

# Computational Systems Biology Deep Learning in the Life Sciences

6.802 20.390 20.490 HST.506

6.874 Area II TQE (AI)

David Gifford  
Lecture 1  
February 4, 2019



<http://mit6874.github.io>

# Your guides



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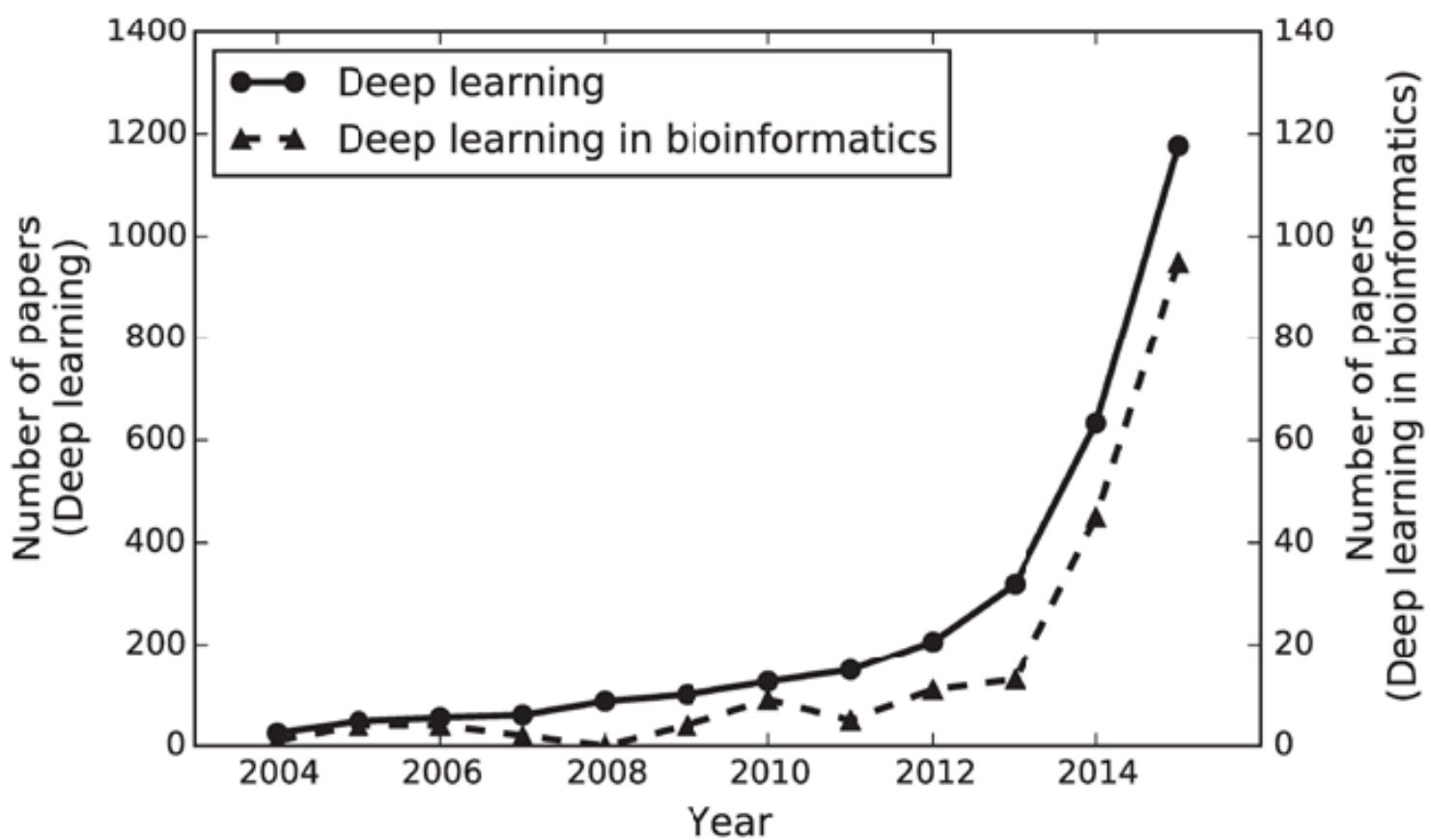
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You should have received the Google  
Cloud coupon URL in your email

Recitations (this week)  
Thursday 4 - 5pm 36-155  
Friday 4 - 5pm 36-155

Office hours are after recitation at 5pm  
in same room  
(PS1 help and advice)

Approximately 8% of deep learning publications are in bioinformatics



## Welcome to a new approach to life sciences research

- Enabled by the convergence of three things
  - Inexpensive, high-quality, collection of large data sets (sequencing, imaging, etc.)
  - New machine learning methods (including ensemble methods)
  - High-performance Graphics Processing Unit (GPU) machine learning implementations
- Result is completely transformative

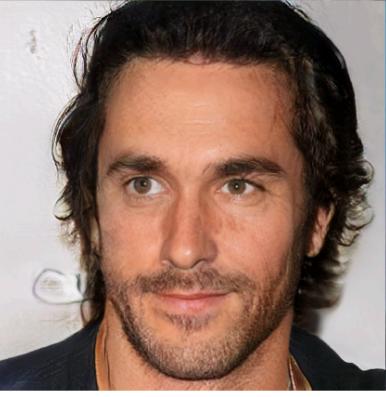
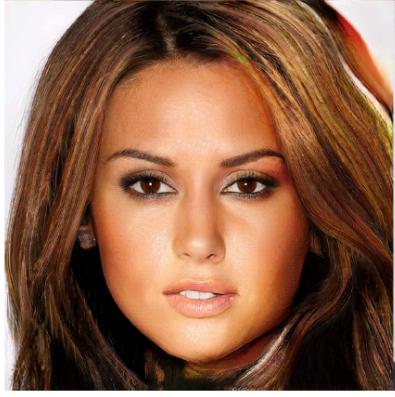


