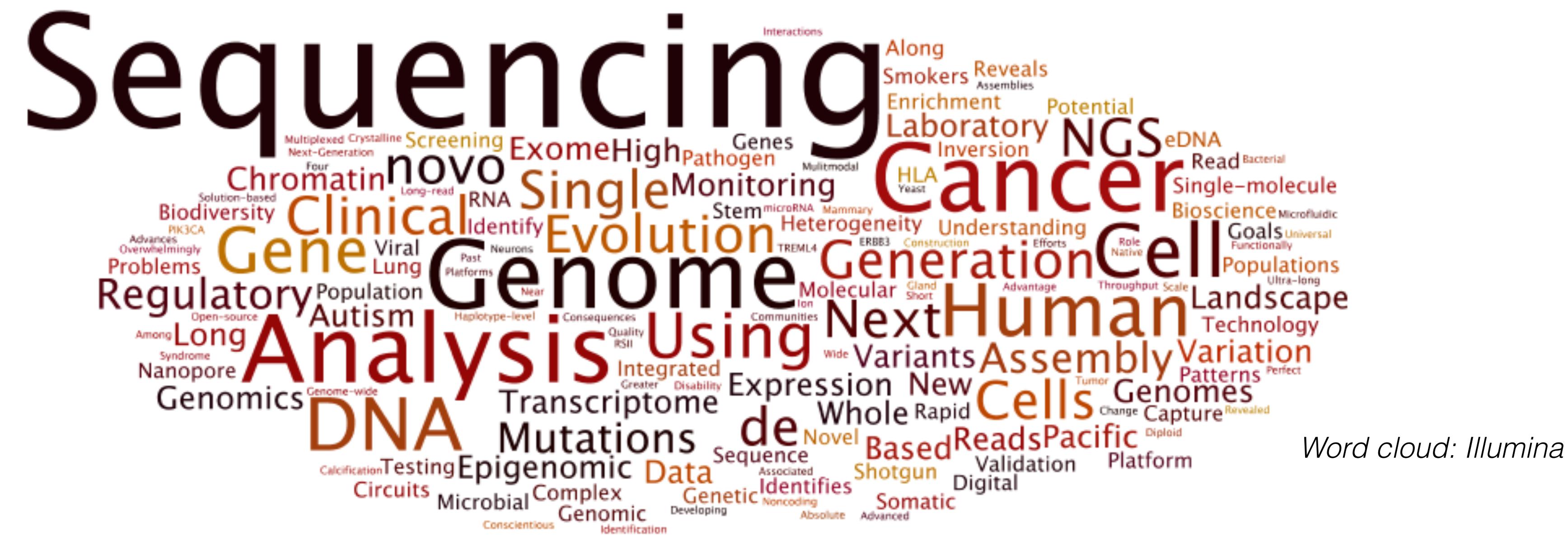


# An overview of genomics and sequencing terminology and practices

Mark Stenglein, GDW



The jargon and terminology associated with genomics and ‘next gen’ sequencing can be confusing and intimidating



The goal of this lecture is to explain and demystify some common jargon  
and explain how sequencing works

There is a glossary available online that explains many of these terms

- [Transcriptome](#)
- [Variant](#)
- [WGS](#)

### 16S

The **16S** ribosomal RNA gene is present in all bacterial and archaeal genomes. This gene is sufficiently conserved that primers that anneal to conserved regions of the gene will amplify essentially any prokaryotic 16S rRNA gene. These PCR products (amplicons) can be sequenced to provide a survey of microbial diversity in a sample.

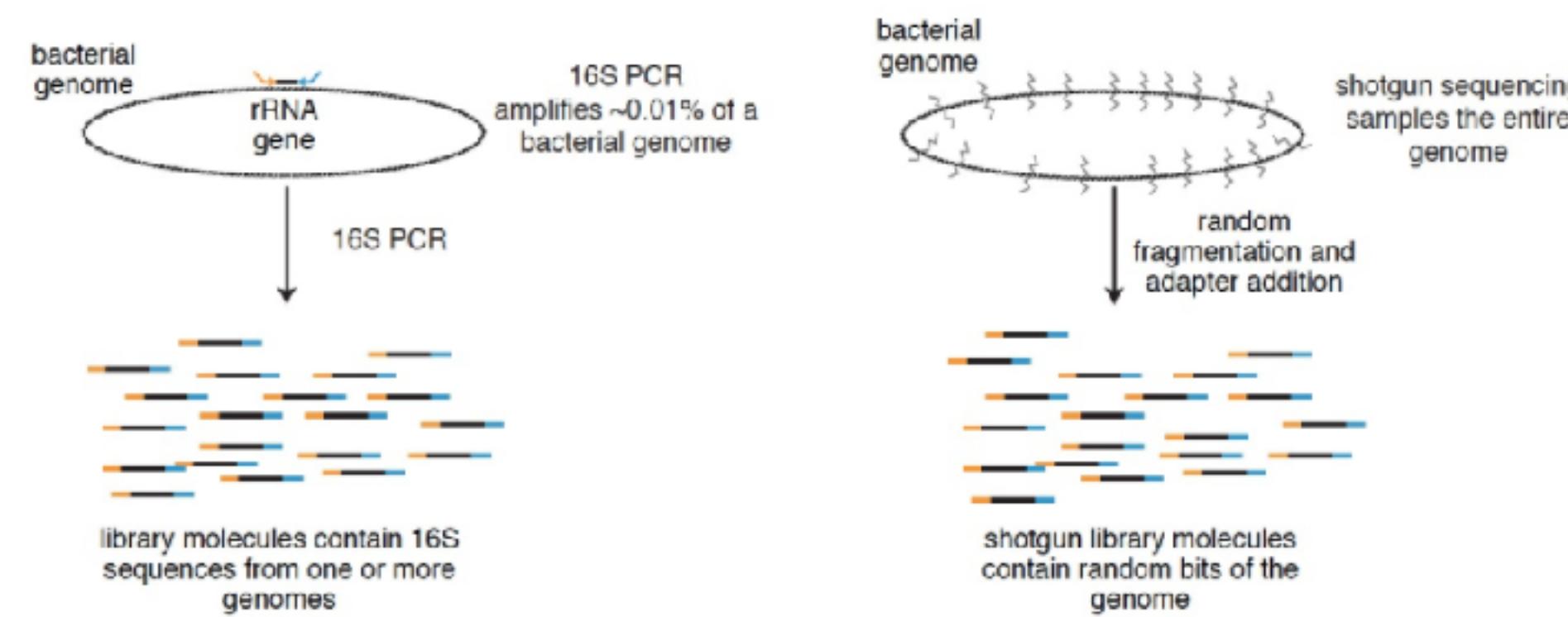


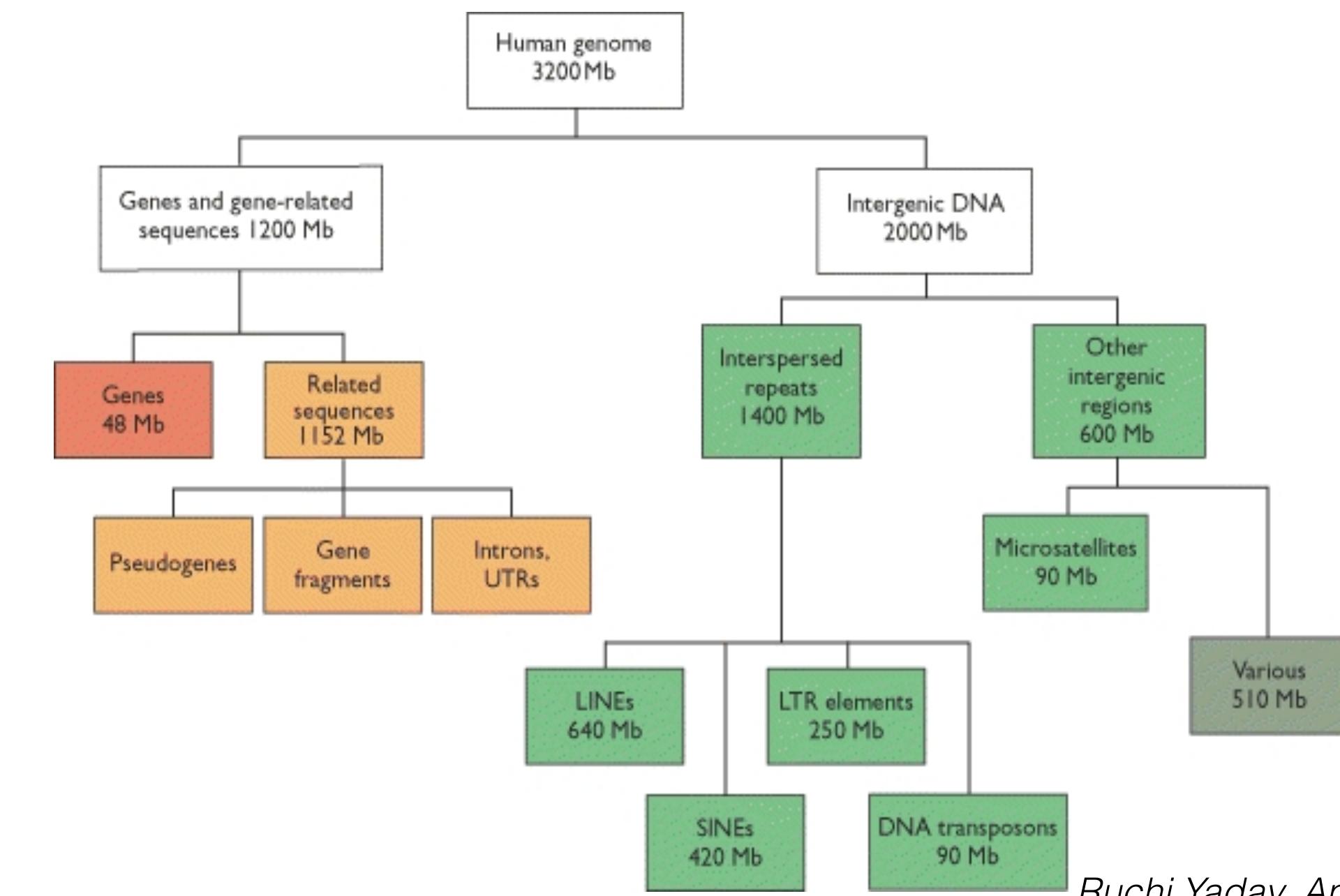
Figure: 16S vs. shotgun sequencing.

### Adapter

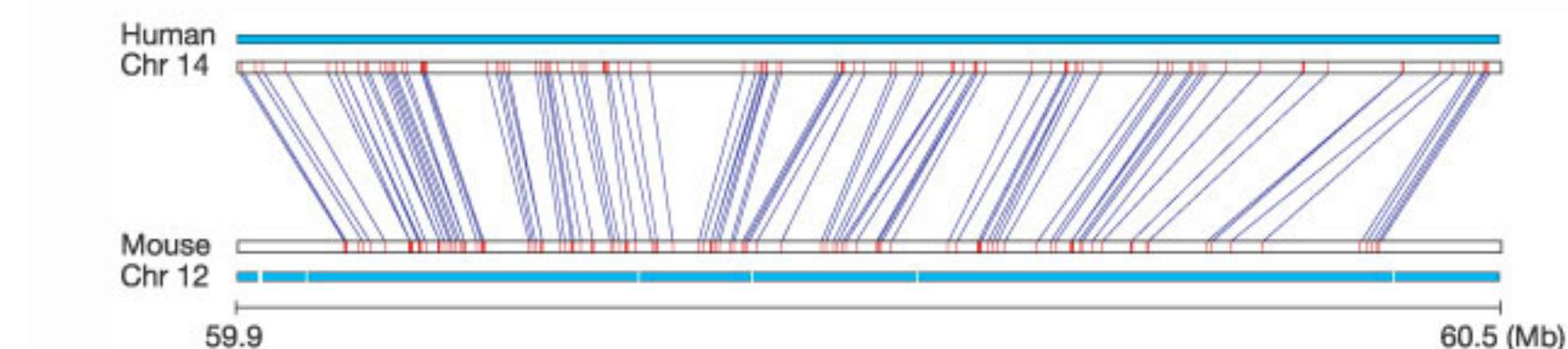
Most NGS instruments require that dsDNA of known sequence be added to the 2 ends of **library** molecules that will be sequenced on the instrument. Adapters can be added in a variety of ways to starting nucleic acid molecules during **library**

# Genomics is the study of any of a number of attributes of genome or genomes

- Genome:
  - size
  - sequence
  - structure / variation
  - evolution
- Gene:
  - structure
  - expression
- Comparative genomics
- Epigenomics
- Metagenomics
- Transcriptomics
- Other -omics: Proteomics/Metabolomics

