

Energy-based Advection Modelling Using Bond Graphs: Additional Material

Peter Gawthrop. peter.gawthrop@unimelb.edu.au

June 22, 2022

Contents

1	Introduction	1
1.1	Import some python code	2
2	Pipe	2
2.1	Chain of compartments	2
2.2	Transfer-function analysis	9
3	Coupled Advection and Transformation	10
3.1	Orifice connection	10
3.2	Pipe connection	14
4	Enzyme-catalysed reaction	19
4.1	Split enzyme-catalysed reaction	20
5	Circular advection	21
5.1	Orifice (Symmetric case): Bond graph model	21
5.1.1	Simulate	22
5.2	Circular advection: pipe connection	23
6	Variable flow rate	32
7	Pharmacokinetics	35
7.1	Parameters: Map78 Table I	44
7.2	Simulation	45
8	Appendix (Material not used in the paper)	48
8.1	Pure time delay simulation of symmetric case	48
8.1.1	Numerical model	48
8.2	Simulation	49
8.2.1	Compare $d=0$ and $d>0$	50
8.3	Numerical computation of chemical/hydraulic interaction (early version)	53

1 Introduction

This notebook generates the figures for the paper: *Energy-based Advection Modelling Using Bond Graphs*. This is a tidied and extended version of the 2021 version.

1.1 Import some python code

The bond graph analysis uses a number of Python modules:

```
[1]: import numpy as np
import matplotlib.pyplot as plt
import IPython.display as disp

## Control toolbox
import control as con
## Bond graph tools
import BondGraphTools as bgt
print('BondGraphTools version', bgt.version)
## Stoichiometric analysis
import stoich as st
## SVG bg representation conversion
import svgBondGraph as sbg
## Modular bond graphs
import modularBondGraph as mbg
## Stoichiometric conversion
import stoichBondGraph as stbg
## Allow reloading of modules
import importlib as imp

## Set quiet=False for verbose output
quiet = True

## Plot parameters
lw = 5
```

Warning - scikit.odes not found. Simulations are disabled.
BondGraphTools version 0.4.6

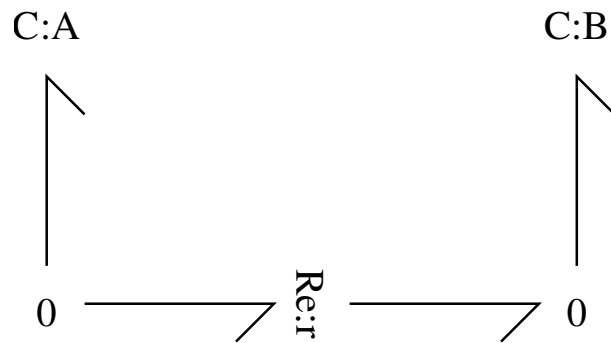
2 Pipe

2.1 Chain of compartments

This is the lumped model of advection considered in Section 3.1. of the paper. Using the modular extension to BGT, a chain of simple $A \rightleftharpoons B$ reactions of length N is created.

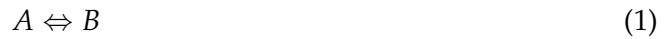
```
[2]: ## Simple A <=> B reaction
sbg.model('AB_abg.svg')
import AB_abg
disp.SVG('AB_abg.svg')
```

[2]:



```
[3]: s0 = st.stoich(AB_abg.model(),quiet=quiet)
     disp.Latex(st.sprintrl(s0))
```

[3]:



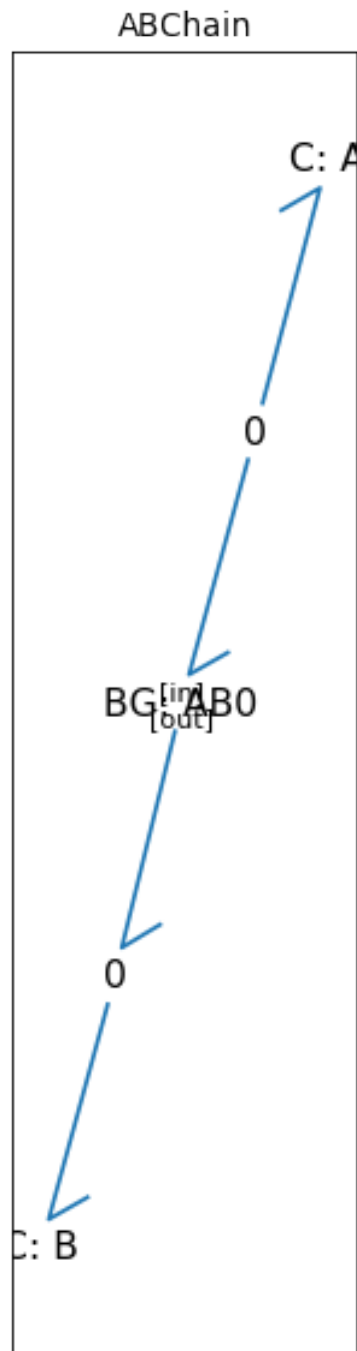
```
[4]: chain = mbg.chain(AB_abg.model(),inport = 'A',outport = 'B',
                       N=1,rename_components=False,quiet=False)
     ss0 = st.stoich(chain,quiet=quiet)
     disp.Latex(st.sprintrl(ss0))
```

Exposing A and B
 Creating ABChain
 Creating ABO from AB within ABChain
 Creating A from A within ABChain
 Creating jA from 013 within ABChain
 Creating B from A within ABChain
 Creating jB from 013 within ABChain

[4]:



```
[5]: bgt.draw(chain)
```



```
[6]: ## Vary number of lumps
N_lumps = [1,5,10,20]
#N_lumps = [1]
Q0 = 2
V_p = 10
delay = V_p/Q0

## Time
t = np.linspace(0,2*delay,100)
```

```

plt.rcParams.update({'font.size': 14})

## Create pipe model Pipe for various numbers of lumps
for N_lump in N_lumps:
    Pipe = mbg.chain(AB_abg.model(),inport = 'A',outport = 'B',
                    ␣
    →N=N_lump+1,rename_components=True,Comps=['R'],quiet=quiet)
    V_i = V_p/N_lump # Volume of lump
    parameter = {}
    UniDir = []
    for i in range(N_lump+1):
        reac = f'r_{i}'
        UniDir.append(reac)
        kap = f'kappa_{reac}'
        parameter[kap] = Q0

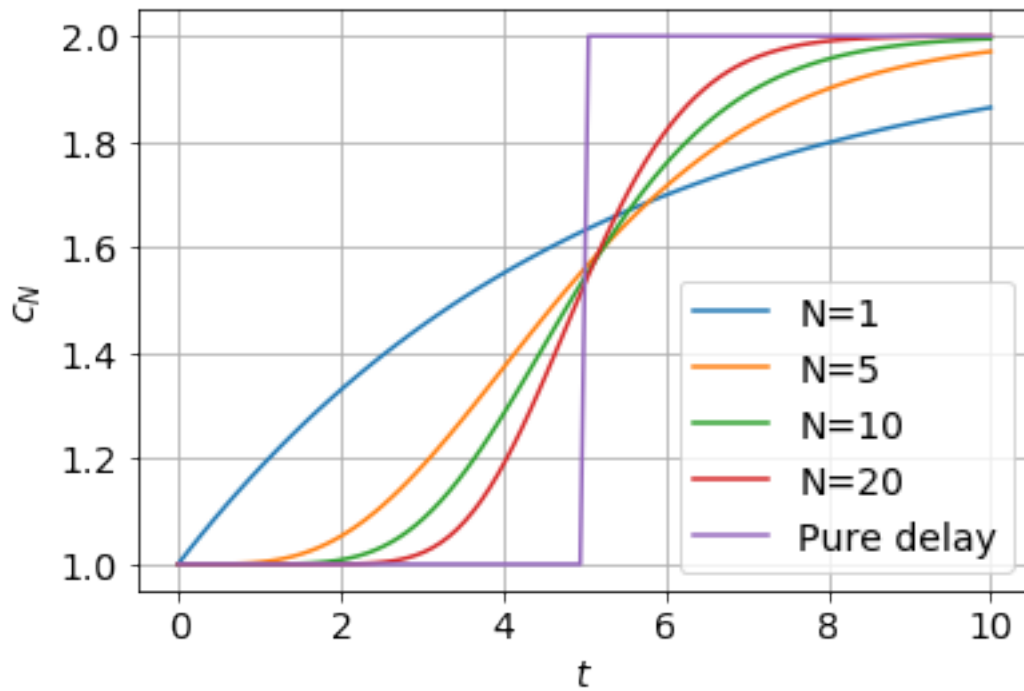
    s = st.stoich(Pipe,UniDir=UniDir, quiet=quiet)
    sc = st.statify(s,chemostats=['A','B'])
    species = s['species']
    disp.Latex(st.sprintrl(s))
    X0 = np.ones(s['n_X'])
    X0[species.index('A')] = 2
    for i in range(N_lump):
        spec = f'IS{i+1}'
        K = f'K_{spec}'
        parameter[K] = 1/V_i
        X0[species.index(spec)] = V_i

    dat = st.sim(s,sc=sc,t=t,parameter=parameter,X0=X0,quiet=quiet)
    r_N = f'r_{N_lump}'
    i_N = species.index(f'IS{N_lump}')
    x_N = dat['X'][:,i_N]

    plt.plot(t,x_N/V_i,label=f'N={N_lump}')

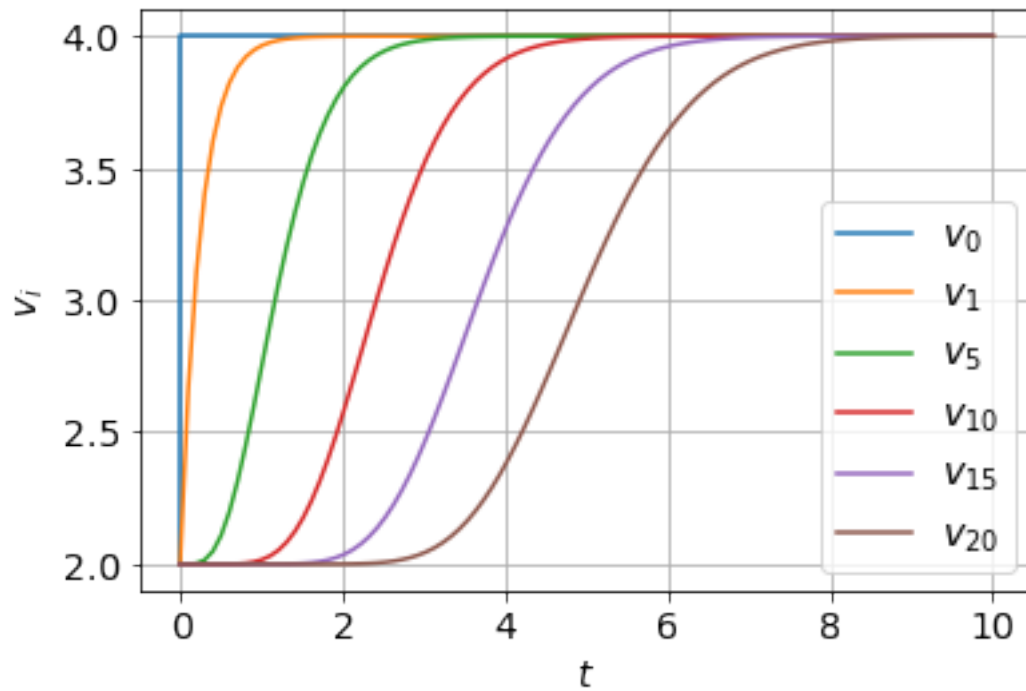
## Pure delay
y_del = 1+np.heaviside(t-delay,1)
plt.plot(t,y_del,label='Pure delay')
plt.grid()
plt.legend()
plt.xlabel('$t$')
plt.ylabel('$c_N$')
plt.savefig('Figs/Pipe.pdf')

```



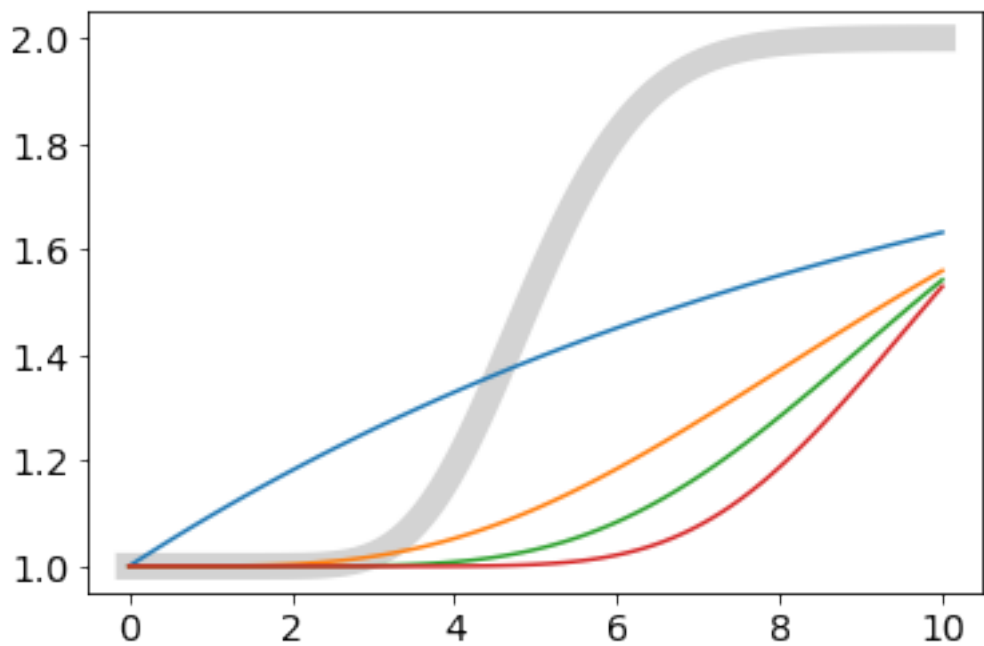
```
[7]: V = dat['V']
reaction = s['reaction']
I = [0,1,5,10,15,20]
for i, reac in enumerate(reaction):
    if i in I:
        v = V[:,i]
        if i == 0:
            vv = np.hstack((2,v))
            tt = np.hstack((0,t))
        else:
            tt = t
            vv = v
        vi = 'v_{' + f'{i}' + '}'
        plt.plot(tt,vv,label=f'${vi}$')

plt.legend()
plt.grid()
plt.xlabel('$t$')
plt.ylabel('$v_i$')
plt.show()
plt.savefig('Figs/Pipe_v.pdf')
```



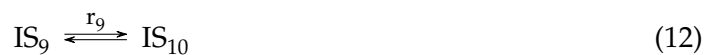
<Figure size 432x288 with 0 Axes>

```
[8]: ## Compare with a constructed transfer function  $1/(s+sT/N)^N$ 
plt.plot(t,x_N/V_i,lw=10,color='lightgrey')
for N_lump in N_lumps:
    V_i = V_p/N_lump
    lump = con.tf(1,[V_i,1])
    lumps = con.tf(1,1)
    for i in range(N_lump):
        lumps = con.series(lumps,lump)
#     print(lumps)
t,y = con.step_response(lumps,T=t)
plt.plot(t,y+1)
```



```
[9]: ## Redo for particular case
N_lump = 10
UniDir = []
for i in range(N_lump+1):
    reac = f'r_{i}'
Pipe = mbg.chain(AB_abg.model(),inport = 'A',outport = 'B',
                 N=N_lump+1,rename_components=True,Comps=['R'],quiet=quiet)
s = st.stoich(Pipe,UniDir=None, quiet=quiet)
s['name'] = 'Pipe_abg'
stbg.model(s)
disp.Latex(st.sprintrl(s,chemformula=True))
```

[9]:





2.2 Transfer-function analysis

This chain of reactions has liner dynamics which are extracted to give a model in the Python Control Toolbox format. The effective timeconstant is computed from:

$$\tau = \sum_{i=1}^N 1/p_i \quad (14)$$

where p_i is the i th pole of the tranfer function relating the input and output of the chain.

```
[10]: imp.reload(st)
      UniDir=[]
      for i in range(N_lump+1):
          UniDir.append(f'r_{i}')
      print(UniDir)
      s = st.stoich(Pipe,UniDir=UniDir, quiet=quiet)
      disp.Latex(st.sprintvl(s))
```

```
['r_0', 'r_1', 'r_2', 'r_3', 'r_4', 'r_5', 'r_6', 'r_7', 'r_8', 'r_9', 'r_10']
```

[10]:

$$v_{r0} = K_A \kappa_{r0} x_A \quad (15)$$

$$v_{r1} = K_{IS1} \kappa_{r1} x_{IS1} \quad (16)$$

$$v_{r2} = K_{IS2} \kappa_{r2} x_{IS2} \quad (17)$$

$$v_{r3} = K_{IS3} \kappa_{r3} x_{IS3} \quad (18)$$

$$v_{r4} = K_{IS4} \kappa_{r4} x_{IS4} \quad (19)$$

$$v_{r5} = K_{IS5} \kappa_{r5} x_{IS5} \quad (20)$$

$$v_{r6} = K_{IS6} \kappa_{r6} x_{IS6} \quad (21)$$

$$v_{r7} = K_{IS7} \kappa_{r7} x_{IS7} \quad (22)$$

$$v_{r8} = K_{IS8} \kappa_{r8} x_{IS8} \quad (23)$$

$$v_{r9} = K_{IS9} \kappa_{r9} x_{IS9} \quad (24)$$

$$v_{r10} = K_{IS10} \kappa_{r10} x_{IS10} \quad (25)$$

```
[11]: sc = st.statify(s,chemostats=['A','B'])
      sys = st.lin(s,sc)
      # print(sys)
      A,B,C,D = con.ssddata(sys)
      sys12 = con.ss(A,B[:,0],C[N_lump,:],0)
      print(con.tf(sys12))
      print(con.pole(sys12))
      tc = np.sum(-1/con.pole(sys12))
      gain = con.dcgain(sys12)
      print('Time constant =',tc)
      t_step,y_step = con.step_response(sys12,T=np.linspace(0,2*tc))
      plt.plot(t_step/tc,y_step.T/gain)
```

