.xlabel('$t$ (min)')
  if conc:
       plt.ylabel('$c/c_L$')
  else:
       plt.ylabel('$x/x_L$')
  plt.tight_layout()
  Savefig(f'Figs/PK_{figname}.pdf')
  plt.show()
```

```
[59]: ## Plot data plotPool()
```







References

W. W. Mapleson. Circulation-time models of the uptake of inhaled anaesthetics and data for quantifying them. British Journal of Anaesthesia, 45:319, 1973.