

Physically-Plausible Parameters

Peter Gawthrop. peter.gawthrop@unimelb.edu.au

January 15, 2021

Contents

1	Introduction	1
1.1	Setup modules	1
1.2	Quadratic programming QP.	2
2	Conversion factor	3
3	Extract Model	4
3.1	Extract stoichiometry	4
3.2	Extract reaction potentials Φ and deduce plausible species potentials ϕ	5
3.3	Extracted reactions and reaction potentials	8
4	Deduce Pathway Flows	8
5	Reaction constants (modified mass-action)	9
5.1	Show computed reaction flows	10
5.2	Show computed chemostat flows	12
5.3	Show pathway flows	12
6	Species constants	13

1 Introduction

This note illustrates an approach to fitting the parameters of a bond graph model to experimental data. Insofar as the parameters are associated with a bond graph, they are *physically-plausible* [Gawthrop et al. \(2020\)](#).

The approach uses a bond-graph derived from a stoichiometric model of *e.coli* [Orth et al. \(2010\)](#) (using a method described elsewhere [Gawthrop \(2020\)](#)) combined with experimental values of *reaction potential*, *reaction flux* and *species concentration* from the literature [Park et al. \(2016\)](#).

1.1 Setup modules

```
[1]: ## Paths
NeedPath=True
if NeedPath:
    import sys
    sys.path += ['/usr/lib/python3/dist-packages']
```

```
[2]: ## Maths library
import numpy as np
import scipy

## BG tools
import BondGraphTools as bgt

## SVG bond graph
import svgBondGraph as sbg

## BG stoichiometric utilities
import stoich as st

## Modular bond graphs
import modularBondGraph as mbg

## Stoichiometric conversion
import CobraExtract as Extract
import stoichBondGraph as stbg

## Potentials
import phiData

## Faraday constant
import scipy.constants as con
F = con.physical_constants['Faraday constant'][0]

## Display
import IPython.display as disp

## Plotting
import matplotlib.pyplot as plt

import copy

## Allow output from within functions
from IPython.core.interactiveshell import InteractiveShell
InteractiveShell.ast_node_interactivity = "all"

import importlib as imp

quiet = True
showMu=True
```

1.2 Quadratic programming QP.

$$\text{minimise } \frac{1}{2}x^T Px + q^T x \quad (1)$$

$$\text{subject to } Gx \leq h \quad (2)$$

$$\text{and } Ax = b \quad (3)$$

In the case considered here, there is no equality constraint and

$$x = \hat{\phi} \quad (4)$$

$$P = NN^T + \mu I_{n_X \times n_X} \quad (5)$$

$$q = (N\Phi)^T \quad (6)$$

$$G = N^T \quad (7)$$

$$h = -\Phi_{min} \quad (8)$$

$\mu > 0$ is required to give a convex QP: in essence it turns a non-unique solution for ϕ into a minimum norm solution.

```
[3]: ## Quadratic programming stuff.
import quadprog

## Function from https://scaron.info/blog/quadratic-programming-in-python.html
def quadprog_solve_qp(P, q, G=None, h=None, A=None, b=None):
    qp_G = .5 * (P + P.T) # make sure P is symmetric
    qp_a = -q
    if A is not None:
        qp_C = -numpy.vstack([A, G]).T
        qp_b = -numpy.hstack([b, h])
        meq = A.shape[0]
    else: # no equality constraint
        qp_C = -G.T
        qp_b = -h
        meq = 0
    return quadprog.solve_qp(qp_G, qp_a, qp_C, qp_b, meq)[0]

## Function to compute phi from Phi subject to Phi>positive number
## NN Reduced N corresponding to known Phi
def quadsolve_phi(N0,N1,Phi0,Phi_min=0.0,mu=1e-10):

    (n_X,n_V) = N1.shape
    print(N1.shape)
    P = 1.0*N0@(N0.T) + mu*np.eye(n_X)
    q = (N0@Phi0).T
    G = 1.0*N1.T
    h = -Phi_min*np.ones((n_V))
    phi = quadprog_solve_qp(P, q, G=G, h=h)
    #Phi = -N.T@phi

    return phi
```

2 Conversion factor

```
[4]: Factor = st.F()/1e6
print(f'To convert from kJ/mol to mV, divide by {1/Factor:4.3}')
```

To convert from kJ/mol to mV, divide by 10.4

3 Extract Model

This example uses the Glycolysis and Pentose Phosphate pathways.