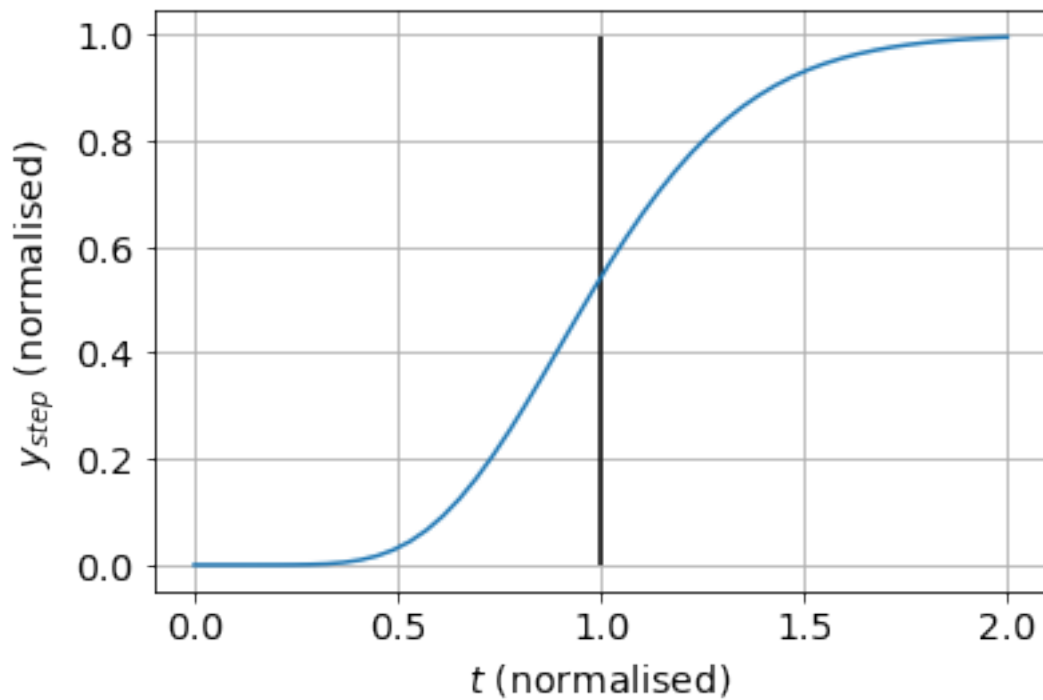
```
plt.vlines(1,0,max(y_step)/gain)
plt.grid()
plt.xlabel('$t$ (normalised)')
plt.ylabel('$y_{step}$ (normalised)')
```

1

 $s^{10} + 10 s^9 + 45 s^8 + 120 s^7 + 210 s^6 + 252 s^5 + 210 s^4 + 120 s^3 + 45 s^2 + 10 s + 1$

$[-1. -1. -1. -1. -1. -1. -1. -1. -1. -1.]$
Time constant = 10.0

[11]: Text(0,0.5,'\$y_{step}\$ (normalised)')



3 Coupled Advection and Transformation

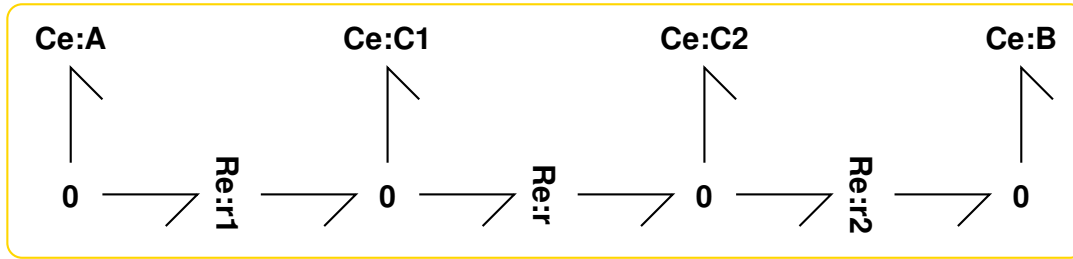
This corresponds to Section 4 of the paper.

3.1 Orifice connection

[12]:

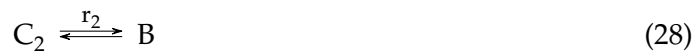
```
##
sbg.model('splitReos_abg.svg')
import splitReos_abg
disp.SVG('splitReos_abg.svg')
```

[12]:



```
[13]: ##Stoichiometry
s = st.stoich(splitReos_abg.model(),quiet=quiet)
chemostats = ['A', 'B']
sc = st.statify(s,chemostats=chemostats)
disp.Latex(st.sprintrl(s,chemformula=True))
```

[13]:



```
[14]: Q0 = 2
t1 = 10
f1 = 10-Q0
t = np.linspace(0,20,100)
Flow = f'{Q0}*(1+[f1]*np.heaviside(t-{t1},0.5))'
#Flow = f'{Q0}'
print(Flow)
## Simulation
species = s['species']
reaction = s['reaction']

X0 = np.ones(s['n_X'])
X0[species.index('A')] = 2
parameter = {}
Kappa = {}
for unidir in [False,True]:
    if unidir:
        UniDir=['r']
    else:
        UniDir=[]

    parameter['kappa_r1'] = parameter['kappa_r2'] = 1
    parameter['kappa_r'] = Q0
    Kappa['r'] = Flow

s = st.stoich(splitReos_abg.model(),UniDir=UniDir, quiet=quiet)
sc = st.statify(s,chemostats=chemostats)
```

```

    dat = st.
    →sim(s,sc=sc,t=t,parameter=parameter,Kappa=Kappa,X0=X0,quiet=quiet)

    X_old = {}
    specPlot = ['C1','C2']
    plt.rcParams.update({'font.size': 18})
    for spec in specPlot:
        x = dat['X'][:,species.index(spec)]
        plt.plot(t,x,label=spec,lw=lw)
        X_old[spec] = x
    plt.grid()
    plt.legend()
    plt.xlabel('$t$')
    plt.ylabel('$x$')
    plt.tight_layout()
    name = f'Figs/splitReos_{unidir}'
    plt.savefig(name+'.pdf')
    plt.show()

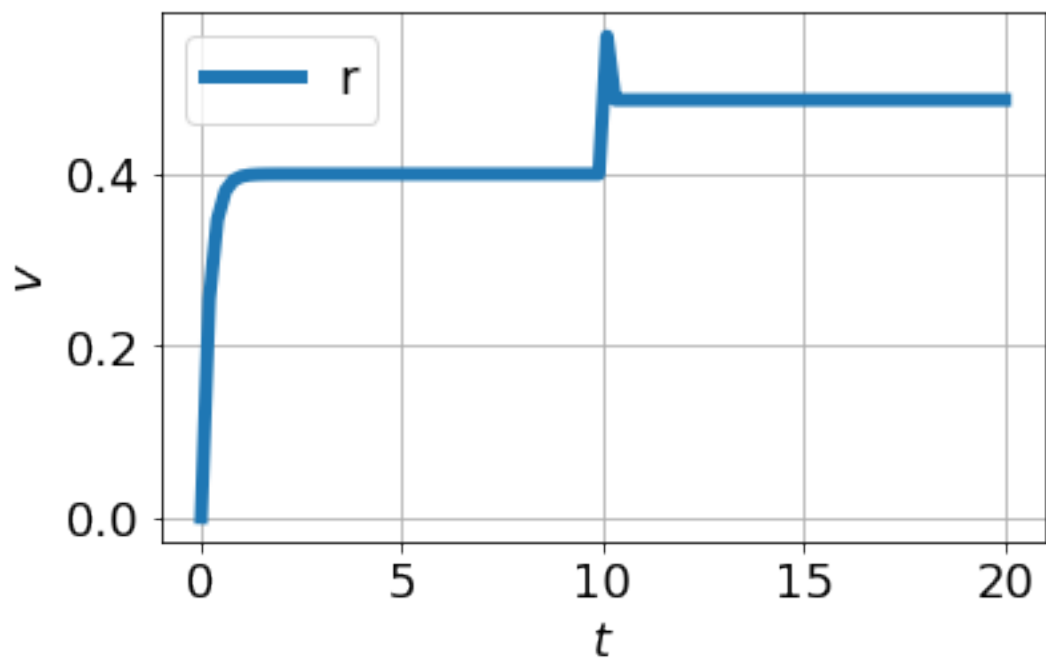
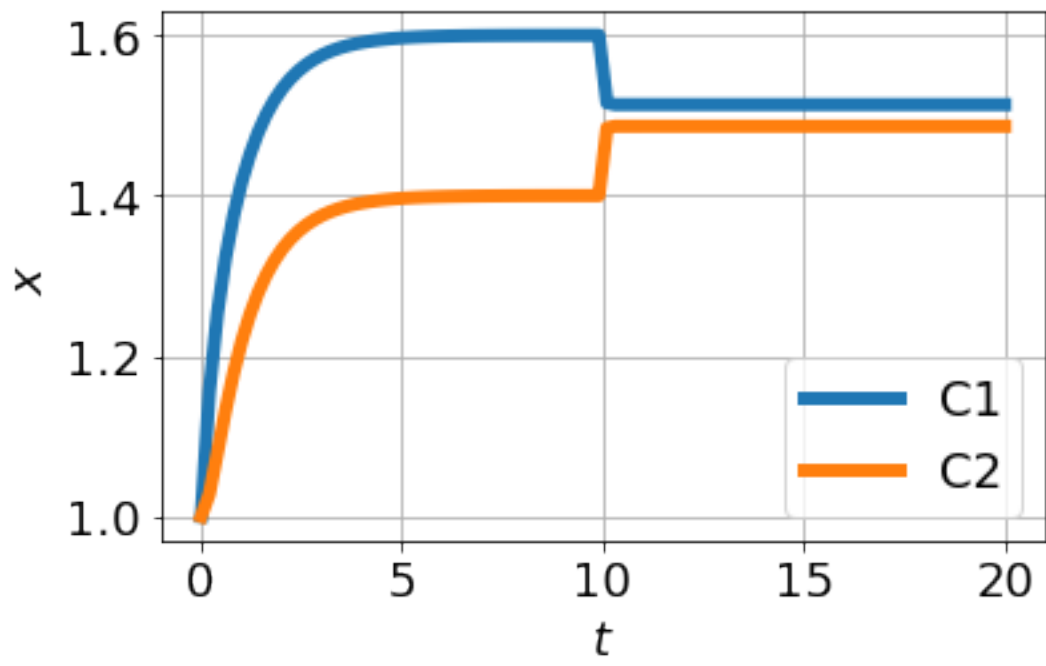
    V_old = {}
    reacPlot = ['r']
    for reac in reacPlot:
        v = dat['V'][:,reaction.index(reac)]
        plt.plot(t,v,label=reac,lw=lw)
        V_old[reac] = v
    plt.grid()
    plt.legend()
    plt.xlabel('$t$')
    plt.ylabel('$v$')
    plt.tight_layout()

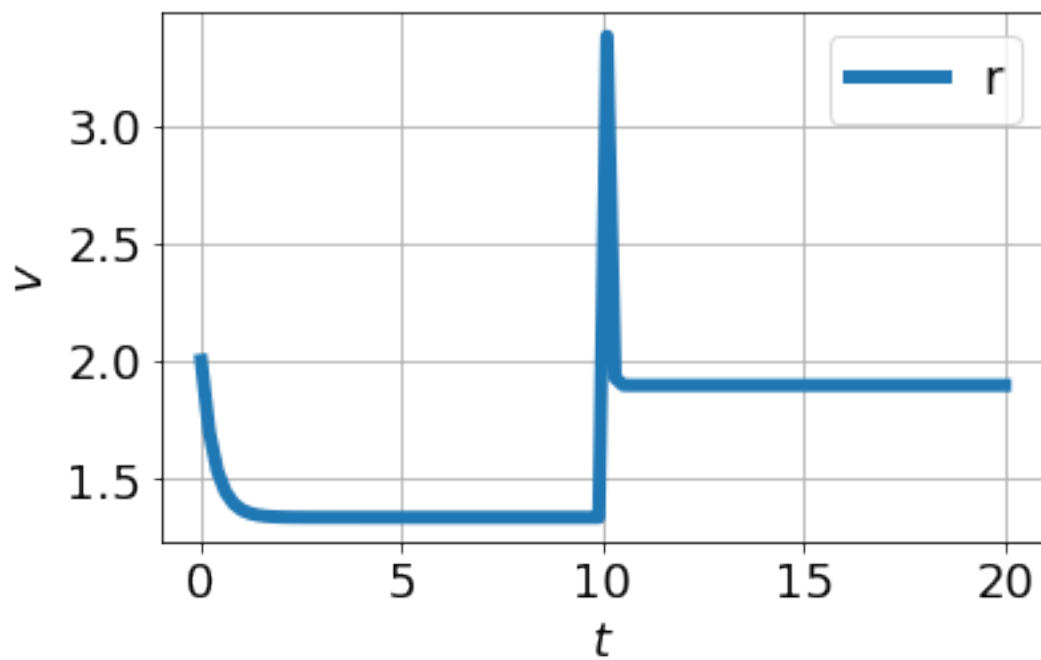
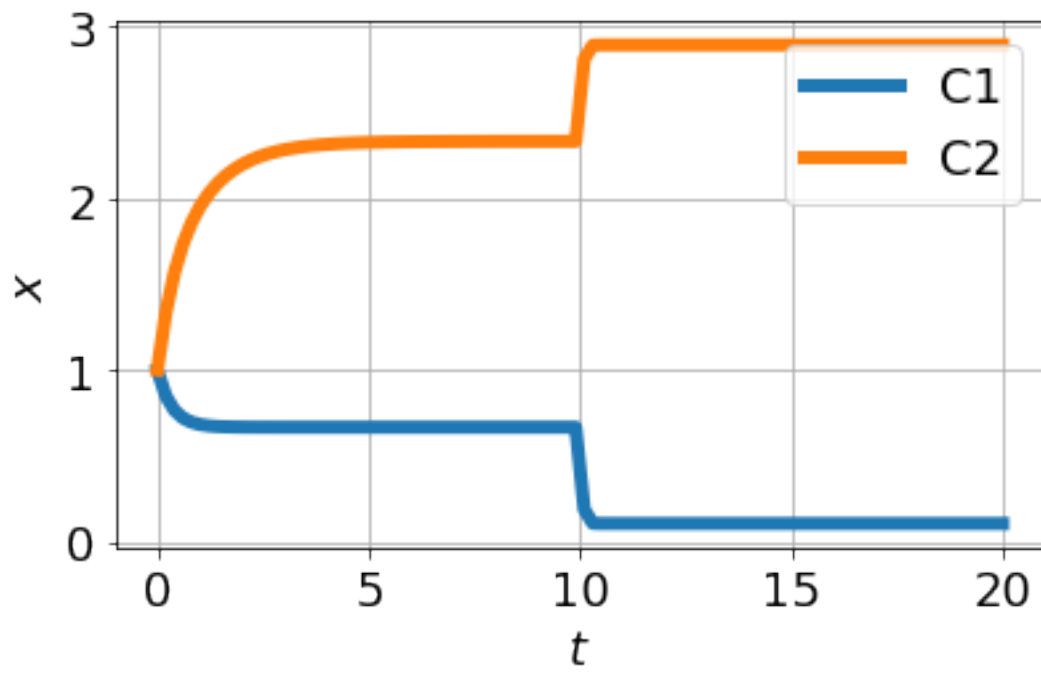
    name = f'Figs/splitReos_{unidir}_v'
    plt.savefig(name+'.pdf')
    plt.show()

x_B_0 = dat['X'][:,species.index('B')]
#     st.plot(s,dat,species=['A','B','C1','C2'])
#     st.plot(s,dat,species=[],filename=f'{name}_v.pdf')

2*(1+8*np.heaviside(t-10,0.5))

```





3.2 Pipe connection

```
[15]: ## Create Pipe component
      N_lump = 5
      V_p = 10
```

```

Pipe = mbg.chain(AB_abg.model(),inport = 'A',outport = 'B',
                 N=N_lump+1,rename_components=True,Comps=['R'],quiet=quiet)
s = st.stoich(Pipe,UniDir=None, quiet=quiet)
s['name'] = 'Pipe_abg'
stbg.model(s)
import Pipe_abg
imp.reload(Pipe_abg)

```

```

[15]: <module 'Pipe_abg' from
      '/home/peterg/WORK/Research/SystemsBiology/Notes/2022/Advection/Pipe_abg.py'>

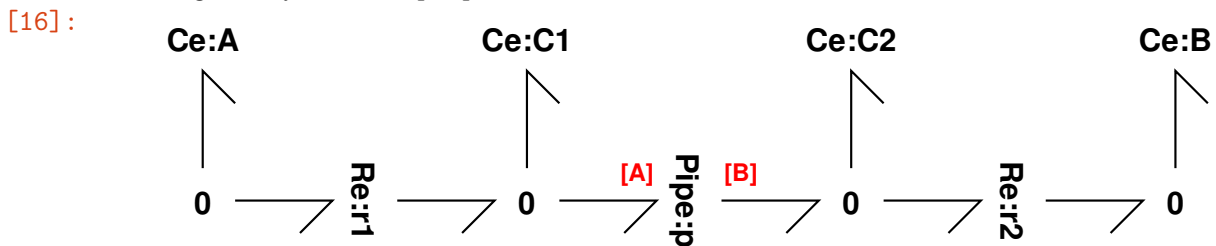
```

```

[16]: ## Split enzyme catalysed Reaction
      sbg.model('splitRep_abg.svg')
      import splitRep_abg
      disp.SVG('splitRep_abg.svg')

