

01 **Chapter 5**
02 **Monte Carlo Modeling of Light Transport**
03 **in Tissue (Steady State and Time of Flight)**

06
07 **Steven L. Jacques**
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13 **5.1 Introduction**
14

15 Monte Carlo simulations are a fundamental and versatile approach toward modeling
16 light transport in tissues. While diffusion theory for light transport is a fast and
17 convenient way to model light transport, it fails when close to sources or boundaries
18 and when absorption is strong compared to scattering; in other words, whenever
19 conditions cause the gradient of fluence rate (or photon concentration) to not be
20 simply linear but to have some curvature. Monte Carlo steps in to treat problems
21 when diffusion theory fails. Figure 5.1 illustrates a Monte Carlo simulation.

22 In the general Monte Carlo simulation, “photons” are inserted into tissue at a
23 location defined by x , y , z coordinates with a trajectory defined by directional cosines
24 (projection of trajectory onto x , y and z axes). The random distance traveled before
25 the photon interacts with the tissue is based upon the selection of a random number
26 [0,1] and the local attenuation coefficient of the medium. At the end of each photon
27 step, the weight of the photon is reduced by absorption. The remaining non-absorbed
28 weight is redirected according to a scattering (or “phase”) function that describes
29 the angular dependence of single scattering by the particular tissue. Once a new
30 trajectory is specified, the photon is again moved a random distance. The details of
31 the photon path, absorption, scattering, reflection, and refraction are described in
32 the following paragraphs.

33 Refraction at mismatched boundaries and even changes in local optical properties
34 can be included in the simulation. The heat generated, S [W/cm³], within a small
35 volume depends upon the total photon weight absorbed in the volume, the total
36 number of photons, and power of the laser beam. Total energy absorbed [J/cm³] can
37 be computed when the energy [J] of the light source is given.

38 The Monte Carlo method is a widely used approach toward sampling probability
39 density functions for simulating a wide range of problems. The first use of the
40

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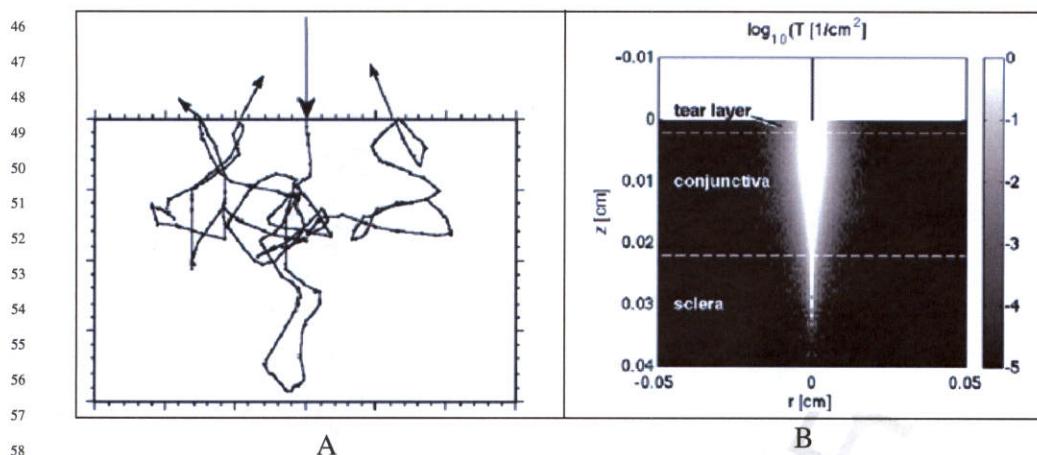


Fig. 5.1 A Monte Carlo simulation of photon transport. **(a)** Movement of a single photon in a light-scattering tissue. When the photon is totally internally reflected at the air/tissue surface, a fraction of the photon weight is allowed to escape as reflectance and the remaining photon weight continues to propagate in the tissue. In this example for the sake of illustration, the third time the photon encounters the surface, the photon is allowed to fully escape. **(b)** Superposition of millions of photons recorded as relative fluence rate, [W/cm^2] per W delivered, which is equivalently stated as transport T [$1/\text{cm}^2$]. The example is the penetration of a CO_2 laser ($10.6 \mu\text{m}$ wavelength) into the conjunctiva of the eye, which illustrates how Monte Carlo can also describe the scattering in a situation where absorption dominates over scattering, a situation not well described by diffusion theory

Monte Carlo method for photon transport in biological materials was Adams and Wilson (1983), which considered isotropic scattering [1]. Keijzer et al. (1987) introduced anisotropic scattering into the Monte Carlo simulation of biological tissues, implementing a simulation that propagated photons using cylindrical coordinates, which introduced the Hop/Drop/Spin nomenclature for organizing the program [2]. Prahl et al. (1989) reformulated the program using photon propagation based on Cartesian coordinates, which made the program much simpler to convey in written form [3]. Wang and Jacques (1993) adapted and augmented the work of Keizer and Prahl to write the Monte Carlo Multi-Layered (MCML) program that considers tissues with many planar layers with different optical properties [4]. MCML is a well-organized program with a simple text input file that the user can modify to specify different problems, which allows various problems to be run without needing to re-compile the program each time. MCML has been widely promulgated via the web as source code [5–6], and modified by various groups to handle a variety of problems. Jacques (1998) reported on using Monte Carlo to specify the point spread function for light in tissue in planar, cylindrical and spherical coordinates from a plane source, line source or point source, respectively [7], using a minimal Monte Carlo program derived from MCML called `mc321.c` that is listed in the Appendix of this chapter for reference and is available on the web [8]. Students have usually modified `mc321.c` to build their own specialized programs. Jacques (2003) prepared a Monte Carlo subroutine, `mcsub.c`, which allows routine calls to Monte Carlo runs

from other programs, discussed in [9], with updated versions listed on the web [10]. Ramella-Roman et al. (2005) modified mc321.c to simulate polarized light transport by propagating the Stokes Vector state of a photon [11], and posted the program on the web [12]. This paragraph does not offer a complete review of all contributions to Monte Carlo simulations, but provides a brief history of the programming code that is most widely used in biomedical optics and is freely available on the web.

A small note: throughout this chapter, simple math equations are mixed with programming language, and often an equal sign, “=”, is used when an assignment, “ \leftarrow ”, is the appropriate symbol. The meaning will be obvious, but for those readers familiar with scientific grammar, the poor grammar is intentional.

5.2 Basic Monte Carlo Sampling

The Monte Carlo simulation of light propagation in tissue requires random selection of photon step size, scattering angle and reflection or transmission at boundaries. This is accomplished by a random number [0,1] assigned to the value of a random variable x , such as photon step size. The relationship is established through the density function $p(x)$ and distribution function $F(x)$ (see Chapter 3). Given $p(x)$, the value of the probability distribution function at a particular value x_1 of the random variable x is

$$F(x_1) = \int_0^{x_1} p(x)dx = \text{function}(x_1) \quad (5.1)$$

This $F(x_1)$ is equated with a random number, RND_1 , in the interval [0,1]:

$$RND_1 = F(x_1) \quad (5.2)$$

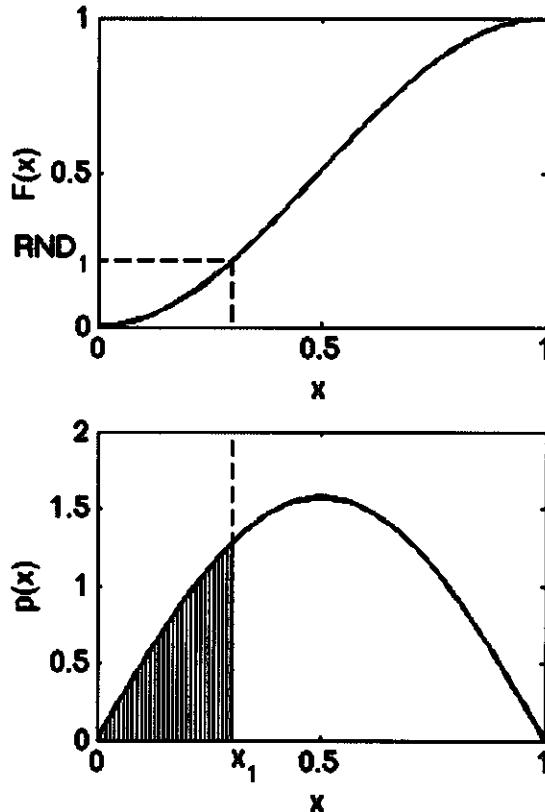
Then Eq. (5.2) is rearranged to solve for x_1 in terms of RND_1 . The resulting expression allows a series of values RND_1 to specify a series of values x_1 . The histogram of x_1 values will conform to the probability density function $p(x)$. Figure 5.2 illustrates the procedure.

This process is illustrated in the following section by the random selection of photon step size, s .

5.2.1 Predicting the Step Size of a Photon

The first example is predicting the step size of a photon between interaction events (absorption or scattering) as the photon multiply scatters within a tissue. A photon takes a step (s [cm]) whose length is exponentially distributed, and depends on the value of the total attenuation coefficient μ_t [cm⁻¹] equal to $\mu_a + \mu_s$, where μ_a is

136 **Fig. 5.2** A random number
 137 generator [0,1] selects a value
 138 RND_1 which is set equal to
 139 the distribution function $F(x)$
 140 which then specifies the value
 141 x_1 . The cross hatched area is
 142 equal to RND_1



161 the absorption coefficient and μ_s is the scattering coefficient. The value $1/\mu_t$ is the
 162 photon's mean free pathlength before either absorption or scattering occurs.

163 The first step is to specify a properly normalized probability density function that
 164 describes the probability of a particular photon step size s :

$$166 \quad 167 \quad p(s) = \exp(-\mu_t s)/\mu_t \quad (5.3)$$

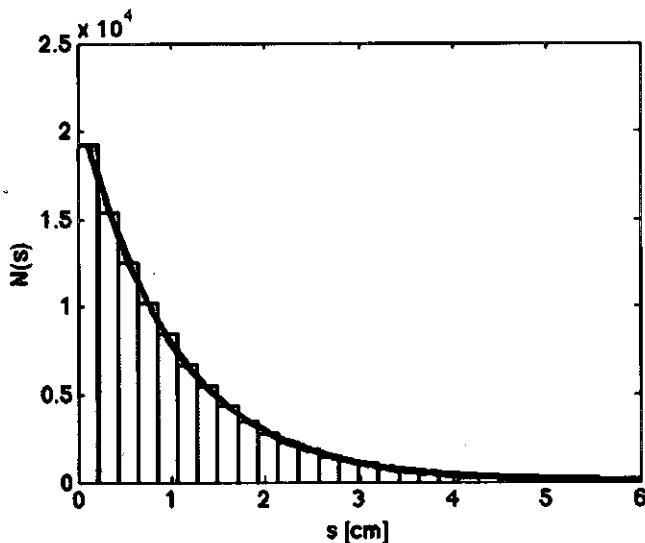
168 such that

$$170 \quad 171 \quad \int_0^\infty p(s)ds = \int_0^\infty \frac{e^{-\mu_t s}}{\mu_t} ds = 1 \quad (5.4)$$

174 The second step is to determine the integral of $p(s)$ evaluated at a particular s_1 ,
 175 which defines the probability distribution function:

$$177 \quad 178 \quad F(s_1) = \int_0^{s_1} p(s)ds = \int_0^{s_1} \frac{e^{-\mu_t s}}{\mu_t} ds = 1 - e^{-\mu_t s_1} \quad (5.5)$$

181 Fig. 5.3 Histogram of step
 182 sizes predicted by 1000
 183 random numbers using Monte
 184 Carlo sampling (Eq. (5.7)).
 185 $N(s)$ is the number of step
 186 sizes per bin over a range of s
 187 value bins. The value of μ_t is
 188 1 cm^{-1} . The bin size of the
 189 histogram is $ds_1 = 0.275 \text{ cm}$.
 190 The curved line is $N(s) =$
 191 $1000 \exp(-\mu_t s_1) ds_1 / \mu_t$



197
 198 The third step is to set $F(x)$ equal to a random number RND_1 .