Notes:

- Reactions RPI, PGK and PGM are reversed to correspond to positive flows.
- The resultant stoichiometric matrix N relates reaction flows (f) to species flows (\dot{x}):

$$\dot{x} = Nf \quad (9)$$

3.1 Extract stoichiometry

```
[5]: sm = Extract.extract(cobraname='textbook',Remove=['_C','_'],
                        negReaction=['RPI','PGK','PGM'], quiet=quiet)
```

```
Extracting stoichiometric matrix from: textbook
Cobra Model name: e_coli_core BondGraphTools name: e_coli_core_abg
Extract.Integer only handles one non-integer per reaction
Multiplying reaction BIOMASS_ECOLIORE ( 12 ) by 0.6684491978609626 to avoid non-integer species 3PG ( 2 )
Multiplying reaction CYTBD ( 15 ) by 2.0 to avoid non-integer species 02 ( 55 )
Multiplying reaction PGK ( 54 ) by -1
Multiplying reaction PGM ( 56 ) by -1
Multiplying reaction RPI ( 65 ) by -1
```

```
[6]: name = 'GlyPPP_abg'
    reaction = []

    ## Glycolysis
    reaction += ['PGI','PFK','FBA','TPI']

    ## Pentose Phosphate
    reaction += ['G6PDH2R','PGL','GND','RPI','TKT2','TALA','TKT1','RPE']

    ss = Extract.choose(sm,reaction=reaction)

    ## Create BG
    ss['name'] = name
    stbg.model(ss)
    import GlyPPP_abg
    imp.reload(GlyPPP_abg)
    s = st.stoich(GlyPPP_abg.model(),quiet=quiet)
```

```
[6]: <module 'GlyPPP_abg' from
    '/home/peterg/WORK/Research/SystemsBiology/Notes/2021/Parameter/GlyPPP_abg.
    ↪py'>
```

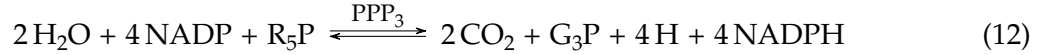
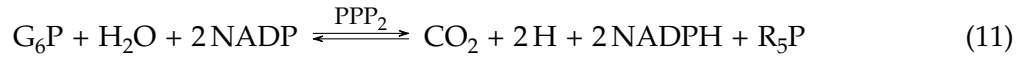
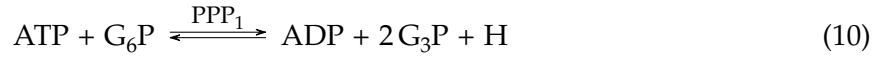
```
[7]: ## Set up chemostats
    chemostats = ['ADP','ATP','H','H2O','NADP','NADPH','CO2']
    chemostats += ['G6P','G3P','R5P']
    chemostats.sort()
```

```
print(chemostats)
sc = st.statify(s,chemostats=chemostats)

sp = st.path(s,sc,pathname='PPP')
print(st.sprintp(sc))
disp.Latex(st.sprintrl(sp,chemformula=True))
```

```
['ADP', 'ATP', 'CO2', 'G3P', 'G6P', 'H', 'H2O', 'NADP', 'NADPH', 'R5P']
3 pathways
0: + PGI + PFK + FBA + TPI
1: + G6PDH2R + PGL + GND + RPI
2: - 2 PGI + 2 G6PDH2R + 2 PGL + 2 GND + TKT2 + TALA + TKT1 + 2 RPE
```

[7]:



```
[8]: print(st.sprintrl(sc,'K'))
disp.Latex(st.sprintrl(sc,'K'))
```

```
\begin{align}
K &= \\
\left(\begin{matrix} 1 & 0 & -2 \\ 1 & 0 & 0 \\ 1 & 0 & 0 \\ 1 & 0 & 0 \\ 0 & 1 & 2 \\ 0 & 1 & 2 \\ 0 & 1 & 2 \\ 0 & 1 & 0 \\ 0 & 0 & 1 \\ 0 & 0 & 1 \\ 0 & 0 & 1 \\ 0 & 0 & 2 \end{matrix}\right) \\
&\end{align}
```

[8]:

$$K = \begin{pmatrix} 1 & 0 & -2 \\ 1 & 0 & 0 \\ 1 & 0 & 0 \\ 1 & 0 & 0 \\ 0 & 1 & 2 \\ 0 & 1 & 2 \\ 0 & 1 & 2 \\ 0 & 1 & 0 \\ 0 & 0 & 1 \\ 0 & 0 & 1 \\ 0 & 0 & 1 \\ 0 & 0 & 2 \end{pmatrix} \quad (13)$$