```

Creating subsystem: Pipe:p



```

[17]: ##Stoichiometry+parameters
      parameter = {}
      Kappa = {}
      UniDir = []
      for i in range(N_lump+1):
          reac = f'p_r_{i}'
          UniDir.append(reac)
          kap = f'kappa_{reac}'
          parameter[kap] = Q0
          Kappa[reac] = Flow

      s = st.stoich(splitRep_abg.model(),UniDir=UniDir,quiet=quiet)
      species = s['species']
      sc = st.statify(s,chemostats=chemostats)

      ## Initial condition
      X0 = np.ones(s['n_X'])
      X0[species.index('A')] = 2

      ## Pipe parameters
      small = 1e-6
      if N_lump>0:

```

```

V_i = V_p/N_lump # Volume of lump
for i in range(N_lump):
    spec = f'p_IS{i+1}'
    K = f'K_{spec}'
    parameter[K] = 1/V_i
    X0[species.index(spec)] = small

# print(parameter)
# print(X0)
# print(parameter)
# print(UniDir)
print(Kappa)

disp.Latex(st.sprintrl(s,chemformula=True))

```

```

{'p_r_0': '2*(1+8*np.heaviside(t-10,0.5))', 'p_r_1':
'2*(1+8*np.heaviside(t-10,0.5))', 'p_r_2': '2*(1+8*np.heaviside(t-10,0.5))',
'p_r_3': '2*(1+8*np.heaviside(t-10,0.5))', 'p_r_4':
'2*(1+8*np.heaviside(t-10,0.5))', 'p_r_5': '2*(1+8*np.heaviside(t-10,0.5))'}

```

[17]:



```

[18]: ## Simulate
dat = st.sim(s,sc=sc,t=t,parameter=parameter,X0=X0,Kappa=Kappa,quiet=quiet)

plt.rcParams.update({'font.size': 18})
for spec in specPlot:
    x = dat['X'][:,species.index(spec)]
    plt.plot(t,x,label=spec,lw=lw)
    plt.plot(t,X_old[spec],color='black',ls='dashed')
plt.grid()
plt.legend()
plt.xlabel('$t$')
plt.ylabel('$x$')
plt.tight_layout()
plt.savefig('Figs/splitRep.pdf')
plt.show()

reaction = s['reaction']

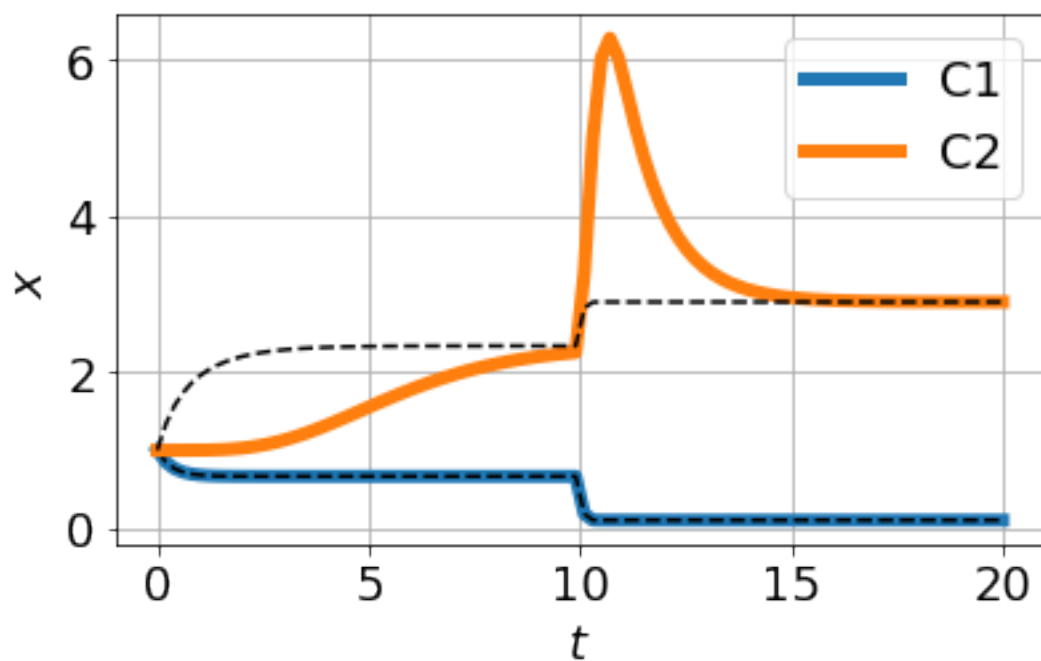
```

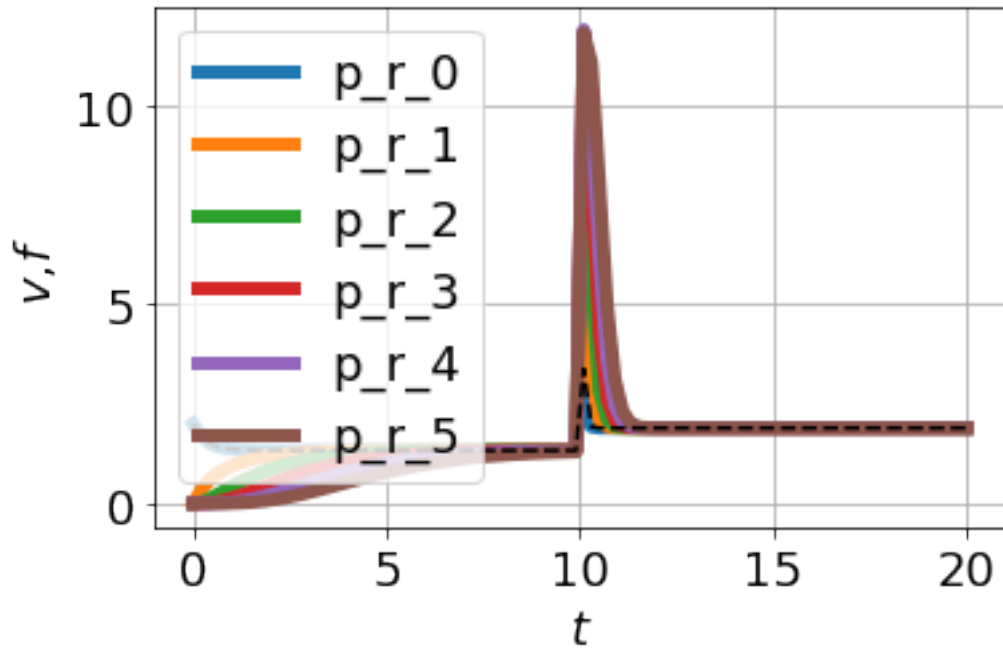


```

for i in range(N_lump+1):
    reac = f'p_r_{i}'
    v = dat['V'][:,reaction.index(reac)]
    plt.plot(t,v,label=reac,lw=lw)
plt.plot(t,V_old['r'],color='black',ls='dashed')
plt.grid()
plt.legend()
plt.xlabel('$t$')
plt.ylabel('$v$, $f$')
plt.tight_layout()
name = f'Figs/splitRep_v'
plt.savefig(name+'.pdf')
plt.show()

```



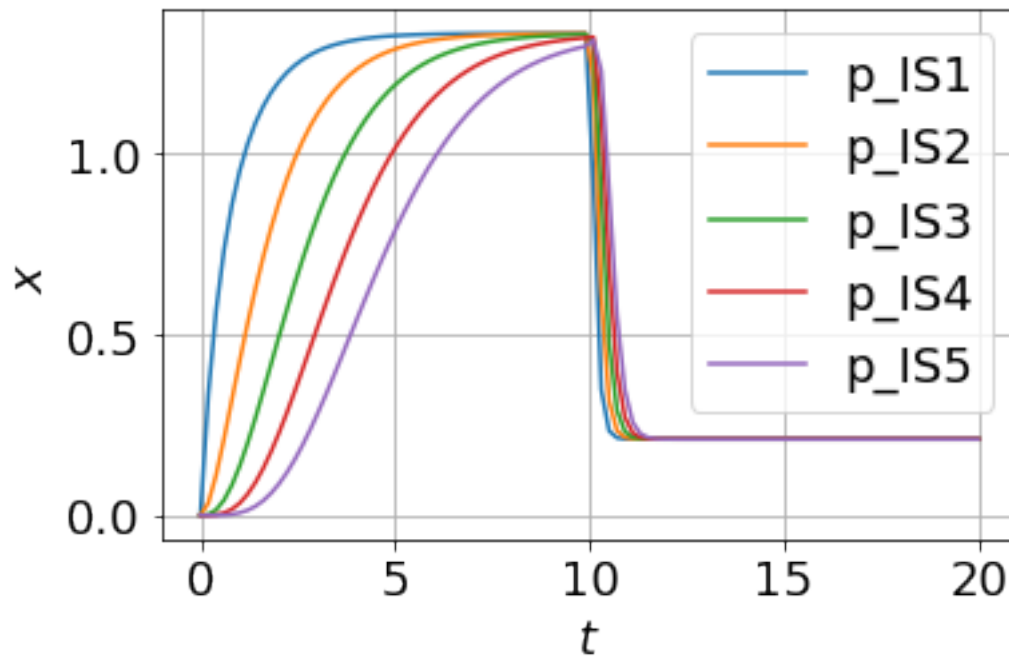


```
[19]: # st.plot(s,dat,reaction=[],species=['A','B','C1','C2'])
```

```
[20]: # st.plot(s,dat,species=[])
```

```
[21]: # st.plot(s,dat,reaction=[],species=['p_IS1'])
```

```
[22]: for i,spec in enumerate(species):
        if 'I' in spec:
            x = dat['X'][:,i]
            plt.plot(t,x,label=spec)
plt.grid()
plt.legend()
plt.xlabel('$t$')
plt.ylabel('$x$')
plt.tight_layout()
plt.savefig('Figs/splitRep_pipe.pdf')
```



4 Enzyme-catalysed reaction

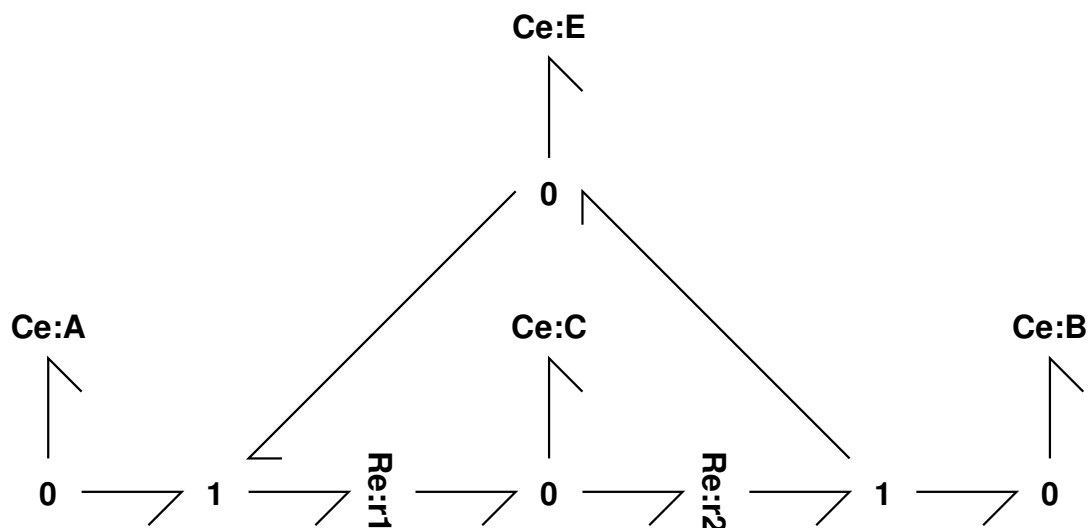
This section reprises the standard bond graph model of an enzyme-catalysed reaction – see [ECR](#). In this case: - **Ce_A** represents the unbound oxygen entering the blood - **Ce_B** represents the unbound oxygen leaving the blood - **Ce_E** represents the unbound haemoglobin - **Ce_C** represents the haemoglobin bound to oxygen

This corresponds to a simple model of haemoglobin with one binding site (like myoglobin). A more detailed model would incorporate four cooperative binding sites – see [Cooperative](#).

[]:

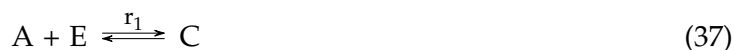
```
[23]: ## Enzyme catalysed reaction
      sbg.model('RE_abg.svg')
      import RE_abg
      disp.SVG('RE_abg.svg')
```

[23]:



```
[24]: ##Stoichiometry
s = st.stoich(RE_abg.model(),quiet=quiet)
chemostats = ['A', 'B']
sc = st.statify(s,chemostats=chemostats)
disp.Latex(st.sprintrl(s,chemformula=True))
```

[24]:

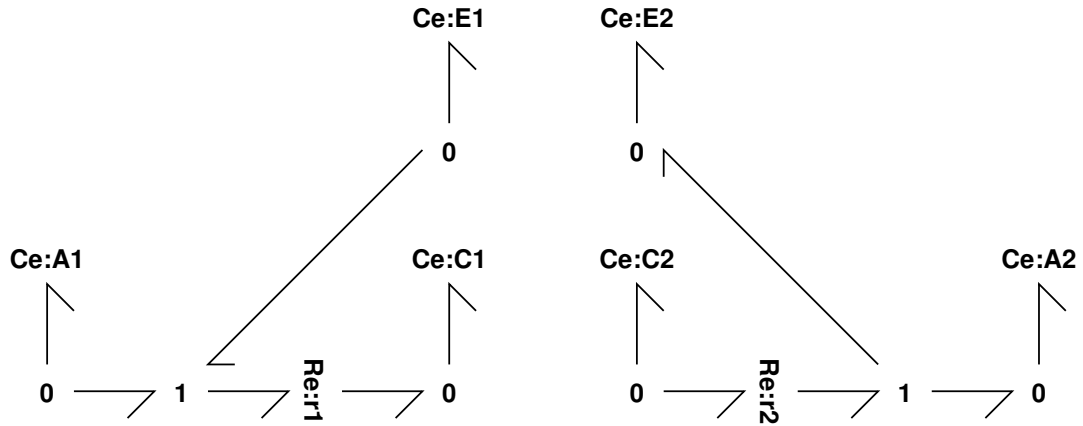


4.1 Split enzyme-catalysed reaction

In this bond graph, the components **Ce_E** and **Ce_E** are split: advection carries the corresponding molecules from one side to the other. An explicit bond graph for this coupling is provided below for the case where the advection time-delay is zero; the bond graph for the general case has not yet been determined.

```
[25]: ## Split enzyme catalysed reaction
sbg.model('splitRE_abg.svg')
import splitRE_abg
disp.SVG('splitRE_abg.svg')
```

[25]:



```
[26]: ##Stoichiometry
ss = st.stoich(splitRE_abg.model(),quiet=quiet)
chemostats = ['A1','A2']
ssc = st.statify(ss,chemostats=chemostats)
disp.Latex(st.sprintrl(ss,chemformula=True))
```

[26]:

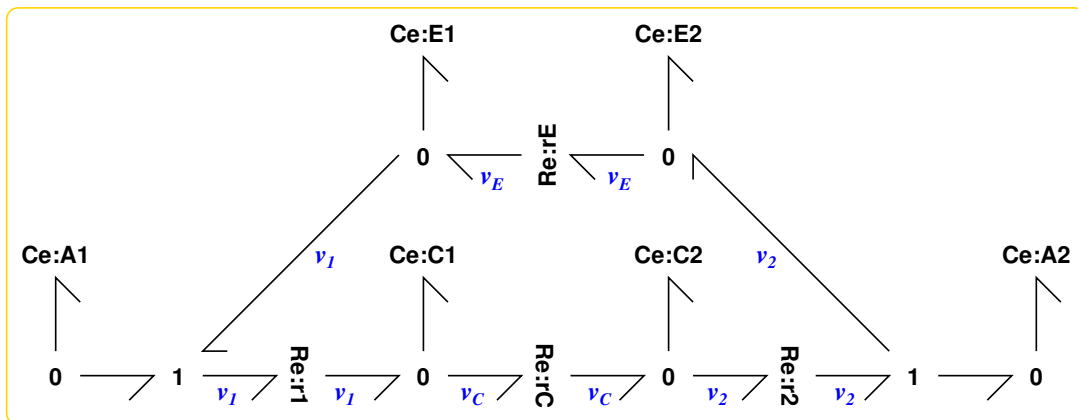


5 Circular advection

5.1 Orifice (Symmetric case): Bond graph model

```
[27]: ## Split enzyme catalysed reaction with symmetric connection
sbg.model('splitREos_abg.svg')
import splitREos_abg
disp.SVG('splitREos_abg.svg')
```

[27]:



```
[28]: ##Stoichiometry
snd = st.stoich(splitREos_abg.model(),quiet=quiet)
chemostats = ['A1','A2']
sndc = st.statify(snd,chemostats=chemostats)
print(snd['species'])
print(snd['reaction'])
disp.Latex(st.sprintrl(snd,chemformula=True))
```

```
['A1', 'A2', 'C1', 'C2', 'E1', 'E2']
['r1', 'r2', 'rC', 'rE']
```

[28]:



5.1.1 Simulate

Note that - the final steady state of the two cases $d = 0$ and $d > 0$ are the same - if the simulation is repeated with $f_0 = 100$, the $x_{C1} \approx x_{C2}$ and $x_{E1} \approx x_{E2}$ as predicted.