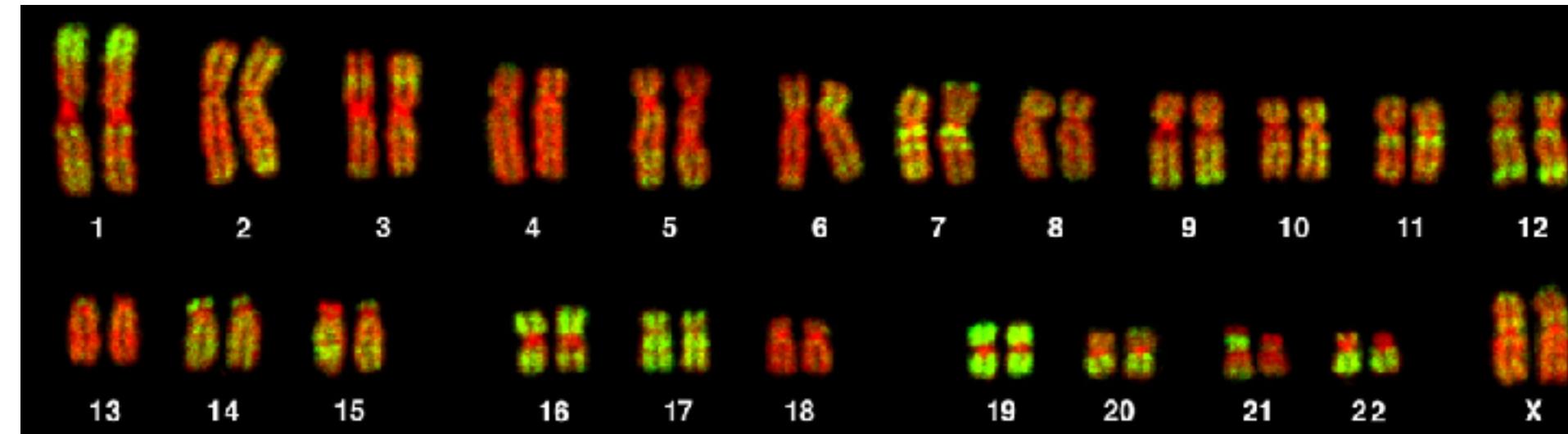


Ruchi Yadav, Amity University



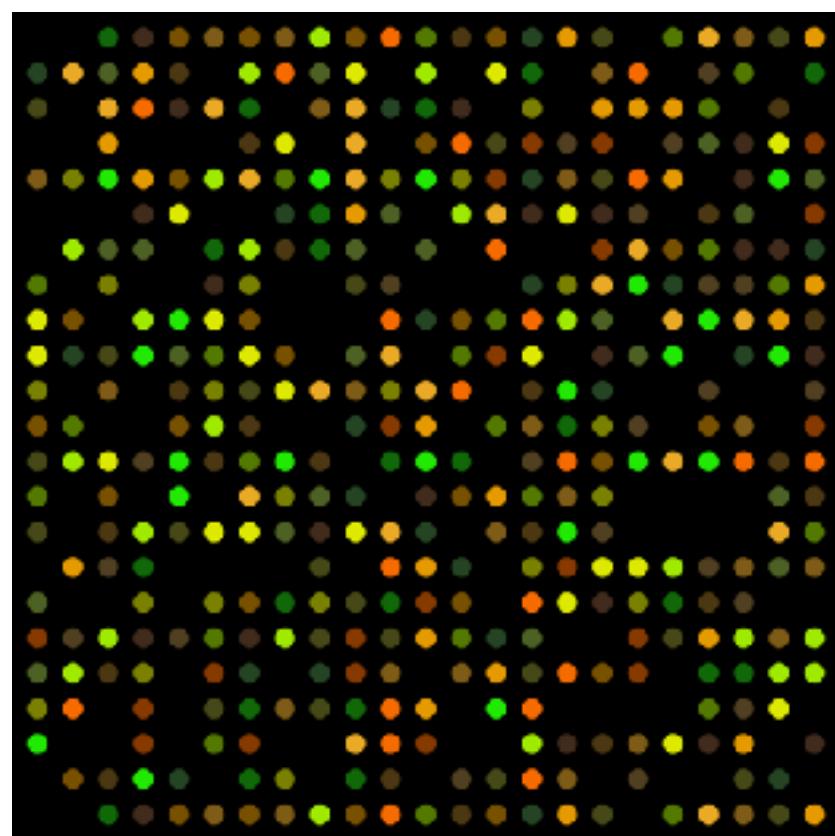
Nature (2002) Mouse Genome

# Genomics isn't the same thing as sequencing, but they're increasingly related

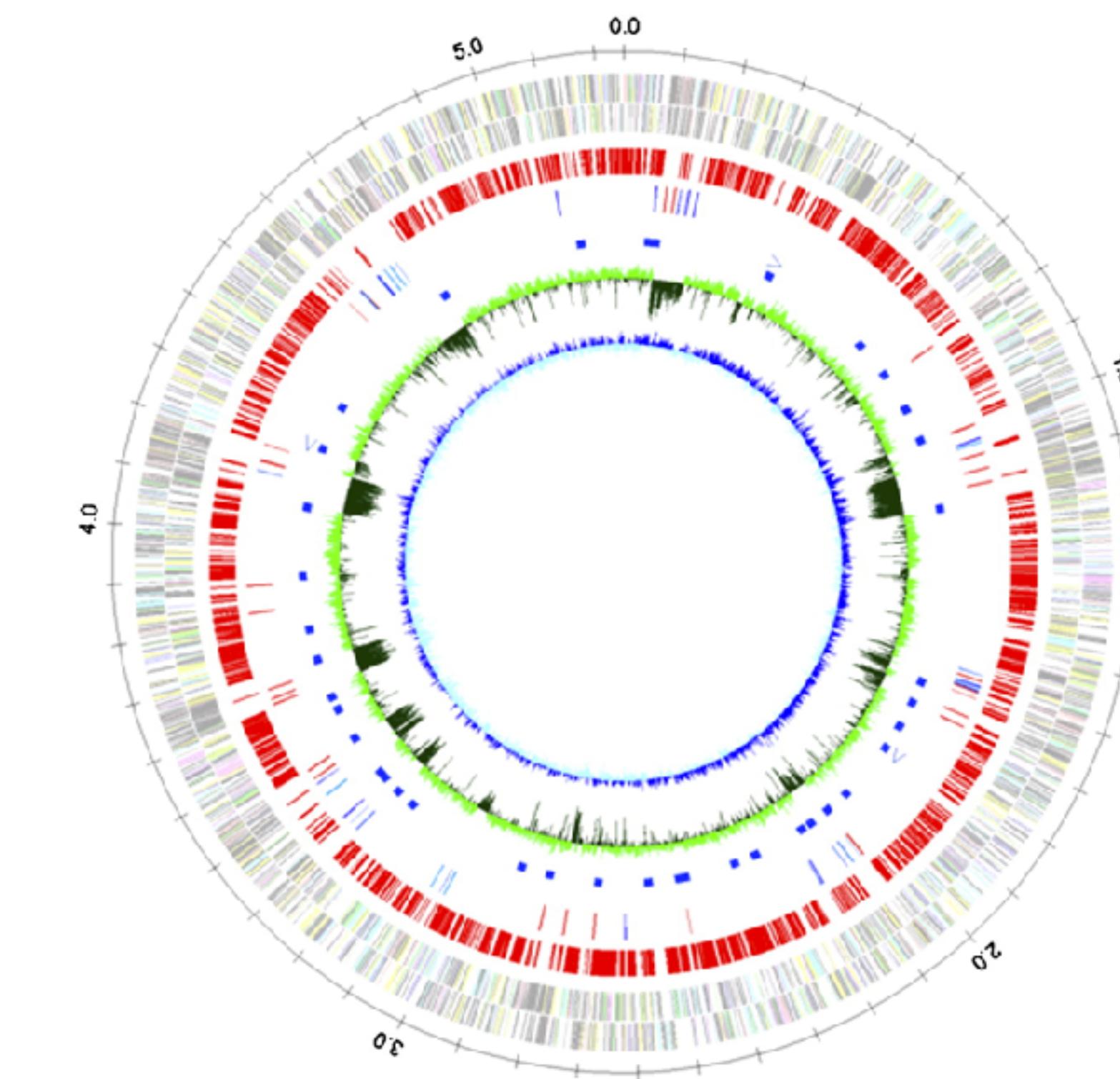


Bolzer et al (2005) PLoS Biol

Microarray

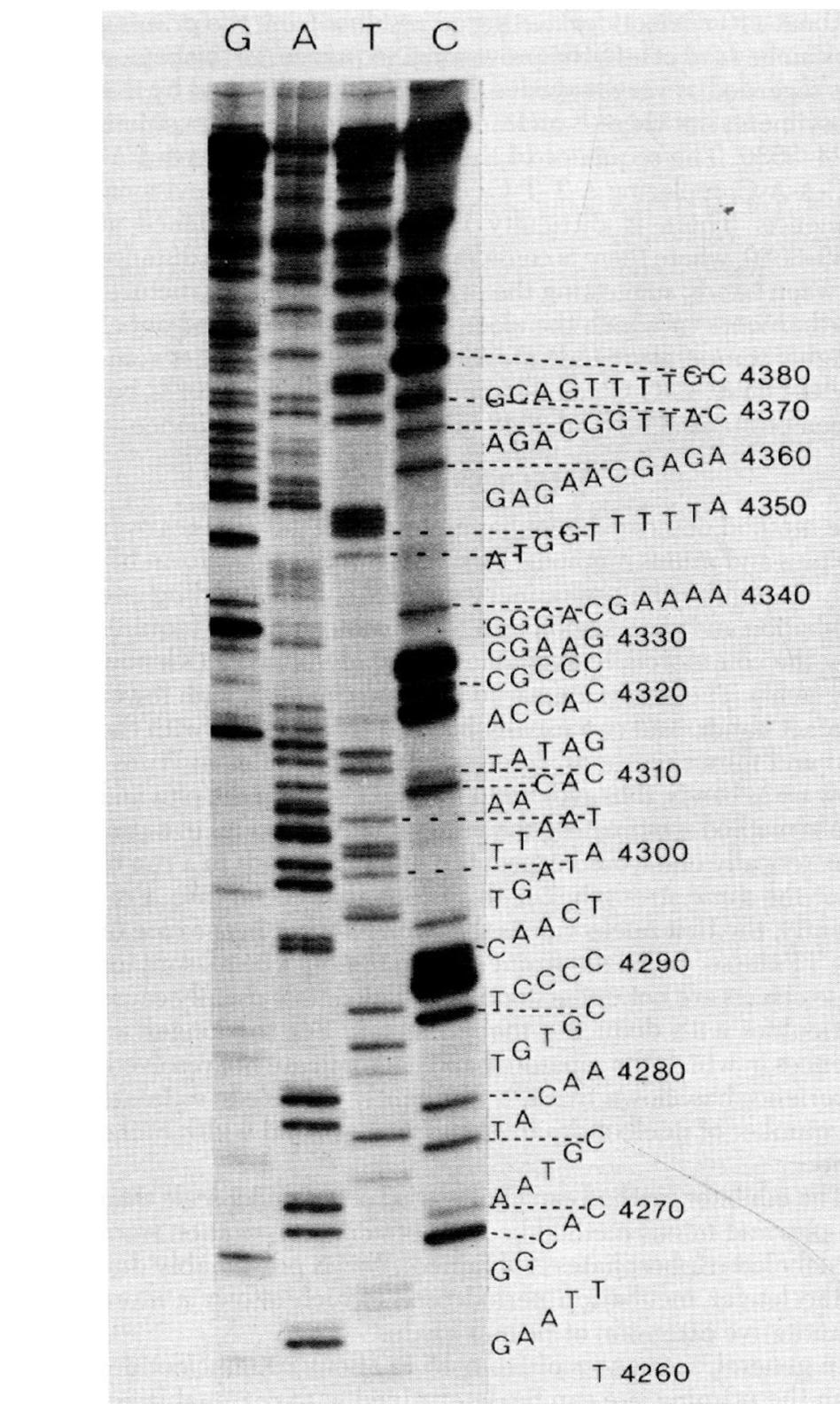
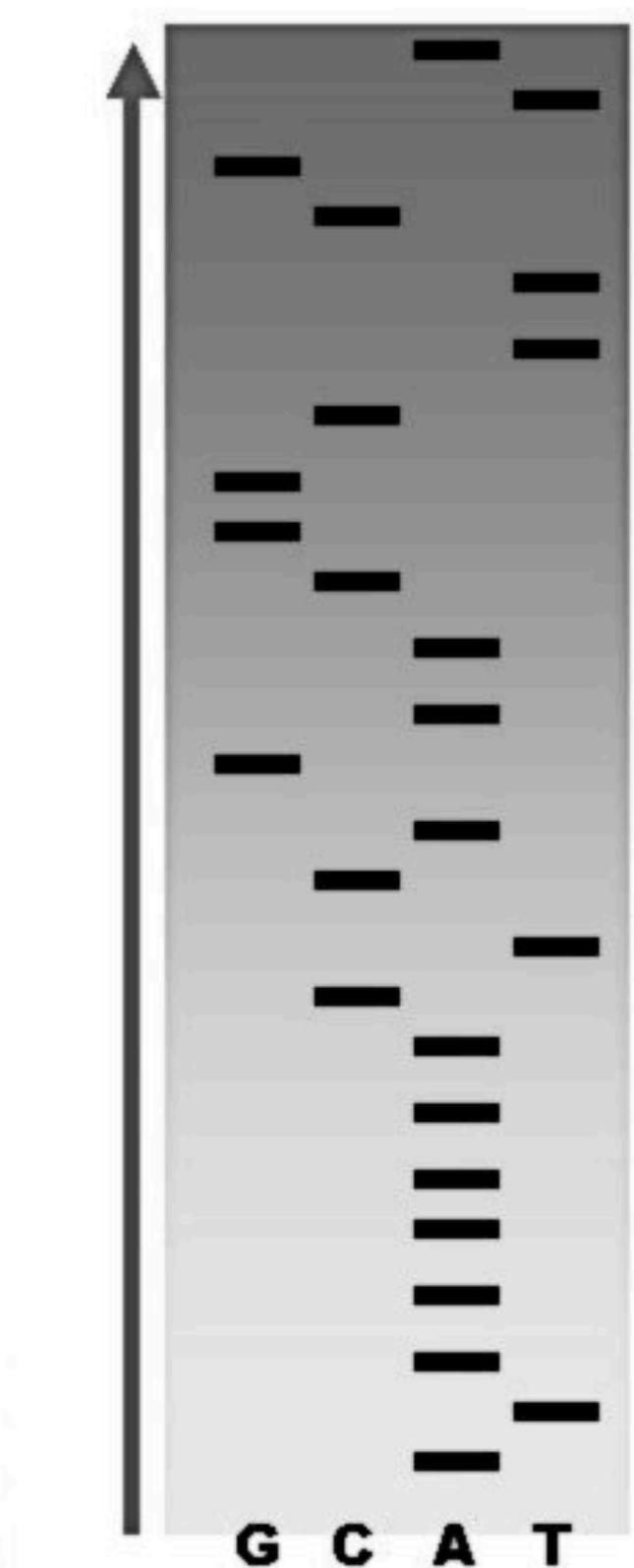
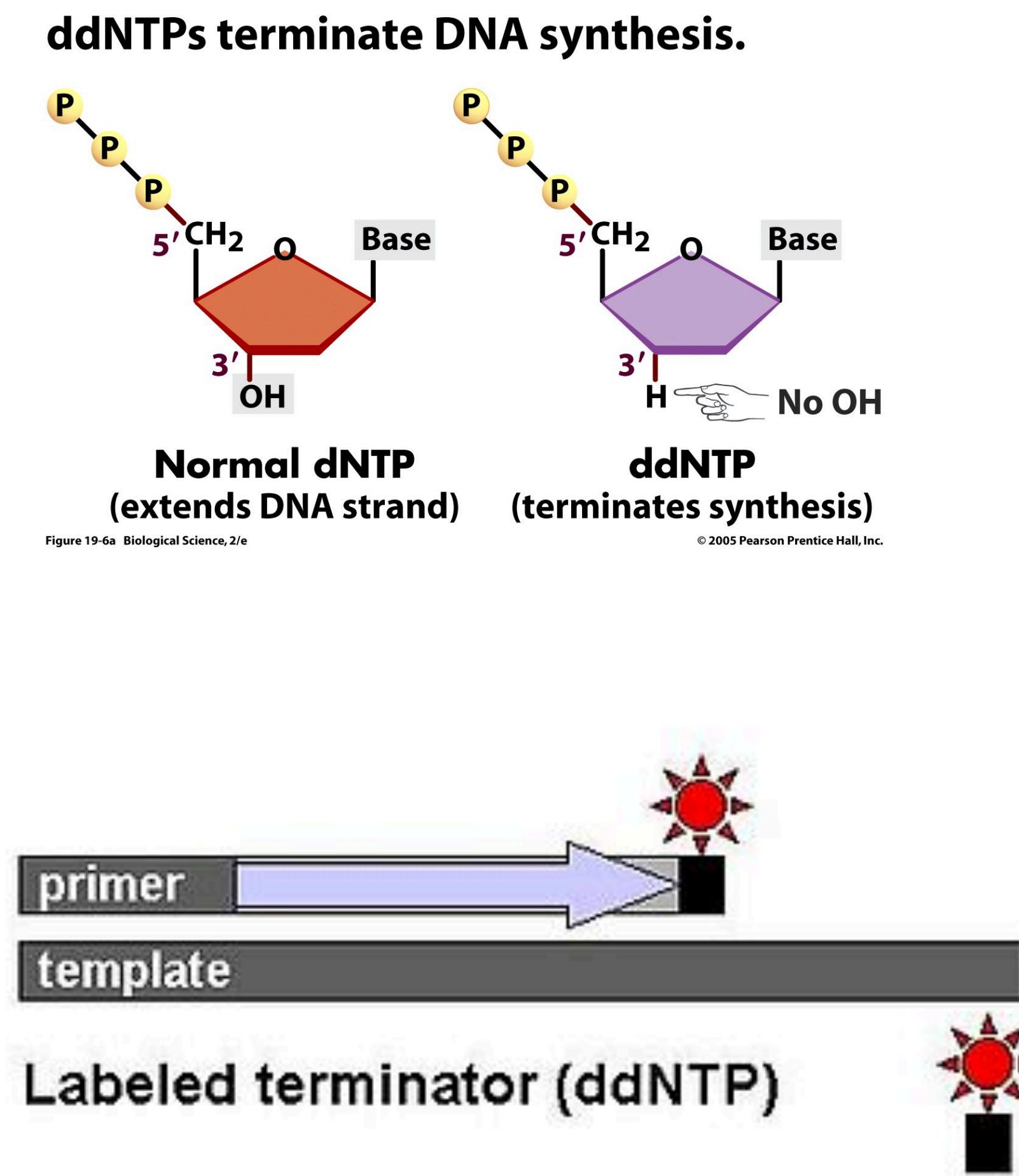