199
 200
$$RND_1 = F(s_1) = 1 - e^{-\mu_t s_1} \quad (5.6)$$

201
 202 and solve for s_1 :

203
 204
$$s_1 = \frac{-\ln(1 - RND_1)}{\mu_t} = \frac{-\ln(RND_1)}{\mu_t} \quad (5.7)$$

205
 206 The terms $(1 - RND_1)$ and (RND_1) are equal in a probabilistic sense because
 207 of the uniform distribution of the random number between $[0,1]$. This is the final
 208 answer. A series of random numbers RND_1 will generate a series of step sizes s_1
 209 that follow the probability density function $p(s)$ of Eq. (5.3).

210 To illustrate, Fig. 5.3 shows a histogram of s_1 values specified by 1000 RND_1
 211 values. The value of μ_t is 1 cm^{-1} . The bin size of the histogram is $ds_1 = 0.275 \text{ cm}$.
 212 Also plotted is the curve $N(s) = 1000 \exp(-\mu_t s_1) ds_1$.

214
 215
 216 **5.2.2 Predicting the Photon Launch Point for a Circular**
 217 **Flat-Field Beam**

218
 219 The second example is predicting where a photon should be launched at a tissue
 220 surface so as to mimic a circular flat-field (i.e., a uniform irradiance) beam of light.
 221 Assume the beam of light is delivered perpendicular to the tissue surface. Also
 222 assume that we are working in cylindrically symmetric Cartesian coordinates. The
 223 Monte Carlo simulation will propagate the photon in 3D using x , y and z coordi-
 224 nates, but we will report out our results only as a function of r and z . Therefore, we
 225 need only specify the radial distance r from the center of the beam where a photon

should enter the tissue, and the photon can be launched along the x axis by letting $x = r$. Assume the beam of light has a radius a .

The number of photons per unit area should be constant. For each radial position r there is an annular ring of area $(2\pi r dr)$. Therefore, the number of photons to be launched at each choice of r should vary as $(2\pi r dr)$. The total area of the light beam is πa^2 . The first step is to specify the properly normalized probability density function $p(r)$:

$$p(r) = \frac{2\pi r}{\pi a^2} = \frac{2}{a^2}r \quad (5.8a)$$

such that

$$\int_0^a p(r)dr = \int_0^a \frac{2}{a^2}rdr = 1 \quad (5.8b)$$

The second step is to integrate this $p(r)$, evaluated for a specific r_1 , and equate to RND_1 .

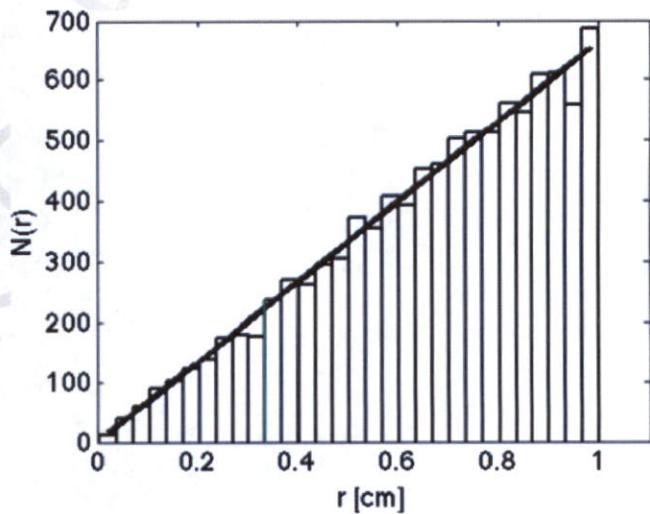
$$\int_0^{r_1} \frac{2}{a^2}rdr = \frac{r_1^2}{a^2} = RND_1 \quad (5.9)$$

The third step is to solve for r_1 in terms of RND_1 :

$$r_1 = a\sqrt{RND_1} \quad (5.10)$$

Now to illustrate, Fig. 5.4 shows the histogram of 1000 r_1 values predicted by 1000 random numbers.

Fig. 5.4 Histogram of radial position of photon launch to achieve a circular flat-field beam of light, perpendicularly illuminating a tissue surface. The histogram is predicted by 1000 random numbers using Monte Carlo sampling (Eq. (5.10)). $N(r)$ is the number of predicted r positions per bin over a range of r value bins. The value of the beam radius a is 1 cm. The bin size of the histogram is $ds_1 = 0.0327$ cm. The diagonal line is $N(s) = (1000 2r dr)/a^2$



271 5.3 The Steady-State Monte Carlo Propagation of Photons 272 in a Tissue 273

274 This section presents the basic form of the steady-state Monte Carlo simulation of
275 photon propagation in tissues with optical scattering and absorption properties. The
276 term steady-state means that a stable distribution of light has been achieved, and
277 there are no time dynamics. Section 5.4 will discuss time-resolved Monte Carlo.

278 It is convenient to consider the Monte Carlo simulation as yielding the fractional
279 density matrix of incident light absorbed, A [1/cm³], in response to one unit of
280 delivered power or energy. The corresponding heat source matrix S [W/cm³] or
281 radiant energy density matrix W [J/cm³] is obtained by scaling A by the beam power
282 P or radiant energy Q , respectively, such that

283

$$284 \quad S = PA \quad (5.11a)$$

285

286 OR

287

$$288 \quad W = QA \quad (5.11b)$$

289

290 where $S(r)$ [W/cm³] is the local density of power deposition at $r = (x, y, z)$ in
291 response to a delivered power P [W], and $W(r)$ [J/cm³] is the local density of energy
292 deposition in response to a delivered energy Q [J].

293 Also, the simulation keeps track of the escaping flux density out of the front sur-
294 face to which light is delivered, R_r [1/cm²], and out of the rear surface, T_r [1/cm²].
295 The A [1/cm³] is converted into the fractional transport $T[iz][ir]$ [1/cm²] within the
296 tissue, where iz and ir are the indices of the array T , such that the fluence rate
297 ϕ [W/cm²] and the fluence ψ [J/cm²] are

298

$$299 \quad \phi(r) = PT(r) \quad (5.12a)$$

300

301

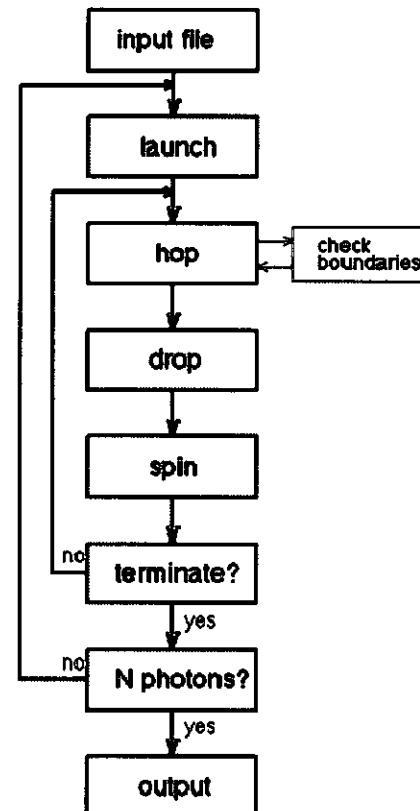
$$302 \quad \psi(r) = QT(r) \quad (5.12b)$$

303 The following sections will present the algorithm for implementing the
304 Monte Carlo simulation. The overall algorithm is described by the flow diagram
305 in Fig. 5.5.

306 307 5.3.1 The Input File

308 The parameters that govern a Monte Carlo simulation must be passed to the pro-
309 gram before it can run. These parameters can be included in the programming at the
310 beginning of a simple Monte Carlo code (as in `mc321.c`, see Appendix), or passed
311 to a Monte Carlo subroutine from another program (as in `mcsup.c` [8, 9]) or read
312 from an input file (as in MCML [4–6]). In Fig. 5.5, these possibilities are referred to
313 as the “input file”.

316 **Fig. 5.5** The flow-diagram
 317 for a steady-state Monte
 318 Carlo simulation



341 In this simple summary of a generic Monte Carlo simulation, a simple homogeneous tissue is specified. The tissue parameters to be specified are:

$$\begin{aligned}
 345 & \mu_a [\text{cm}^{-1}] \text{ absorption coefficient of tissue} \\
 346 & \mu_s [\text{cm}^{-1}] \text{ scattering coefficient of tissue} \\
 347 & g [-] \text{ anisotropy of scattering of tissue} \\
 348 & n [-] \text{ refractive index of tissue}
 \end{aligned} \tag{5.13}$$

349 where $[-]$ denotes [dimensionless]. The above parameters specify an infinite optically homogeneous medium, and the simulation will track the 3D movement of a photon over an infinite spatial range. To specify boundaries, for example the front and rear surfaces of a slab of tissue, the following parameters should be specified:

$$\begin{aligned}
 355 & D [\text{cm}] \text{ thickness of the tissue slab} \\
 356 & n_f [-] \text{ refractive index of external medium outside front of tissue} \\
 357 & n_r [-] \text{ refractive index of external medium outside rear of tissue}
 \end{aligned} \tag{5.14}$$

358 The position $z = 0$ is aligned with the front surface of the tissue slab, and the rear surface is aligned with $z = D$. As the program runs, the history of the photon

is recorded by deposition of “photon weight” into spatially distributed bins, either $A(x, y, z)$ for full 3-D accumulations or $A(r, z)$ for circular symmetric laser beams. In the example of this chapter, the bins are denoted $A[iz][ir]$ [photon weight/bin]. The grid of bins for recording this A is independent of the photon propagation, which is implemented in x, y, z coordinates. Once a photon is absorbed at x, y, z , its position in this example is recorded as z , where $r = \sqrt{x^2 + y^2}$, and some “photon weight” is deposited in $A[iz][ir]$. The parameters to be specified for recording output are:

368

369 N_z [-] number of z bins (depth)
 370 N_r [-] number of r bins (radial position)
 371 dz [cm] size of z bins
 372 dr [cm] size of r bins

373

374 where the bins cover the extent of the tissue,

375

$$376 \quad dz = \frac{D}{N_z} \quad (5.15)$$

378

379 Finally, the number of photons that will be launched must be specified,

380

$$381 \quad N_{\text{photons}}$$
 [-] number of photons to be launched (5.17)

383 Typically about 10^4 – 10^8 photons are launched, depending on how much com-
 384 putation time is appropriate or available. The run time will vary with the optical
 385 properties. As will be discussed in later sections, each photon will take an average
 386 step size of $1/(\mu_s + \mu_a)$ and at each step the photon weight will drop via multipli-
 387 cation by the fraction $\mu_s/(\mu_s + \mu_a)$ called the albedo. The weight is initially 1 upon
 388 launching the photon and drops until it reaches a threshold value, THRESHOLD,
 389 which is typically 10^{-4} . The time required to reach THRESHOLD is proportional
 390 to the number of steps, N_{steps} , taken by the photon

391

$$392 \quad \text{THRESHOLD} = \left(\frac{\mu_s}{\mu_s + \mu_a} \right)^{N_{\text{steps}}} \quad (5.18)$$

394

395 and the number of steps required for each photon is

396

$$397 \quad N_{\text{steps}} = \frac{\ln(\text{THRESHOLD})}{\ln\left(\frac{\mu_s}{\mu_s + \mu_a}\right)} \quad (5.19)$$

400

401 As the absorption μ_a decreases, N_{steps} increases. As the scattering μ_s increases,
 402 N_{steps} increases. The values of g and n have no effect. Therefore, the time required
 403 to run N_{photons} is

404

405

$$t = N_{\text{photons}} t_{\text{per.step}} N_{\text{steps}} \quad (5.20)$$

406 where $t_{\text{per.step}}$ is the time required per photon step in the simulation. One may
 407 run several example simulations with known values of THRESHOLD, μ_s , μ_a , and
 408 N_{photons} , and record the time required for each simulation. Then fit the data to solve
 409 for $t_{\text{per.step}}$.