The two chemostats corresponding to A1 and A2 are:

$$x_{A1} = \begin{cases} 1 & \text{if } t < 1 \\ 2 & \text{if } t \geq 1 \end{cases} \quad (45)$$

$$x_{A2} = 1 \quad (46)$$

```
[29]: ## Simulate split ECR with no delay
i_chemo = []
for chemo in chemostats:
    i_chemo.append(snd['species'].index(chemo))
# print(i_chemo)
## Time
tt = np.linspace(0, 20,10000)

## Parameters
parameter = {}
parameter['kappa_rC'] = Q0
parameter['kappa_rE'] = Q0

# parameter['K_E'] = 0.5
# parameter['K_C'] = 0.5
## Chemostats
t_0 = 0
X_chemo = {'A1':f'1 + np.heaviside(t-{t_0},1)', 'A2':'1'}

## Simulate
dat = st.sim(snd,sc=sndc,t=tt,parameter=parameter,X_chemo=X_chemo,quiet=False)
```

```

XX0 = dat['X'][:,2:6]
Legend = snd['species'][2:6]
V0 = dat['V'][:,0:2]
dX0 = dat['dX'][:,i_chemo]

```

Setting kappa_rC to 2

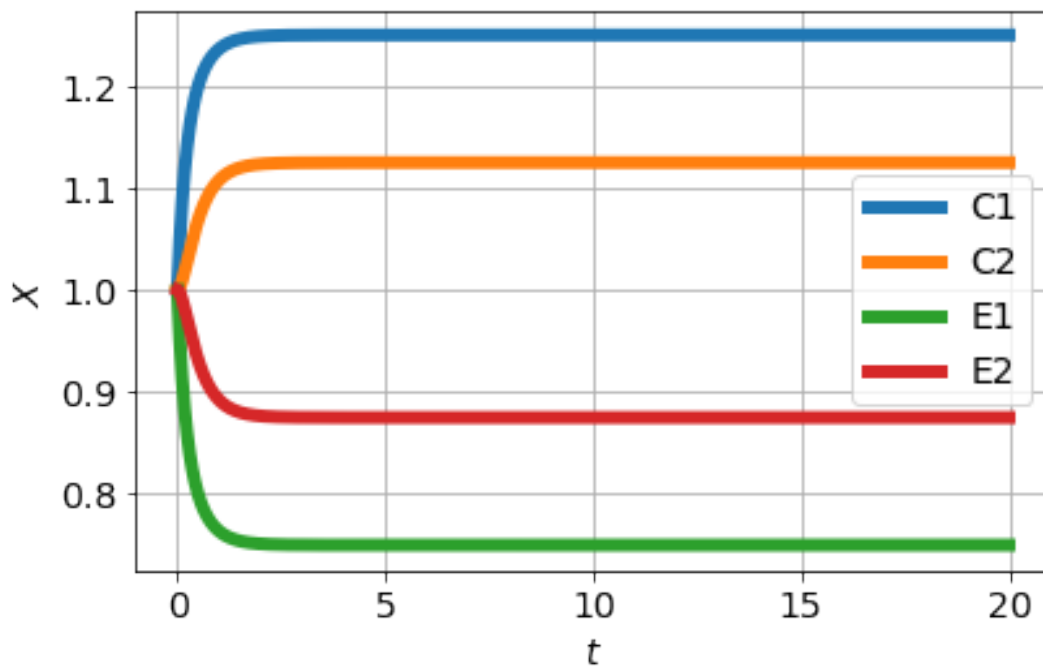
Setting kappa_rE to 2

```

[30]: ## Plot
plt.rcParams.update({'font.size': 14})
## Orifice
plt.plot(tt,XX0,lw=lw)

plt.legend(Legend)
plt.grid()
plt.xlabel('$t$')
plt.ylabel('$X$')
plt.tight_layout()
plt.savefig('Figs/OrificeSim.pdf')

```



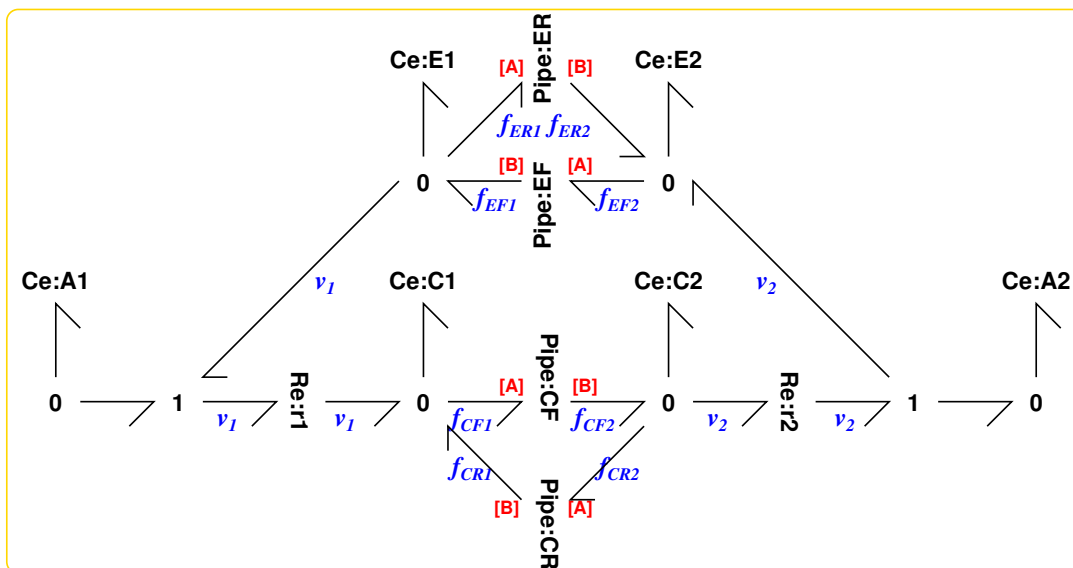
5.2 Circular advection: pipe connection

```

[31]: disp.SVG('splitREp_abg.svg')

```

[31]:



```
[32]: def CircularAdvection(N_lump=5,Flow=f'{Q0}', quiet=True):

    ## Create Pipe component
    ## Note: N_lump = 0 corresponds to orifice case.
    Pipe = mbg.chain(AB_abg.model(),inport = 'A',outport = 'B',
                    ⊥
    →N=N_lump+1,rename_components=True,Comps=['R'],quiet=quiet)
    s = st.stoich(Pipe,UniDir=None, quiet=quiet)
    s['name'] = 'Pipe_abg'
    stbg.model(s)

    ## Set uni directional Re components
    UniDir = []
    for i in range(N_lump+1):
        for comp in ['EF','ER','CF','CR']:
            ud = f'{comp}_r_{i}'
            UniDir.append(ud)

    # print(UniDir)
    ## Create stoichiometry
    sbg.model('splitREp_abg.svg')
    import Pipe_abg
    imp.reload(Pipe_abg)
    import splitREp_abg
    imp.reload(splitREp_abg)
    sPipe = st.stoich(splitREp_abg.model(),UniDir=UniDir,quiet=quiet)

    if not quiet:
        disp.Latex(st.sprintrl(sPipe,chemformula=True))

    ## Setup parameters
    parameter = {}
```



```

Kappa = {}

## Assume K=KK is same for all species
## Note that in the simulation code K corresponds to KK/V_i
KK = 1

for i in range(N_lump+1):
    for comp in ['EF', 'ER', 'CF', 'CR']:
        kap = f'{comp}_r_{i}'
        Kappa[kap] = f'({Flow})/{KK}'
        #parameter[kap] = Q0
        if i>0:
            V_i = V_p/N_lump
            KO = KK/V_i
            KOF = KO
            KOR = KO
            K = f'K_{comp}_IS{i}'
            if 'F' in comp:
                parameter[K] = KOF
            else:
                parameter[K] = KOR

## Setup initial states
X0 = []
small = 1e-6
for spec in sPipe['species']:
    # print(spec)
    if 'IS' in spec:
        K = f'K_{spec}'
        Xi = 1/parameter[K]
# Xi = small
    else:
        Xi = 1
    X0.append(Xi)

X0 = np.array(X0)

# print(Kappa)
# print(sPipe['reaction'])
## Simulate
chemostats=['A1', 'A2']
i_chemo = []
for chemo in chemostats:
    i_chemo.append(sPipe['species'].index(chemo))
# print(i_chemo)
sPipec = st.statify(sPipe, chemostats=chemostats)
dat = st.
→sim(sPipe, sc=sPipec, t=tt, parameter=parameter, Kappa=Kappa, X0=X0, X_chemo=X_chemo, quiet=quiet)

return sPipe, dat, i_chemo

```

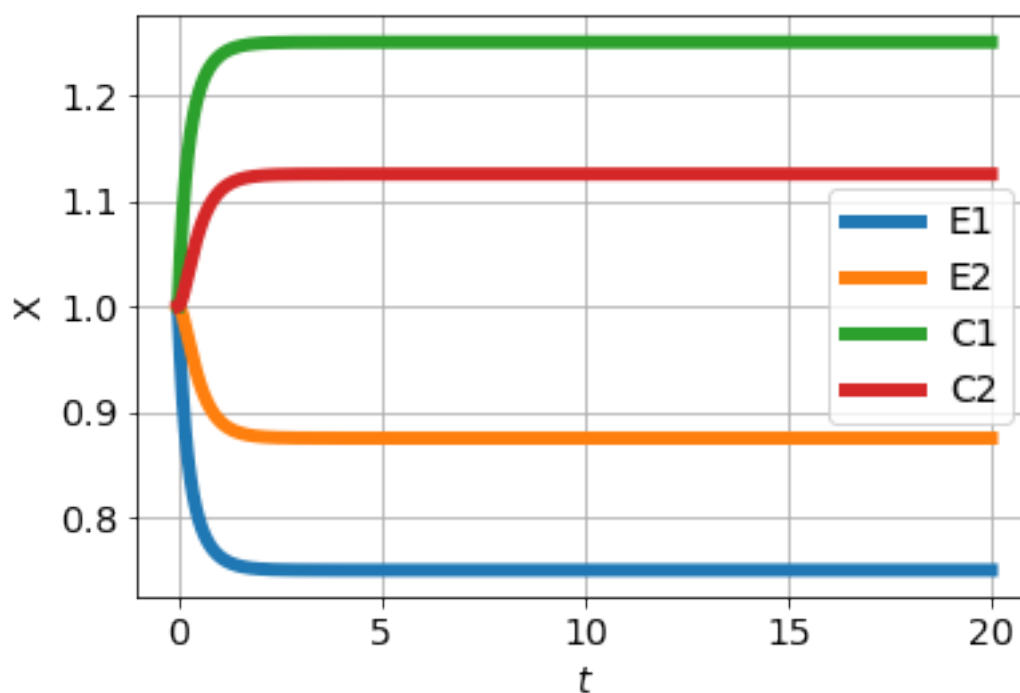
```
[33]: #Orifice (Pipe with no lumps)
N_lump = 0
sPipe0,dat0,i_chemo0 = CircularAdvection(N_lump=N_lump)

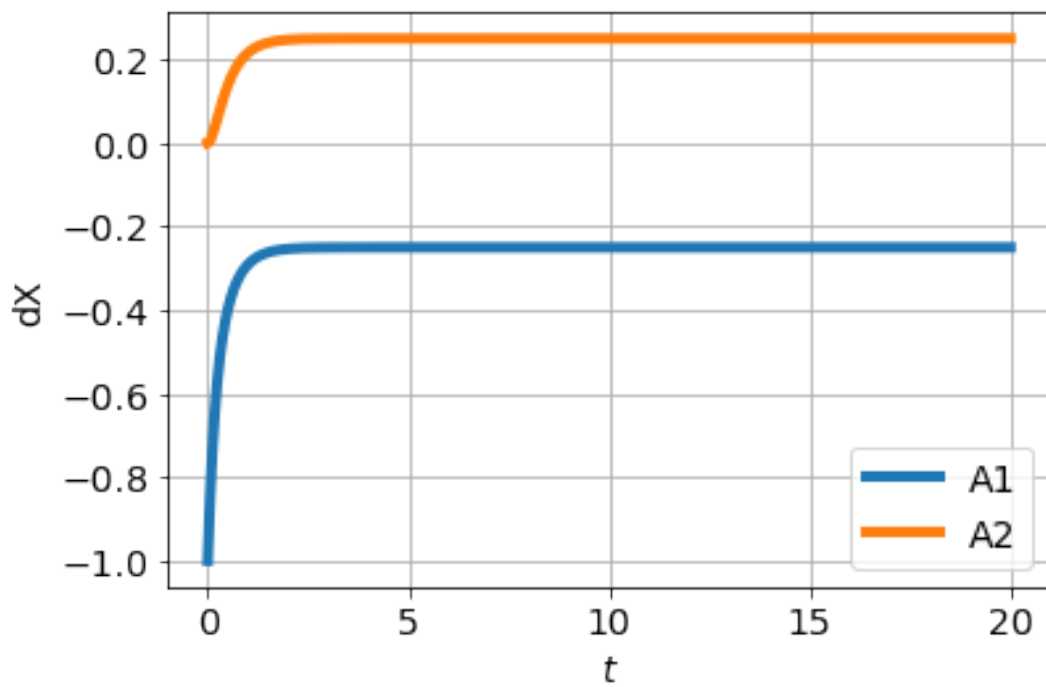
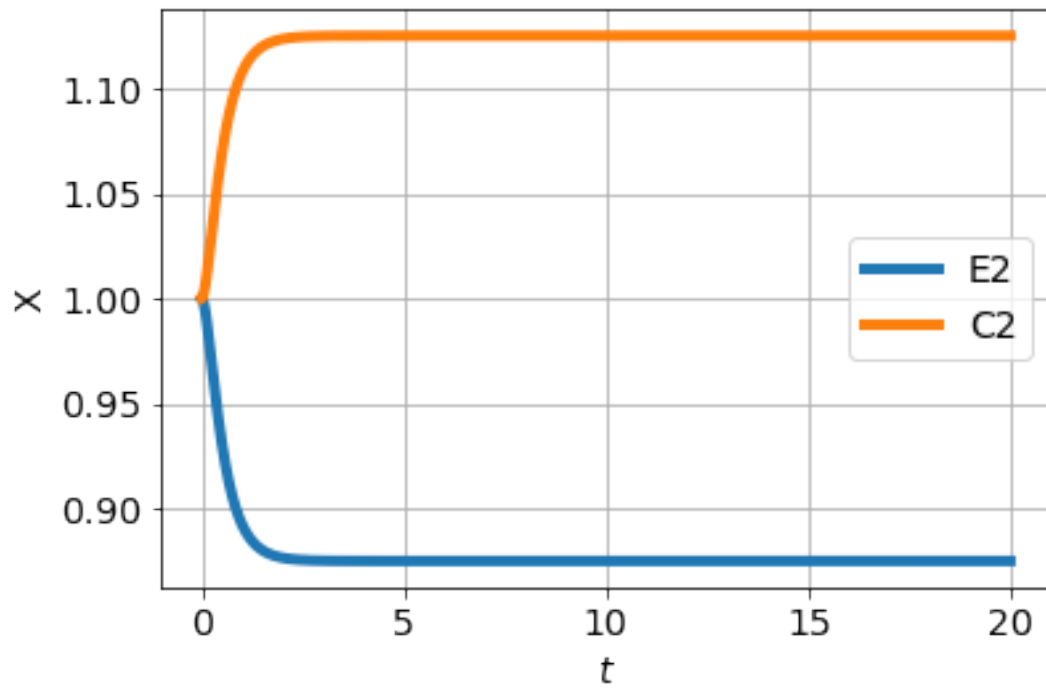
print('N_lump =', N_lump)

plt.rcParams.update({'font.size': 14})

st.plot(sPipe0,dat0,species=['E1','E2','C1','C2'], reaction = [],lw=5)
#st.plot(sPipe,dat,species=[], reaction = ['PipeEF_r_0','PipeCF_r_0'])
st.plot(sPipe0,dat0,species=['E2','C2'], reaction = [])
st.plot(sPipe0,dat0,dX=True,species=['A1','A2'], reaction = [])
```

Creating subsystem: Pipe:CF
 Creating subsystem: Pipe:CR
 Creating subsystem: Pipe:EF
 Creating subsystem: Pipe:ER
 N_lump = 0





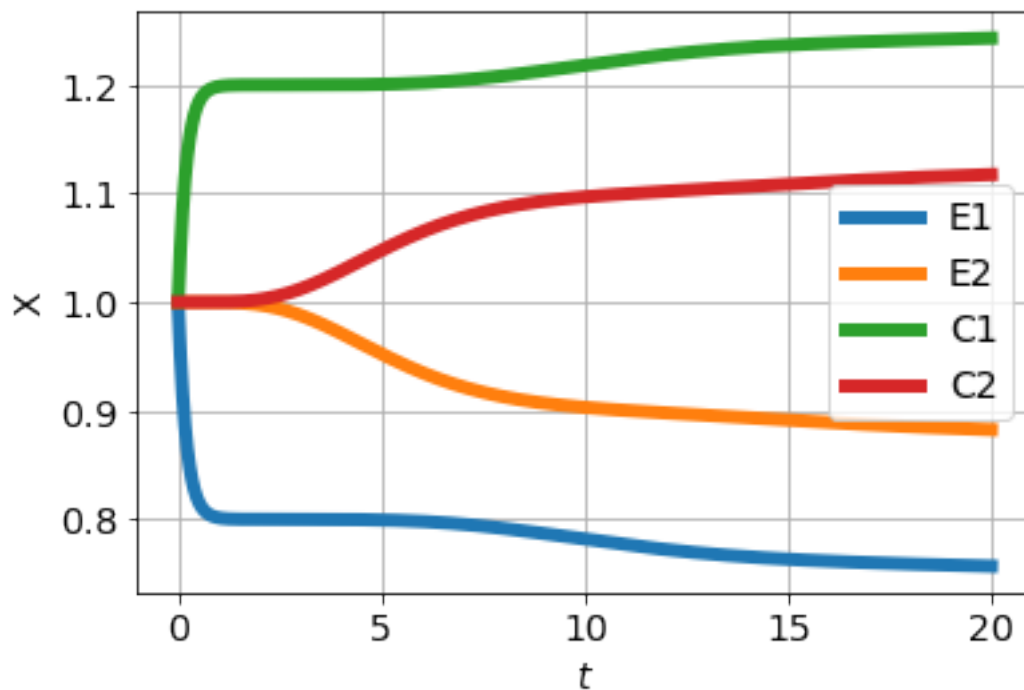
```
[34]: #Pipe
N_lump = 5
sPipe, dat, i_chemo = CircularAdvection(N_lump=N_lump)
```