Wikimedia commons



Nakazawa et al (2009) Genome Research

# Sanger Sequencing (1977): sequencing 1 target at a time



Slide courtesy Dan Sloan. Image credits: Sanger et al (1977) and Wikipedia

Next generation sequencing (NGS) ~ deep sequencing ~ high throughput sequencing (HTS)

All simultaneously sequence many molecules in parallel

## Short read sequencing

- Millions of reads
- Relatively short: ~50-300 nt (Illumina)
- Relative low error rates
- Illumina has virtually all of the market share



MiSeq

## Long read sequencing

- Fewer, longer reads
- >1 kb (PacBio), up to 100s of kb (Oxford Nanopore)
- Relative high error rates

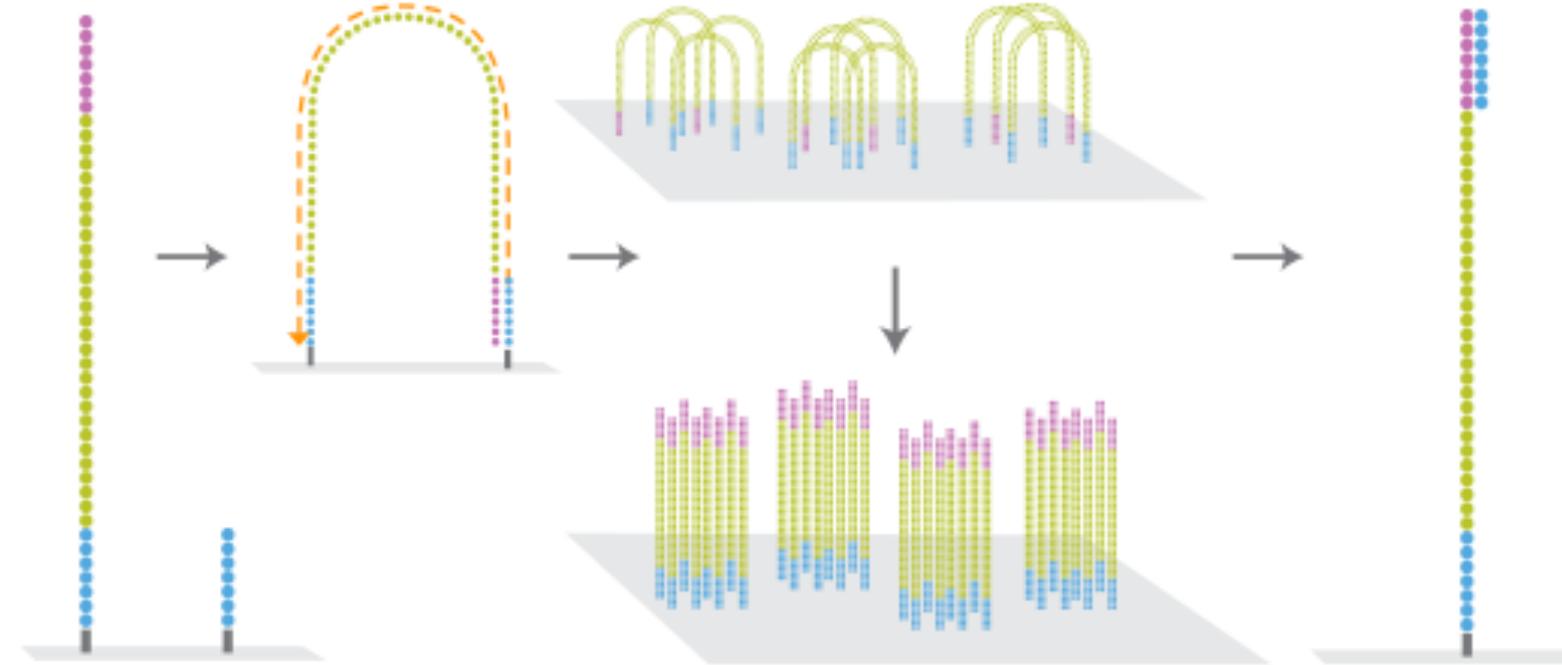
Oxford Nanopore MinION



PacBio RS-II

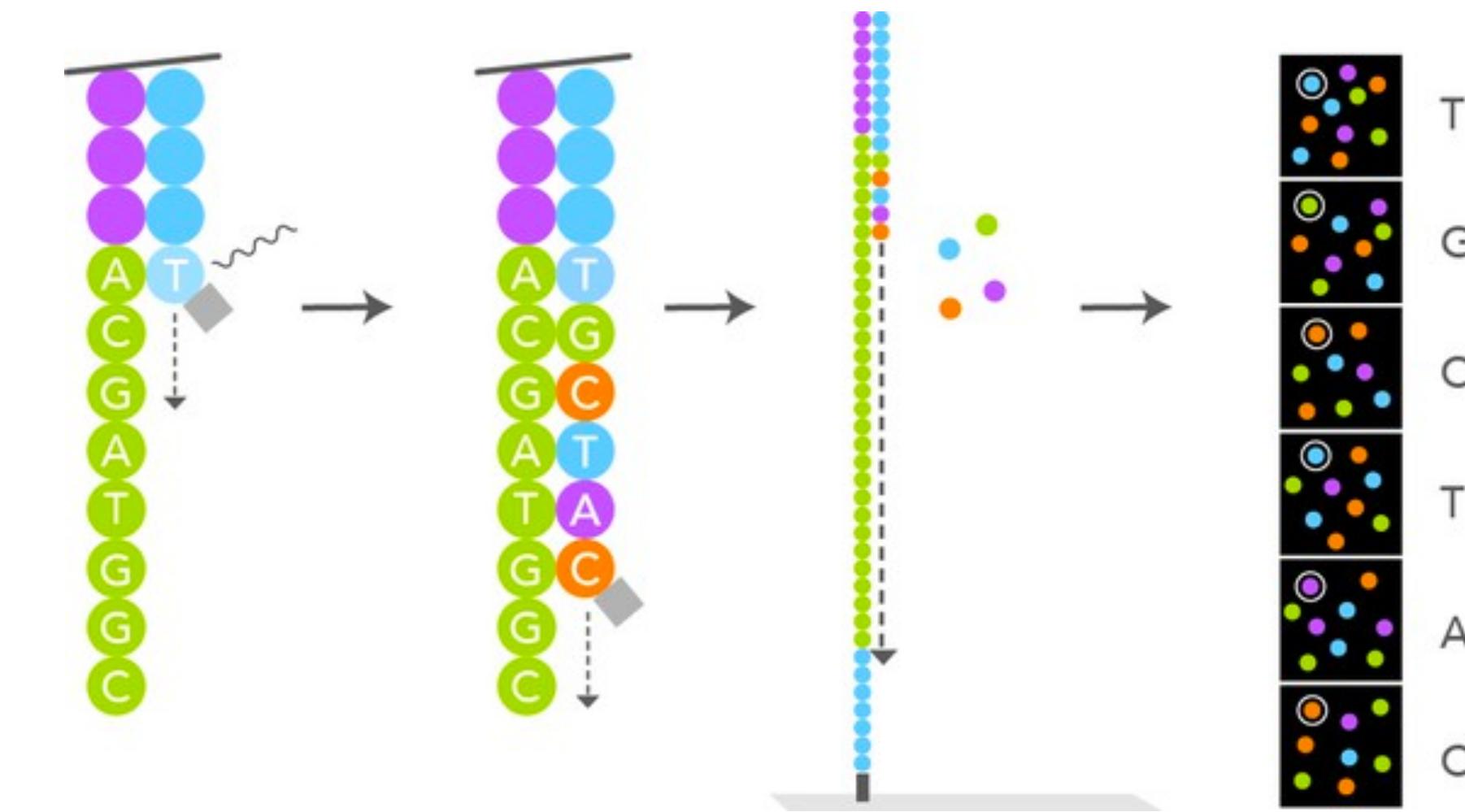


# Illumina instruments use sequencing by synthesis (SBS)



Millions of clusters per flow cell

Each cluster contains 1000s of clonal copies of a library molecule



Library molecules are sequenced by primer extension reactions that incorporate chain-terminated, fluorescent nucleotides