410

$$411 \quad t_{\text{per.step}} = \frac{t}{N_{\text{photons}} N_{\text{steps}}} \quad (5.21)$$

413

414 For example, if it takes 23.0 s to run 10^5 photons with optical properties $\mu_a =$
 415 1 cm^{-1} , $\mu_s = 100 \text{ cm}^{-1}$, using THRESHOLD = 10^{-4} , then $t_{\text{per.step}}$ is 249 ns. Now,
 416 if one desires a simulation that takes exactly $t = 10 \text{ min}$ (600 s) for properties $\mu_a =$
 417 0.4 cm^{-1} and $\mu_s = 65 \text{ cm}^{-1}$, then the choice of N_{photons} is

418

$$419 \quad N_{\text{photons}} = \frac{t}{t_{\text{per.step}} N_{\text{steps}}} = \frac{t}{t_{\text{per.step}} \ln(\text{THRESHOLD})} \frac{\ln\left(\frac{\mu_s}{\mu_s + \mu_a}\right)}{420}$$

$$421 \quad = \frac{600 \text{ s}}{249 \times 10^{-9}} \frac{\ln\left(\frac{65}{65 + 0.4}\right)}{\ln(10^{-4})} = 1.60 \times 10^6 \quad (5.22)$$

426

427 So one would launch 1.60 million photons for a 10-min simulation using the
 428 above optical properties. In this manner, the choice of N_{photons} for a given simulation
 429 can be made on the basis of available time, regardless of the optical properties in the
 430 simulation.

431

Now the Monte Carlo simulation can proceed to run.

432

433

434 5.3.2 Launching Photons

435

436 Photons are launched with an initial weight (w) set to 1.0. As the photon propagates,
 437 this weight will be decremented as “photon weight” is “dropped” into the
 438 bins $A[iz][ir]$, where iz and ir are index pointers to the bins for the depth z and radial
 439 position r , respectively. A large number of photons will be launched (N_{photons}) and
 440 later at the end of the program all the weight deposited in $A[iz][ir]$, as well as any
 441 weight that escaped the tissue as transmission or reflectance, will be normalized by
 442 N_{photons} , such that the final results are reported as the fraction of all delivered light
 443 that is either absorbed, reflected or transmitted.

444

445 The position of the photon is specified in Cartesian coordinates, (x, y, z) , and all
 446 the propagation is done in full 3D. The recording of $A[iz][ir]$ is in cylindrical coor-
 447 dinates, but one could implement $A[iz][ix][iy]$ if desired. If one is using 100×100
 448 bins for $A[iz][ir]$, one needs only to fill 10^4 bins with photon weight. If one is using
 449 $100 \times 100 \times 100$ bins for $A[iz][ix][iy]$, one needs to fill 10^6 bins with photon
 450 weight. Since the signal-to-noise in any bin is roughly proportional to \sqrt{A}/A , it takes
 many more photons to attain good signal-to-noise filling $A[iz][ix][iy]$ than filling.

451 $A[iz][ir]$. This is why cylindrical coordinates are often chosen for use. But there is
 452 no reason not to record data as $A[iz][ix][iy]$. The choice of recording does not affect
 453 the photon propagation.

454 The trajectory of the photon will be specified by the trajectory cosines
 455 (ux, uy, uz):

$$\begin{aligned} 457 \quad ux &= \cos \theta \cos \varphi \\ 458 \quad uy &= \cos \theta \sin \varphi \\ 459 \quad uz &= \sin \theta \end{aligned} \tag{5.23}$$

460 where θ is the angle that the trajectory makes with respect to the z axis, and φ is the
 461 angle that the trajectory makes with the x axis.

462 So let's launch some photons.

463 In this example, photons are launched perpendicular to the tissue and enter the tissue
 464 exactly at the origin $(x, y, z) = (0, 0, 0)$. The photon position and trajectory are

$$\begin{aligned} 470 \quad x &= 0 \\ 471 \quad y &= 0 \\ 472 \quad z &= 0 \end{aligned} \tag{5.24a}$$

473 and

$$\begin{aligned} 476 \quad ux &= 0 \\ 477 \quad uy &= 0 \\ 478 \quad uz &= 1 \end{aligned} \tag{5.24b}$$

479 The photon is directed straight downward, so $uz = 1$. The values of ux and uy are
 480 zero because no component of the trajectory is directed in the x or y directions.

481 The case of collimated launch as a flat-field beam, i.e., uniform irradiance, where
 482 the beam has a radius a , was discussed in Section 5.2.2. Photons can be placed along
 483 the x axis according to Eq. (5.10) with $y = 0, z = 0$.

486 **5.3.2.2 Isotropic Point Source**

487 To launch a photon isotropically, i.e., with no preferential direction, at a position
 488 located within the tissue, for example at $x = 0, y = 0$, and $z = 0.1$ cm, the launch
 489 specification is:

$$\begin{aligned} 491 \quad x &= 0 \\ 492 \quad y &= 0 \\ 493 \quad z &= 0.1 \end{aligned} \tag{5.25a}$$

496 and

497

$$\cos \theta = 2RND - 1$$

498

$$\sin \theta = \sqrt{(1 - \cos^2 \theta)}$$

499

$$\varphi = 2\pi RND$$

500

501 if ($\varphi < \pi$)

502

$$\sin \varphi = \sqrt{(1 - \cos^2 \varphi)}$$

503

504 else

505

$$\sin \varphi = -\sqrt{(1 - \cos^2 \varphi)}$$

506

507 such that

508

$$ux = \sin \theta \cos \varphi$$

509

$$uy = \sin \theta \sin \varphi$$

510

$$uz = \cos \theta$$

511

512 The z position is 0.1 cm. The trajectory (ux, uy, uz) is randomly selected such that

513

$$\sqrt{ux^2 + uy^2 + uz^2} = 1 \quad (5.25c)$$

514

515 It is necessary for the total length of the trajectory vector to be unity.

516

517 The photon is launched at $x = 0$ and $y = 0$ because we wish to retain cylindrical
518 symmetry. If one launched at any other x, y position the cylindrical symmetry would
519 be broken. Then the results would have to be recorded as $A[iz][ix][iy]$. The results
520 recorded as $A[iz][ir]$ would respond as if photons were launched in a ring of radius
521 $\sqrt{x^2 + y^2}$.

522

523 5.3.2.3 Collimated Gaussian Beam

524

525 Consider a collimated Gaussian beam that irradiates a tissue, which has a $1/e$ radius
526 of b . The probability density function for the radial position of launch is

527

$$p(r) = \frac{e^{-(r/b)^2}}{\pi b} 2\pi r \quad (5.27)$$

528

529 which can be sampled using the Monte Carlo sampling method by

530

$$r = b\sqrt{-\ln(RND)} \quad (5.28)$$

531

532

533 To launch a Gaussian beam when using cylindrically symmetric results, one can
534 launch the photon at $x = r$. The launch parameters are:

541 $x = b\sqrt{-\ln(RND)}$
 542 $y = 0$ (5.29a)
 543 $z = 0$

544
545 and
546

547 $ux = 0$
 548 $uy = 0$ (5.29b)
 549 $uz = 1$

5.3.2.4 Focused Gaussian Beam

553 Consider the same Gaussian beam but focused to a focal point within the tissue at
 554 depth z_{focus} . Let the focus have a radial Gaussian distribution with a $1/e$ radius of w .
 555 Then the launch parameters are calculated:

556 $x = w\sqrt{-\ln(RND)}$
 557 $y = 0$ (5.30a)
 558 $z = 0$

559
560 and
561

562 $x_{\text{focus}} = w\sqrt{-\ln(RND)} \text{ sign}(2RND - 1)$
 563 $\text{temp} = \sqrt{(x - x_{\text{focus}})^2 + z_{\text{focus}}^2}$ (5.30b)
 564 $\sin \theta = -(x - x_{\text{focus}})/\text{temp}$
 565 $\cos \theta = z_{\text{focus}}/\text{temp}$

566
567 such that
568

569 $ux = \sin \theta$
 570 $uy = 0$ (5.30c)
 571 $uz = \cos \theta$

572 There is a tendency in Monte Carlo simulations using cylindrical coordinates to
 573 not get sufficient photons in the bins along the z axis, because the size of each bin
 574 is $(2\pi r dr dz)$, so the size of bins near $r = 0$ is very small, and the likelihood of
 575 photon deposition in such a small bin is quite low. During this focused Gaussian
 576 launch, one should launch toward both $+x_{\text{focus}}$ and $-x_{\text{focus}}$ positions, which forces
 577 the photons to cross the z axis. Doing this causes the bins along the central axis to
 578 not be so neglected, and is accomplished by the extra term $\text{sign}(2 RND - 1)$ in the
 579 expression for x_{focus} . The function $\text{sign}()$ has a value of $+1$ or -1 .

586 **5.3.3 Hop**

587

588 Now the photon is launched along a trajectory, and takes a step along this trajectory.
 589 The standard procedure for taking the step is described as the “Standard Hop” in
 590 Section 5.3.3.1. But if the tissue has a front and/or rear boundary, then a second step
 591 to “Check boundaries” is taken, as described in Section 5.3.3.2, and if the photon
 592 is attempting to escape the tissue, a procedure is used to decide whether the photon
 593 escapes or is reflected back into the tissue.

594

595 **5.3.3.1 Standard Hop**

596

597 The step size of the photon’s step (or hop) must be determined. The step size is
 598 calculated:

599

600 $s = -\ln(RND)/\mu_t$ (5.31)

601

602 as was described in Section 3.2.1. Now the position of the photon is updated:

603

604 $x = x + s u_x$
 605 $y = y + s u_y$
 606 $z = z + s u_z$ (5.32)

607

608

609 **5.3.3.2 Check Boundaries**

610 As part of the Hop step, there is a side box labeled “check boundaries” in Fig. 5.5,
 611 which is used when there are front and surface boundaries in the problem. This
 612 boundary check is denoted by a side box to emphasize that it is part of the Hop step.

613 As the photon moves toward the front surface of the tissue and attempts to
 614 cross the boundary to escape the tissue, there is a possibility that the photon will
 615 be reflected by the surface boundary where the air/tissue interface (or *external*
 616 medium/tissue interface) presents a mismatch in refractive indices ($n_f \neq n$ or
 617 $n_r \neq n$, for front and rear boundaries, respectively). The method chosen here for
 618 handling the boundary is to let a fraction of the photon weight escape the tissue as
 619 observable reflectance, and let the remaining fraction of photon weight reflect back
 620 into the tissue and continue to propagate.