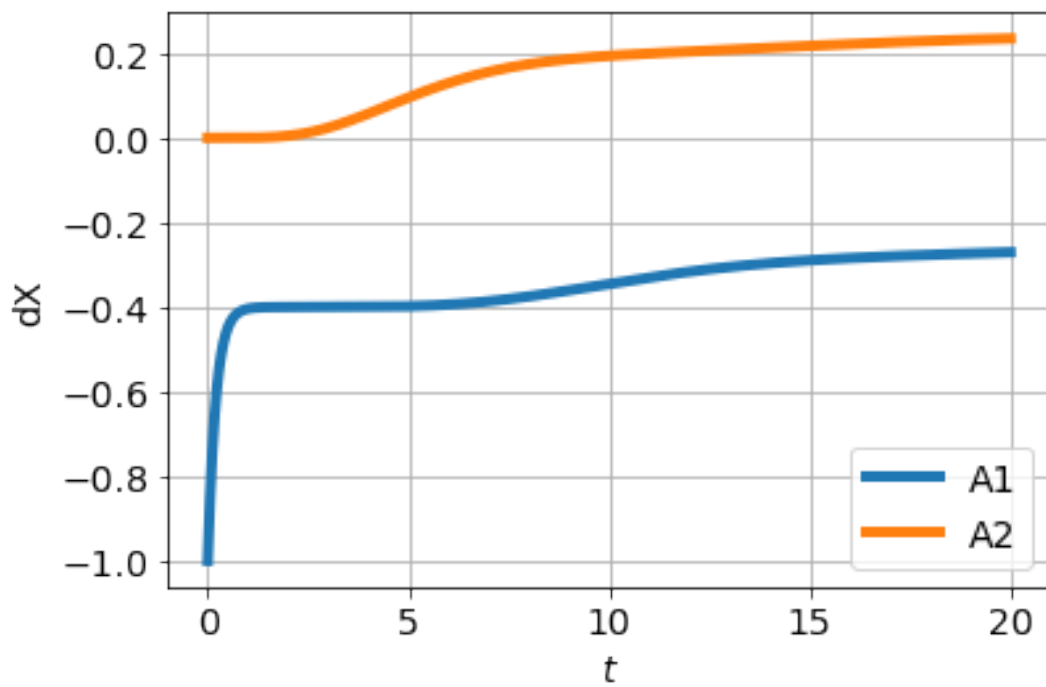
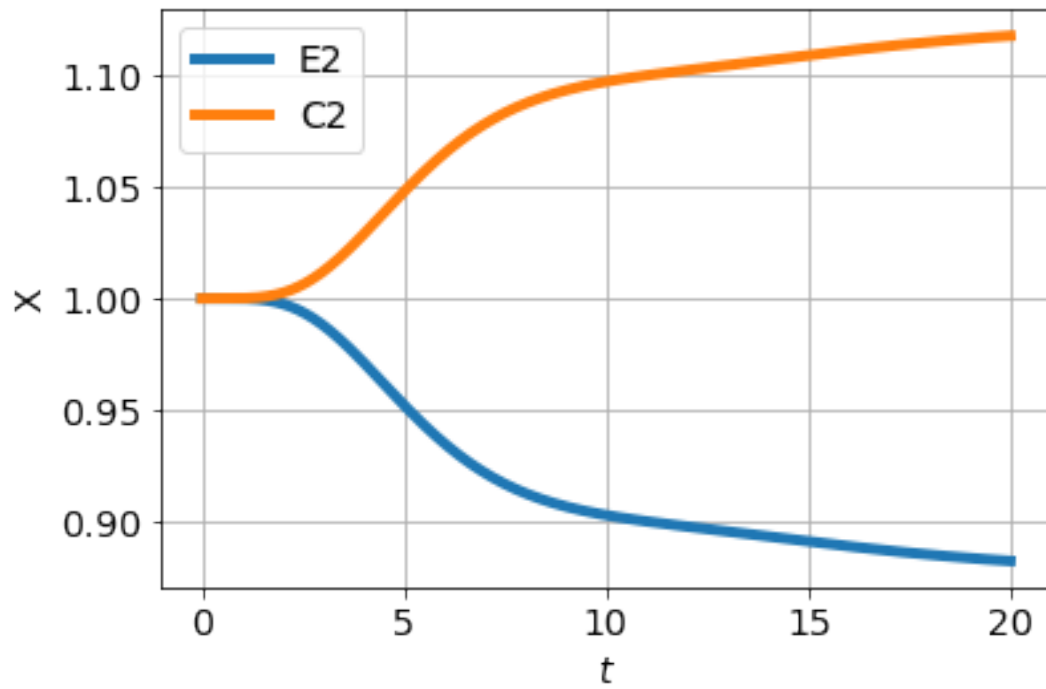
```

print('N_lump =', N_lump)
st.plot(sPipe,dat,species=['E1','E2','C1','C2'], reaction = [],lw=lw)
#st.plot(sPipe,dat,species=[], reaction = ['PipeEF_r_0','PipeCF_r_0'])
st.plot(sPipe,dat,species=['E2','C2'], reaction = [])
st.plot(sPipe,dat,dX=True,species=['A1','A2'], reaction = [])

```

Creating subsystem: Pipe:CF
 Creating subsystem: Pipe:CR
 Creating subsystem: Pipe:EF
 Creating subsystem: Pipe:ER
 N_lump = 5





```
[35]: ## Extract data
XXPipe = dat['X'][:,2:6]
VPipe = dat['V'][:,[0,N_lump+1]]
dXPipe = dat['dX'][:,i_chemo]
```

```

dXPipe0 = dat0['dX'][:,i_chemo]

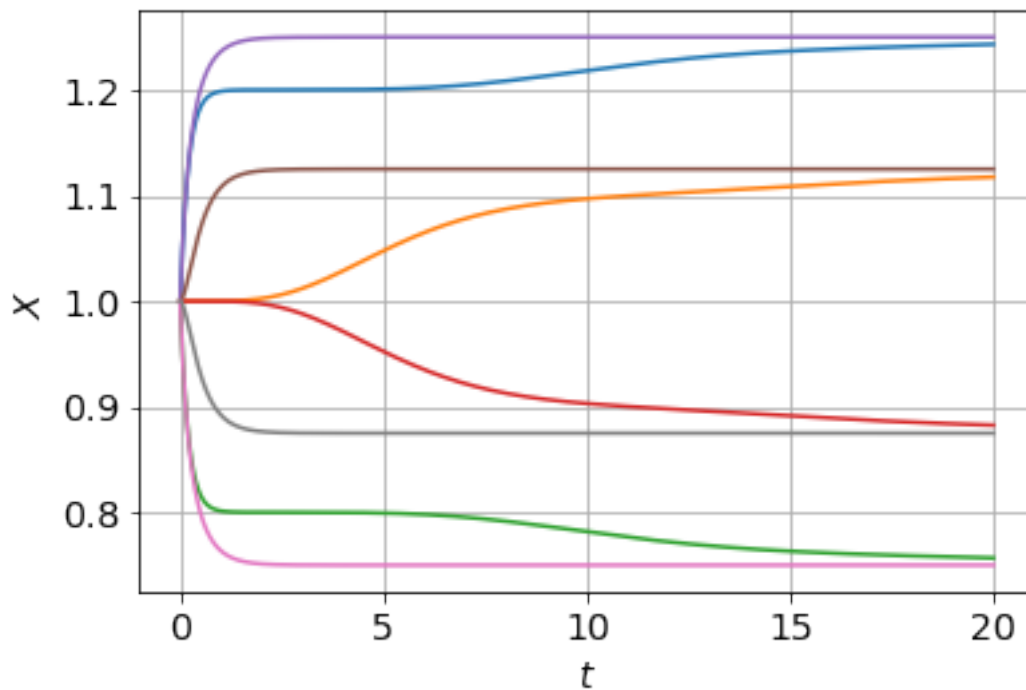
## Plot and compare
## Pipe
plt.plot(tt,XXPipe)

## Orifice
plt.plot(tt,XX0)

plt.grid()
plt.xlabel('$t$')
plt.ylabel('$X$')

```

[35]: Text(0,0.5,'\$X\$')



```

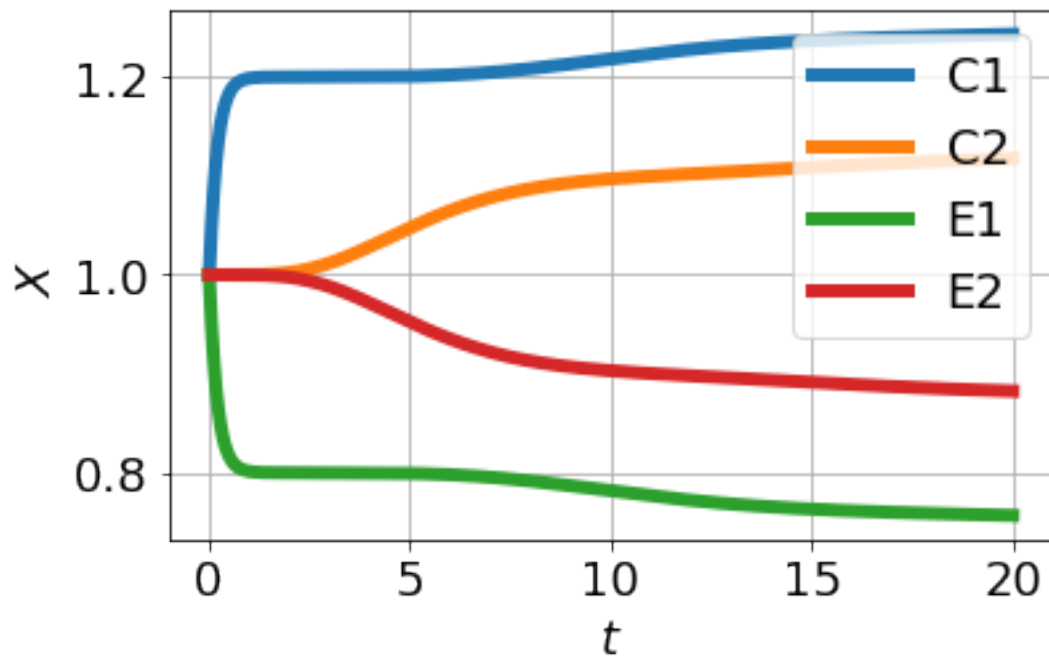
[36]: ## Plot

## Pipe
## Font
plt.rcParams.update({'font.size': 18})
plt.plot(tt,XXPipe,lw=lw)

plt.legend(Legend)
plt.grid()
plt.xlabel('$t$')
plt.ylabel('$X$')
plt.tight_layout()

```

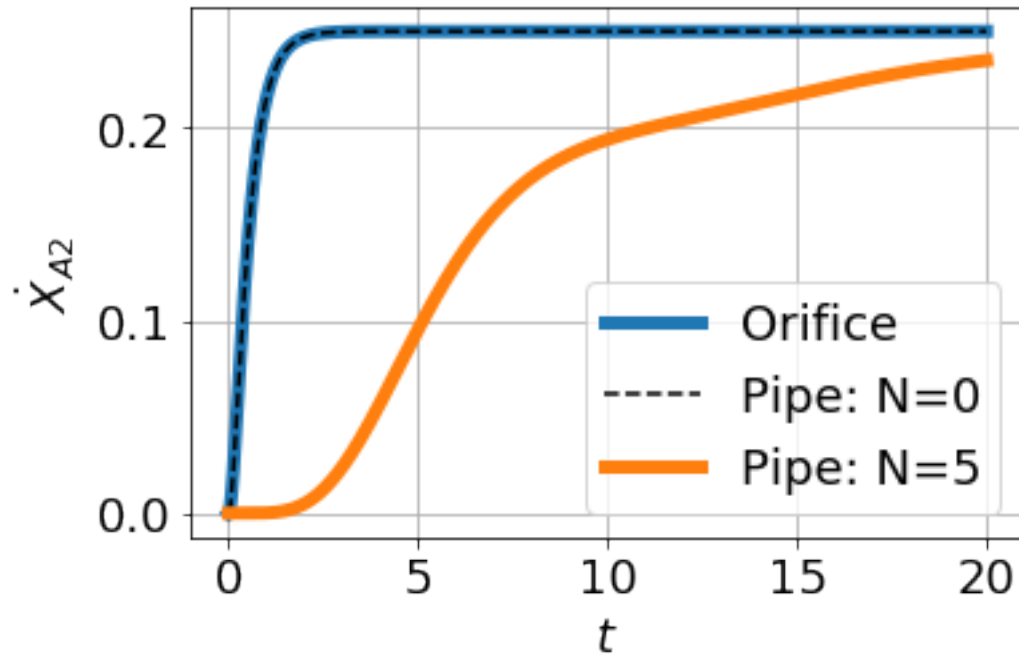
```
plt.savefig('Figs/PipeSim.pdf')
```



```
[37]: ## Plot Chemostat flows
      ## Orifice
      plt.plot(tt,dX0[:,1],label='Orifice',lw=lw)

      ## Pipe
      plt.plot(tt,dXPipe0[:,1],label='Pipe: N=0',linestyle='dashed',color='black')
      plt.plot(tt,dXPipe[:,1],label=f'Pipe: N={N_lump}',lw=lw)

      plt.grid()
      plt.legend()
      plt.xlabel('$t$')
      plt.ylabel('$\dot{X}_{A2}$')
      plt.tight_layout()
      plt.savefig('Figs/PipeSim_flow.pdf')
```

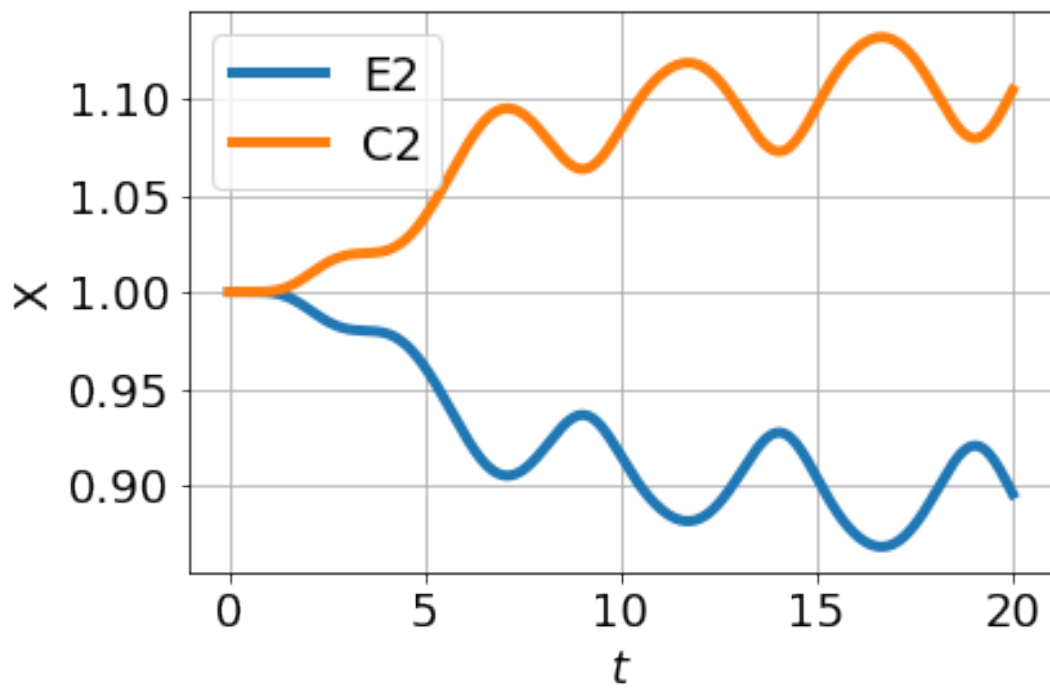
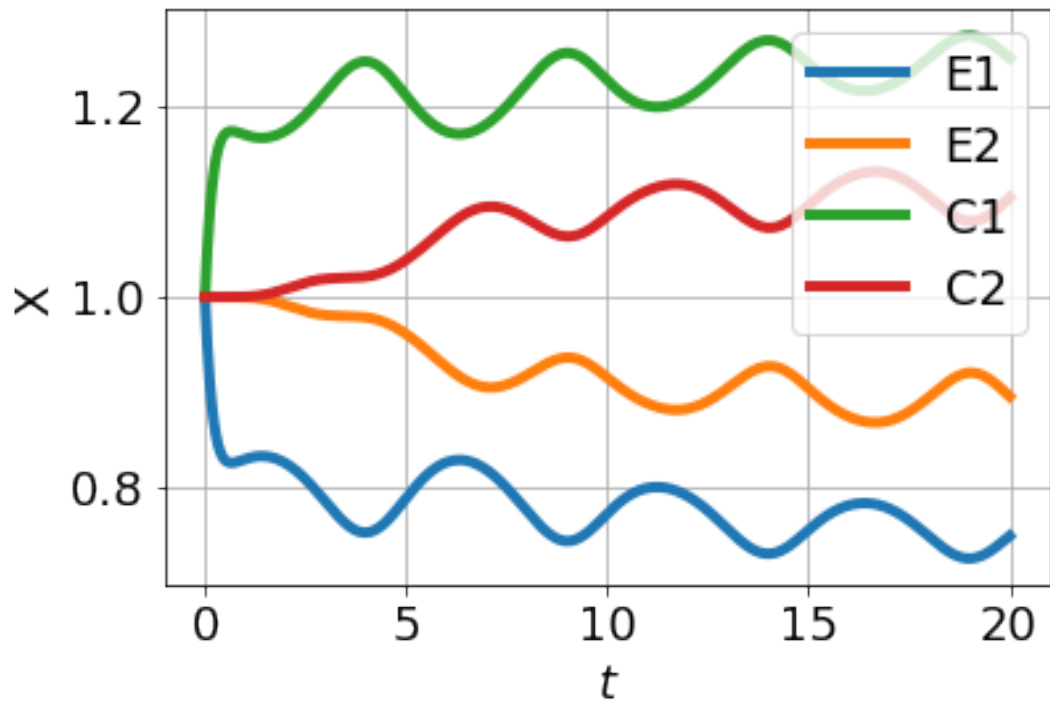