real raw Illumina sequencing data

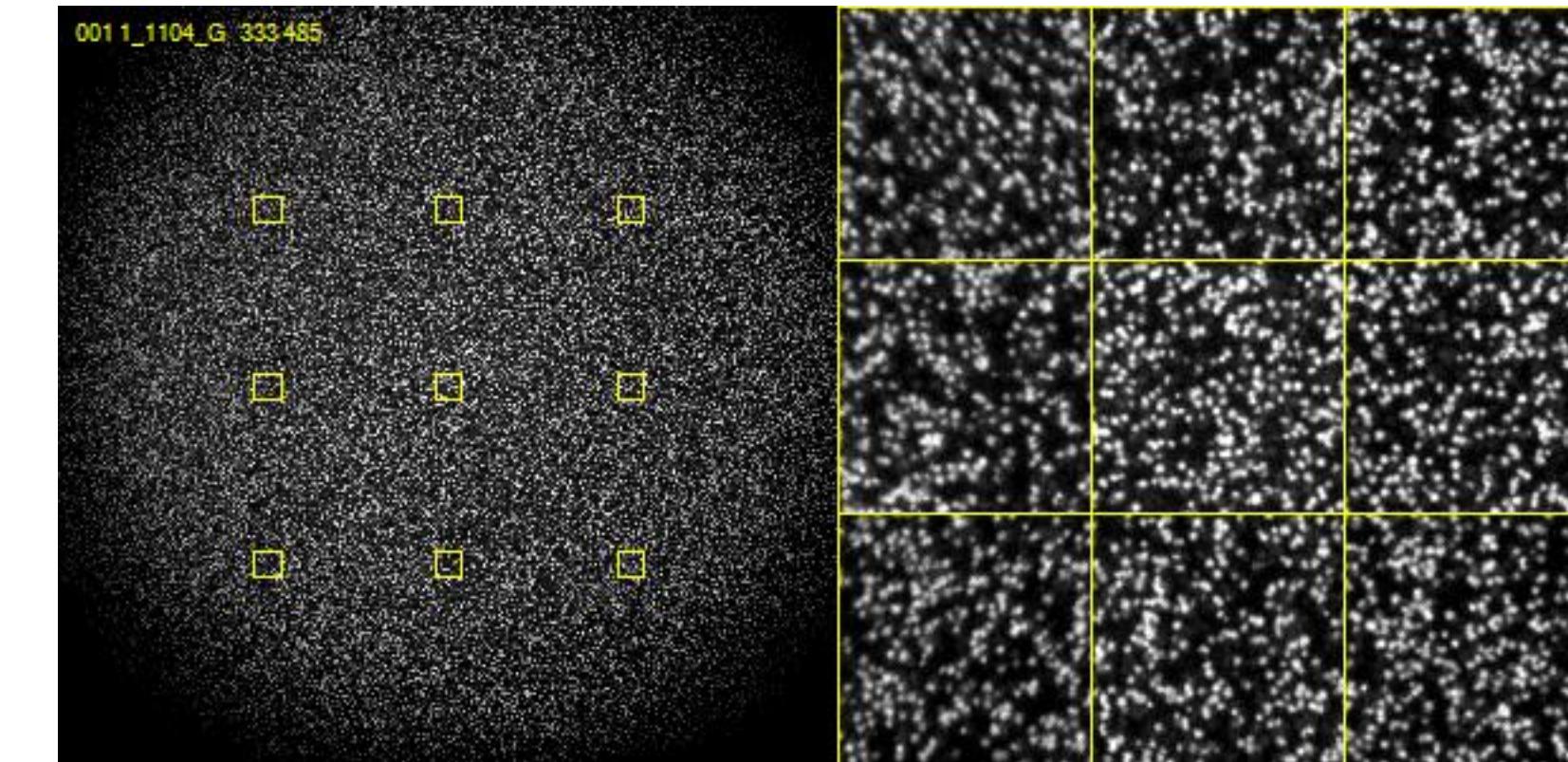
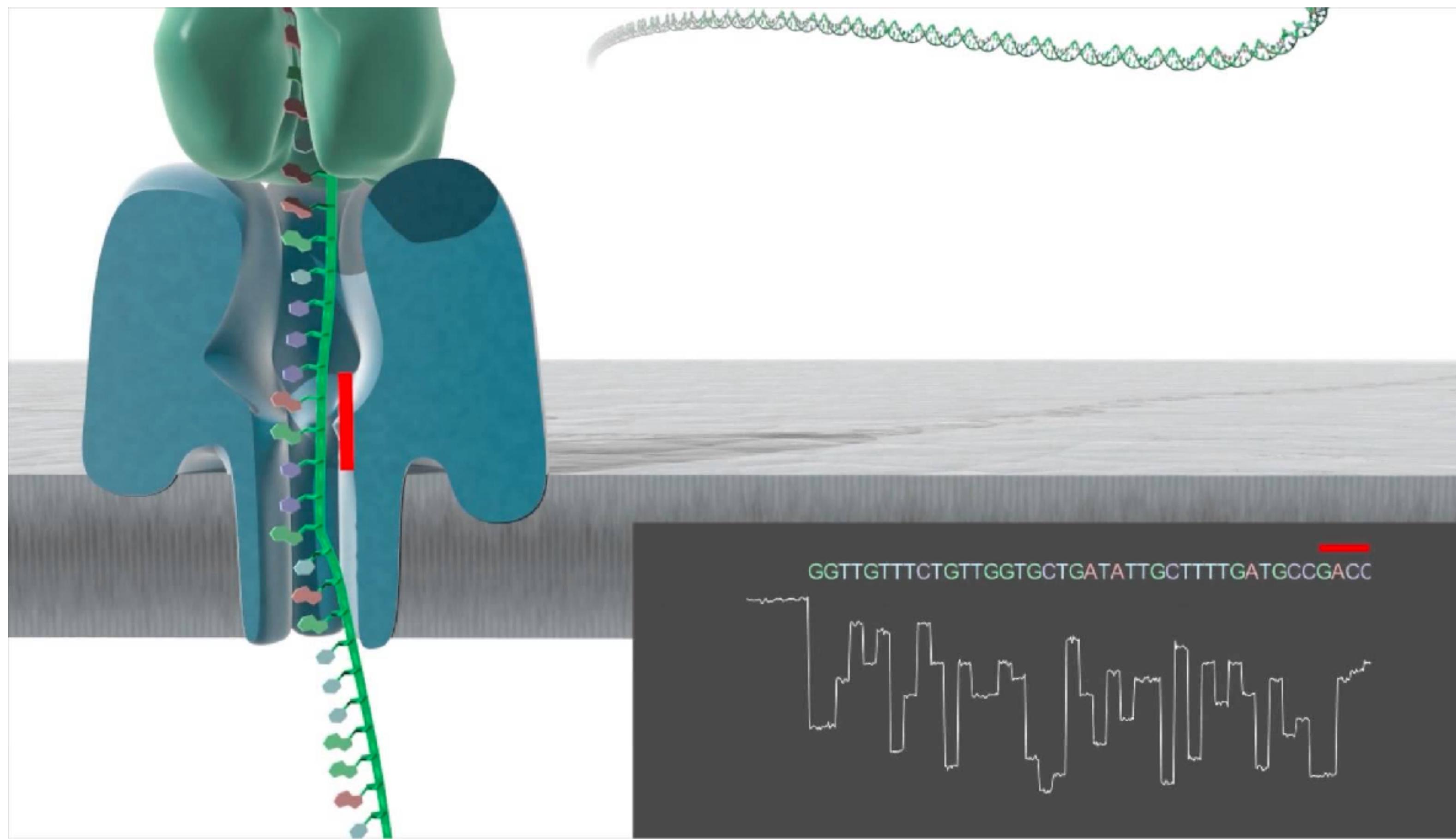


Image credit: Illumina

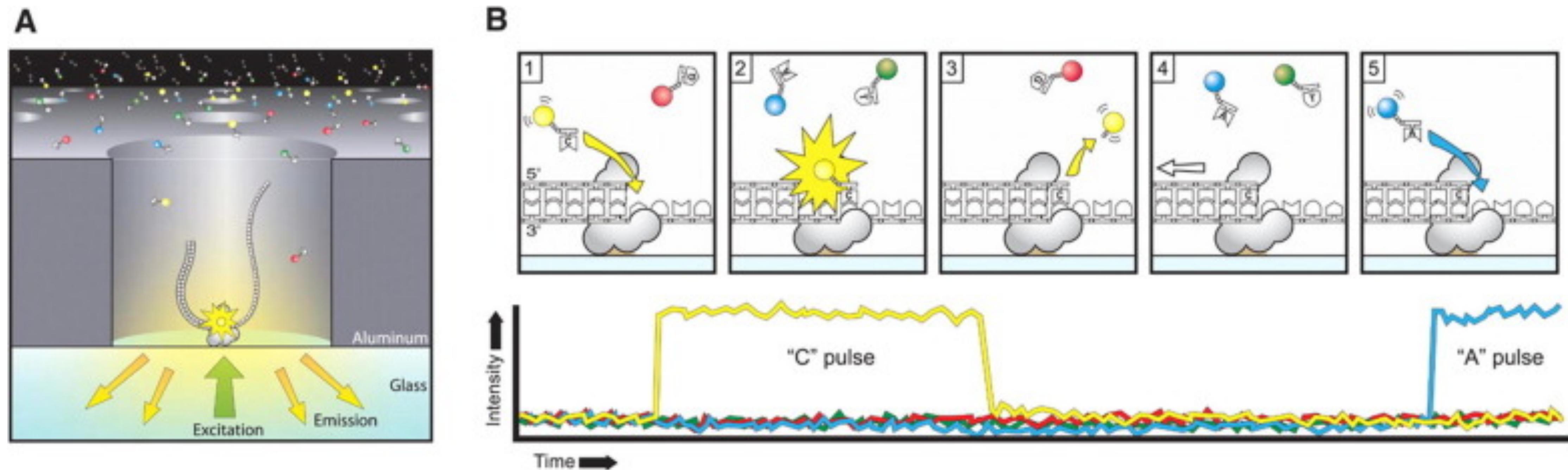
# Long read sequencers sequence single molecules



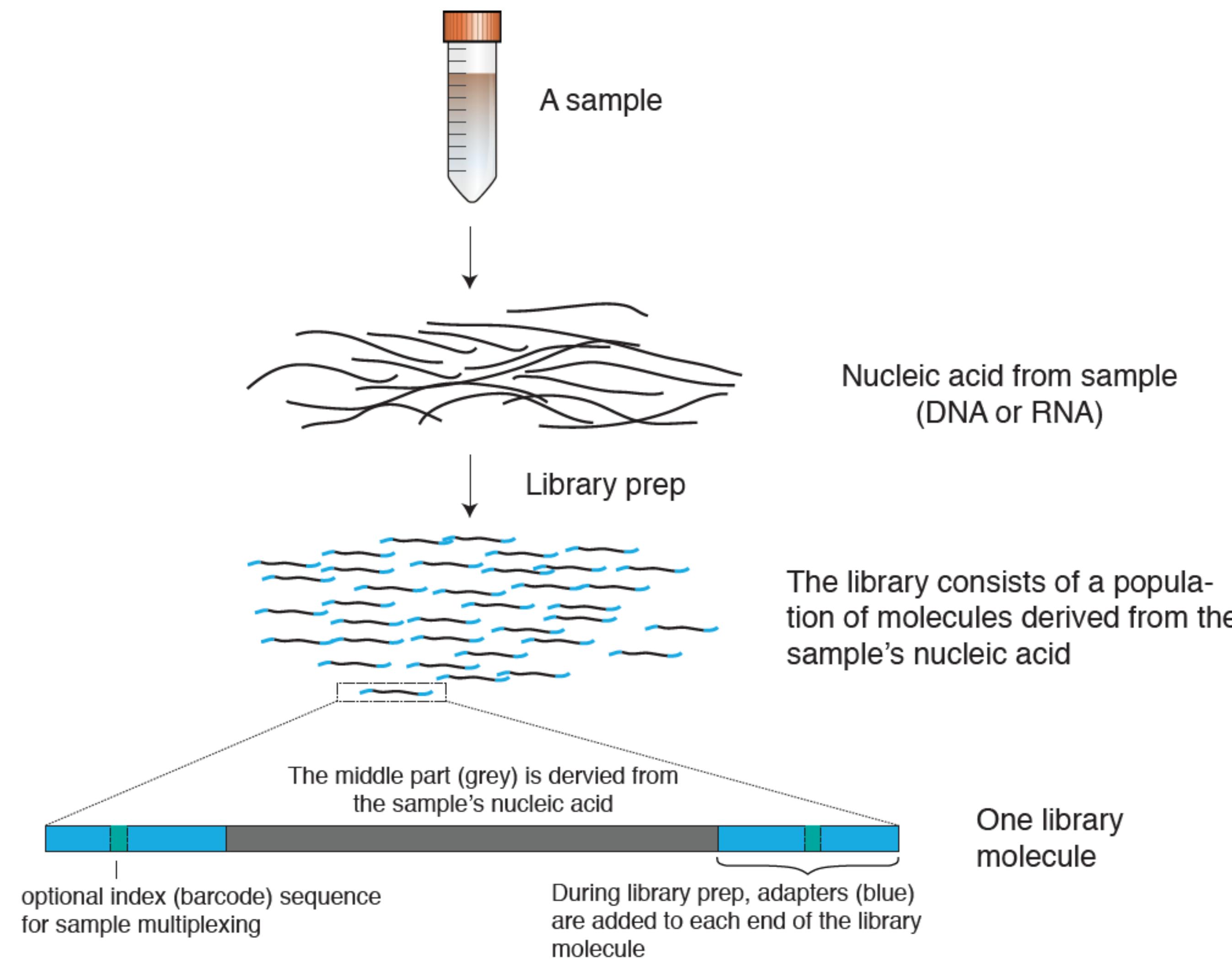
Nanopore sequencing: Much longer reads, but with much higher error rates

