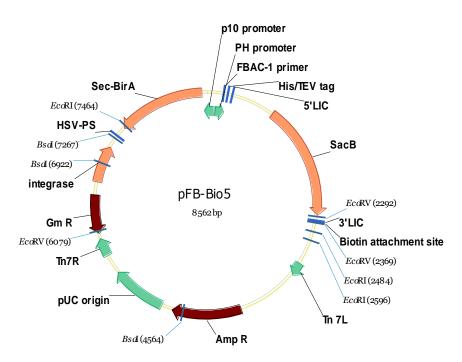
Vector information sheet.

Vector Name	pFB-Bio5
Source	Opher Gileadi and Claire Strain-Damerell
Sequence accession/link	

Description	Baculovirus transfer vector with His ₆ tag in 22-aa N-terminal fusion peptide, with TEV protease cleavage site and C-terminal biotin attachment site. The vector also includes a secreted
	version of <i>birA</i> under the control of the p10 promoter for <i>in vivo</i> biotinylation. Includes sites for LIC cloning, and a "stuffer"
	fragment that includes the SacB gene, allowing negative
	selection of transformed bacteria on 5% sucrose.

Antibiotic resistance	Ampicillin, 100 μg/ml				
Promoter	Polyhedrin				
Cloning	LIC. (vector treated with BseRI, then with T4 DNA polymerase in presence of dGTP)				
Initiation codon	Supplied in PCR primer				
N-terminal fusion – seq.	MGHHHHHHSSGVDLGTENLYFQ*SM (* - TEV cleavage site)				
N-terminal fusion – MW	2630 Da including Met (2411.8 Da removed by TEV cleavage)				
C-terminal fusion – seq.	SSKGGYGLNDIFEAQKIEWHE				
C-terminal fusion – MW	2408.6 Da				
Termination codons	supplied in PCR primer				
Protease cleavage	TEV (cleaves His-tag only)				
Additional features	Tn7 sequences for in vivo recombination into bacmid DNA in DH10Bac (using InVitrogen's Bac-to-bac system).				
Preferred host	Initial transformation into any cloning strain, then transform purified plasmid into DH10Bac to generate recombinant bacmid DNA				
5' sequencing primer	FBAC1: TATTCATACCGTCCCACCA				
3' sequencing primer	FBAC2: GGGAGGTTTTTTAAAGCAAGTAAA				



Opher Gileadi and Claire Strain-Damerell, Structural Genomics Consortium (SGC), Oxford. Email: opher.gileadi@sgc.ox.ac.uk / claire.strain-damerell@sgc.ox.ac.uk

Cloning region in the vector:

Fbacl AATAAAAAAA CCTATAAATA TTCCGGATTA TTCATACCGT CCCACCATCG GGCGCGCTTA TTATTTTTTT GGATATTAT AAGGCCTAAT AAGTATGGCA GGGTGGTAGC CCGCGCGAAT M G H H H H H H S S G V D L G AGAAGGAGAT ATACTATGGG CCACCATCAT CATCATCATT CTTCTGGTGT AGATCTGGGT TCTTCCTCTA TATGATACCC GGTGGTAGTA GTAGTAAG AAGACCACA TCTAGACCCA BSeRI T E N L Y F Q 301 ACCGAGAACC TGTACTTCCA ATCCATAAGC TAGCTTCTCC TCCTGAAAGA TCCATAACTT TGGCTCTTGG ACATGAAGGT TAGGTATTCG ATCGAAGAGG AGGACTTTCT AGGTATTGAA LIC5' --Sacb linker--

				BseRI			
				~~~~~			
2281	AAAATGCCGA	TATCCTATTG	GCATTGACGT	CAGGTGGCAC	TTTTCGAGGA	GTTTACTAGT	
	TTTTACGGCT	ATAGGATAAC	CGTAACTGCA	GTCCACCGTG	AAAAGCTCCT	CAAATGATCA	
	S K G	G Y G I	LNDI	FEA	OKII	E W H E ·	
2341	AAGTAAAGGT	GGATACGGCC	TGAATGATAT	CTTTGAAGCG	CAGAAGATTG	AATGGCATGA	
2011	1110111111001	CCTATGCCGG					
	TICATITECA	CCIAIGCCGG	ACTIACIATA	GAAACIICGC	GICTICIAAC	IIACCGIACI	
	птсэ						
	. * *						
0401		03 03 3 00 mom	3.0000003.300	000000000000000000000000000000000000000	03 03 mommoo	3 3 CCEC3 C3 3	
2401		GAGAACCTCT					
	TACTATTCGT	CTCTTGGAGA	TGAAGGTTAG	CCCAGACTTG	CTGTAGAAGC	TTCGAGTCTT	
2461	AATTGAATGG	CACTGAGGAT	CCGAATTCGA	GCTCCGTCGA	CAATTGGGCA	GAGAACCTCT	
	TTAACTTACC	GTGACTCCTA	GGCTTAAGCT	CGAGGCAGCT	GTTAACCCGT	CTCTTGGAGA	
2521	ACTTCCAATC	GCACCATCAT	CACCATCACC	ATCACCACCA	TGATTACAAG	GATGACGACG	
	TGAAGGTTAG	CGTGGTAGTA	GTGGTAGTGG	TAGTGGTGGT	ACTAATGTTC	CTACTGCTGC	
2581	ATAAGTGAGG	ATCCGAATTC	GAGCTCCGTC	GACAAGCTTG	TCGAGAAGTA	CTAGAGGATC	
	TATTCACTCC	TAGGCTTAAG	CTCGAGGCAG	CTGTTCGAAC	AGCTCTTCAT	GATCTCCTAG	
		FBac2					
		(					
2641	מ שממ שר מכיר	ATACCACATT	тстасасстт			CCCACACCTC	
2011		TATGGTGTAA					
	TATIAGICGG	TATGGIGIAA	ACAICICCAA	AAIGAACGAA	ATTITITE	DADDIDIDDD	

## Primers for LIC cloning:

Upstream: add TACTTCCAATCC<u>ATG</u> to the 5' end (ATG in-frame with the desired coding sequence).

Downstream: add TATCCACCTTTACTGCT to 5' end of downstream primer. Do not add termination codon as this will remove the biotin attachment site.