De novo Assembly and Comparative Genomics

SENCKENBERG

world of biodiversity

Lasallia pustulata

15 million read pairs

(2x250 bp)

sequence & measure read statistics

FLASH [2]

observed (black) and fitted

Data set characteristics

(blue) insert size distribution

ART [5]

WGS simulation

Simulated WGS reads

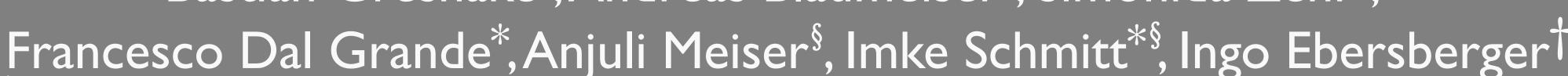
Merge reads simulated from either reference genomes to form L. pustulata twin sets with

varying coverage ratios for the two genomes. Assemble each set with 6 different assemblers.

thallus

on Eukaryotic Species Mixtures

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Summary

Mutualistic symbiotic relationships are found across organisms of all complexity. In extreme instances, as in some lichens, the interaction appears so close that the participating organisms grow only poorly – or even not at all - when cultivated in isolation. This renders mutualistic symbionts valuable objects to study the genomic basis of adaptation and co-evolution. The close interdependence in such communities, however, confounds genomic studies. In many cases separate sequencing of the participating organisms is not feasible, leaving metagenomics approaches as the method of choice. Here we address how and to what extent eukaryotic genomes can be reconstructed from such data.

Cladonia grayi [3]

Reference Genomes

Assembler Selection

Concatenate contigs of draft assembly

into one pseudo-chromosome each

1. Assembler Evaluation with Simulated Twin Sets [1] 3. Sequencing the L. pustulata metagenome

Asterochloris sp. [4]

SPAdes

de Bruijn Graph

String Graph Assembler

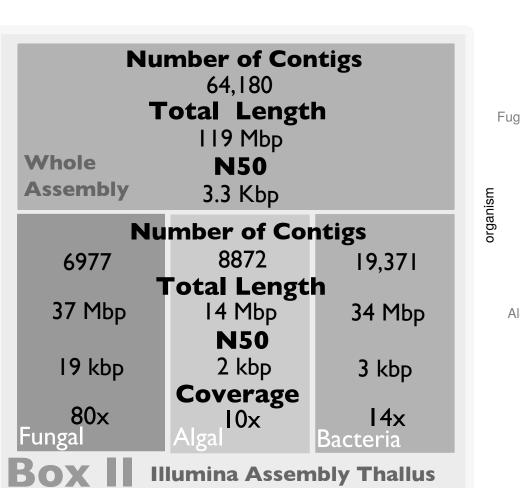
Omega

Pilot Study

Box II summarizes the assembly the metagenome skimming data (IA) with MIRA.

A comparison to the twin set (2) analysis hints at unexpected issues with the reconstruction of the algal genome.

A qPCR analysis of the lichen thallus reveals a highly biased fungal-to-algal genome ratio of 15:1.



We use in silico-generated data sets to sound out the performance of different assembly paradigms on

Whole Genome Shotgun (WGS) data from eukaryotic species mixtures. On this basis we have begun

reconstructing the metagenome of the lichen Lasallia pustulata. Using a hybrid sequencing approach, that

combines Illumina short read and PacBio long read data, we have assembled the genome of the mycobiont

and a major fraction of the algal photobiont. We integrate this data with genome sequences of closely

related non-lichenized fungi as a first step towards analyzing how lichenization affects genome evolution.