3.2 Extract reaction potentials Φ and deduce plausible species potentials ϕ .

Because of the energetic constraints implied by the bond graph, the reaction potentials Φ are related to the species potentials ϕ by

$$\Phi = -N^T \phi \quad (14)$$

Typically, there are more species than reactions and so N has more rows than columns. Given the reaction potentials Φ , the species potentials can be estimated using the *pseudo inverse* N^\dagger of $-N^T$:

$$\hat{\phi} = N^\dagger \Phi \quad (15)$$

Notes:

- In general $\hat{\phi} \neq \phi$ but is physically plausible insofar as $-N^T \hat{\phi} = \Phi$.

```
[9]: def getPhi(s,Phi_hyd=0.5,phi_6PGL=None,quadprog=False):
    """Extract phi for given system using
    Reaction potentials from ParRubXu16"""

    ## Reaction potentials from ParRubXu16
    PHI = phiData.Phi_ParRubXu16_Measured()

    # Phenotype = 'Mammalian'
    # Phenotype = 'Yeast'
    Phenotype = 'Ecoli'
    Phi_reac = PHI[Phenotype]

    Phi = np.zeros((len(s['reaction']),1))
    N = copy.copy(s['N'])
    N_0 = None
    N_1 = None
    Phi_0 = []
    for j, reac in enumerate(s['reaction']):
        if (reac in Phi_reac.keys()) and not np.isnan(Phi_reac[reac]):
            Phi_0.append(Phi_reac[reac])
            if N_0 is None:
                N_0 = N[:,j]
            else:
                N_0 = np.vstack((N_0,N[:,j]))
        else:
            if N_1 is None:
                N_1 = N[:,j]
            else:
                N_1 = np.vstack((N_1,N[:,j]))

    Phi_0 = np.array(Phi_0)
    #print(N_1)

    ## Compute Phi
    N_0 = N_0.T
    N_1 = N_1.T

    n_X,n_V = N_0.shape
    print(f'Extracting {n_X} values of phi from {n_V} values of Phi')

    if quadprog:
        phi = quadsolve_phi(N_0,N_1,Phi_0,Phi_min=1e-3,mu=1e-10)
    else:
```

```

    ## Compute Phi using pseudo inverse
    pinvNT = scipy.linalg.pinv(N_0.T)
    phi = -pinvNT@Phi_0

    if phi_6PGL is not None:
        ## Reset 6PGL
        i_6PGL = s['species'].index('6PGL')
        phi[i_6PGL] = phi_6PGL
        print (f'Resetting phi_6GPL to {int(1e3*phi[i_6PGL])} mV' )

    ## Sanity check
    Phi_new = -N_0.T@phi
    err = np.linalg.norm(Phi_new-Phi_0)
    print(f'Phi error = {int(err*1000)}mV\n')

    Phi = -N.T@phi

    return Phi,phi,Phi_0,Phi_reac

```

```

[10]: Phi_,phi_est_,Phi_0,Phi_reac_ = getPhi(s,quadprog=False)
print('Minimum Phi = ', int(round(np.min(1e3*Phi_))), 'mV')

```

Extracting 19 values of phi from 10 values of Phi

Phi error = 0mV

Minimum Phi = -3 mV

```

[11]: Phi,phi_est,Phi_0,Phi_reac = getPhi(s,quadprog=True)
print('Minimum Phi = ', int(round(np.min(1e3*Phi))), 'mV')

print('\nChange in phi')
for i,spec in enumerate(s['species']):
    change = int(1e3*(phi_est[i]-phi_est_[i]))
    if not change==0:
        print(f'{i} {spec}\t {change}')

print('\nChange in Phi')
for i, reac in enumerate(s['reaction']):
    change = int(round(1e3*(Phi[i]-Phi_[i])))
    if not change == 0:
        print(f'{i} {reac}\t {change} {int(round(1e3*Phi[i]))}␣
→{int(round(1e3*Phi_[i]))}')

```

Extracting 19 values of phi from 10 values of Phi

(19, 2)