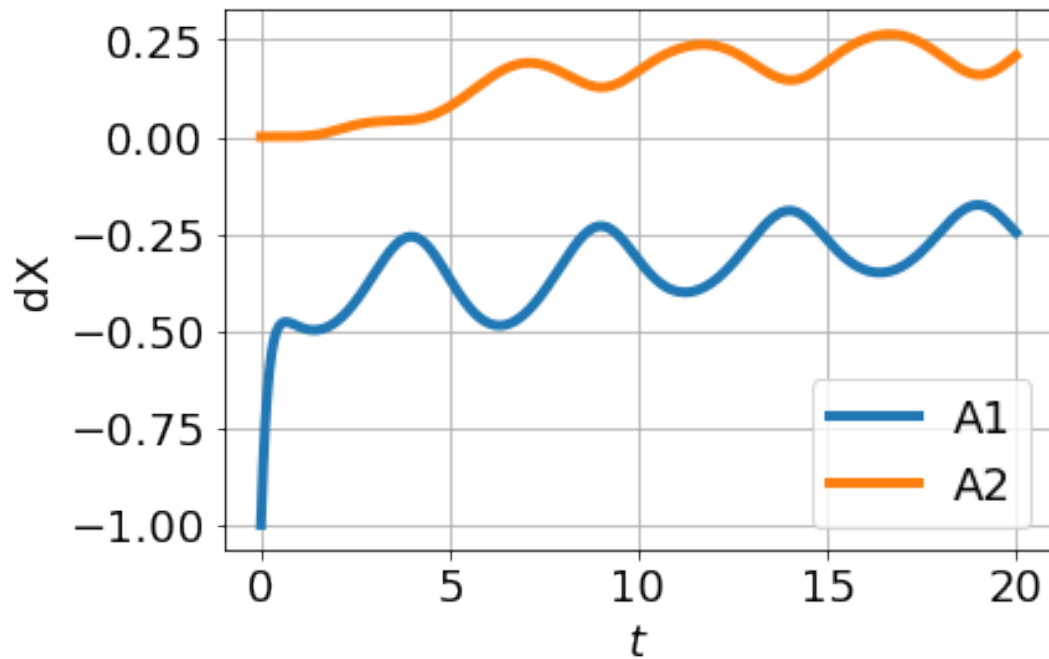
6 Variable flow rate

```
[38]: N_lump = 5
Period = 5
Flow = f'({Q0})*(1+0.5*np.sin(2*np.pi*t/{Period}))'
print('Flow:', Flow)
sPipe, dat, i_chemo = CircularAdvection(N_lump=N_lump, Flow=Flow)

print('N_lump =', N_lump)
st.plot(sPipe, dat, species=['E1', 'E2', 'C1', 'C2'], reaction = [])
#st.plot(sPipe, dat, species=[], reaction = ['PipeEF_r_0', 'PipeCF_r_0'])
st.plot(sPipe, dat, species=['E2', 'C2'], reaction = [])
st.plot(sPipe, dat, dX=True, species=['A1', 'A2'], reaction = [])
```

```
Flow: (2)*(1+0.5*np.sin(2*np.pi*t/5))
Creating subsystem: Pipe:CF
Creating subsystem: Pipe:CR
Creating subsystem: Pipe:EF
Creating subsystem: Pipe:ER
N_lump = 5
```

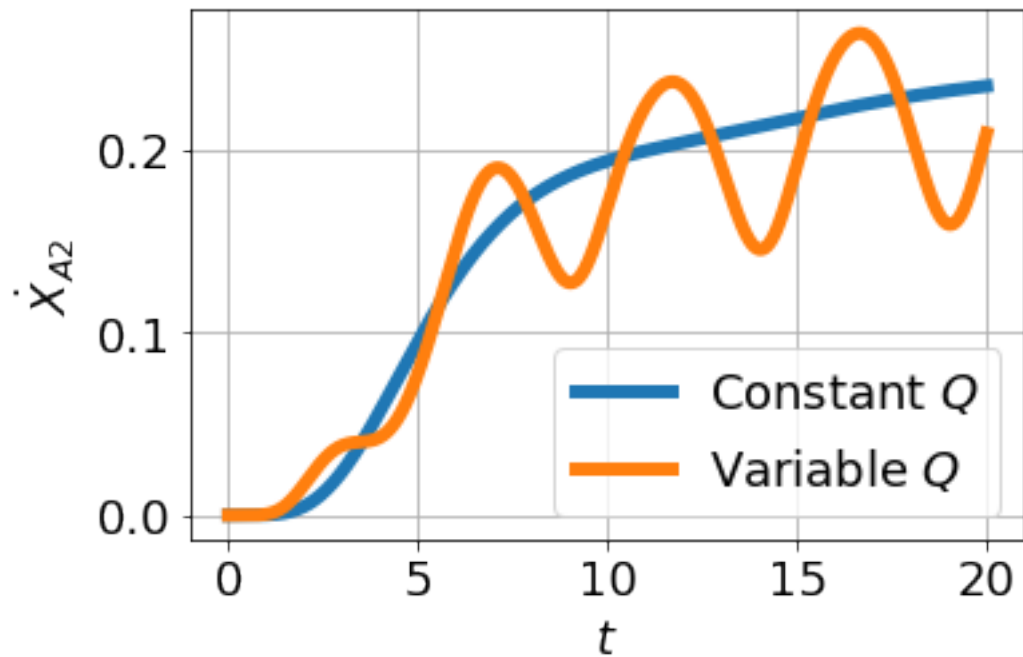





```
[39]: ## Extract data
XXPipev = dat['X'][:,2:6]
VPipev = dat['V'][:,[0,N_lump+1]]
dXPipev = dat['dX'][:,i_chemo]

## Plot Chemostat flows
## Pipe
## Font
plt.rcParams.update({'font.size': 18})

plt.plot(tt,dXPipe[:,1],label='Constant $Q$',lw=1w)
plt.plot(tt,dXPipev[:,1],label='Variable $Q$',lw=1w)
#plt.plot(tt,Q/4,ls='dashed',label='$Q/4$')
plt.grid()
plt.legend()
plt.xlabel('$t$')
plt.ylabel('$\dot{X}_{A2}$')
plt.tight_layout()
plt.savefig('Figs/PipeSim_flow_vary.pdf')
```



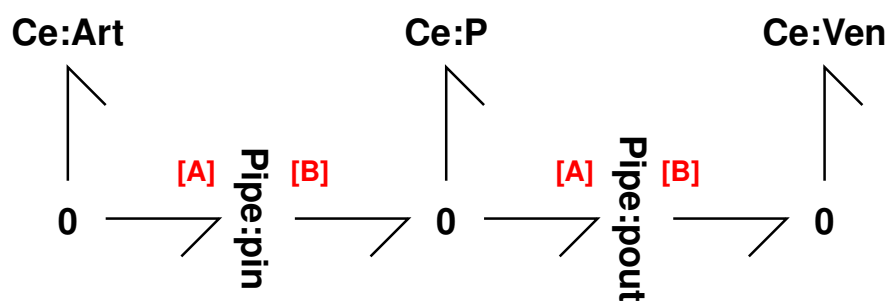
7 Pharmacokinetics

The data is taken from [Mapleson \(1973\)](#).

```
[40]: # sbg.model('Lung_abg.svg')
      # disp.SVG('Lung_abg.svg')
```

```
[41]: disp.SVG('Pool_abg.svg')
```

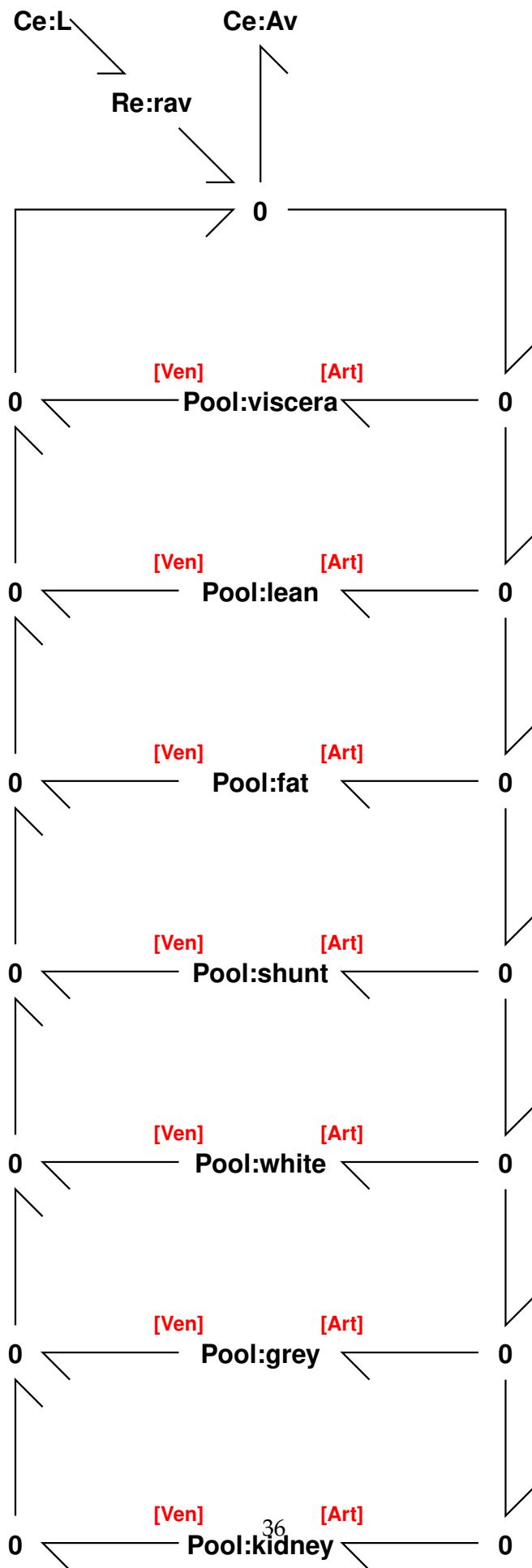
[41]:



[]:

```
[42]: disp.SVG('PK_abg.svg')
```

[42]:



```

[43]: def Pool(N_lump=0, quiet=True):

    ## Create Pipe component
    ## Note: N_lump = 0 corresponds to orifice case.
    Pipe = mbg.chain(AB_abg.model(), inport = 'A', outport = 'B',
                     ⌋
    → N=N_lump+1, rename_components=True, Comps=['R'], quiet=quiet)
    s = st.stoich(Pipe, UniDir=None, quiet=quiet)
    s['name'] = 'Pipe_abg'
    stbg.model(s)

    ## Set unidirectional Re components
    UniDir = []
    for i in range(N_lump+1):
        for comp in ['pin', 'pout']:
            ud = f'{comp}_r_{i}'
            UniDir.append(ud)

    # print(UniDir)
    ## Create stoichiometry
    sbg.model('Pool_abg.svg')
    import Pipe_abg
    imp.reload(Pipe_abg)
    import Pool_abg
    imp.reload(Pool_abg)
    sPool = st.stoich(Pool_abg.model(), UniDir=UniDir, quiet=quiet)

    if not quiet:
        disp.Latex(st.sprintrl(sPool, chemformula=True))

    ## Flatten
    stbg.model(sPool, filename='Pool_abg')

    return UniDir

def setPool(N_lump=0, Flow=f'{Q0}', quiet=True):
    ## Setup parameters
    parameter = {}
    Kappa = {}

    ## Assume K=KK is same for all species
    ## Note that in the simulation code K corresponds to KK/V_i
    KK = 1

    for i in range(N_lump+1):
        for comp in ['pin', 'pout']:
            kap = f'{comp}_r_{i}'

```

```

Kappa[kap] = f'({Flow})/{KK}'
#parameter[kap] = Q0
if i>0:
    V_i = V_p/N_lump
    KO = KK/V_i
    KOF = KO
    KOR = KO
    K = f'K_{comp}_IS{i}'
    if 'f' in comp:
        parameter[K] = KOF
    else:
        parameter[K] = KOR

## Setup initial states
X0 = []
for spec in sPool['species']:
    # print(spec)
    if 'IS' in spec:
        K = f'K_{spec}'
        Xi = 1/parameter[K]
    else:
        Xi = 1
    X0.append(Xi)

X0 = np.array(X0)

return parameter,X0

```

```

[44]: N_lump = 5
UniDir = Pool(N_lump=N_lump)
import Pool_abg
imp.reload(Pool_abg)
sPool = st.stoich(Pool_abg.model(),UniDir=UniDir,quiet=quiet)
print(sPool['name'])
print(UniDir)
disp.Latex(st.sprintrl(sPool,chemformula=True))

```

```

Creating subsystem: Pipe:pin
Creating subsystem: Pipe:pout
Pool

```

```

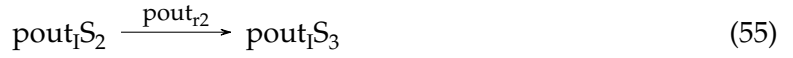
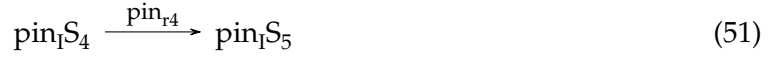
['pin_r_0', 'pout_r_0', 'pin_r_1', 'pout_r_1', 'pin_r_2', 'pout_r_2', 'pin_r_3',
'pout_r_3', 'pin_r_4', 'pout_r_4', 'pin_r_5', 'pout_r_5']

```

```

[44]:

```



```
[45]: import Pool_abg
imp.reload(Pool_abg)
sbg.model('PK_abg.svg')
import PK_abg
imp.reload(PK_abg)
pools = ['viscera', 'lean', 'fat', 'shunt', 'white', 'grey', 'kidney']
PK_UniDir = []
for pool in pools:
    for ud in UniDir:
        PKud = f'{pool}_{ud}'
        PK_UniDir.append(PKud)
# print(PK_UniDir)
sPK = st.stoich(PK_abg.model(), UniDir=PK_UniDir, quiet=quiet)
```

```
Creating subsystem: Pool:fat
Creating subsystem: Pool:grey
Creating subsystem: Pool:kidney
Creating subsystem: Pool:lean
Creating subsystem: Pool:shunt
Creating subsystem: Pool:viscera
Creating subsystem: Pool:white
```

```
[46]: disp.Latex(st.sprintrl(sPK, chemformula=True))
```

```
[46]:
```

$$Av \xrightarrow{\text{fat}_p \text{in}_{r0}} \text{fat}_p \text{in}_I S_1 \quad (59)$$

$$\text{fat}_p \text{in}_I S_1 \xrightarrow{\text{fat}_p \text{in}_{r1}} \text{fat}_p \text{in}_I S_2 \quad (60)$$

$$\text{fat}_p \text{in}_I S_2 \xrightarrow{\text{fat}_p \text{in}_{r2}} \text{fat}_p \text{in}_I S_3 \quad (61)$$

$$\text{fat}_p \text{in}_I S_3 \xrightarrow{\text{fat}_p \text{in}_{r3}} \text{fat}_p \text{in}_I S_4 \quad (62)$$

$$\text{fat}_p \text{in}_I S_4 \xrightarrow{\text{fat}_p \text{in}_{r4}} \text{fat}_p \text{in}_I S_5 \quad (63)$$

$$\text{fat}_p \text{in}_I S_5 \xrightarrow{\text{fat}_p \text{in}_{r5}} \text{fat}_p \quad (64)$$

$$\text{fat}_p \xrightarrow{\text{fat}_p \text{out}_{r0}} \text{fat}_p \text{out}_I S_1 \quad (65)$$

$$\text{fat}_p \text{out}_I S_1 \xrightarrow{\text{fat}_p \text{out}_{r1}} \text{fat}_p \text{out}_I S_2 \quad (66)$$

$$\text{fat}_p \text{out}_I S_2 \xrightarrow{\text{fat}_p \text{out}_{r2}} \text{fat}_p \text{out}_I S_3 \quad (67)$$

$$\text{fat}_p \text{out}_I S_3 \xrightarrow{\text{fat}_p \text{out}_{r3}} \text{fat}_p \text{out}_I S_4 \quad (68)$$

$$\text{fat}_p \text{out}_I S_4 \xrightarrow{\text{fat}_p \text{out}_{r4}} \text{fat}_p \text{out}_I S_5 \quad (69)$$

$$\text{fat}_p \text{out}_I S_5 \xrightarrow{\text{fat}_p \text{out}_{r5}} Av \quad (70)$$

$$Av \xrightarrow{\text{grey}_p \text{in}_{r0}} \text{grey}_p \text{in}_I S_1 \quad (71)$$

$$\text{grey}_p \text{in}_I S_1 \xrightarrow{\text{grey}_p \text{in}_{r1}} \text{grey}_p \text{in}_I S_2 \quad (72)$$

$$\text{grey}_p \text{in}_I S_2 \xrightarrow{\text{grey}_p \text{in}_{r2}} \text{grey}_p \text{in}_I S_3 \quad (73)$$

$$\text{grey}_p \text{in}_I S_3 \xrightarrow{\text{grey}_p \text{in}_{r3}} \text{grey}_p \text{in}_I S_4 \quad (74)$$

$$\text{grey}_p \text{in}_I S_4 \xrightarrow{\text{grey}_p \text{in}_{r4}} \text{grey}_p \text{in}_I S_5 \quad (75)$$

$$\text{grey}_p \text{in}_I S_5 \xrightarrow{\text{grey}_p \text{in}_{r5}} \text{grey}_p \quad (76)$$