*Image: Oxford Nanopore*

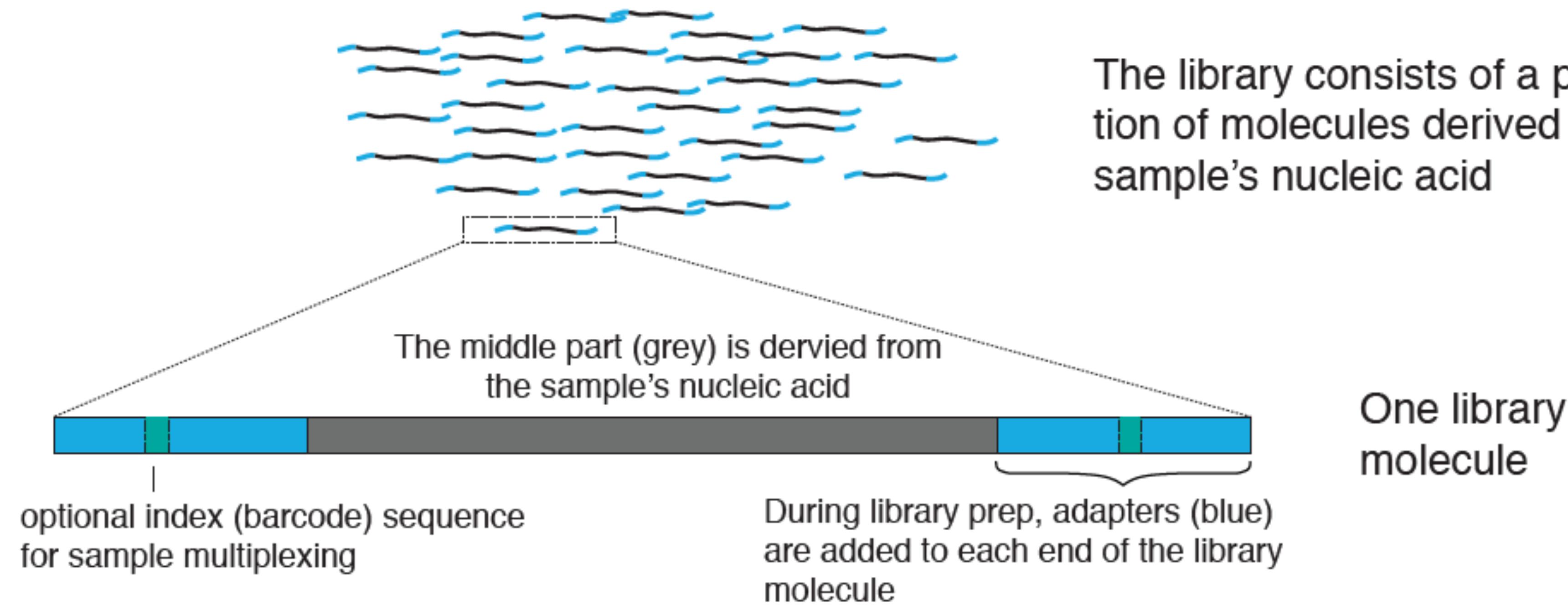
# PacBio (Illumina) SMRT sequencing is the other main long-read technology



# Library prep converts nucleic acids into a form suitable to be sequenced

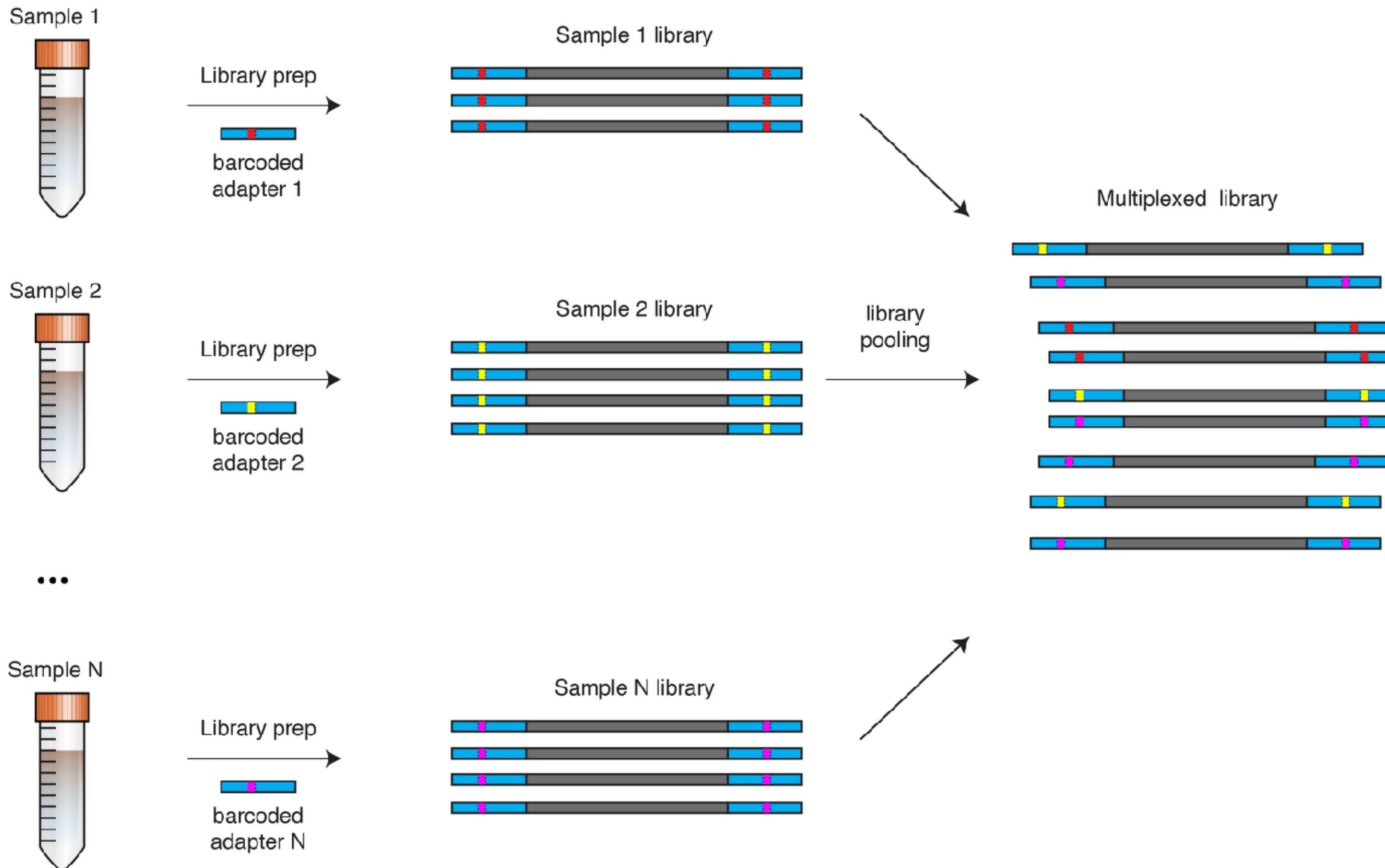


# Library prep converts nucleic acids into a form suitable to be sequenced



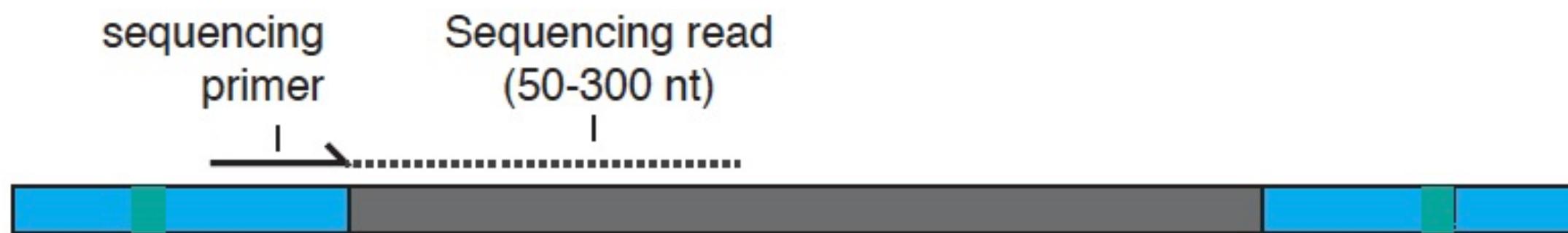
An example Illumina library molecule

# Barcodes (or indexes) allow sample multiplexing



# Illumina sequencing produces 1-4 reads per library molecule

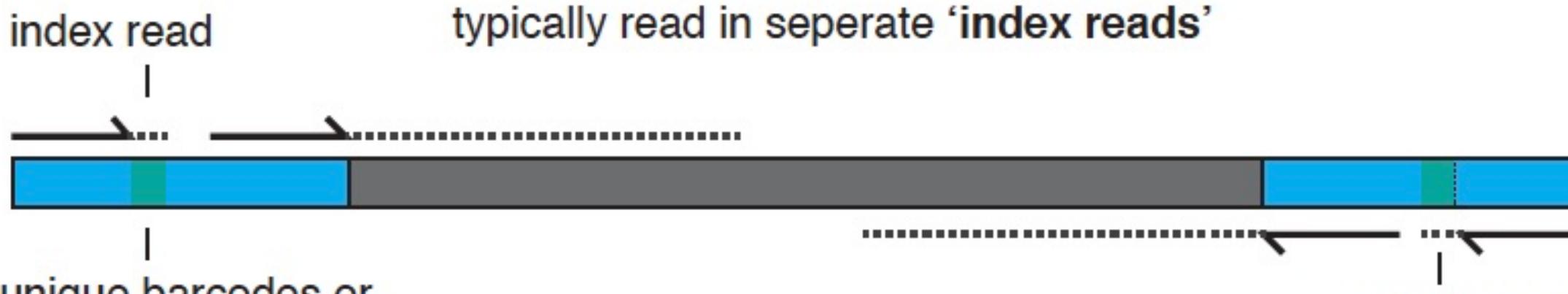
In **single end sequencing**, a library molecule is sequenced from one end



In **paired end sequencing**, a library molecule is sequenced from both ends



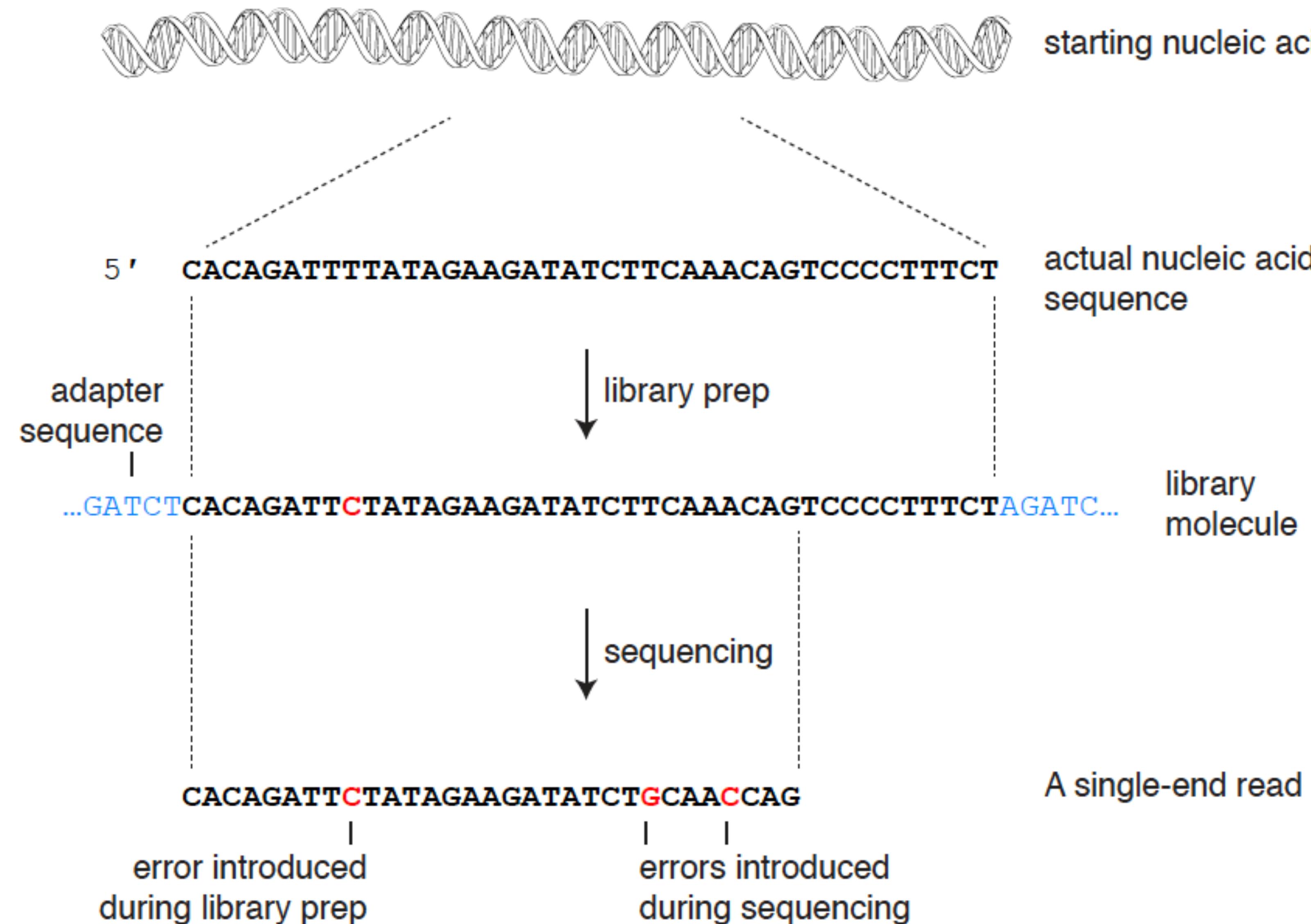
The library molecule's barcodes (indexes) are typically read in separate 'index reads'



unique barcodes or barcode pairs can be used to differentiate multiplexed samples