621 After taking the step, the position of the photon is checked to see if the photon is
 622 still within the tissue or has escaped the tissue:

623

624 If $z < 0$
 625 photon is trying to escape
 626 else
 627 photon still within tissue

628

629 If the photon is trying to escape, then a partial step is taken along the escaping
 630 trajectory that will just reach the boundary surface. The size of the partial step s_1 is

631 $s_1 = \text{abs}(z/u_z)$ (5.34)

632 where $\text{abs}()$ denotes the absolute value function. First, the photon retracts the full
633 step it took in Hop, which led to escape,

635 $x = x - s u_x$
636 $y = y - s u_y$ (5.35a)
637 $z = z - s u_z$

639 and then the photon takes the partial step s_1 ,

641 $x = x + s_1 u_x$
642 $y = y + s_1 u_y$ (5.35b)
643 $z = z + s_1 u_z$

645 Now the photon is located at the boundary surface. Next, a decision is made about
646 how much of the photon weight escapes or how much is reflected.

648 The probability of reflectance at the boundary is a function of the angle of inci-
649 dence, encoded as the value u_z , and the refractive indices n_1 and n , where n_1 denotes
650 the external medium and equals either n_f or n_r for the front and rear boundaries,
651 respectively. Consider a photon striking the front surface. The internal reflectance,
652 R_i , is calculated using the Fresnel reflection equation:

653 $R_i = \frac{\sin \theta^2 (\sin \theta_1 \cos \theta_2 - \cos \theta_1 \sin \theta_2)^2}{2}$
654 $\times \frac{((\cos \theta_1 \cos \theta_2 + \sin \theta_1 \sin \theta_2)^2 + (\cos \theta_1 \cos \theta_2 - \sin \theta_1 \sin \theta_2)^2)}{((\sin \theta_1 \cos \theta_2 + \cos \theta_1 \sin \theta_2)^2 (\cos \theta_1 \cos \theta_2 + \sin \theta_1 \sin \theta_2)^2)}$ (5.36)

658 where

n_1	refractive index of incident medium	$= n_{\text{tissue}}$
n_2	refractive index of transmitted medium	$= n_f$
$\cos \theta_1$	incident trajectory	$= u_z$
$\sin \theta_1$	incident trajectory	$= (1 - u_z^2)^{1/2}$
$\sin \theta_2$	transmitted trajectory	$= \sin \theta_1 (n_1/n_2)$
$\cos \theta_2$	transmitted trajectory	$= (1 - \sin^2 \theta_1)^{1/2}$

668 Once the reflectance R_i is computed, a fraction $(1 - R_i)$ of the current pho-
669 ton weight is allowed to escape and the remaining fraction of weight is reflected
670 back into the tissue to continue propagating. The escape is recorded by placing its
671 remaining weight in the reflectance array bin, $R_r[ir]$,

674 $R_r = R_r[ir] + (1 - R_i)w$ (5.37)

676 where the radial position of escape is r :

$$\begin{aligned} 678 \quad r &= \sqrt{x^2 + y^2} \\ 679 \quad ir &= \text{round}(r/dr) + 1 \end{aligned} \quad (5.38)$$

681 The function $\text{round}(r/dr)$ denotes taking the integer value of the value r/dr , round-
682 ing down, such that a photon somewhere within the location of one bin will be
683 assigned to that particular bin. (Note: In this summary, ir extends from 1 to N_z , and
684 ir is not allowed to equal zero. Sometimes, programs let ir extend from 0 to N_z-1 .
685 Not mentioned here but important in programming is that a check should be made
686 that ir does not exceed N_r , the allocated size of $Rr[ir]$.)

687 The remaining fraction of the photon weight is reflected by the boundary,

$$689 \quad w = R_i w \quad (5.39)$$

690 and the trajectory with respect to the z axis is reversed,

$$693 \quad u_z = -u_z \quad (5.40)$$

695 The ux and uy components of the trajectory are not changed. Then the photon
696 position is updated by taking the remaining portion of the original step, which equals
697 $s - s_1$:

$$\begin{aligned} 699 \quad y &= (s - s_1)ux \\ 700 \quad y &= (s - s_1)uy \\ 701 \quad z &= (s - s_1)uz \end{aligned} \quad (5.41)$$

702 Some of the photon weight has escaped the boundary and some has been reflected
703 by the boundary back into the tissue to continue propagating.

705 The procedure for testing if the photon is attempting to escape the tissue through
706 its rear surface boundary and contributing to $T[iz][ir]$, and for modifying the posi-
707 tion and trajectory accordingly, is very similar to the above procedure for the front
708 surface, and is not outlined here. If one wished to place other boundaries in the
709 problem, like a lateral cylindrical boundary as if the tissue were held within a tube
710 (e.g., a glass test tube, or a pipe), this “check boundaries” box is the proper place
711 within the program to implement such special boundaries.

712 713 5.3.4 Drop

716 Arriving at its new position, the photon must interact with the tissue. Upon interac-
717 tion, a fraction μ_a/μ_t of the photon's weight is absorbed and the remaining μ_s/μ_t
718 fraction of the photon's weight is scattered and continues to propagate. The absorbed
719 fraction is placed in the bin that encloses the current photon position. This process
720 is summarized by the following calculation steps:

$$\begin{aligned}
 721 \quad r &= \sqrt{x^2 + y^2} \\
 722 \quad ir &= \text{round}(r/dr) + 1 \\
 723 \quad iz &= \text{round}(z/dz) + 1 \\
 724 \quad A[iz][ir] &= A[iz][ir] + w(\mu_a/\mu_t) \\
 725 \quad w &= w(\mu_s/\mu_t)
 \end{aligned} \tag{5.42}$$

728 where ir and iz are index values and dr and dz are bin width and depth, respectively.
 729 This finishes the absorption event.

730 Note that ir and iz should not be allowed to exceed N_r and N_z , respectively, lest
 731 one exceed the allocated size of $A[iz][ir]$. Always check this. Often, the last bin is
 732 used as an overflow bin and any photon weight that is deposited outside the array is
 733 simply accumulated in the last iz, ir bin. At the end of the simulation, the values of
 734 weight in $Rr[ir]$, $Tr[ir]$ and $A[iz][ir]$ are the fraction of total delivered photon weight.
 735 The sum of $Rr[ir]$, $Tr[ir]$ and $A[iz][ir]$ over all bins divided by N_{photons} will equal
 736 unity. In Section 5.3.7, the proper normalization of these fractional weights by the
 737 size of the bins will convert Rr and Tr into the fraction of delivered power or energy
 738 escaping per unit surface area and convert A into the fraction of delivered power or
 739 energy deposited per unit volume. But the values of Rr , Tr and A determined from
 740 the overflow bins are meaningless after normalization.

741 Next, is the scattering event.

743 5.3.5 Spin

745 The photon is scattered into a new trajectory according to some scattering function.
 746 The two angles of scatter are θ and φ , the deflection and azimuthal scattering
 747 angles, respectively. This section describes how to calculate the new trajectory after
 748 sampling the probabilities for θ and φ .

749 The most commonly used function for the deflection angle θ is the Henyey-Greenstein (HG) function (1941), which was proposed for describing the scattering
 750 of light from distant galaxies by galactic dust. The original paper does not offer
 751 any explanation for the function, but simply asserts its use. But the HG function
 752 is actually very interesting. The function is here expressed as a function of the
 753 angle of deflection, θ , and the anisotropy of scattering, g . Using the definitions in
 754 Chapter 3 (Section 3.4.3.1) where the probability density functions for $p(\varphi)$ and $p(\theta)$
 755 are considered independently:

$$758 \quad p(\theta) = \frac{1}{2} \frac{1 - g^2}{(1 + g^2 - 2g \cos \theta)^{3/2}} \tag{5.43}$$

761 which has the properties that

$$763 \quad \int_0^\pi p(\theta) 2\pi \sin \theta d\theta = 1 \tag{5.44a}$$

766 and

$$\int_0^{\pi} p(\theta) \cos \theta 2\pi \sin \theta d\theta = g \quad (5.44b)$$

772 The last equation is the definition of g . Hence, the HG function is an identity with
 773 respect to the definition of g . If you choose a value g to define $p(\theta)$ using Eq. (5.43),
 774 the definition of g in Eq. (5.44b) will yield exactly g .

775 The Monte Carlo sampling of the HG function is specified by the following
 776 sequence of calculations:

$$\cos(\theta) = \frac{1 + g^2 - \left(\frac{1 - g^2}{(1 - g + 2gRND)^{3/2}} \right)^2}{2g} \quad (5.45)$$

782 If g is 0, then use $\cos \theta = 2 RND - 1$. If g is 1.0, then simply let $\cos \theta = 1.0$.
 783 Otherwise, use the Monte Carlo sampling in Eq. (5.45).

784 The azimuthal angle is calculated:

$$\varphi = 2\pi RND \quad (5.46)$$

789 To update the trajectory based on the values of $\cos \theta$ and φ specified using random
 790 numbers, use the following calculations:

$$\begin{aligned} 792 \sin \theta &= \sqrt{(1 - \cos \theta^2)} \\ 793 \text{temp} &= \sqrt{(1 - u_z^2)} \\ 794 ux &= \sin \theta (u_x u_z \cos \varphi - u_y \sin \varphi) / \text{temp} + u_x \cos \theta \\ 795 uy &= \sin \theta (u_y u_z \cos \varphi - u_x \sin \varphi) / \text{temp} + u_y \cos \theta \\ 796 uz &= -\sin \theta \cos \varphi \text{temp} + u_z \cos \theta \end{aligned} \quad (5.47a)$$

800 If the trajectory is extremely close to alignment with the z axis, i.e. nearly
 801 (u_z , u_y , u_z) = (0, 0, ±1), do not use Eq. (5.24) above, but instead use:
 802

$$\begin{aligned} 803 ux &= \sin \theta \cos \varphi \\ 804 uy &= \sin \theta \sin \varphi \\ 805 \text{if } u_z \geq 0 & \\ 806 uz &= \cos \theta \\ 807 \text{else} & \\ 808 uz &= -\cos \theta \end{aligned} \quad (5.47b)$$

811 Finally, one updates the trajectory:

$$\begin{aligned} 812 \quad ux &= uxx \\ 813 \quad uy &= uyy \\ 814 \quad uz &= uzz \\ 815 \end{aligned}$$

816 The photon is now oriented along a new trajectory, and ready to take a new step
 817 *s* (see Fig. 5.5).

818 There are alternative scattering functions. Mie theory is an important scattering
 819 function to consider. Perhaps an experiment has yielded a particular scattering func-
 820 tion, and one wishes to run a simulation using this function. This chapter will not
 821 discuss these alternatives, but as long as the criteria of Eqs. (5.44a, b) are followed,
 822 most any scattering function for the deflection angle θ can be used. Sometimes
 823 the scattering function does not lend itself to a solution of $\cos\theta$ in terms of a ran-
 824 dom number, as in Eq. (5.3). Also, sometimes one wishes to consider an azimuthal
 825 scattering angle φ that depends on the deflection angle θ , as in Mie scattering of
 826 polarized light. In such cases, the “rejection method” is a useful approach [10].
 827

828

829

830 5.3.6 Terminate?