Phi error = 0mV

Minimum Phi = 0 mV

Change in phi

1 6PGL 1
12 H2O 1

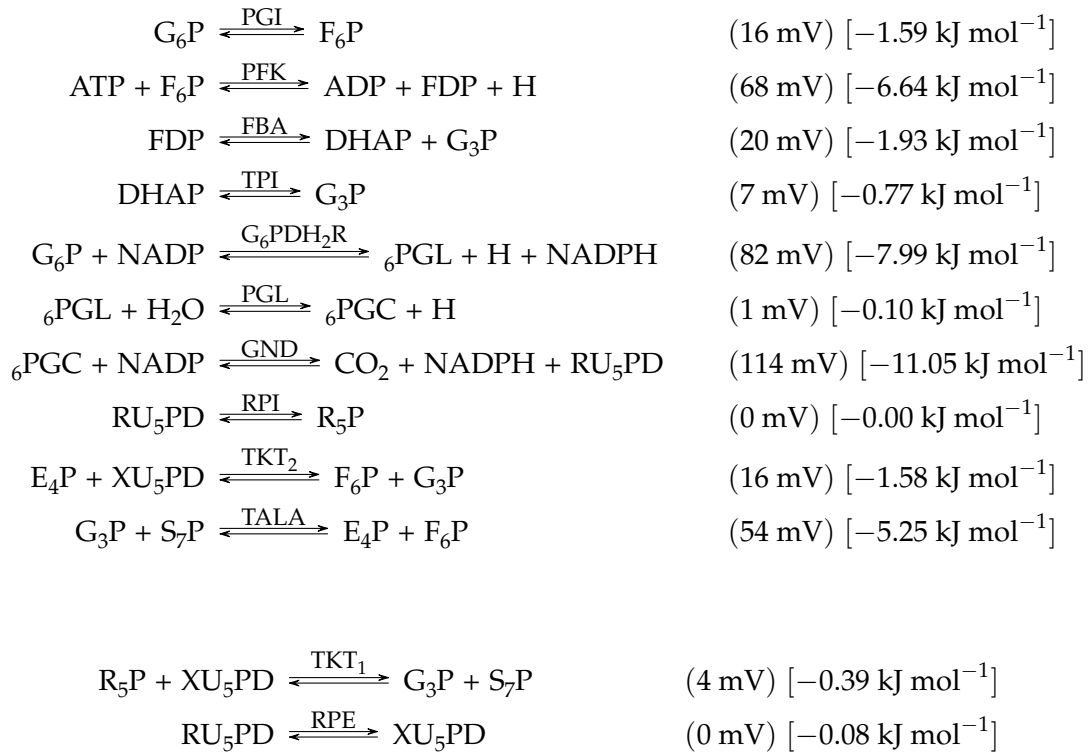
Change in Phi

5 PGL 4 1 -3

3.3 Extracted reactions and reaction potentials

```
[12]: disp.Latex(st.sprintrl(s,chemformula=True,Phi=Phi,units=['mV','kJ'],
    ↪,showMu=showMu))
```

[12]:



4 Deduce Pathway Flows

From basic stoichiometric analysis, steady-state flows can be written as:

$$f = K_p f_p \quad (16)$$

$$\text{where } K_p N^{cd} = 0 \quad (17)$$

Note that the *pathway matrix* K_p is dependent on the choice of chemostats.

Given a set of experimental flows f , an estimate \hat{f}_p of f_p can be obtained from the *least-squares* formula:

$$(K_p^T K_p) \hat{f}_p = K_p^T f \quad (18)$$

Notes:

- v_p is a n_p vector containing the pathways flows
- $(K_p^T K_p)$ is a square $n_p \times n_p$ matrix where n_p is the number of pathways

- If some flows are not measured, the corresponding rows of K_p are deleted
- the reaction flows (including the missing ones) can be estimated from $\hat{f} = K_p \hat{f}_p$.
- the estimated chemostat flows are given by the non-zero elements of

$$\hat{x} = N\hat{f} \quad (19)$$

[13]: `def PathwayFlux(K, reaction, Reaction, flux):`

```

    #KK = st.singleRemove(K)
    KK = K
    Kp = None
    Flux = {}
    reac_known = []
    #flux = phiData.ParRubXu16_flux()
    for i, reac in enumerate(reaction):
        if reac in flux.keys():
            reac_known.append(reac)
            fi = flux[reac]
            #Ki = np.abs(KK[i,:])
            Ki = KK[i,:]
            #print(reac, Ki)
            if Kp is None:
                Kp = Ki
                f = fi
            else:
                Kp = np.vstack((Kp, Ki))
                f = np.vstack((f, fi))
    #print(Kp)

    if Kp is not None:
        #print(f)
        f0 = np.linalg.solve(Kp.T@Kp, Kp.T@f)
        for i, Reac in enumerate(Reaction):
            Flux[Reac] = f0[i][0]
        #print(f0)
        f_est = Kp@f0
        #print(Kp@f0-f)

    error = np.linalg.norm(f_est-f)/len(f)
    print(f'Flux error = {error:.2e}')

    return Flux, f0, f_est, f, reac_known