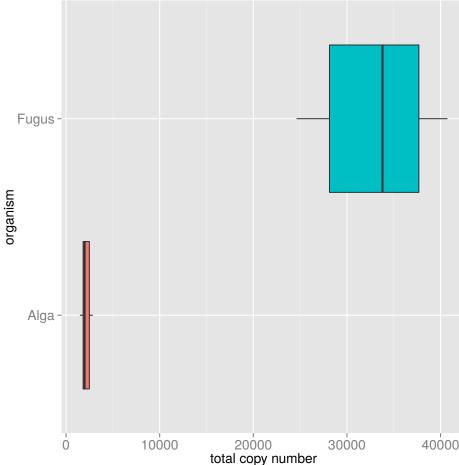
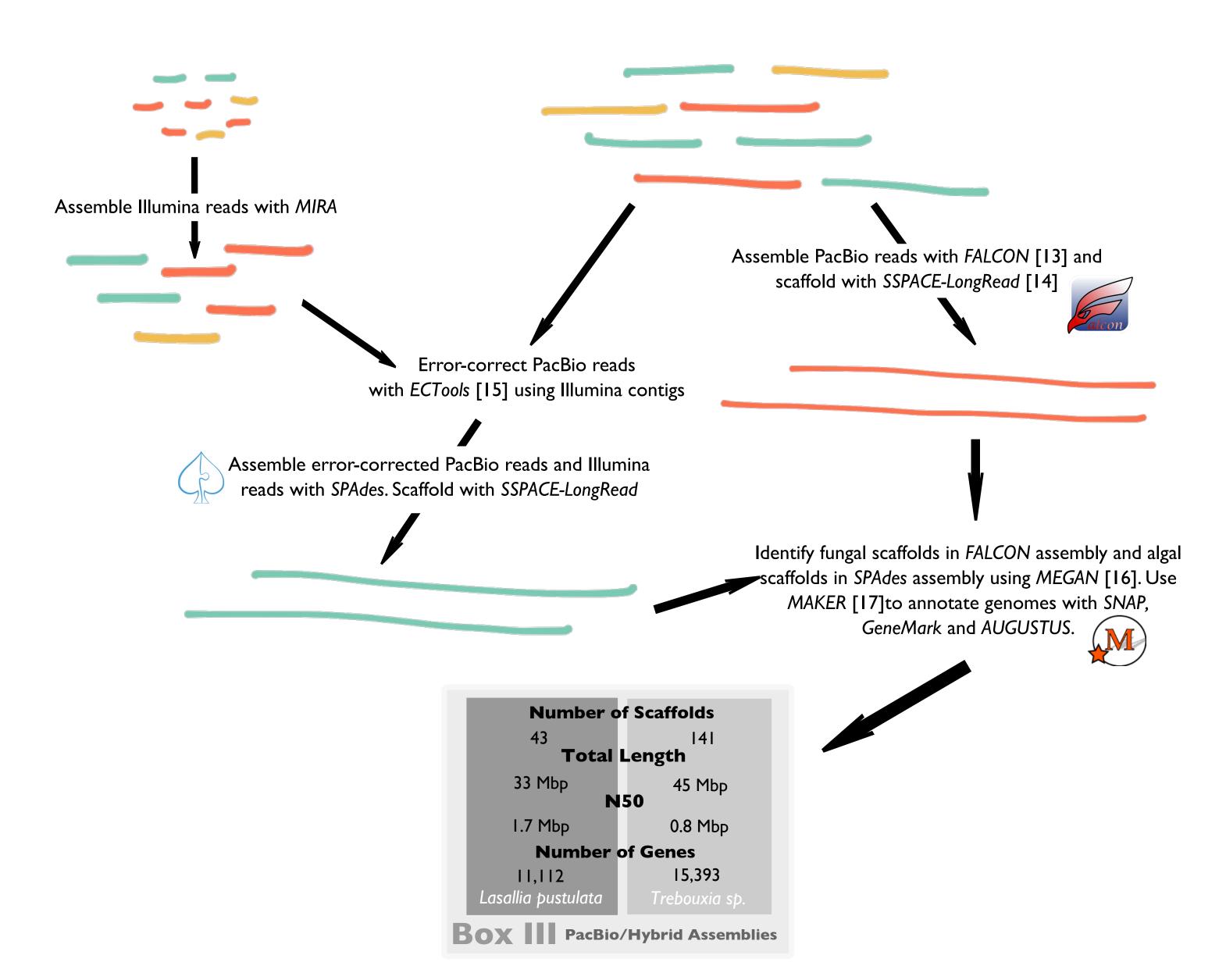


Figure 1: qPCR results for a fungal and an algal single copy. The fungal: algal ratio is around 15:1.