831 The photon will continue propagating as its weight becomes progressively smaller.
 832 How can one stop the photon yet properly conserve energy? The “Roulette Method”
 833 is used to terminate the photon. A threshold value (THRESHOLD) is chosen, typi-
 834 cally 10^{-4} . When the photon’s weight drops below this threshold value, the roulette
 835 procedure is employed. A random number (RND) is generated; and if this random
 836 number is less than a small fraction called CHANCE, typically 0.10, then the photon
 837 weight is increased by dividing w by CHANCE. For CHANCE = 0.10, this would
 838 be a 10-fold increase in w . Otherwise, the photon is terminated. Consequently, 9
 839 out of 10 times the photon is terminated, but 1 out of 10 times the photons weight
 840 is increased 10-fold and the photon continues to propagate. The result is that pho-
 841 tons are usually terminated, but energy is conserved by the occasional surviving
 842 photon being given extra weight. Since millions of photons are run, the statistically
 843 averaged result is correct. In summary, the roulette method is implemented by the
 844 following:
 845

```
846
847     if(w < THRESHOLD)
848         if(RND <= CHANCE)
849             w = w/CHANCE
850         else
851             terminate the photon
852
853
```

854 Once the photon is terminated, a new photon can be launched. One checks
 855 to see if the total number of photons has already reached the maximum number

(N_{photons}) requested by the input file. If not, then a new photon is launched. If yes, the simulation is complete, and it is time to prepare the results for output.

5.3.7 Normalizing Results for Output

Now, all the photons (N_{photons}) have been run, and it is time to save the output of the simulation. The data has been stored in the array A , as $A[iz][ir]$ or perhaps $A[iz][ix][iy]$, in units of [photon weight/bin]. Either way, the key parameter is the volume $V [\text{cm}^3]$ associated with each bin. For $A[iz][ir]$, the volumes vary with the value of ir ,

$$V = 2\pi(ir - 0.5)dr^2dz \quad (5.50\text{a})$$

and for $A[iz][ix][iy]$, the volumes are all equal,

$$V = dx dy dz \quad (5.50\text{b})$$

Then the values A [photon weight/bin] are normalized by the appropriate V and by the value N_{photons} to yield the absorbed fraction, A [$1/\text{cm}^3$], for each pixel:

$$A[ir, iz] = \frac{A[ir, iz]}{V[ir, iz]N_{\text{photons}}} \quad (5.51)$$

The fractional transport, T [$1/\text{cm}^2$], is then calculated as

$$T = \frac{A}{\mu_a} \quad (5.52)$$

Recall from Eqs. (5.12a, b) that fluence rate ϕ [W/cm^2] equals the incident power P [W] times T , $\phi = PT$, and the fluence ψ [J/cm^2] equals the incident energy Q [J] times T , $\psi = QT$.

Keep in mind that the values of A , V and μ_a are specific to each bin, when calculating $T[iz][ir]$ or $T[iz][ix][iy]$. In this summary of a simple implementation of the Monte Carlo method, the μ_a was assumed to be uniform, as well as the scattering properties, so every bin had the same value of μ_a . But in MCML, for example, bins at different depths can have different values of μ_a .

The light fluxes that have escaped at the front and rear surface boundaries are similarly normalized, but in this case the surface area AREA rather than the volume V is used. The value of $\text{AREA}[ir]$ is $2\pi(ir - 0.5)dr^2$. The array of escaping photons, $R_r[ir]$ [photon weight/bin], is converted to the fractional escaping flux density, $R_r[ir][1/\text{cm}^2]$, by the expression:

$$R_r = \frac{R_r}{\text{AREA } N_{\text{photons}}} \quad (5.53)$$

This completes the discussion of the steady-state Monte Carlo simulation. The output is the fractional transport, $T[iz][ir][1/\text{cm}^2]$, and the fractional escaping flux

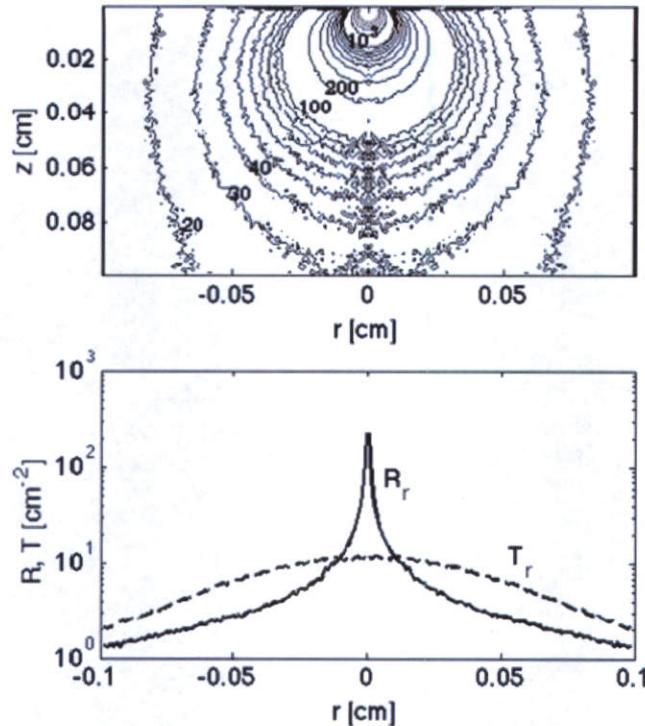


Fig. 5.6 Example steady-state Monte Carlo simulation. The optical properties were $\mu_a = 1 \text{ cm}^{-1}$, $\mu_s = 100 \text{ cm}^{-1}$, $g = 0.90$, $n = 1.4$, for a 1-mm-thick tissue slab with air/tissue boundaries at front and rear surfaces. There were 10^6 photons propagated during a run time of 3 min 49 s on a laptop computer (2 GHz processor). (a) Iso- T contours for 20, 30, 40, 100, 200 and 10^3 1/cm^2 . T denotes the fractional transport. The noise is evident along the central axis near $r = 0$ because the bins are smaller and collect fewer photons. Running more photons improves the signal-to-noise. (b) The reflectance (R_r) and transmission (T_r) as flux densities of escape [$1/\text{cm}^2$] versus radial position r at front and rear surfaces, respectively

densities, $R_r[ir][1/\text{cm}^2]$ and $T_r[ir][1/\text{cm}^2]$. This discussion ends with one example calculation shown in Fig. 5.6, showing T , R_r and T_r for a 1-mm-thick slab of tissue, with light delivered as a pencil beam of collimated light at the origin: $(x, y, z) = (0, 0, 0)$, $(ux, uy, uz) = (0, 0, 1)$.

5.4 Time Resolved Monte Carlo Propagation

Time-resolved Monte Carlo simulation is almost identical to the steady-state simulation discussed above, except for some minor changes. There are actually several ways to implement time-resolved Monte Carlo, and this section shows one approach that illustrates the basic idea.

In this example, the photon is allowed to propagate with no absorption, and the total path of the photon is accumulated after each step s throughout the propagation:

$$L = L + s \quad (5.54)$$

946 Since there is no absorption, $\mu_a = 0$ and $\mu_t = \mu_s$. Since the speed of light is c ,
 947 the time duration of the propagation is:

$$949 \quad t = \frac{L}{c} \quad (5.55)$$

951 where c is the speed of light in the tissue, $c = c_o/n$ where c_o is the speed of light in
 952 vacuo. The time for a photon to escape out the front surface is determined from its
 953 total pathlength L at the moment of escape divided by c .

955 Assume that one is interested in a set of 10 time-points, $t[it]$, where $it = 1 - 10$ is
 956 an index that refers to the desired time-point. Let the first time-point, $t[1]$, be 100 ps.
 957 Propagation is allowed to continue until the next photon step causes L to exceed the
 958 pathlength corresponding to 100 ps,

$$960 \quad L + s > t[it]/c \quad (5.56)$$

962 where $t[1] = 100$ ps for this example. At this point, a partial step size, s_1 , is taken,
 963

$$964 \quad s_1 = t[it]/c - L \quad (5.57)$$

966 The photon is now located exactly at the time-point of 100 ps. The current photon
 967 weight w is deposited in the bin $A[iz][ir][it]$, where $[it]$ selects a full 2-D $A[iz][ir]$
 968 array associated with each particular time point. The entire photon weight ($w = 1$)
 969 is placed in the local bin, but the weight of the photon is not decremented. The bin
 970 $A[iz][ir][it]$ takes a snapshot of the photon's location and weight, but does not affect
 971 the photon. There is no absorption. The photon is allowed to continue propagating.
 972 The remainder of the step size, $s - s_1$, is taken by the photon. The photon continues
 973 to propagate as usual, until a next step causes L to exceed the 2nd time point, $t[2]$.
 974 The process of taking a partial step, depositing w into $A[ir][iz][it]$ without changing
 975 the w of the photon, completing the step, and resuming propagation is executed.
 976 The process continues until L passes the last desired time point, then the photon is
 977 terminated and a second photon is launched.

978 During propagation, when a photon strikes one of the boundaries, a fraction
 979 $(1 - R_i)$ of the current photon weight will escape, and the photon weight will decre-
 980 ment. The new photon weight, $R_i w$, will internally reflect and continue propagating.
 981 The escaping photon weight will be placed in the bin $R_r[ir][jt]$,

$$983 \quad R_r[ir][jt] = R_r[ir][jt] + (1 - R_i)w \quad (5.58)$$

985 where jt is an index that encodes the time of escape, and may be divided into
 986 equal time steps, dt [s], that cover the time duration of interest. The current jt is
 987 computed:

$$989 \quad jt = \text{round}(t/dt) + 1 \quad (5.59)$$

991 As an example of evenly divided time bins, if the time duration of interest was
 992 from 10 ps to 1 ns, then dt would be 10 ps and jt would extend from 1 to 100. An
 993 alternative approach is to have progressively larger dt bin sizes, so the time base can
 994 extend from very short times to very long times, which is not discussed here.

995 Finally, after N_{photons} have been launched and terminated, it is time to normalize
 996 the bins A and R_r for final output. For each time point, $[it]$, conservation of energy
 997 in terms of photon weight is summarized:

$$999 \quad \frac{1}{N_{\text{photons}}} \left(\sum_{iz} \sum_{ir} A[iz][ir][it] + \sum_{jt=1}^{it} R_r[ir][jt] \right) = 1 \quad (5.60)$$

1002 The above calculation is not routinely needed, but is only a check that energy is
 1003 conserved. The final normalization is

$$1006 \quad T = c \frac{A}{V N_{\text{photons}}} \quad (5.61)$$

1009 which has units of $[1/(\text{cm}^2 \text{ s})]$. The time-resolved fluence rate, $[\text{W}/\text{cm}^2]$ in response
 1010 to an impulse of energy Q [J] delivered at time zero is:

$$1012 \quad \phi(t) = QT \quad (5.62)$$

1014 The escaping flux density, $R_r[1/(\text{cm}^2 \text{ s})]$, is normalized:

$$1016 \quad R_r = \frac{R_r}{\text{AREA } N_{\text{photons}} dt} \quad (5.63)$$

1019 Both $A[iz][ir][it]$ and $R_r[ir][it]$ will have good signal-to-noise in regions near the
 1020 source where most photons spend their time, and poor signal-to-noise in regions far
 1021 from the source. It is difficult to get good results far from the source even when a
 1022 large number of photons are launched. Usually, time-resolved Monte Carlo simula-
 1023 tion is used to specify results close to a source, and time-resolved diffusion theory
 1024 is used to specify results at far distances from the source.