```

5 Reaction constants (modified mass-action)

The modified mass-action formula is [Gawthrop et al. \(2020\)](#):

$$f = \kappa \left(\exp \frac{\Phi^f}{\alpha V_N} - \exp \frac{\Phi^r}{\alpha V_N} \right) \quad (20)$$

Thus an estimate for κ can be computed as:

$$\hat{\kappa} = \frac{\hat{f}}{f_0} \quad (21)$$

$$\text{where } f_0 = \left(\exp \frac{\Phi^f}{\alpha V_N} - \exp \frac{\Phi^r}{\alpha V_N} \right) \quad (22)$$

```
[14]: def reactionConstant(s,phi_est,f_est,alpha=1):

    V_N = st.V_N()

    ## Extract stoichiometry
    N = s['N']
    Nf = s['Nf']
    Nr = s['Nr']
    reaction = s['reaction']

    ## Compute Phis from estimated phi
    Phi_ = -N.T@phi_est
    Phi_f = Nf.T@phi_est
    Phi_r = Nr.T@phi_est

    ## Compute normalised flow rates
    f0 = (np.exp(Phi_f/(alpha*V_N)) - np.exp(Phi_r/(alpha*V_N)))
    #print(f0)
    parameter = {}
    for i,react in enumerate(reaction):
        kap = f_est[i][0]/f0[i]
        parameter[f'kappa_{react}'] = kap
        #print(f'{react}: \tPhi = {int(Phi_[i]*1000)}mV, \tf_est = \t{f_est[i][0]:.2e}, \tkappa = {kap:.2}')
    return parameter
```

5.1 Show computed reaction flows

```
[15]: Reaction = ['PPP1','PPP2','PPP3']
#print(s['reaction'])
## Reaction flows
Flux = phiData.ParRubXu16_flux()
flux = Flux['Ecoli']

## Normalise flux wrt PGI = 100
flux_PGI = flux['PGI']
for reac in flux.keys():
    flux[reac] *= 100/flux_PGI

fluxp,f0,f_est,f,reaction = PathwayFlux(sc['K'],s['reaction'],Reaction,flux)
```

```

## Reaction constants
f_est = sc['K']@f0
parameter = reactionConstant(s,phi_est,f_est)

#f_est = sc['K']@f0
j=0

print('\n\n% LaTeX table')
print('\nhline')
print('Reaction &t $ \Phi$ &t $ \hat{\Phi}$ &t $f$ & $ \hat{f}$ &
→$ \hat{\kappa}$\')
print('\nhline')
for i, reac in enumerate(s['reaction']):
    if reac in flux.keys():
        ff = f'{int(round(f[j][0]))}'
        j += 1
    else:
        ff = '--'
    if reac in Phi_reac.keys():
        PP = f'{int(round(1e3*Phi_reac[reac]))}'
    else:
        PP = '--'
    kappa = 'kappa_'+reac
    print(f'{reac} &t {PP} &t {int(round(1e3*Phi[i]))} &t {ff} &
→{int(round(f_est[i][0]))} & {parameter[kappa]:.2} \')
print('\nhline')

```

Flux error = 3.10e-01

```

%% LaTeX table
\hline
Reaction &          $ \Phi$ &          $ \hat{\Phi}$ &  $f$ & $ \hat{f}$ &
$ \hat{\kappa}$\
\hline
PGI &    16 &    16 &    100 & 99 & 2.6e+02 \
PFK &    69 &    69 &    104 & 105 & 9.1e+01 \
FBA &    20 &    20 &    106 & 105 & 2.7e+02 \
TPI &     8 &     8 &    105 & 105 & 5.9e+02 \
G6PDH2R &    -- &    -- &    83 &  -- & 19 & 7.8 \
PGL &    -- &    1 &    -- & 19 & 4.9e+02 \
GND &   115 &   115 &    19 & 19 & 2.1 \
RPI &     0 &     0 &    13 & 13 & 7e+03 \
TKT2 &    16 &    16 &     2 & 3 & 1.5e+01 \
TALA &    54 &    54 &    -- & 3 & 2.8 \
TKT1 &     4 &     4 &     5 & 3 & 1.5e+01 \
RPE &     1 &     1 &     6 & 6 & 1.6e+02 \
\hline