index read

Reads are sub-sequences of the starting nucleic acid that often contain errors

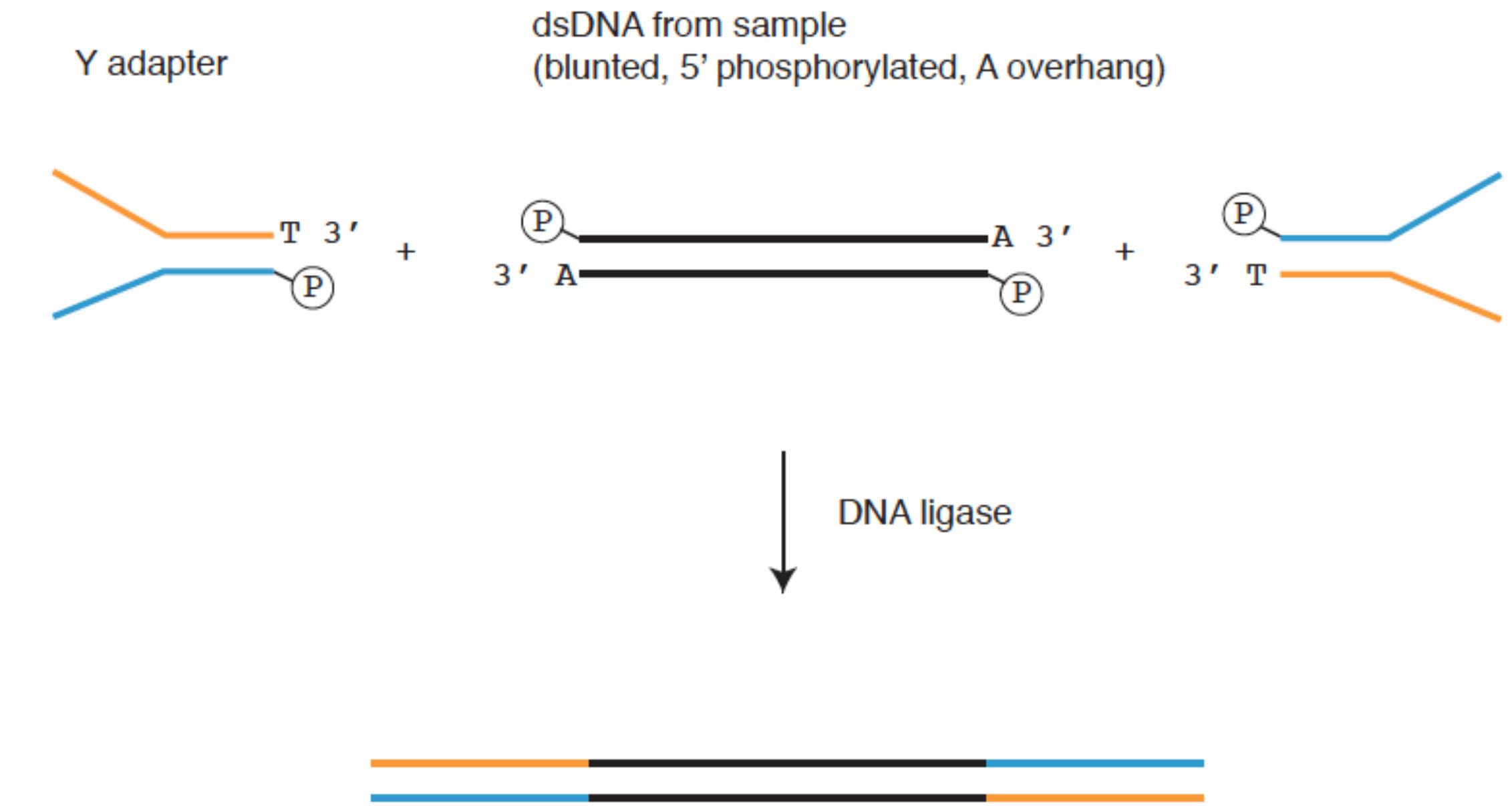


# There are many good ways to make sequencing libraries

## Common Illumina library prep steps (not always included and not always in this order)

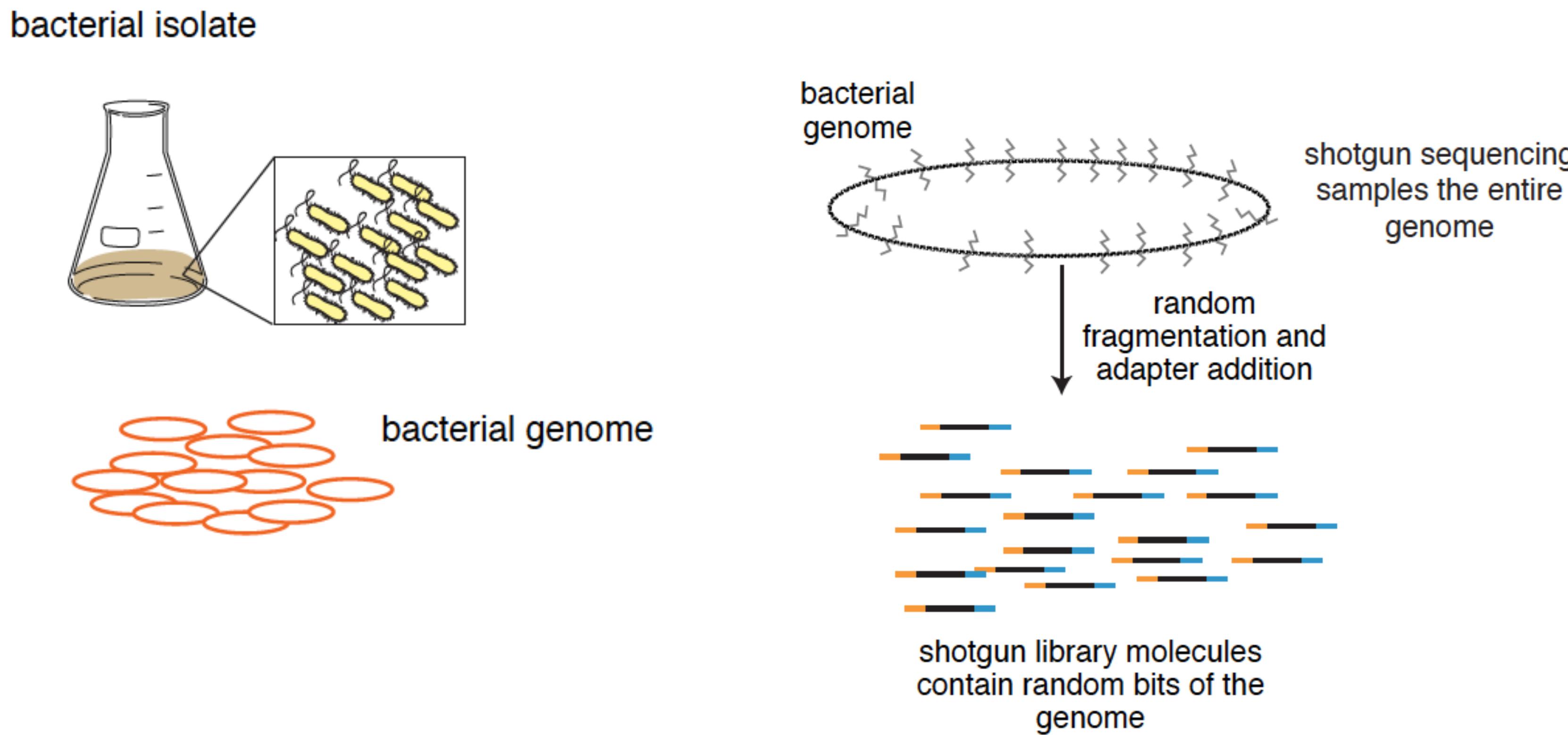
- Nucleic acid isolation
- Enrichment (of nucleic acid subtypes you want) or subtraction (of those you don't want)
- Nucleic acid fragmentation
- Conversion of RNA into dsDNA (for RNA sequencing)
- Addition of adapters to ends of library molecules, possibly with barcodes for multiplexing
- Library amplification
- Pooling of multiplexed samples
- Library QC / quantification

Adapters can be added to sample-derived dsDNA by ligation



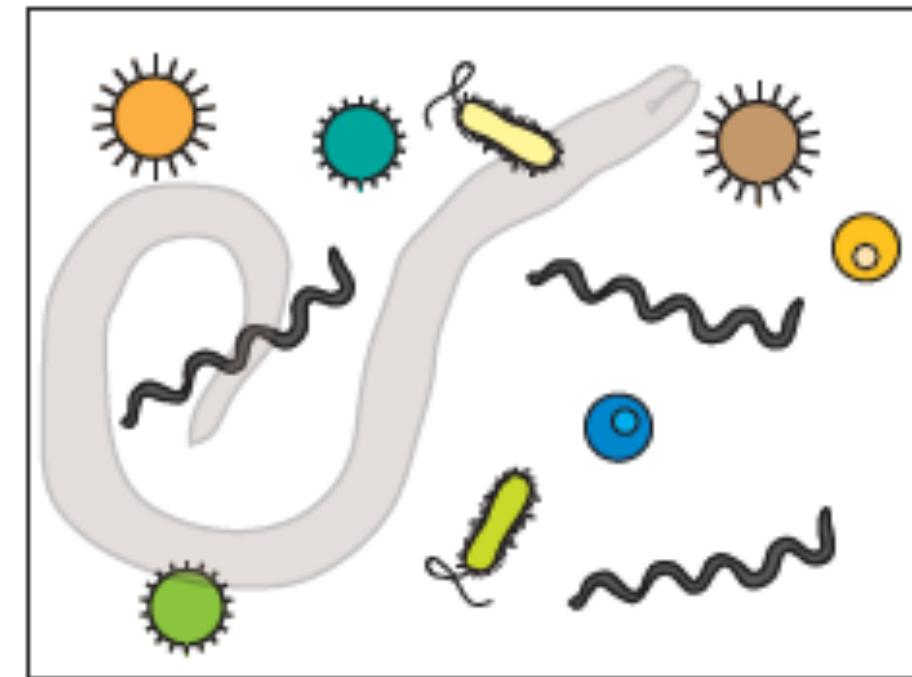
# How you make a library determines what type of sequencing you're doing

For instance, if you make a 'shotgun library' from a single organism, you're doing whole genome sequencing (WGS)



# **Metagenomic** sequencing involves sequencing of genomes from more than one organism

soil community



Could make a 16S or a shotgun library from these genomes

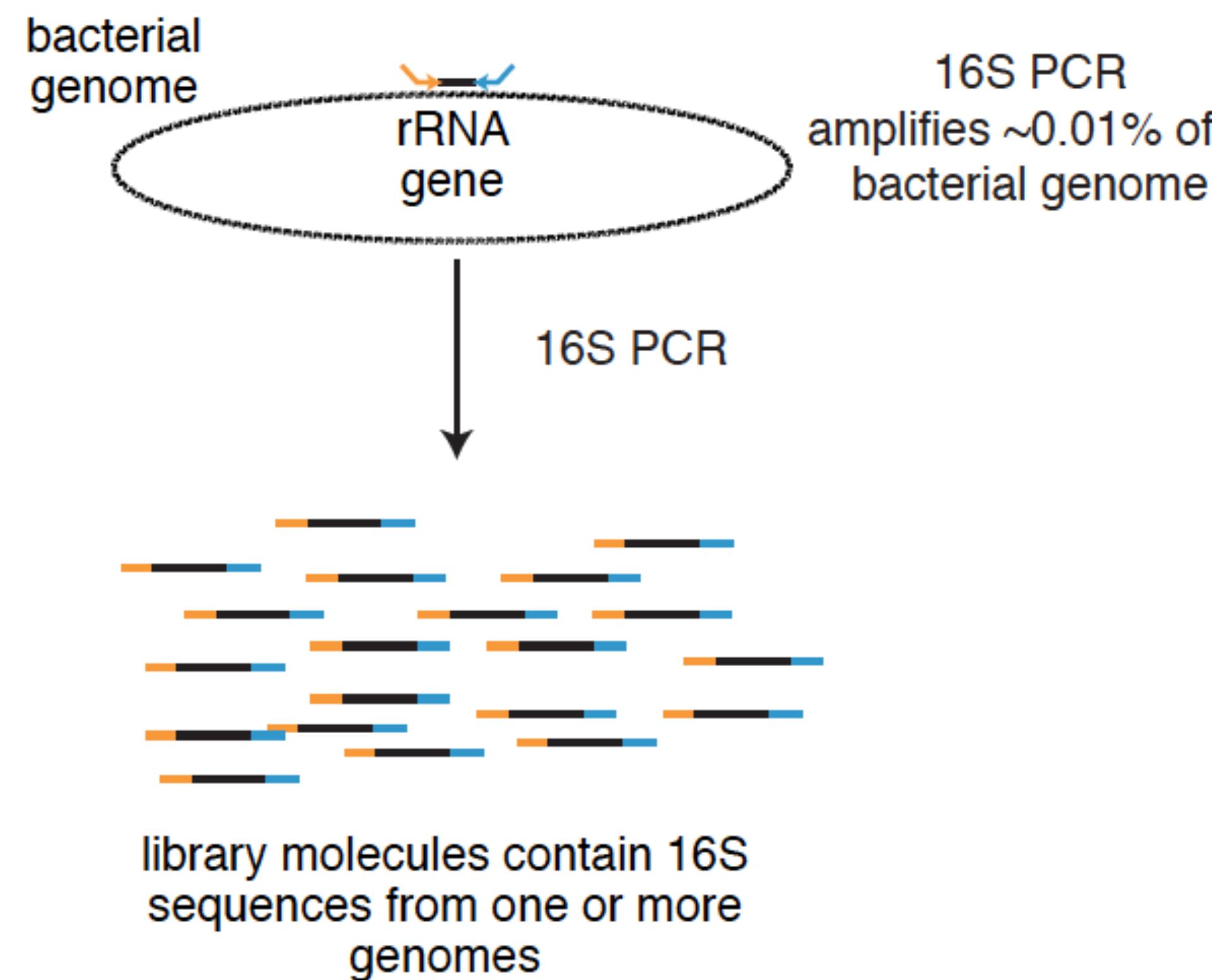
Sequencing of RNAs from a complex sample like this is metatranscriptomics

soil 'metagenome'