$$\text{grey}_p \xrightarrow{\text{grey}_p \text{out}_{r0}} \text{grey}_p \text{out}_I S_1 \quad (77)$$

$$\text{grey}_p \text{out}_I S_1 \xrightarrow{\text{grey}_p \text{out}_{r1}} \text{grey}_p \text{out}_I S_2 \quad (78)$$

$$\text{grey}_p\text{out}_I S_2 \xrightarrow{\text{grey}_p\text{out}_{r2}} \text{grey}_p\text{out}_I S_3 \quad (79)$$

$$\text{grey}_p\text{out}_I S_3 \xrightarrow{\text{grey}_p\text{out}_{r3}} \text{grey}_p\text{out}_I S_4 \quad (80)$$

$$\text{grey}_p\text{out}_I S_4 \xrightarrow{\text{grey}_p\text{out}_{r4}} \text{grey}_p\text{out}_I S_5 \quad (81)$$

$$\text{grey}_p\text{out}_I S_5 \xrightarrow{\text{grey}_p\text{out}_{r5}} \text{Av} \quad (82)$$

$$\text{Av} \xrightarrow{\text{kidney}_p\text{in}_{r0}} \text{kidney}_p\text{in}_I S_1 \quad (83)$$

$$\text{kidney}_p\text{in}_I S_1 \xrightarrow{\text{kidney}_p\text{in}_{r1}} \text{kidney}_p\text{in}_I S_2 \quad (84)$$

$$\text{kidney}_p\text{in}_I S_2 \xrightarrow{\text{kidney}_p\text{in}_{r2}} \text{kidney}_p\text{in}_I S_3 \quad (85)$$

$$\text{kidney}_p\text{in}_I S_3 \xrightarrow{\text{kidney}_p\text{in}_{r3}} \text{kidney}_p\text{in}_I S_4 \quad (86)$$

$$\text{kidney}_p\text{in}_I S_4 \xrightarrow{\text{kidney}_p\text{in}_{r4}} \text{kidney}_p\text{in}_I S_5 \quad (87)$$

$$\text{kidney}_p\text{in}_I S_5 \xrightarrow{\text{kidney}_p\text{in}_{r5}} \text{kidney}_p \quad (88)$$

$$\text{kidney}_p \xrightarrow{\text{kidney}_p\text{out}_{r0}} \text{kidney}_p\text{out}_I S_1 \quad (89)$$

$$\text{kidney}_p\text{out}_I S_1 \xrightarrow{\text{kidney}_p\text{out}_{r1}} \text{kidney}_p\text{out}_I S_2 \quad (90)$$

$$\text{kidney}_p\text{out}_I S_2 \xrightarrow{\text{kidney}_p\text{out}_{r2}} \text{kidney}_p\text{out}_I S_3 \quad (91)$$

$$\text{kidney}_p\text{out}_I S_3 \xrightarrow{\text{kidney}_p\text{out}_{r3}} \text{kidney}_p\text{out}_I S_4 \quad (92)$$

$$\text{kidney}_p\text{out}_I S_4 \xrightarrow{\text{kidney}_p\text{out}_{r4}} \text{kidney}_p\text{out}_I S_5 \quad (93)$$

$$\text{kidney}_p\text{out}_I S_5 \xrightarrow{\text{kidney}_p\text{out}_{r5}} \text{Av} \quad (94)$$

$$\text{Av} \xrightarrow{\text{lean}_p\text{in}_{r0}} \text{lean}_p\text{in}_I S_1 \quad (95)$$

$$\text{lean}_p\text{in}_I S_1 \xrightarrow{\text{lean}_p\text{in}_{r1}} \text{lean}_p\text{in}_I S_2 \quad (96)$$

$$\text{lean}_p\text{in}_I S_2 \xrightarrow{\text{lean}_p\text{in}_{r2}} \text{lean}_p\text{in}_I S_3 \quad (97)$$

$$\text{lean}_p\text{in}_I S_3 \xrightarrow{\text{lean}_p\text{in}_{r3}} \text{lean}_p\text{in}_I S_4 \quad (98)$$

$$\text{lean}_p \text{in}_I S_4 \xrightarrow{\text{lean}_p \text{in}_{r4}} \text{lean}_p \text{in}_I S_5 \quad (99)$$

$$\text{lean}_p \text{in}_I S_5 \xrightarrow{\text{lean}_p \text{in}_{r5}} \text{lean}_p \quad (100)$$

$$\text{lean}_p \xrightarrow{\text{lean}_p \text{out}_{r0}} \text{lean}_p \text{out}_I S_1 \quad (101)$$

$$\text{lean}_p \text{out}_I S_1 \xrightarrow{\text{lean}_p \text{out}_{r1}} \text{lean}_p \text{out}_I S_2 \quad (102)$$

$$\text{lean}_p \text{out}_I S_2 \xrightarrow{\text{lean}_p \text{out}_{r2}} \text{lean}_p \text{out}_I S_3 \quad (103)$$

$$\text{lean}_p \text{out}_I S_3 \xrightarrow{\text{lean}_p \text{out}_{r3}} \text{lean}_p \text{out}_I S_4 \quad (104)$$

$$\text{lean}_p \text{out}_I S_4 \xrightarrow{\text{lean}_p \text{out}_{r4}} \text{lean}_p \text{out}_I S_5 \quad (105)$$

$$\text{lean}_p \text{out}_I S_5 \xrightarrow{\text{lean}_p \text{out}_{r5}} \text{Av} \quad (106)$$

$$\text{Av} \xrightarrow{\text{shunt}_p \text{in}_{r0}} \text{shunt}_p \text{in}_I S_1 \quad (107)$$

$$\text{shunt}_p \text{in}_I S_1 \xrightarrow{\text{shunt}_p \text{in}_{r1}} \text{shunt}_p \text{in}_I S_2 \quad (108)$$

$$\text{shunt}_p \text{in}_I S_2 \xrightarrow{\text{shunt}_p \text{in}_{r2}} \text{shunt}_p \text{in}_I S_3 \quad (109)$$

$$\text{shunt}_p \text{in}_I S_3 \xrightarrow{\text{shunt}_p \text{in}_{r3}} \text{shunt}_p \text{in}_I S_4 \quad (110)$$

$$\text{shunt}_p \text{in}_I S_4 \xrightarrow{\text{shunt}_p \text{in}_{r4}} \text{shunt}_p \text{in}_I S_5 \quad (111)$$

$$\text{shunt}_p \text{in}_I S_5 \xrightarrow{\text{shunt}_p \text{in}_{r5}} \text{shunt}_p \quad (112)$$

$$\text{shunt}_p \xrightarrow{\text{shunt}_p \text{out}_{r0}} \text{shunt}_p \text{out}_I S_1 \quad (113)$$

$$\text{shunt}_p \text{out}_I S_1 \xrightarrow{\text{shunt}_p \text{out}_{r1}} \text{shunt}_p \text{out}_I S_2 \quad (114)$$

$$\text{shunt}_p \text{out}_I S_2 \xrightarrow{\text{shunt}_p \text{out}_{r2}} \text{shunt}_p \text{out}_I S_3 \quad (115)$$

$$\text{shunt}_p \text{out}_I S_3 \xrightarrow{\text{shunt}_p \text{out}_{r3}} \text{shunt}_p \text{out}_I S_4 \quad (116)$$

$$\text{shunt}_p \text{out}_I S_4 \xrightarrow{\text{shunt}_p \text{out}_{r4}} \text{shunt}_p \text{out}_I S_5 \quad (117)$$

$$\text{shunt}_p \text{out}_I S_5 \xrightarrow{\text{shunt}_p \text{out}_{r5}} \text{Av} \quad (118)$$

$$Av \xrightarrow{\text{viscera}_p \text{in}_{r0}} \text{viscera}_p \text{in}_I S_1 \quad (119)$$

$$\text{viscera}_p \text{in}_I S_1 \xrightarrow{\text{viscera}_p \text{in}_{r1}} \text{viscera}_p \text{in}_I S_2 \quad (120)$$

$$\text{viscera}_p \text{in}_I S_2 \xrightarrow{\text{viscera}_p \text{in}_{r2}} \text{viscera}_p \text{in}_I S_3 \quad (121)$$

$$\text{viscera}_p \text{in}_I S_3 \xrightarrow{\text{viscera}_p \text{in}_{r3}} \text{viscera}_p \text{in}_I S_4 \quad (122)$$

$$\text{viscera}_p \text{in}_I S_4 \xrightarrow{\text{viscera}_p \text{in}_{r4}} \text{viscera}_p \text{in}_I S_5 \quad (123)$$

$$\text{viscera}_p \text{in}_I S_5 \xrightarrow{\text{viscera}_p \text{in}_{r5}} \text{viscera}_p \quad (124)$$

$$\text{viscera}_p \xrightarrow{\text{viscera}_p \text{out}_{r0}} \text{viscera}_p \text{out}_I S_1 \quad (125)$$

$$\text{viscera}_p \text{out}_I S_1 \xrightarrow{\text{viscera}_p \text{out}_{r1}} \text{viscera}_p \text{out}_I S_2 \quad (126)$$

$$\text{viscera}_p \text{out}_I S_2 \xrightarrow{\text{viscera}_p \text{out}_{r2}} \text{viscera}_p \text{out}_I S_3 \quad (127)$$

$$\text{viscera}_p \text{out}_I S_3 \xrightarrow{\text{viscera}_p \text{out}_{r3}} \text{viscera}_p \text{out}_I S_4 \quad (128)$$

$$\text{viscera}_p \text{out}_I S_4 \xrightarrow{\text{viscera}_p \text{out}_{r4}} \text{viscera}_p \text{out}_I S_5 \quad (129)$$

$$\text{viscera}_p \text{out}_I S_5 \xrightarrow{\text{viscera}_p \text{out}_{r5}} Av \quad (130)$$

$$Av \xrightarrow{\text{white}_p \text{in}_{r0}} \text{white}_p \text{in}_I S_1 \quad (131)$$

$$\text{white}_p \text{in}_I S_1 \xrightarrow{\text{white}_p \text{in}_{r1}} \text{white}_p \text{in}_I S_2 \quad (132)$$

$$\text{white}_p \text{in}_I S_2 \xrightarrow{\text{white}_p \text{in}_{r2}} \text{white}_p \text{in}_I S_3 \quad (133)$$

$$\text{white}_p \text{in}_I S_3 \xrightarrow{\text{white}_p \text{in}_{r3}} \text{white}_p \text{in}_I S_4 \quad (134)$$

$$\text{white}_p \text{in}_I S_4 \xrightarrow{\text{white}_p \text{in}_{r4}} \text{white}_p \text{in}_I S_5 \quad (135)$$

$$\text{white}_p \text{in}_I S_5 \xrightarrow{\text{white}_p \text{in}_{r5}} \text{white}_p \quad (136)$$

$$\text{white}_p \xrightarrow{\text{white}_p \text{out}_{r0}} \text{white}_p \text{out}_I S_1 \quad (137)$$

$$\text{white}_p \text{out}_I S_1 \xrightarrow{\text{white}_p \text{out}_{r1}} \text{white}_p \text{out}_I S_2 \quad (138)$$

$$\text{white}_p \text{out}_I S_2 \xrightarrow{\text{white}_p \text{out}_{r2}} \text{white}_p \text{out}_I S_3 \quad (139)$$

$$\text{white}_p \text{out}_I S_3 \xrightarrow{\text{white}_p \text{out}_{r3}} \text{white}_p \text{out}_I S_4 \quad (140)$$

$$\text{white}_p \text{out}_I S_4 \xrightarrow{\text{white}_p \text{out}_{r4}} \text{white}_p \text{out}_I S_5 \quad (141)$$

$$\text{white}_p \text{out}_I S_5 \xrightarrow{\text{white}_p \text{out}_{r5}} Av \quad (142)$$

$$L \xleftrightarrow{\text{rav}} Av \quad (143)$$

```
[47]: # disp.Latex(st.sprintvl(sPK))
```

7.1 Parameters: Map78 Table I

```
[48]: ## Parameters: Volume
Volume={}
Volume['lung'] = 0.6
Volume['Lout'] = 0.3
Volume['Lin'] = 0.3
Volume['viscera'] = 6.2
Volume['lean'] = 39.2
Volume['fat'] = 12.2
Volume['shunt'] = 1e-6
Volume['white'] = 0.0007
Volume['grey'] = 0.0007
Volume['kidney'] = 0.0007

Volume['arterial'] = 1.4
Volume['venous'] = 4.0
```

```
[49]: ## Parameters: Flow
FlowFraction={}
FlowFraction['viscera'] = 0.63
FlowFraction['lean'] = 0.131
FlowFraction['fat'] = 0.04
FlowFraction['shunt'] = 0.199
FlowFraction['white'] = 0.000022
FlowFraction['grey'] = 0.000086
FlowFraction['kidney'] = 0.000432
```

```
[50]: ## Parameters: Partition coefficient
PartitionCoefficient = {}
for pool in pools:
    if pool in ['fat']:
        PartitionCoefficient[pool] = 1.40
    else:
        PartitionCoefficient[pool] = 0.46
print(PartitionCoefficient)
```

```
{'viscera': 0.46, 'lean': 0.46, 'fat': 1.4, 'shunt': 0.46, 'white': 0.46,
'grey': 0.46, 'kidney': 0.46}
```

```
[51]: Q = 6.48/60 # lit/sec Table II
Delay = {}
Delay_min = {}
for blood in ['arterial', 'venous']:
    Delay[blood] = Volume[blood]/Q
    Delay_min[blood] = Delay[blood]/60
    print(f'Delay ({blood}) = {Delay[blood]:.2f} sec = {Delay_min[blood]:0.
→2f} min')
```

```

parameter = {}

## Lung diffusion parameters
parameter['kappa_rav'] = 0.5
parameter['K_L'] = 1
parameter['K_Av'] = 0.05

## Pool parameters
for pool in pools:

    ## Pool Ce:P
    K = f'K_{pool}'
    par = 1/(Volume[pool]*PartitionCoefficient[pool])
    #   frac = VolumeFraction[pool]
    #   frac = 0.5 # FIXME
    parameter[f'{K}_P'] = par

    ## Pool pipes RA components
    for i in range(N_lump+1):
        for inout in ['in','out']:
            kappa = f'kappa_{pool}_p{inout}_r_{i}'
            parameter[kappa] = Q*FlowFraction[pool]

    ## Pool pipes C components
    if N_lump>0:
        for i in range(N_lump):
            for inout in ['in','out']:
                K = f'K_{pool}_p{inout}_IS{i+1}'
                if inout in ['in']:
                    V_i = FlowFraction[pool]*Volume['arterial']/N_lump
                else:
                    V_i = FlowFraction[pool]*Volume['venous']/N_lump
                parameter[K] = (1/V_i)

#print(parameter)

```

Delay (arterial) = 12.96 sec = 0.22 min

Delay (venous) = 37.04 sec = 0.62 min

7.2 Simulation

```

[52]: ## Simulation
t = np.linspace(0,360,1000)
# t = np.linspace(0,1000,1000)
# t = np.linspace(0,1000,1000)
t_ss = np.linspace(0,10000,1000)
t_1 = 120

```

```

## Initial conditions
x_min = 1e-6
X0 = x_min*np.ones(sPK['n_X'])

## Chemostats
chemostats = ['L']
scPK = st.statify(sPK,chemostats=chemostats)
X_chemo = {'L':f'{x_min} + 1*(t<{t_1})'}
print(X_chemo)

## Simulate - steady state
dat_ss = st.sim(sPK,sc=scPK,t=t_ss,X0=X0,parameter=parameter)
X_ss = dat_ss['X'][-1,:]
#print(X_ss)

## Simulate
dat = st.sim(sPK,sc=scPK,t=t,X0=X_ss,parameter=parameter,X_chemo=X_chemo)

```

```
{'L': '1e-06 + 1*(t<120)'}
```

```

[53]: def plotPool(plot_pools = ['white','grey','kidney'],figname='sim',conc=True):
    ## Extract state
    species = sPK['species']
    X = dat['X']

    ## Font
    plt.rcParams.update({'font.size': 18})

    ## Compute concentration, and plot
    for pool in plot_pools:
        if pool in ['L','Av']:
            name = f'{pool}'
            par = f'K_{pool}'
            vol = 1/parameter[par]
        else:
            name = f'{pool}_P'
            vol = Volume[pool]
    #    print(name)
    i_pool = species.index(name)

    if conc:
        y = X[:,i_pool]/vol
    else:
        y = X[:,i_pool]

    plt.plot(t/60,y,label=pool,lw=lw)

    plt.legend()
    plt.grid()
    plt.xlabel('$t$ (min)')

```

```

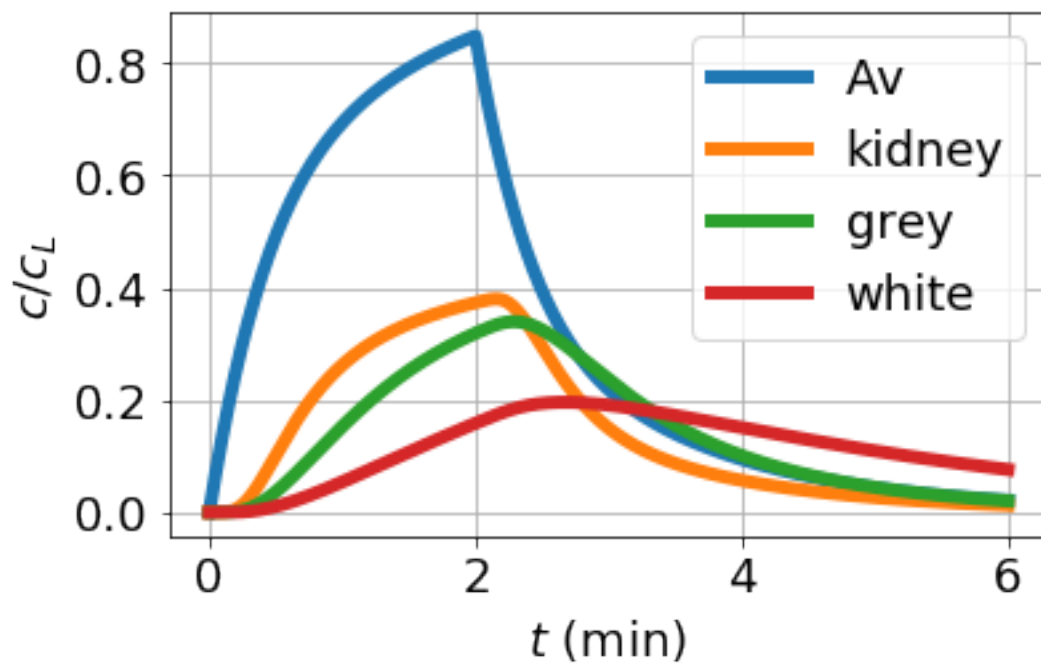
if conc:
    plt.ylabel('$c/c_L$')
else:
    plt.ylabel('$x/x_L$')
plt.tight_layout()
plt.savefig(f'Figs/PK_{figname}.pdf')
plt.show()