```

5.2 Show computed chemostat flows

```
[16]: dx_est = s['N']@f_est

print('\n\n%% LaTeX table')
print('\nhline')
print('Chemostat &\t flow \\\\'')
print('\nhline')
for i,spec in enumerate(s['species']):
    if spec in chemostats:
        print(f'{spec} &\t {int(round(dx_est[i][0]))} \\\\'')
print('\nhline')
```

```
%% LaTeX table
\hline
Chemostat &      flow \\\
\hline
ADP &      105 \\\
ATP &     -105 \\\
CO2 &       19 \\\
G3P &      213 \\\
G6P &     -119 \\\
H &       144 \\\
H2O &     -19 \\\
NADP &     -39 \\\
NADPH &      39 \\\
R5P &       10 \\\
\hline
```

5.3 Show pathway flows

```
[17]: print('\n\n%% LaTeX table')
print('\nhline')
print('Pathway &\t  $\hat{f}_p$  \\\\'')
print('\nhline')
for reac in fluxp.keys():
    print(f'{reac} &\t {int(round(fluxp[reac]))} \\\\'')
print('\nhline')
```

```
%% LaTeX table
\hline
Pathway &       $\hat{f}_p$  \\\
\hline
PPP1 &      105 \\\
PPP2 &       13 \\\
PPP3 &        3 \\\
\hline
```

6 Species constants

$$K = \frac{\exp \phi}{x^{\circ}} = \frac{\exp \phi}{V c^{\circ}} \quad (23)$$

```
[18]: #imp.reload(phiData)

print('\n\n% LaTeX table')
print('\nhline')
print('Species & \t $\hat{\phi}^mV$ & $c$ & $\hat{K}$ \\\')
print('\nhline')

concentration = phiData.ParRubXu16_conc()
#concentration['H'] = 1e-7

for i,spec in enumerate(s['species']):
    if spec in concentration.keys():
        conc = 1e3*concentration[spec]
        K_spec = np.exp(phi_est[i])/conc
        print(f'{spec} & {int(round(1e3*phi_est[i]))} & \t{conc:.2} & \t{K_spec:.4} \\\')
    # else:
    #     print(f'{spec} & {phi_est[i]:.2} & -- & -- \\\')

print('\nhline')
```

```
%% LaTeX table
\hline
Species &          $\hat{\phi}^mV$ & $c$ & $\hat{K}$ \\\
\hline
6PGC & 29 & 0.017 & 62.4 \\\
ADP & -27 & 0.57 & 1.711 \\\
ATP & 27 & 4.7 & 0.22 \\\
CO2 & -30 & 7.6 & 0.1272 \\\
DHAP & -10 & 1.6 & 0.6075 \\\
E4P & -27 & 0.01 & 94.47 \\\
F6P & -21 & 0.097 & 10.1 \\\
FDP & -8 & 1.5 & 0.6528 \\\
G3P & -18 & 0.14 & 6.967 \\\
G6P & -5 & 0.68 & 1.474 \\\
NADP & 30 & 0.028 & 36.29 \\\
NADPH & -30 & 0.065 & 14.83 \\\
R5P & 5 & 0.028 & 35.4 \\\
RU5PD & 5 & 0.0053 & 190.8 \\\
S7P & 24 & 0.018 & 56.58 \\\
XU5PD & 5 & 0.03 & 33.6 \\\
\hline
```

References

- Peter J Gawthrop. Energy-based Feedback Control of Biomolecular Systems with Cyclic Flow Modulation. Available at arXiv:2007.14762, July 2020.
- Peter J. Gawthrop, Peter Cudmore, and Edmund J. Crampin. Physically-plausible modelling of biomolecular systems: A simplified, energy-based model of the mitochondrial electron transport chain. *Journal of Theoretical Biology*, 493:110223, 2020. ISSN 0022-5193. doi: 10.1016/j.jtbi.2020.110223.
- J. Orth, R. Fleming, and B. Palsson. Reconstruction and use of microbial metabolic networks: the core escherichia coli metabolic model as an educational guide. *EcoSal Plus*, 2010. doi: 10.1128/ecosalplus.10.2.1.
- Junyoung O. Park, Sara A. Rubin, Yi-Fan Xu, Daniel Amador-Noguez, Jing Fan, Tomer Shlomi, and Joshua D. Rabinowitz. Metabolite concentrations, fluxes and free energies imply efficient enzyme usage. *Nat Chem Biol*, 12(7):482–489, Jul 2016. ISSN 1552-4450. doi: 10.1038/nchembio.2077.