Hybrid Assembly: Short-Read meets Long-Read Using PacBio 2,705,256 polymerase reads with a read N50 of 15kb were sequenced. A 250 bp mate pair library (5kb inserts) of 15 million reads was sequenced using Illumina MiSeq.

To cope with the coverage differences we pursued two different assembly strategies, targeting the fungal and the algal genome respectively. For high coverage data PacBio-only assemblies are state of the art, lowcoverage data require hybrid assemblies using Illumina and PacBio data [12].



4. Does Lichenization Facilitate Gene Loss?

Ancestral Gene Set To investigate lineage specific gene loss,

the Last Common Ancestor (LCA) gene set of the Pezizomycotina

was reconstructed using OMA [18] (Figure 2). In total 12,595

Absence of LCA Genes For 1,357 groups genes were only

found in 7 species. In 1/3 or these groups the L. pustulata ortholog

is missing, hinting that these genes are lost as a consequence of

orthologous groups were formed (Figure 3).

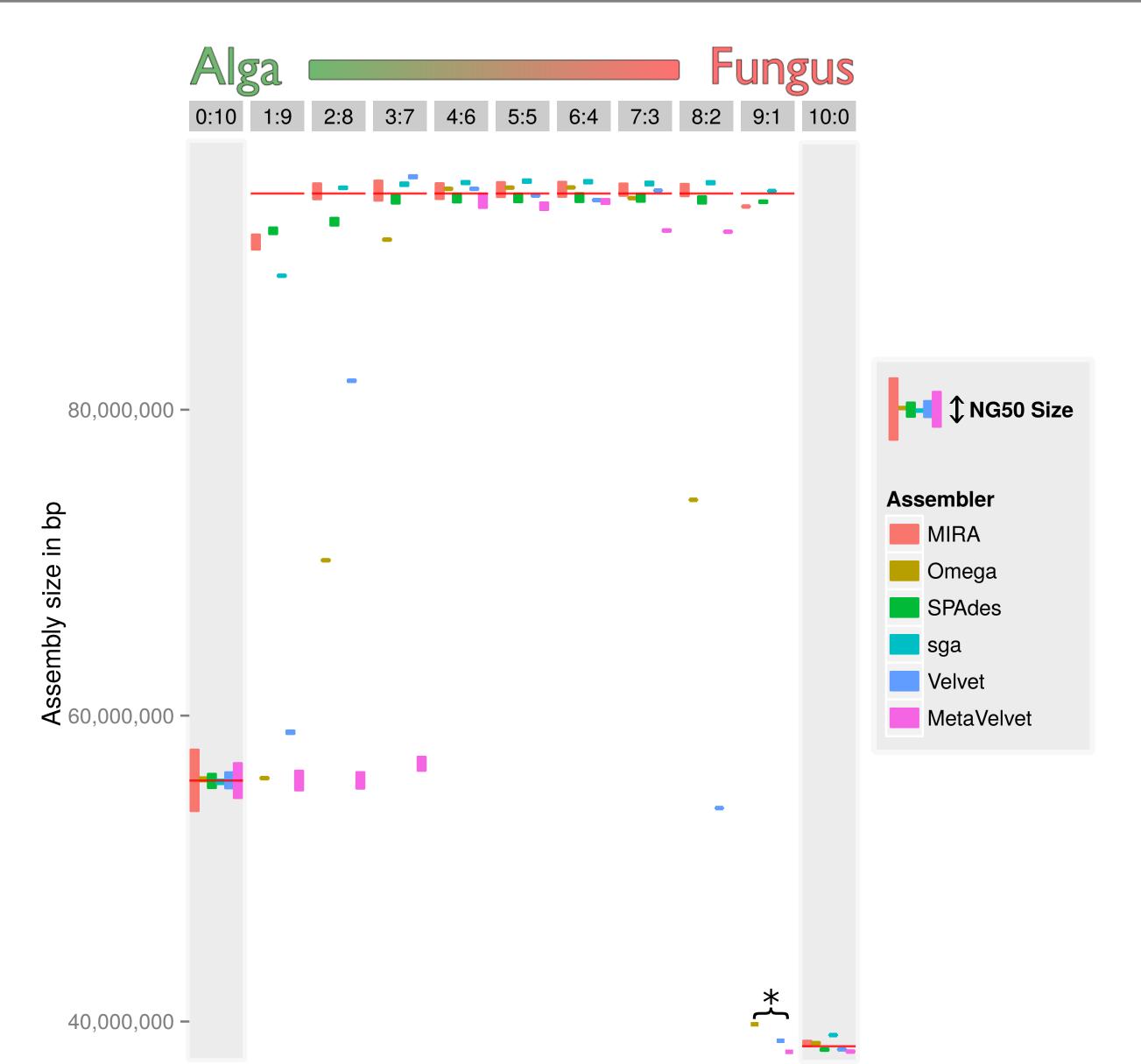
lichenization (Figure 4).