1025 Now that the Monte Carlo simulation is completed, absorption can be added to
 1026 the problem. The attenuation due to absorption is specified by Beer's law which
 1027 says that photon survival equals $\exp(-\mu_a ct)$, since the ct equals the photon's total
 1028 pathlength L at any particular time t . Therefore,

$$1030 \quad \phi(t) = QT(t)e^{-\mu_a ct} \quad (5.64)$$

1032 and

$$1034 \quad R_r(t) = \frac{R_r(t)}{\text{AREA } N_{\text{photons}} dt} e^{-\mu_a ct} \quad (5.65)$$

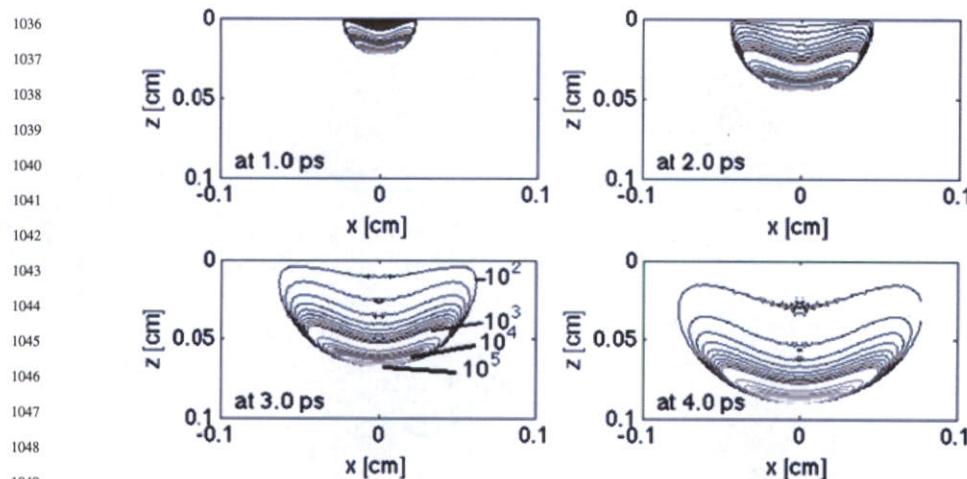


Fig. 5.7 Time-resolved propagation of a collimated laser impulse launched at $r = 0$, $z = 0$ into a 1-mm thickness of a standard tissue. The thickness represents ~ 10 optical depths for optical properties of $\mu_a = 1 \text{ cm}^{-1}$, $\mu_s = 100 \text{ cm}^{-1}$, $g = 0.90$, $n = 1.4$. The four time points are 1, 2, 3 and 4 ps. The maps are iso- T contours, where T is the time-resolved fractional transport [$1/(\text{cm}^2 \text{ s})$]. The influence of μ_a is minor, for example, $\exp(-\mu_a c t)$ is only 0.92 when $t = 4 \text{ ps}$

In this way, any value of μ_a can be introduced to learn its influence on the time-resolved distribution of $T(t)$ or $R_r(t)$.

To illustrate time-resolved Monte Carlo, an example of snapshots of $T[iz][ir]$ at 4 time-points is presented in Fig. 5.7, which illustrates the early movement of an incident laser pulse into a tissue. The narrow beam laser pulse broadens with time due to scattering.

Figure 5.8 shows an optical fiber embedded within a tissue and terminating at a depth of 500 μm , delivering light toward the surface. As the light reaches the surface, the time-resolved escape of fractional flux density, $R_r(t, r)[1/(\text{cm}^2 \text{ s})]$ is shown.

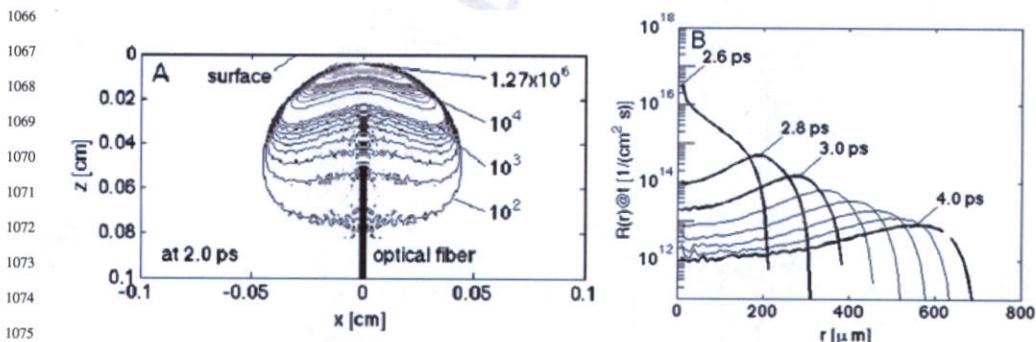


Fig. 5.8 Time-resolved escape of a laser pulse from within tissue. The optical properties were $\mu_s = 100 \text{ cm}^{-1}$, $g = 0.90$, $n_{\text{tissue}} = 1.33$. The impulse is delivered toward the surface from an optical fiber terminated at a depth of 500 μm within the tissue and pointed toward the surface. (a) Impulse at time 2 ps, shown as $T[1/(\text{cm}^2 \text{ s})]$, maximum is 1.27×10^6 . (b) The escaping fractional flux density versus radial position at different times, $R_r(t, r)[1/(\text{cm}^2 \text{ s})]$

5.5 Converting Time-Resolved Results to Frequency-Domain

When the intensity of a light source is modulated at very high frequencies, the ability of the frequency of modulation to transport to some position of observation is described as *frequency domain* light transport. Time-resolved information generated by a time-resolved Monte Carlo simulation can be converted by Fourier Transform into frequency domain information.

Consider the time-resolved escape of fractional flux density, $R_r(t, r)[1/(cm^2 s)]$, when light is delivered as a collimated impulse to position $(r, z) = (0, 0)$ on a tissue surface. The light enters the tissue, but due to scattering begins to escape from the tissue after some delay. This time-resolved $R_r(t, r)$ is shown in Fig. 5.9a, for a typical tissue. As the position of observation moves from 1 mm to 6 mm distance from the source, the time delay before onset of escaping flux increases, and the amount of light escaping decreases.

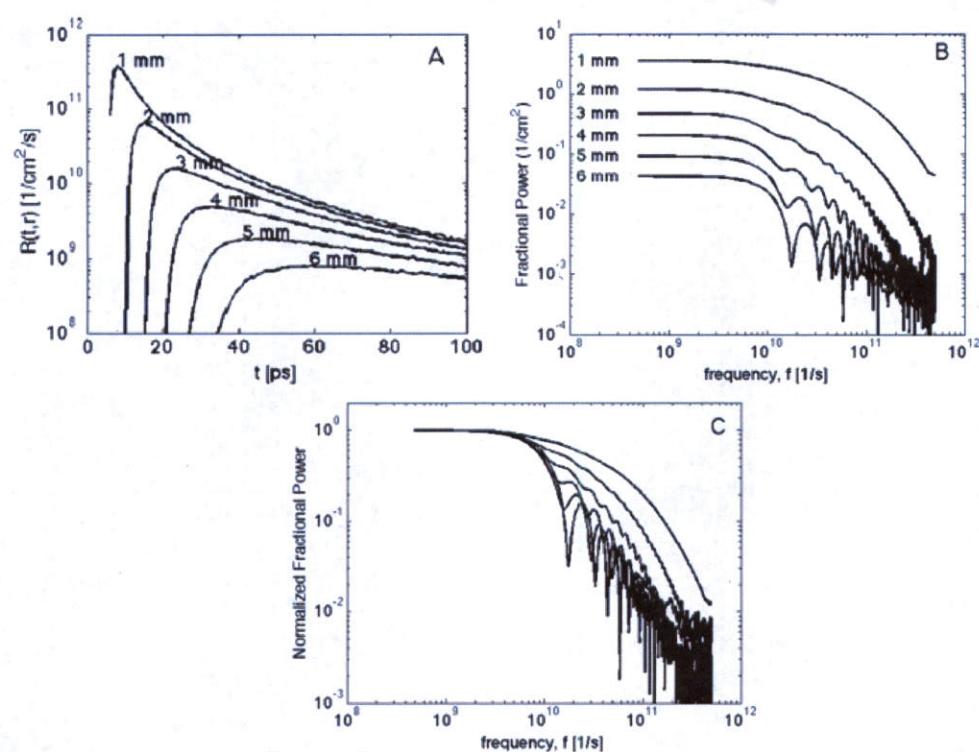


Fig. 5.9 Frequency domain light transport, showing how light modulated at different frequencies and delivered as a collimated beam to position $(r, z) = (0, 0)$, will escape from the tissue as a function of radial position. (a) The time-resolved escape of light from the tissue, $R_r[1/(cm^2 s)]$ for $r = 1 - 6$ mm. (b) The fractional power spectrum [$1/cm^2$] versus frequency f [1/s]. The optical properties of the tissue were $\mu_a = 1\text{cm}^{-1}$, $\mu_s = 100\text{ cm}^{-1}$, $g = 0.90$, $n = 1.4$, and 10^7 photons were launched during a 35-min simulation. The data at the higher frequencies for the most distant radial positions were based on low photon weights and show artifactual oscillations. (c) Normalized fractional power versus frequency

1126 The corresponding frequency domain information is obtained by using a Fast
 1127 Fourier Transform (*fft*) to convert the time-resolved $R_r(t, r)$ escaping at a particular
 1128 radial position r , denoted as $R_r(t)$, into the frequency domain:

1129

1130

$$F(f) = \text{abs}(\text{fft}(R_r(t)dt)) \quad (5.66)$$

1131

1132 where dt is the time-step of the time-resolved data. The absolute value converts the
 1133 imaginary values generated by the *fft()* into real values that correspond to the power
 1134 spectrum, expressed as the fractional power $F [1/cm^2]$. In other words, if the source
 1135 was modulated at a frequency $f [1/s]$, the function F would specify the fractional
 1136 escaping flux, $R_r[1/cm^2]$, that was still modulated at frequency f . Figure 5.9b shows
 1137 this power spectrum. The limiting values toward low frequencies of modulation
 1138 correspond to the steady-state reflectance $R_r(r)[1/cm^2]$, which is why the factor dt
 1139 was included in the above equation.

1140 To illustrate the above equation more specifically, the equivalent programming
 1141 code written in MATLABTM notation is listed:

1142

```
1143 mua = 1; % absorption coefficient [cm^-1]
1144 mus = 100; % scattering coefficient [cm^-1]
1145 g = 0.90; % anisotropy of scattering [dimensionless]
1146 n = 1.5; % refractive index [dimensionless]
1147 dt = 1e-12; % time step of time-resolved data [s], in this case dt = 1 ps.
1148 t = (1:100)'*dt; % time base of Monte Carlo data [s], up to 100 ps
1149 r = (1:100)'/100*1.0; % radial position of Monte Carlo data [cm], up to 1 cm
1150 Rr = getRrMonteCarlo(t,r,mua,mus,g,n); % get Monte Carlo data [1/cm^2/s],
1151 not shown
1152 N = 2048; % adds zeros to end of time-resolved data, for padding the transform
1153 f = (1:N/2)'/N/dt; % the x-axis frequency of the power spectrum
1154 ir = 10; % selects one radial position r(ir)
1155 F = abs(fft(Rr(:,ir)*dt, N)); % Rr(:,ir) is the time-resolved Rr(t) at r(ir)
1156 plot(f, F) % the plot command yields the power spectrum in [1/cm^2]
```

1157

1158 The original data $R_r(t, r)$ has 100 time-points, but the *fft()* operates best when the
 1159 number of data points is a multiple of 2. Therefore, zeros are added to the end of
 1160 the data, which is called *padding*. Adding more zeros causes the result to have more
 1161 points, so the curves look smoother. In this case 1948 zeros were added to yield a
 1162 final 2048 data points. The above program yielded Fig. 5.9b. Figure 5.9c normalizes
 1163 the data by the DC value of R_r , so as to emphasize the shape of the power spectra.

