

## CSC 314, Exam II Review

**Note:** This review is not comprehensive, but contains several practice problems to help prepare for Exam II. In addition to these practice problems, you should make sure to understand all labs assigned since the first exam, as well as the cancer bio project.

### 1. Sequence Database questions

Look at the GenBank entry with accession number NM\_001185098.1 to answer the questions below:

- What is the length of this sequence?
- When was the sequence last modified?
- How many exons does this gene have?
- What position marks the beginning and end of the poly-adenylation signal sequence, and what is this sequence?
- What are the first nine nucleotides in the coding sequence (CDS), and the corresponding amino acids (you can give the 1-letter amino acid codes)?

### 2. Sequence Alignments

For (a) and (b), use a linear gap penalty of 4 points, a match score of +5 points, and a mismatch score of -1 point.

- Find the optimal global alignment and optimal global alignment score for the words *handy* and *say*. You must show your dynamic programming matrix to receive credit.
- Find the optimal local alignment and optimal local alignment score between the words *stars* and *that*. You must show your dynamic programming matrix to receive credit.
- Using the BLOSUM-62 matrix, a gap opening penalty of 5, and a gap extension penalty of 1, find the score of the *semiglobal* alignment given below (Recall that semiglobal alignments do not penalize gaps at the beginning or end of the alignment).

```
F R I D A - - Y
- - P - A R T Y
```

### Coding questions

3. The regular expression corresponding to a standard coding sequence (CDS) is given by: "ATG(?:[ACTG]{3})+?(?:TAG|TAA|TGA)"

Note: it is not necessary to understand the regular expression (this is much more advanced than the examples covered in class). But for completeness, the regular expression can be interpreted as follows:

- ATG – the start codon, ATG
- (?:[ACTG]{3})+?- any number of codons (any three nucleotides) (non-greedy)
- (?:TAG|TAA|TGA)– any of the stop codons

Suppose that a file called *sequences.fasta* contains a large number of sequences in FASTA format. Write a python script that generates a list of the sequences that contain at least one possible CDS.

4. Suppose that the header of a FASTA sequence is of the form: *Human\_GeneName* (e.g., *Human\_TP53*). If the object *ids* contains a list of FASTA headers, write python statements that extract the corresponding gene names, which are stored in a list.
5. Suppose that the file “contacts.txt” contains a person’s last name, first name, and e-mail address separated by tabs, as in the format below. Note that the first row is a header.

Last	First	E-mail
Dancik	Garrett	<a href="mailto:dancikg@easternct.edu">dancikg@easternct.edu</a>
Smith	Joe	<a href="mailto:smithj@gmail.com">smithj@gmail.com</a>
McGuire	Madison	<a href="mailto:MadMcG@gmail.com">MadMcG@gmail.com</a>

Write a python script that reads in this file and creates a new file named “emails.txt” that contains *only* the e-mail addresses (i.e., it does not contain the header).

6. **BLAST**. The protein sequence for the *C. elegans* gene F55F8.2 can be found at accession NP\_491652. BLAST this protein to find similar proteins in humans (limit your analysis to *Homo sapiens*). Note: according to Biomart, the protein DDX24 is a human ortholog of this worm gene, based on the fact that there is a high similarity.
- a. What is the % identity and % similarity of the alignment between F55F8.2 and DDX24?
  - b. What pfam domains are present in this protein? Based on these domains, what do you infer about the function of F55F8.2?