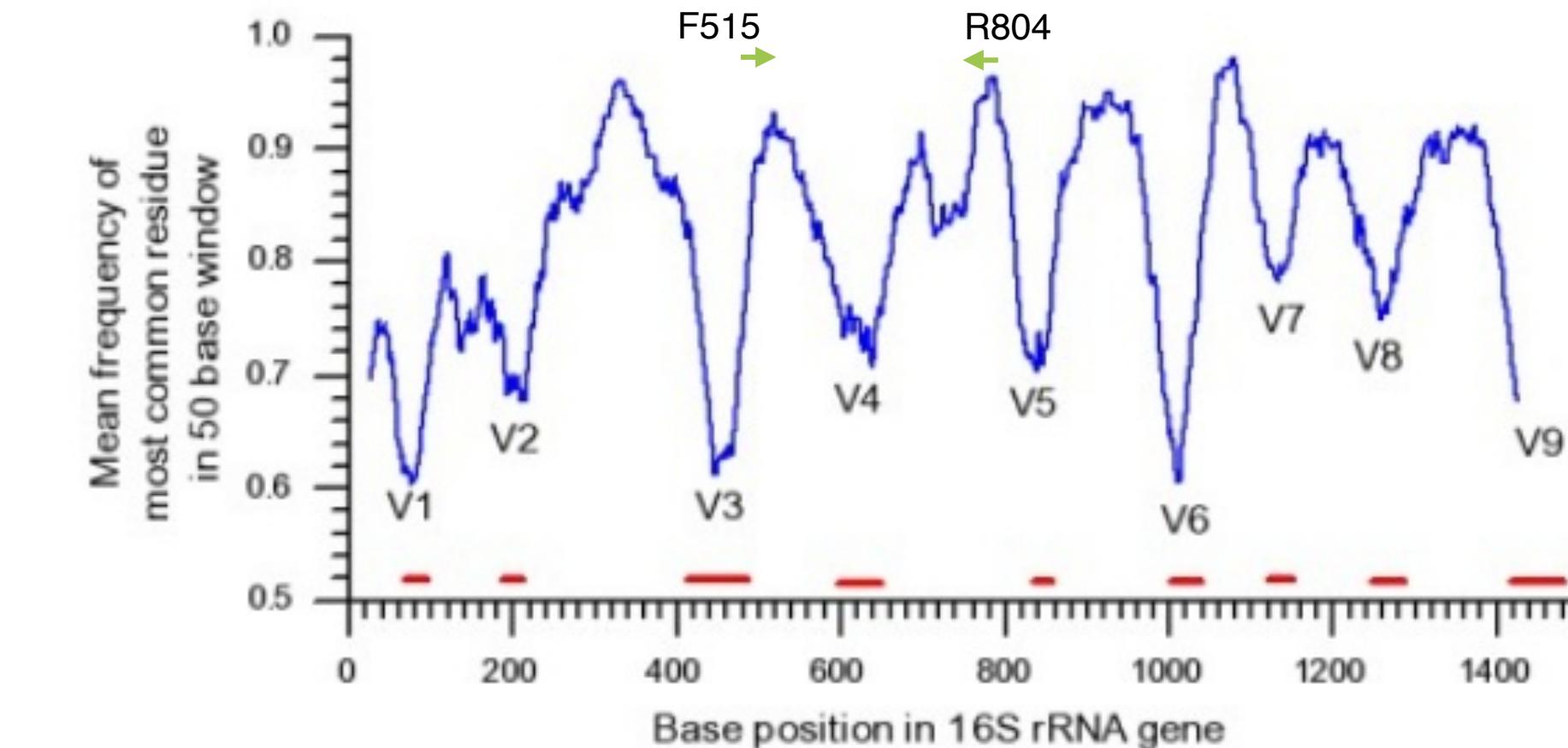


# Microbiome sequencing often means **16S rRNA** sequencing

16S sequencing is one type of  
'amplicon sequencing'

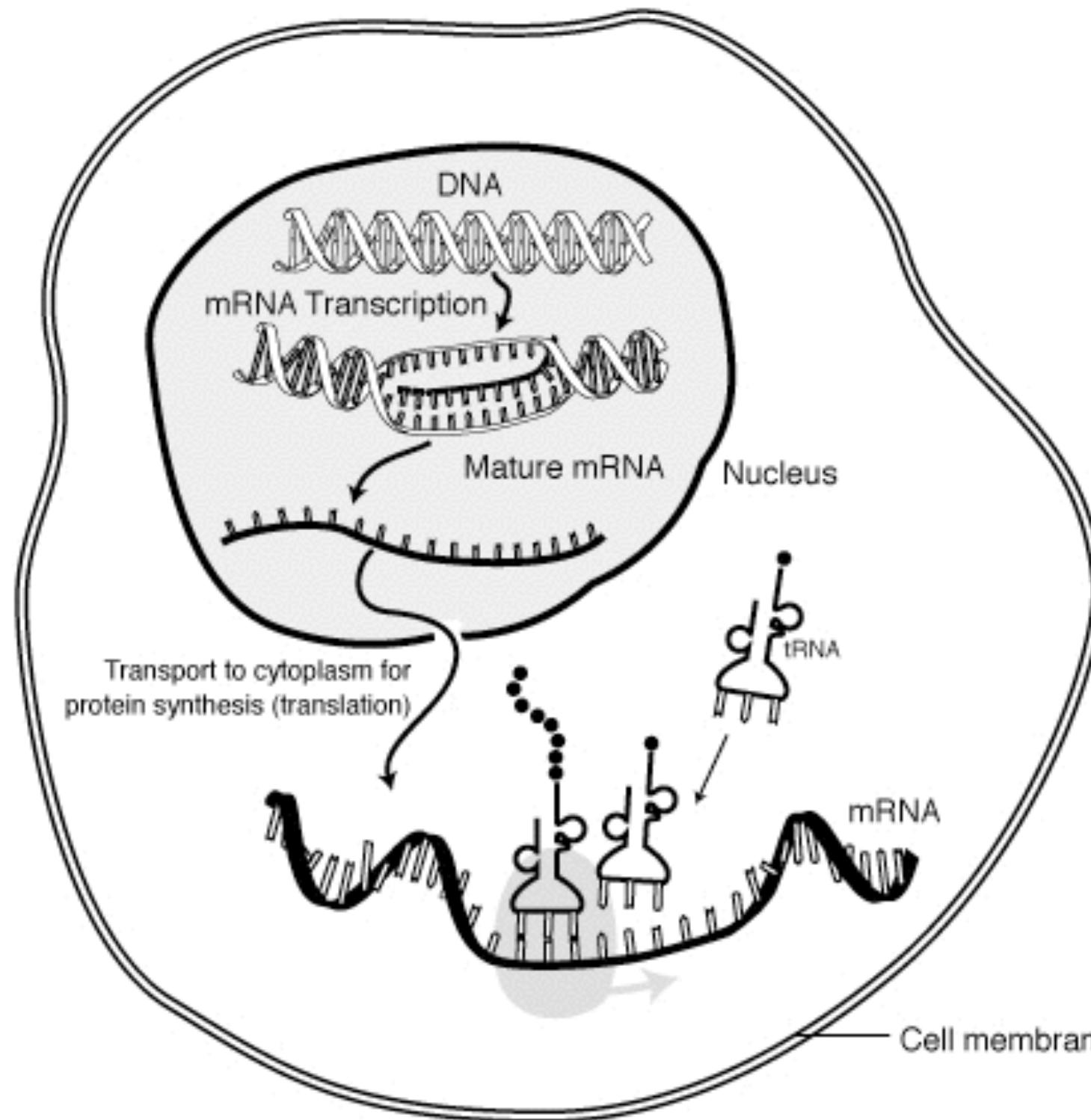


16S rRNA genes have highly conserved regions flanking variable regions



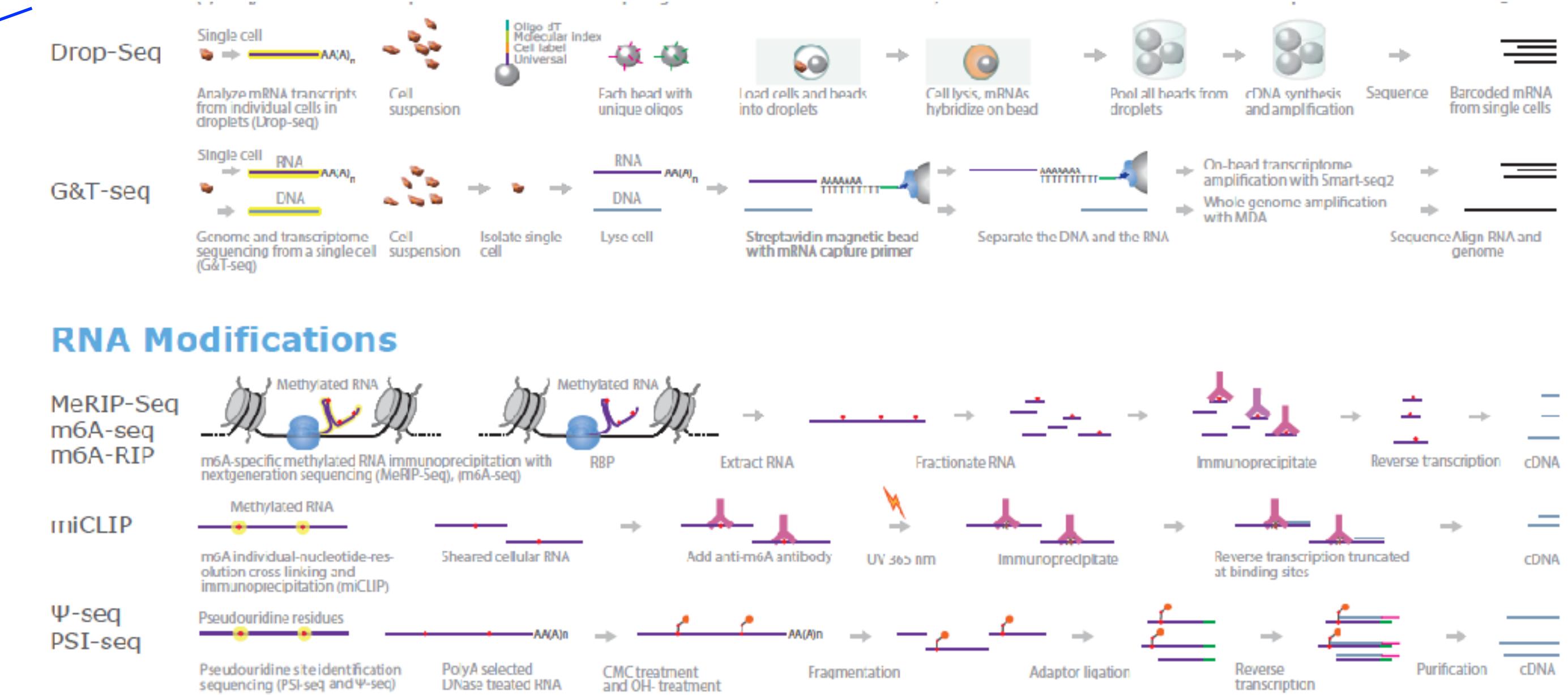
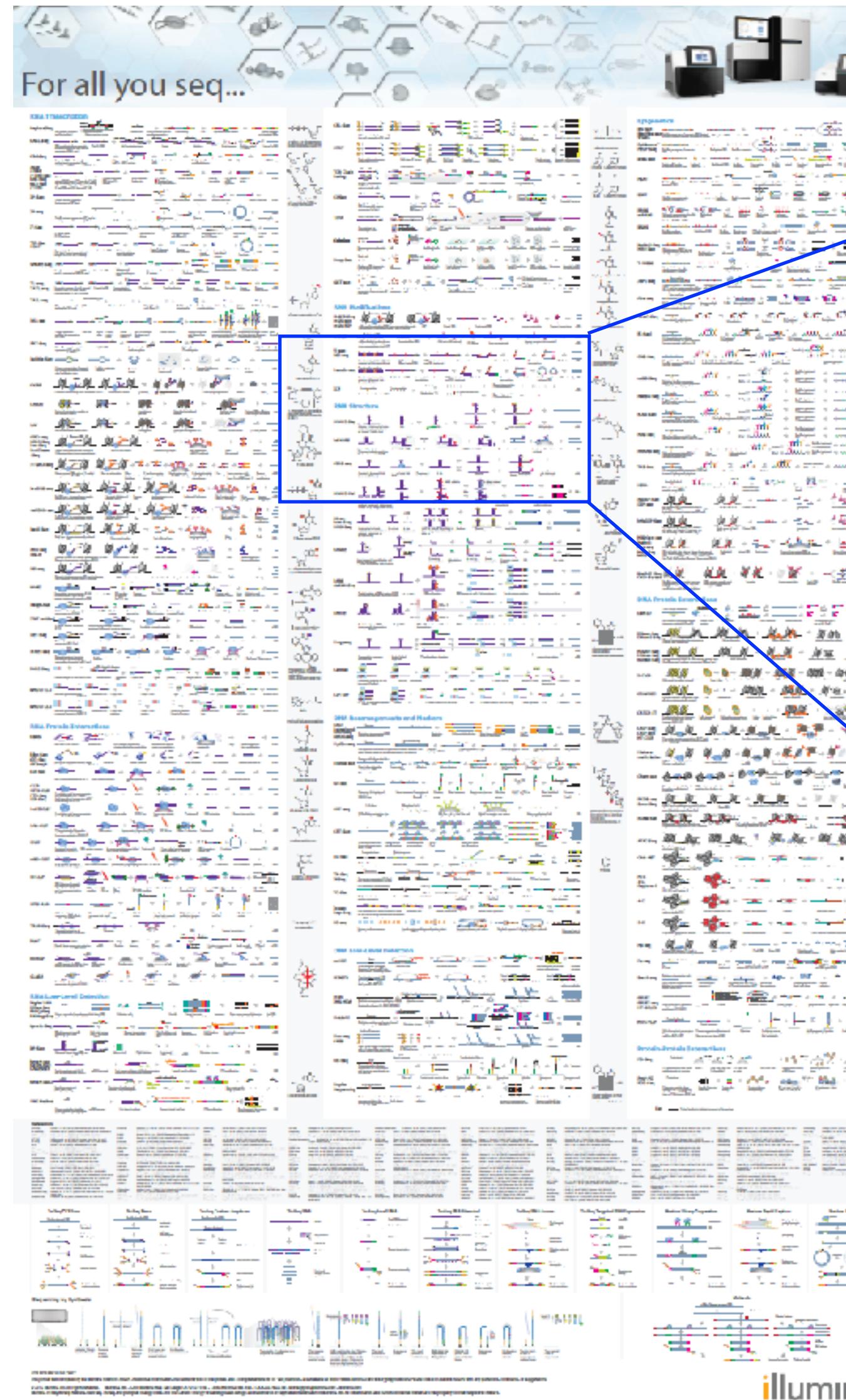
# How you make a library determines what type of sequencing you're doing