1164 Note in this example that light escaping at 1 mm from the source has higher
 1165 frequency content than light escaping at 6 mm from the source. In practical fre-
 1166 quency domain measurements, the detection of light transport is usually made at
 1167 least 10 mm from the source, where diffusion theory suffices to describe light
 1168 transport, and the frequency content of interest is in the hundreds of MHz [1/s].
 1169 This example shows how time-resolved Monte Carlo data can be converted to the
 1170 frequency domain to address questions where diffusion theory is inadequate.

1171 **5.6 Summary**

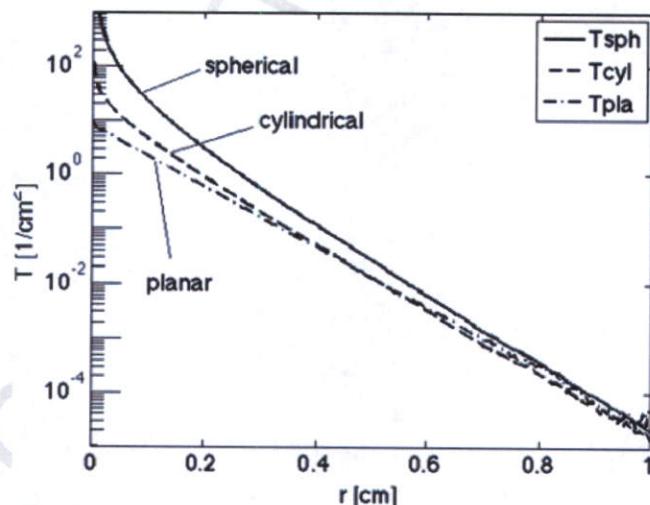
1172
 1173 Monte Carlo simulations are a relatively simple and flexible method for exploring
 1174 the behavior of light transport in biological tissues or other media with average
 1175 absorption and scattering properties. The simulations are like experiments involving
 1176 a number of photons (N_{photons}) and hence the simulations take time to execute. Some
 1177 simulations take only a minute, and others take hours or even days. If you are trying
 1178 to fill small bins with rare photons, the simulations take a lot of time. But common
 1179 problems take about 10 min or less to get a good result.

1180 The strength of Monte Carlo simulations is to treat situations where diffusion
 1181 theory, or some other analytic expression of light transport, fails. It is not the proper
 1182 tool for every job.

1183
 1184 **5.7 Appendix: mc321.c**

1185 A simplest version of a Monte Carlo simulation, called `mc321.c`, is listed below.
 1186 The program can be downloaded from the web [8], but since websites may change,
 1187 the program is listed here as an archival example, and as an easy reference while
 1188 reading this chapter. This program does not consider escape across boundaries, but
 1189 shows the basic format of photon propagation, recording and normalization. The
 1190 output of this program is plotted in Fig. 5.10:
 1191

1192 **Fig. 5.10** The data in
 1193 `mc321.out`, produced by
 1194 `mc321.c`, plotted using a
 1195 separate graphics program.
 1196 The spherical transport from
 1197 a point source, the cylindrical
 1198 transport from a line source,
 1199 and the planar transport from
 1200 a planar source are shown
 1201 versus the distance, r , from
 1202 the source



1203
 1204
 1205
 1206
 1207
 1208
 1209
 1210
 1211 /*****
 1212 * mc321.c , in ANSI Standard C programming language
 1213 *
 1214 * Monte Carlo simulation yielding spherical, cylindrical, and planar
 1215 * responses to an isotropic point source (equivalent to a plane source,

```

1216 *      line source, and point source, respectively) in an infinite homogeneous
1217 *      medium with no boundaries. This program is a minimal Monte Carlo
1218 *      program scoring photon distributions in spherical, cylindrical,
1219 *      and planar shells.
1220 *
1221 *      by Steven L. Jacques based on prior collaborative work
1222 *      with Lihong Wang, Scott Prahl, and Marleen Keijzer.
1223 *      partially funded by the NIH (R29-HL45045, 1991-1997) and
1224 *      the DOE (DE-FG05-91ER617226, DE-FG03-95ER61971, 1991-1999).
1225 *
1226 *      A published report illustrates use of the program:
1227 *      S. L. Jacques: "Light distributions from point, line, and plane
1228 *      sources for photochemical reactions and fluorescence in turbid
1229 *      biological tissues," Photochem. Photobiol. 67:23-32, 1998.
1230 *****/
1231
1232 #include <math.h>
1233 #include <stdio.h>
1234
1235 #define Nbins          500
1236 #define Nbinspl        501
1237 #define PI            3.1415926
1238 #define LIGHTSPEED    2.997925E10 /* in vacuo speed of light [cm/s] */
1239 #define ALIVE          1           /* if photon not yet terminated */
1240 #define DEAD           0           /* if photon is to be terminated */
1241 #define THRESHOLD     0.01        /* used in roulette */
1242 #define CHANCE         0.1         /* used in roulette */
1243 #define COS90D        1.0E-6
1244 /* If cos(theta) <= COS90D, theta >= PI/2 - 1e-6 rad. */
1245 #define ONE_MINUS_COSZERO 1.0E-12
1246 /* If 1-cos(theta) <= ONE_MINUS_COSZERO, fabs(theta) <= 1e-6 rad. */
1247 /* If 1+cos(theta) <= ONE_MINUS_COSZERO, fabs(PI-theta) <= 1e-6 rad. */
1248 #define SIGN(x)        ((x)>=0 ? 1:-1)
1249 #define InitRandomGen (double) RandomGen(0, 1, NULL)
1250 /* Initializes the seed for the random number generator. */
1251 #define RandomNum       (double) RandomGen(1, 0, NULL)
1252 /* Calls for a random number from the random number generator. */
1253
1254 /* DECLARE FUNCTION */
1255 double RandomGen(char Type, long Seed, long *Status);
1256 /* Random number generator */
1257 main() {
1258
1259 /* Propagation parameters */
1260 double x, y, z;    /* photon position */

```

```

1261 double ux, uy, uz; /* photon trajectory as cosines */
1262 double uxx, uyy, uzz; /* temporary values used during SPIN */
1263 double s; /* step sizes. s = -log(RND)/mus [cm] */
1264 double costheta; /* cos(theta) */
1265 double sintheta; /* sin(theta) */
1266 double cospsi; /* cos(psi) */
1267 double sinpsi; /* sin(psi) */
1268 double psi; /* azimuthal angle */
1269 double i_photon; /* current photon */
1270 double W; /* photon weight */
1271 double absorb; /* weighted deposited in a step due to absorption */
1272 short photon_status; /* flag = ALIVE=1 or DEAD=0 */
1273
1274 /* other variables */
1275 double Csph[Nbinspl]; /* spherical photon concentration CC[ir=0..100] */
1276 double Ccyl[Nbinspl]; /* cylindrical photon concentration CC[ir=0..100] */
1277 double Cpla[Nbinspl]; /* planar photon concentration CC[ir=0..100] */
1278 double Fsp; /* fluence in spherical shell */
1279 double Fcyl; /* fluence in cylindrical shell */
1280 double Fpla; /* fluence in planar shell */
1281 double mua; /* absorption coefficient [cm^-1] */
1282 double mus; /* scattering coefficient [cm^-1] */
1283 double g; /* anisotropy [-] */
1284 double albedo; /* albedo of tissue */
1285 double nt; /* tissue index of refraction */
1286 double Nphotons; /* number of photons in simulation */
1287 short NR; /* number of radial positions */
1288 double radial_size; /* maximum radial size */
1289 double r; /* radial position */
1290 double dr; /* radial bin size */
1291 short ir; /* index to radial position */
1292 double shellvolume; /* volume of shell at radial position r */
1293 double CNT; /* total count of photon weight summed over all bins */
1294
1295 /* dummy variables */
1296 double rnd; /* assigned random value 0-1 */
1297 short i, j; /* dummy indices */
1298 double u, temp; /* dummy variables */
1299 FILE* target; /* point to output file */
1300
1301 **** INPUT
1302 Input the optical properties
1303 Input the bin and array sizes
1304 Input the number of photons
1305 ****/

```

```

1306 mua      = 1.673; /* cm^-1 */
1307 mus      = 312.0; /* cm^-1 */
1308 g        = 0.90;
1309 nt       = 1.33;
1310 Nphotons = 10000; /* set number of photons in simulation */
1311 radial_size = 2.0; /* cm, total range over which bins extend */
1312 NR       = Nbins; /* set number of bins. */
1313 /* IF NR IS ALTERED, THEN USER MUST ALSO ALTER THE ARRAY DECLARATION TO A
1314     SIZE = NR+1. */
1315 dr       = radial_size/NR; /* cm */
1316 albedo   = mus/(mus + mua);
1317
1318 /****** INITIALIZATIONS
1319 *****/
1320 i_photon = 0;
1321 InitRandomGen;
1322 for (ir=0; ir<=NR; ir++) {
1323     Csphe[ir] = 0;
1324     Ccyl[ir] = 0;
1325     Cpla[ir] = 0;
1326 }
1327
1328
1329 /****** RUN
1330     Launch N photons, initializing each one before propagation.
1331 *****/
1332 do {
1333
1334 /****** LAUNCH
1335     Initialize photon position and trajectory.
1336     Implements an isotropic point source.
1337 *****/
1338 i_photon += 1; /* increment photon count */
1339 if ( fmod(i_photon, Nphotons/10) == 0)
1340     printf("%0.0f%% done\n", i_photon/Nphotons*100);
1341
1342 W = 1.0; /* set photon weight to one */
1343 photon_status = ALIVE; /* Launch an ALIVE photon */
1344
1345 x = 0; /* Set photon position to origin. */
1346 y = 0;
1347 z = 0;
1348
1349 /* Randomly set photon trajectory to yield an isotropic source. */
1350 costheta = 2.0*RandomNum - 1.0;