```

```

[54]: ## Plot data
plotPool(plot_pools = ['Av', 'kidney', 'grey', 'white'], conc=True)

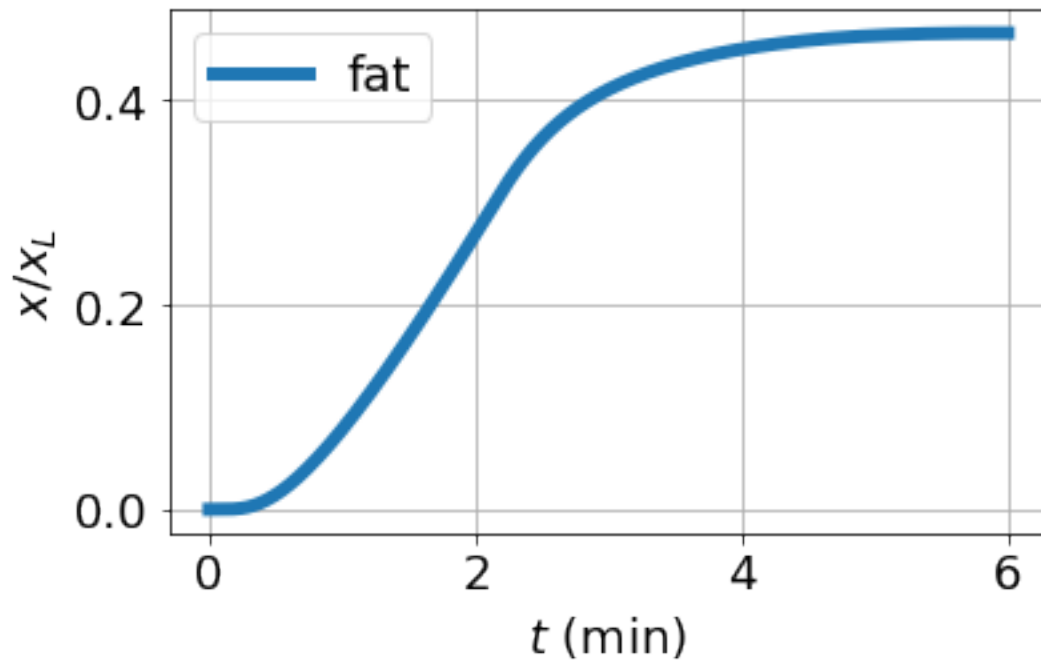
```



```

[55]: plotPool(plot_pools = ['fat'], conc=False, figname='fat')

```



8 Appendix (Material not used in the paper)

8.1 Pure time delay simulation of symmetric case

8.1.1 Numerical model

As, at the moment, a bg component with delay is not available, the case with delay is modelled directly using equations suitable for the [ddeint](#) Python package.

```
[56]: ## Vary the chemostats
def X_chemostat(t,delay):
    x_A1 = 1 + np.heaviside(t-t_0,1)
    x_A2 = 1

    return x_A1,x_A2
```

```
[57]: ## Delay-differential equation solver
from ddeint import ddeint

## Model
def model(X, t, delay=1, Q0=1):
    """
    Model for case with non-zero delay
    See the ddeint Python package
    """

    ## Extract the state and delayed state components
    x_E1, x_C1, x_E2, x_C2 = X(t)
```



```

xd_E1,xd_C1,xd_E2,xd_C2 = X(t-delay)

## Set chemostats
x_A1,x_A2 = X_chemostat(t,delay)

## Upstream flows
f1 = x_E1*x_A1 - x_C1
F_E1 = (x_E1 - xd_E2)*Q0
F_C1 = (x_C1 - xd_C2)*Q0

## Upstream state derivatives
dx_E1 = -f1 - F_E1
dx_C1 = f1 - F_C1

## Downstream flows
f2 = x_C2 - x_E2*x_A2
F_E2 = (x_E2 - xd_E1)*Q0
F_C2 = (x_C2 - xd_C1)*Q0

## Downstream state derivatives
dx_E2 = f2 - F_E2
dx_C2 = -f2 - F_C2

## State derivative
dX = np.array([dx_E1,dx_C1,dx_E2,dx_C2])

return dX

```

8.2 Simulation

```

[58]: ## Main parameters
delay = 5
Q0 = 1

## Initial state (ddeint form)
x0 = (1)*np.array([1,1,1,1])
initial = lambda t: x0

e0 = sum(x0)
print("e0 =",e0)

## Simulate delayed model
XX = ddeint(model, initial, tt, fargs=(delay,Q0,))
XX1 = XX[:,0:4]
e0_sim = sum(XX.T)

## Plot all 8 states
plt.plot(tt, XX)
plt.grid()

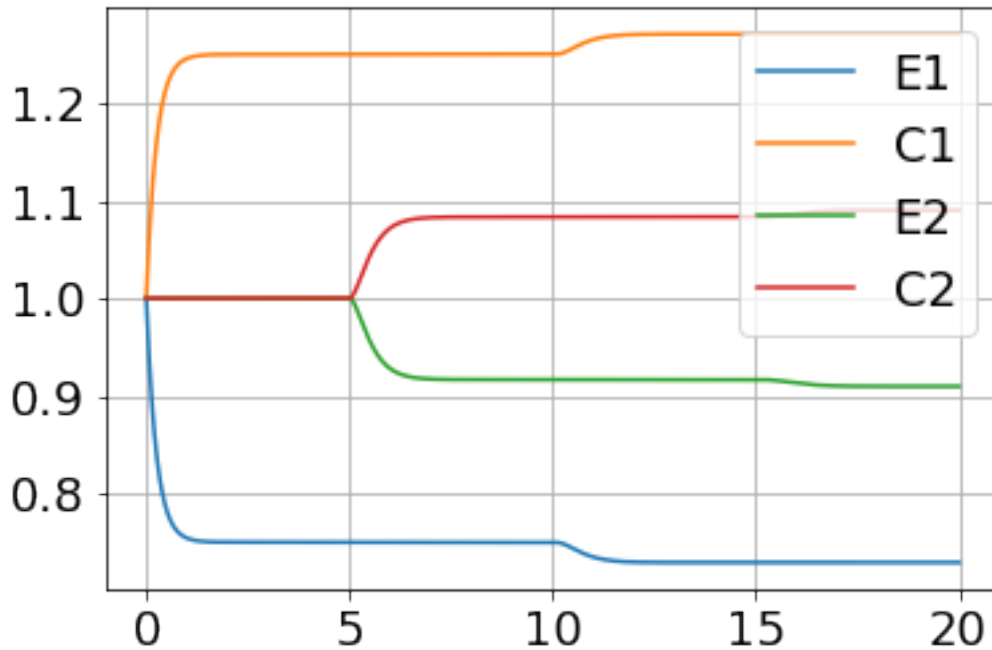
```

```

Legend = ['E1', 'C1', 'E2', 'C2', 'E12', 'C12', 'E21', 'C21']
plt.legend(Legend)
plt.show()
print('Final states:', XX[-1,:])
print('Sum of final states:', sum(XX[-1,:]))

```

e0 = 4



Final states: [0.72916561 1.27083439 0.90972036 1.09027964]

Sum of final states: 4.0

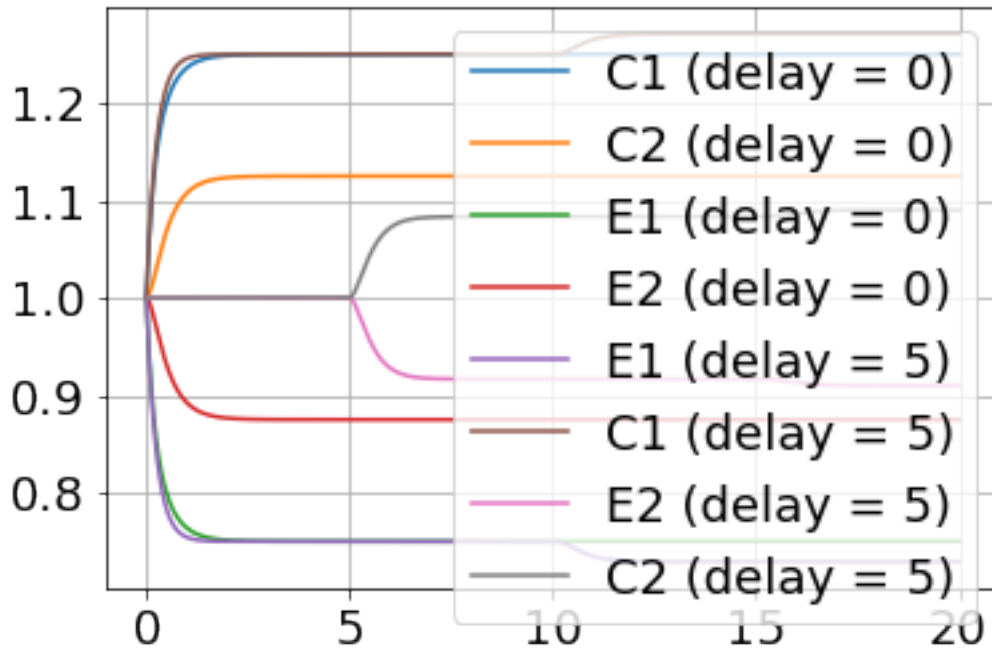
8.2.1 Compare d=0 and d>0

```

[59]: ## Plot states
Delay = ' (delay = '+str(delay)+' )'
Delay0 = ' (delay = 0) '
Legend = []
for i in range(2,6):
    Legend.append(snd['species'][i]+Delay0)
Legend1 = ['E1', 'C1', 'E2', 'C2']
for Leg in Legend1:
    Legend.append(Leg+Delay)
#Legend = snd['species'][2:6]
plt.plot(tt, XX0, tt, XX1)
plt.grid()

plt.legend(Legend)
plt.show()

```



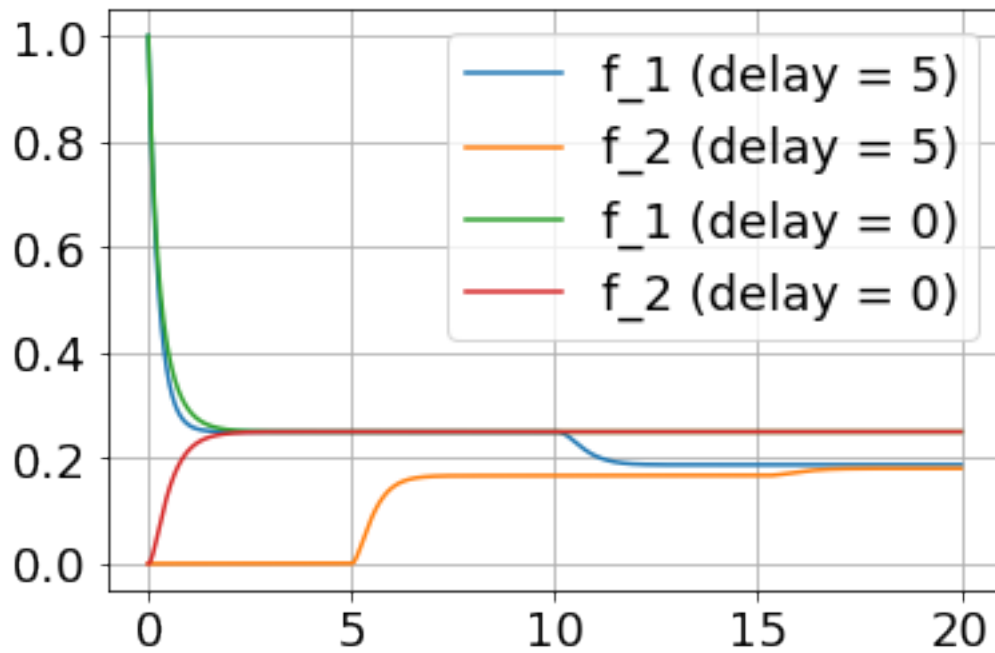
```
[60]: ## Plot flows
x_E1 = XX1[:,0]
x_C1 = XX1[:,1]
x_E2 = XX1[:,2]
x_C2 = XX1[:,3]
x_A,x_B = X_chemostat(tt,delay)
print(x_A,x_B)
f1 = x_E1*x_A - x_C1
f2 = x_C2 - x_E2*x_B

plt.plot(tt,f1, tt,f2, tt,V0)
plt.grid()

Legend = ['f_1'+Delay, 'f_2'+Delay, 'f_1'+Delay0, 'f_2'+Delay0]
plt.legend(Legend)
plt.show()

print('Final flows',f1[-1],f2[-1])
```

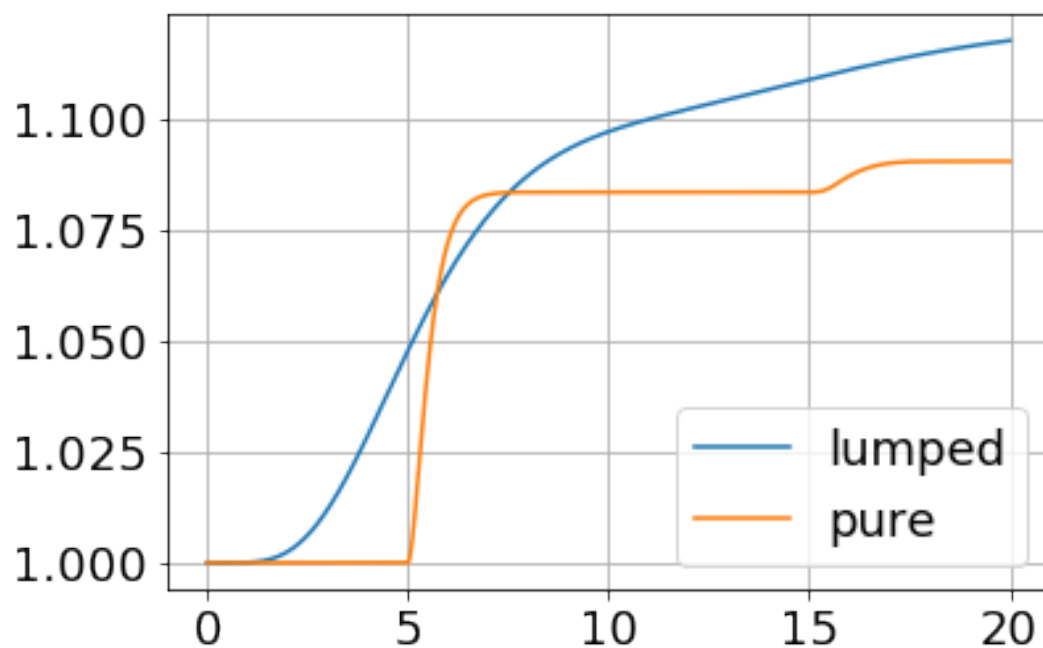
```
[2. 2. 2. ... 2. 2. 2.] 1
```



Final flows 0.1874968285755121 0.18055928632873708

```
[61]: plt.plot(tt,XXPipe[:,1],label='lumped')
plt.plot(tt,XX1[:,3],label='pure')
plt.grid()
plt.legend()
```

[61]: <matplotlib.legend.Legend at 0x7f6d0e024af0>



8.3 Numerical computation of chemical/hydraulic interaction (early version)

```
[62]: imp.reload(st)
R = st.R()
print(f'R = {R:4.2f} J/mol/K')
RT = st.RT()
print(f'RT = {RT/1000:4.2f} kJ/mol')

## Concentration ratio rho (example value)
rho = 2
print(f'rho = {rho}')

## Change in chemical potential
dmu = RT*np.log(rho)
print(f'Delta mu = {dmu:4.2f} J/mol')

## Typical haemoglobin conc
c_gl = 150 # g/l - Bionumbers: Hemoglobin concentration in human blood
mass = 64e3 # Daltons=g/mol - https://en.wikipedia.org/wiki/Hemoglobin
c_moll = c_gl/mass # mol/l
c_molm3 = c_moll*1e3 # mol/m^3
# print(f'concentration: {c_molm3} mol/m^3')
#c = 10 # mol/m3
c = c_molm3 # mol/m3
print(f'c = {c:0.2} mol/m3')
dP = c*dmu
print(f'Delta P = {(dP/1000):0.1f} kPa')

## Typical systolic blood pressure 120mmHg = 16 kPa (https://en.wikipedia.org/
→wiki/Blood_pressure)
BP = 16e3 # Pa
print(f'Typical blood pressure = {BP/1000} kPa')
```

```
R = 8.31 J/mol/K
RT = 2.58 kJ/mol
rho = 2
Delta mu = 1787.44 J/mol
c = 2.3 mol/m3
Delta P = 4.2 kPa
Typical blood pressure = 16.0 kPa
```

```
[ ]:
```

References

W. W. Mapleson. Circulation-time models of the uptake of inhaled anaesthetics and data for quantifying them. *British Journal of Anaesthesia*, 45:319, 1973.