If you make a library from mRNA, that is RNA-Seq (transcriptome sequencing)



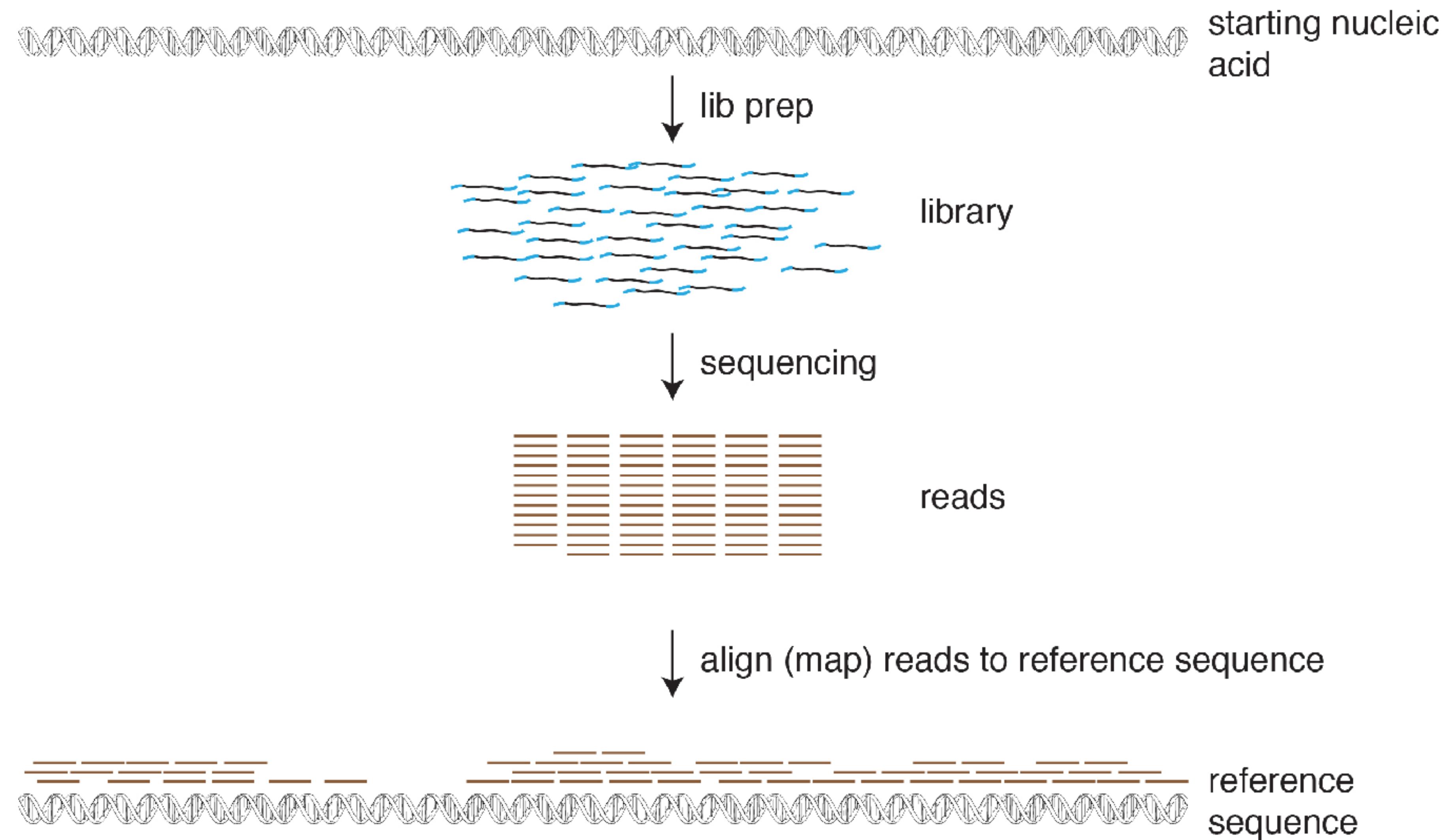
The abundance of reads from a particular mRNA is proportional to that mRNA's abundance in the cell

There are **5 bajillion** ways to make libraries and to do sequencing (all have names that end in -seq)

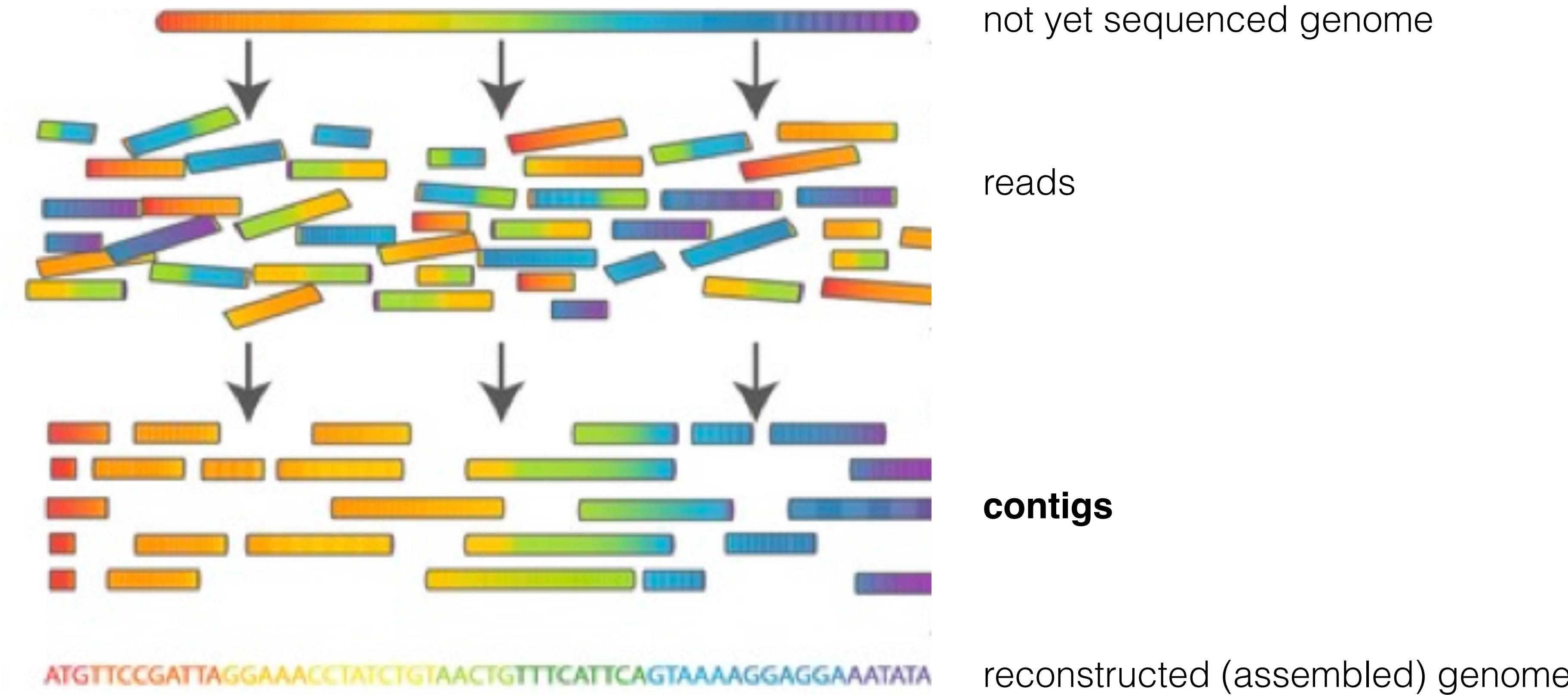


They're all variations on a few themes. Don't let it overwhelm you. Most sequencing is of a few simple types, and it's better to focus on the Biology and experimental design.

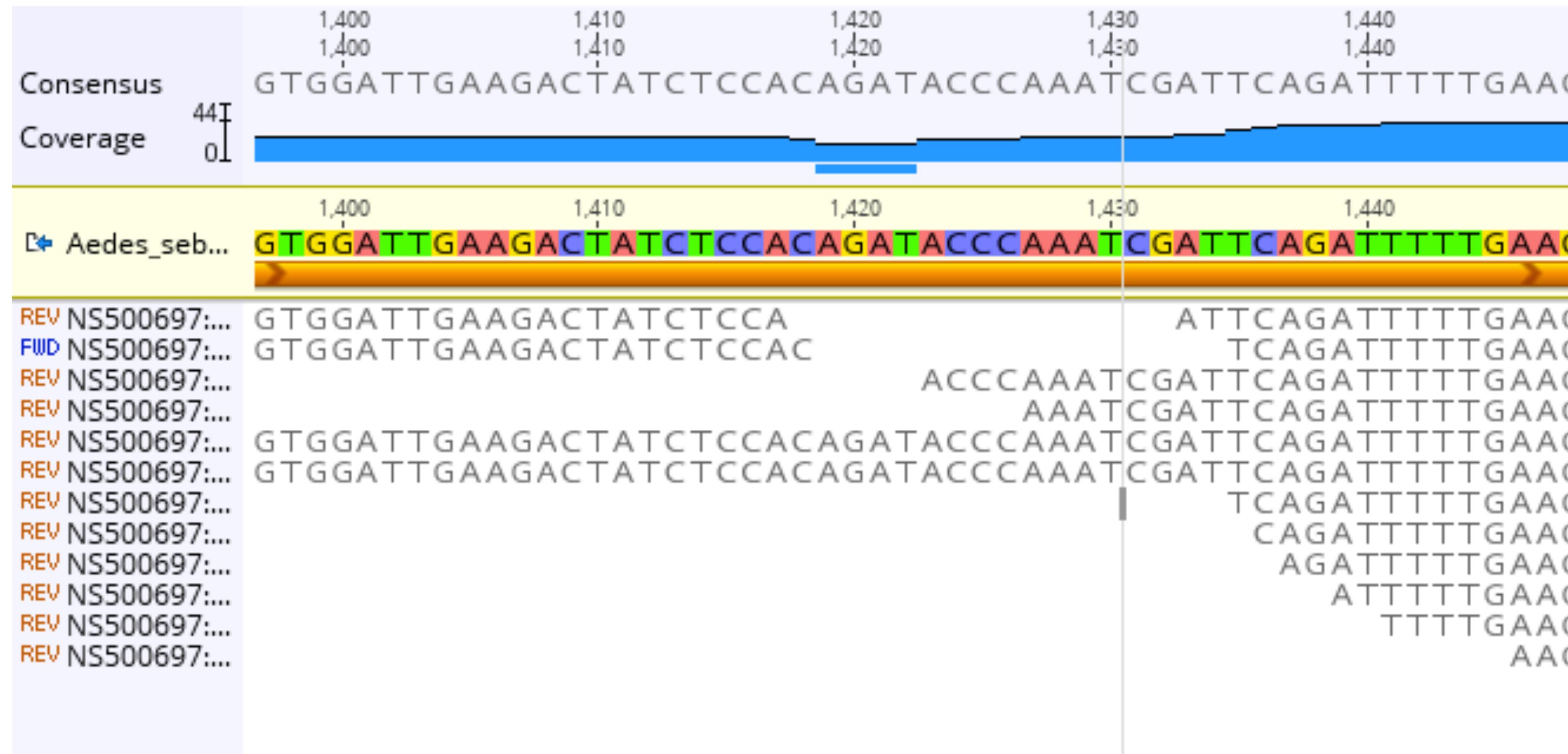
**Mapping** is the process by which sequencing reads are aligned to the region of a genome from which they derive.



(De novo) **assembly** is the process of trying to reconstruct a genome sequence from reads



**Coverage** is the number of individual reads that support a particular nucleotide in an assembled (reconstructed) sequence or that align to a particular nucleotide in a reference sequence



coverage is often referred to as  
'depth' or 'depth of coverage'

## Questions?

Is there genomics or sequencing jargon about which you're not certain?