```

```

1351 sintheta = sqrt(1.0 - costheta*costheta); /* sintheta is always positive */
1352 psi = 2.0*PI*RandomNum;
1353 ux = sintheta*cos(psi);
1354 uy = sintheta*sin(psi);
1355 uz = costheta;
1356
1357 /* HOP_DROP_SPIN_CHECK
1358 Propagate one photon until it dies as determined by ROULETTE.
1359 *****/
1360 do {
1361
1362 /****** HOP
1363 Take step to new position
1364 s = step size
1365 ux, uy, uz are cosines of current photon trajectory
1366 *****/
1367 while ((rnd = RandomNum) <= 0.0); /* yields 0 < rnd <= 1 */
1368 s = -log(rnd)/(mua + mus); /* Step size. Note: log() is base e */
1369 x += s * ux; /* Update positions. */
1370 y += s * uy;
1371 z += s * uz;
1372 /****** DROP
1373 Drop photon weight (W) into local bin.
1374 *****/
1375 absorb = W*(1 - albedo); /* photon weight absorbed at this step */
1376 W -= absorb; /* decrement WEIGHT by amount absorbed */
1377
1378 /* spherical */
1379 r = sqrt(x*x + y*y + z*z); /* current spherical radial position */
1380 ir = (short)(r/dr); /* ir = index to spatial bin */
1381 if (ir >= NR) ir = NR; /* last bin is for overflow */
1382 Cph[ir] += absorb; /* DROP absorbed weight into bin */
1383
1384 /* cylindrical */
1385 r = sqrt(x*x + y*y); /* current cylindrical radial position */
1386 ir = (short)(r/dr); /* ir = index to spatial bin */
1387 if (ir >= NR) ir = NR; /* last bin is for overflow */
1388 Ccyl[ir] += absorb; /* DROP absorbed weight into bin */
1389
1390 /* planar */
1391 r = fabs(z); /* current planar radial position */
1392 ir = (short)(r/dr); /* ir = index to spatial bin */
1393 if (ir >= NR) ir = NR; /* last bin is for overflow */
1394 Cpla[ir] += absorb; /* DROP absorbed weight into bin */
1395

```

```

1396  /**** SPIN
1397      Scatter photon into new trajectory defined by theta and psi.
1398      Theta is specified by cos(theta), which is determined
1399      based on the Henyey-Greenstein scattering function.
1400      Convert theta and psi into cosines ux, uy, uz.
1401  *****/
1402  /* Sample for costheta */
1403  rnd = RandomNum;
1404  if (g == 0.0)
1405      costheta = 2.0*rnd - 1.0;
1406  else {
1407      double temp = (1.0 - g*g)/(1.0 - g + 2*g*rnd);
1408      costheta = (1.0 + g*g - temp*temp)/(2.0*g);
1409  }
1410  sintheta = sqrt(1.0 - costheta*costheta); /* sqrt() is faster than sin(). */
1411  /* Sample psi. */
1412  psi = 2.0*PI*RandomNum;
1413  cospsi = cos(psi);
1414  if (psi < PI)
1415      sinpsi = sqrt(1.0 - cospsi*cospsi); /* sqrt() is faster than sin(). */
1416  else
1417      sinpsi = -sqrt(1.0 - cospsi*cospsi);
1418
1419  /* New trajectory. */
1420  if (1 - fabs(uz) <= ONE_MINUS_COSZERO) { /* close to perpendicular. */
1421      uxx = sintheta * cospsi;
1422      uyy = sintheta * sinpsi;
1423      uzz = costheta * SIGN(uz); /* SIGN() is faster than division. */
1424  }
1425  else { /* usually use this option */
1426      temp = sqrt(1.0 - uz * uz);
1427      uxx = sintheta * (ux * uz * cospsi - uy * sinpsi) / temp + ux * costheta;
1428      uyy = sintheta * (uy * uz * cospsi + ux * sinpsi) / temp + uy * costheta;
1429      uzz = -sintheta * cospsi * temp + uz * costheta;
1430  }
1431
1432  /* Update trajectory */
1433  ux = uxx;
1434  uy = uyy;
1435  uz = uzz;
1436
1437  /**** CHECK ROULETTE
1438  If photon weight below THRESHOLD, then terminate photon using Roulette
1439  technique.
1440

```

```
1441     Photon has CHANCE probability of having its weight increased by factor of
1442         1/CHANCE,
1443         and 1-CHANCE probability of terminating.
1444     *****/
1445     if (W < THRESHOLD) {
1446         if (RandomNum <= CHANCE)
1447             W /= CHANCE;
1448         else photon_status = DEAD;
1449     }
1450 } /* end STEP_CHECK_HOP_SPIN */
1451 while (photon_status == ALIVE);
1452
1453 /* If photon dead, then launch new photon. */
1454 } /* end RUN */
1455 while (i_photon < Nphotons);
1456
1457 /***** SAVE
1458     Convert data to relative fluence rate [cm^-2] and save to file called
1459     "mcmin321.out".
1460 */
1461 target = fopen("mc321.out", "w");
1462
1463 /* print header */
1464 fprintf(target, "number of photons = %f\n", Nphotons);
1465 fprintf(target, "bin size = %5.5f [cm] \n", dr);
1466 fprintf(target, "last row is overflow. Ignore.\n");
1467
1468 /* print column titles */
1469 fprintf(target, "r [cm] \t Fsph [1/cm2] \t Fcyl [1/cm2] \t Fpla [1/cm2]\n");
1470
1471 /* print data: radial position, fluence rates for 3D, 2D, 1D geometries */
1472 for (ir=0; ir<=NR; ir++) {
1473     /* r = sqrt(1.0/3 - (ir+1) + (ir+1)*(ir+1))*dr; */
1474     r = (ir + 0.5)*dr;
1475     shellvolume = 4.0*PI*r*r*dr; /* per spherical shell */
1476     Fsph = Csph[ir]/Nphotons/shellvolume/mua;
1477     shellvolume = 2.0*PI*r*dr; /* per cm length of cylinder */
1478     Fcyl = Ccyl[ir]/Nphotons/shellvolume/mua;
1479     shellvolume = dr; /* per cm2 area of plane */
1480     Fpla = Cpla[ir]/Nphotons/shellvolume/mua;
1481     fprintf(target, "%5.5f \t %4.3e \t %4.3e \t %4.3e \n", r, Fsph, Fcyl, Fpla);
1482 }
1483
1484 fclose(target);
1485
```

```

1486 } /* end of main */

1487

1488 /* SUBROUTINES */

1489

1490 /***** RandomGen *****
1491 *      RandomGen
1492 *      A random number generator that generates uniformly
1493 *      distributed random numbers between 0 and 1 inclusive.
1494 *      The algorithm is based on:
1495 *      W.H. Press, S.A. Teukolsky, W.T. Vetterling, and B.P.
1496 *      Flannery, "Numerical Recipes in C," Cambridge University
1497 *      Press, 2nd edition, (1992).
1498 *      and
1499 *      D.E. Knuth, "Seminumerical Algorithms," 2nd edition, vol. 2
1500 *      of "The Art of Computer Programming", Addison-Wesley, (1981).
1501 *
1502 *      When Type is 0, sets Seed as the seed. Make sure 0<Seed<32000.
1503 *      When Type is 1, returns a random number.
1504 *      When Type is 2, gets the status of the generator.
1505 *      When Type is 3, restores the status of the generator.
1506 *
1507 *      The status of the generator is represented by Status[0..56].
1508 *
1509 *      Make sure you initialize the seed before you get random
1510 *      numbers.
1511 ****/
1512 #define MBIG 1000000000
1513 #define MSEED 161803398
1514 #define MZ 0
1515 #define FAC 1.0E-9
1516
1517 double RandomGen(char Type, long Seed, long *Status){
1518     static long i1, i2, ma[56]; /* ma[0] is not used. */
1519     long      mj, mk;
1520     short     i, ii;
1521
1522     if (Type == 0) {           /* set seed. */
1523         mj = MSEED - (Seed < 0 ? -Seed : Seed);
1524         mj %= MBIG;
1525         ma[55] = mj;
1526         mk = 1;
1527         for (i = 1; i <= 54; i++) {
1528             ii = (21 * i) % 55;
1529             ma[ii] = mk;
1530             mk = mj - mk;
1531             if (mk < MZ)

```

```

1531     mk += MBIG;
1532     mj = ma[ii];
1533 }
1534 for (ii = 1; ii <= 4; ii++) {
1535     for (i = 1; i <= 55; i++) {
1536         ma[i] -= ma[1 + (i + 30) % 55];
1537         if (ma[i] < MZ)
1538             ma[i] += MBIG;
1539     }
1540     i1 = 0;
1541     i2 = 31;
1542 } else if (Type == 1) /* get a number. */
1543     if (++i1 == 56)
1544         i1 = 1;
1545     if (++i2 == 56)
1546         i2 = 1;
1547     mj = ma[i1] - ma[i2];
1548     if (mj < MZ)
1549         mj += MBIG;
1550     ma[i1] = mj;
1551     return (mj * FAC);
1552 } else if (Type == 2) /* get status. */
1553     for (i = 0; i < 55; i++)
1554         Status[i] = ma[i + 1];
1555     Status[55] = i1;
1556     Status[56] = i2;
1557 } else if (Type == 3) /* restore status. */
1558     for (i = 0; i < 55; i++)
1559         ma[i + 1] = Status[i];
1560     i1 = Status[55];
1561     i2 = Status[56];
1562 } else
1563     puts("Wrong parameter to RandomGen().");
1564 return (0);
1565 }
1566 #undef MBIG
1567 #undef MSEED
1568 #undef MZ
1569 #undef FAC
1570

```

References

1. Wilson BC and Adam G. A Monte Carlo model for the absorption and flux distributions of light in tissue. *Med. Phys.* 10:824–830 (1983).
2. Keijzer M, Jacques SL, Prahl SA, and Welch AJ. Light distributions in artery tissue: Monte Carlo simulations for finite-diameter laser beams. *Lasers Surg. Med.* 9:148–154 (1989).

- 1576 3. Prahl, SA, Keijzer, Jacques SL, and Welch AJ. A Monte Carlo model of light propagation in
1577 tissue. In: G Müller and D Sliney (eds) *Dosimetry of laser radiation in medicine and biology*,
1578 SPIE Series, Vol. IS 5, pp. 102–111 (1989).
- 1579 4. Wang L-H, Jacques SL, and Zheng L-Q. MCML – Monte Carlo modeling of photon transport
in multi-layered tissues. *Comput. Methods Programs Biomed.*, 47:131–146 (1995).
- 1580 5. Jacques, SL, <http://omlc.ogi.edu/software/mc/mcml>, Oregon Health & Science University,
1581 2010. This site includes a 178-page manual on MCML. Also, a convolution program, CONV,
1582 is available for convolving the point spread functions generated by MCML.
- 1583 6. Wang, LV, <http://labs.seas.wustl.edu/bme/Wang/mc.html>, Washington University in St. Louis,
2010. Alternative site for obtaining MCML, CONV and the manual.
- 1584 7. Jacques SL. Light distributions from point, line, and plane sources for photochemical
1585 reactions and fluorescence in turbid biological tissues. *Photochem. Photobiol.* 67:23–32
(1998).
- 1587 8. Jacques, SL, <http://omlc.ogi.edu/software/mc/mc321>, Oregon Health & Science University,
2010. This site lists the minimal Monte Carlo program mc321.m, used in Ref. 7.
- 1588 9. Jacques SL. Monte Carlo simulations of fluorescence in turbid media, Ch. 6. In: MA Mycek
1589 and BW Pogue (eds) *Handbook of biomedical fluorescence*. Marcel-Dekker, New York, NY
(2003).
- 1591 10. Jacques, SL, <http://omlc.ogi.edu/software/mc/mcsub>, Oregon Health & Science University,
1592 2010. This site lists the subroutine mcsub() that can be called by c programs to run a Monte
1593 Carlo simulation.
- 1594 11. Ramella-Roman JC, Prahl SA, and Jacques SL. Three Monte Carlo programs of polarized
light transport into scattering media: part I. *Opt. Express* 13(12):4420–4438(2005).
- 1595 12. Ramella-Roman JC, Prahl SA, and Jacques SL, <http://omlc.ogi.edu/software/mc/polarizedlight>,
1596 Oregon Health & Science University, 2010. This site lists the program for Monte Carlo
1597 simulation of polarized light, reported in Ref. 11.
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of Laser-Irradiated Tissue**

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