2. Assembly Results of the Twin Sets

1:9

3:7

0:10



Assembly results for the 11 twin sets. Bars are centered at total assembly length, red lines indicate reference lengths. Height of bars shows the NG50 size. Assemblies marked with an asterisk cover less than 50% of the reference length. A default height was used in those instances.

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[1] Greshake B, Zehr S, Dal Grande F et al. Mol Ecol Res (2015) epub ahead of print [2] Magoc T and Salzberg S. Bioinformatics (2011) 27 (21):2957-63 [3] http://genome.jgi.doe.gov/Clagr2/ [4] http://genome.jgi.doe.gov/Astpho2/

all 8 species were found. For 1357 groups only 7 species were found.

[5] Huang W, Li L, Myers JR, Marth GT. Bioinformatics (2012) 28 (4):593-594 [6] Zerbino DR and Birney E. Genome Research (2008) 18:821-829. [7] Namiki T, Hachiya T, Tanaka H, Sakakibara Y. Nucleic Acids Res, (2012) 40(20), e155

[8] Bankevich A, Nurk S, Antipov D et al. Journal of Computational Biology (2012) 19(5):455-477 [9] http://sourceforge.net/projects/mira-assembler/ [10] Haider B, Ahn T, Bushnell B et al. Bioinformatics (2014) btu395

[11] Simpson JT and Durbin R. Bioinformatics (2010) 26 (12): i367-i373

[12] Mike Schatz, PAG 2014 (http://schatzlab.cshl.edu/presentations/2014-01-14.PAG.Single% 20Molecule%20Assembly.pdf) [13] https://github.com/PacificBiosciences/FALCON [14] Boetzer M and Pirovano W. BMC Bioinformatics (2014) 15:211

genes missing in LCA core set

Figure 2: Tree of 8 species used for creating the LCA gene set.

[15] https://github.com/jgurtowski/ectools [17] Campbell MS, Holt C, Moore B, Yandell M.

[18] http://omabrowser.org/standalone/

S.sclerotiorum

many orthologous groups as any other species.

Figure 3: Results of the orthology prediction with OMA. For 1153 groups Figure 4: Distribution of genes missing in the LCA set. Lasallia pustulata is missing in twice as



asallia pustulata Lecanoromycetes

Eurotiomycetes

Sordariomyceta





[16] Huson DH, Mitra S, Ruscheweyh H et al. Genome Research (2011) 21: 1552-1560 Curr Protoc Bioinformatics (2014) 48:4.11.1-4.11.39

References