# Alternative MIT subjects

- 6.047 / 6.878 Computational Biology: Genomes, Networks, Evolution
- 6.S897/HST.956: Machine Learning for Healthcare (2:30pm 4-270)
- 8.592 Statistical Physics in Biology
- 7.09 Quantitative and Computational Biology
- 7.32 Systems Biology
- 7.33 Evolutionary Biology: Concepts, Models and Computation
- 7.57 Quantitative Biology for Graduate Students
- 18.417 Introduction to Computational Molecular Biology
- 20.482 Foundations of Algorithms and Computational Techniques in Systems Biology

Machine Learning is the ability to improve on  
a task with more training data

- Task T to be performed
  - Classification, Regression, Transcription, Translation, Structured Output, Anomaly Detection, Synthesis, Imputation, Denoising
- Measured by Performance Measure P
- Trained on Experience E (Training Data)

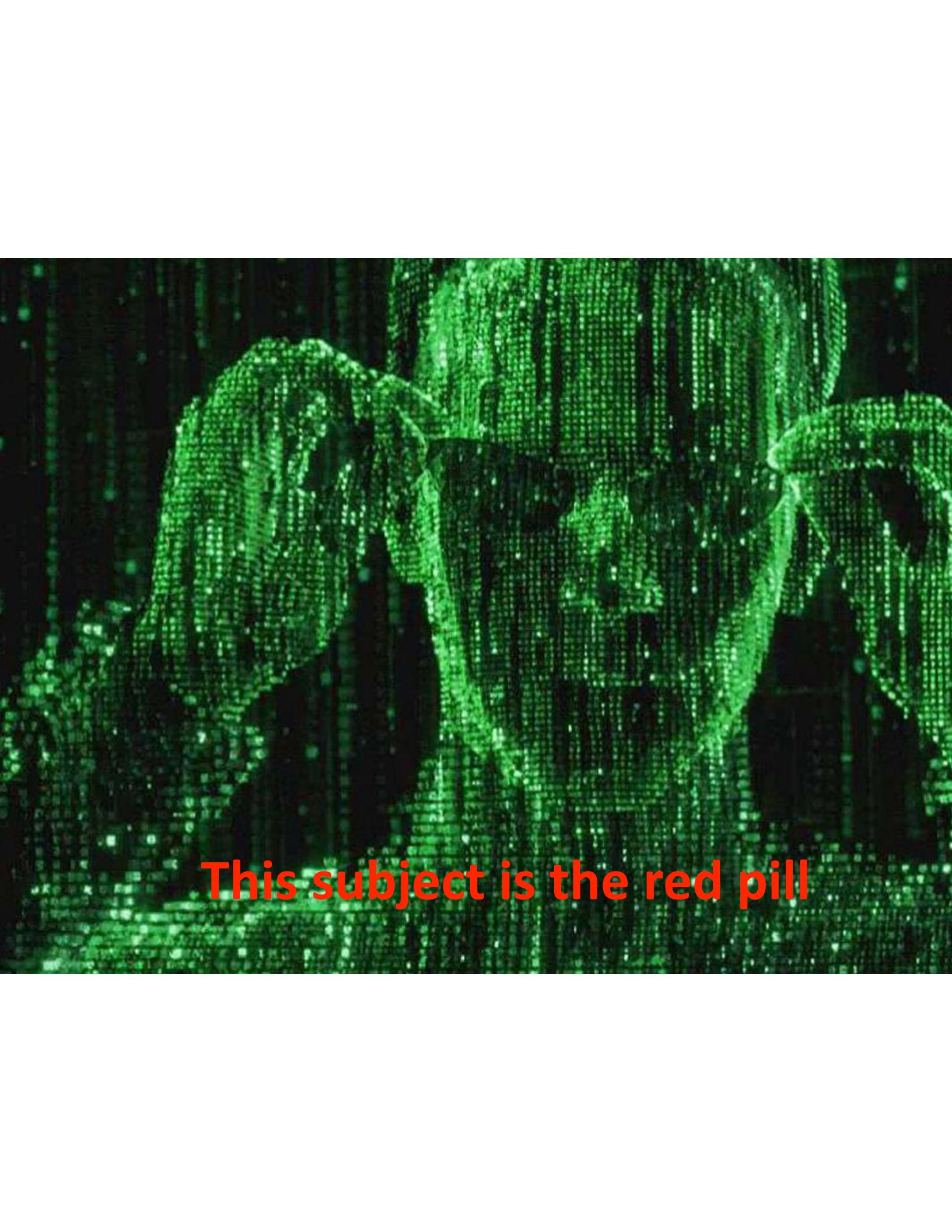


# Synthetic Celebrities



Trained on 30,000 images from CelebA-HQ

<https://arxiv.org/abs/1710.10196>

A close-up photograph of a person's face, where every pixel is replaced by a single green binary digit ('1'). The person has dark hair and is looking slightly to the right.

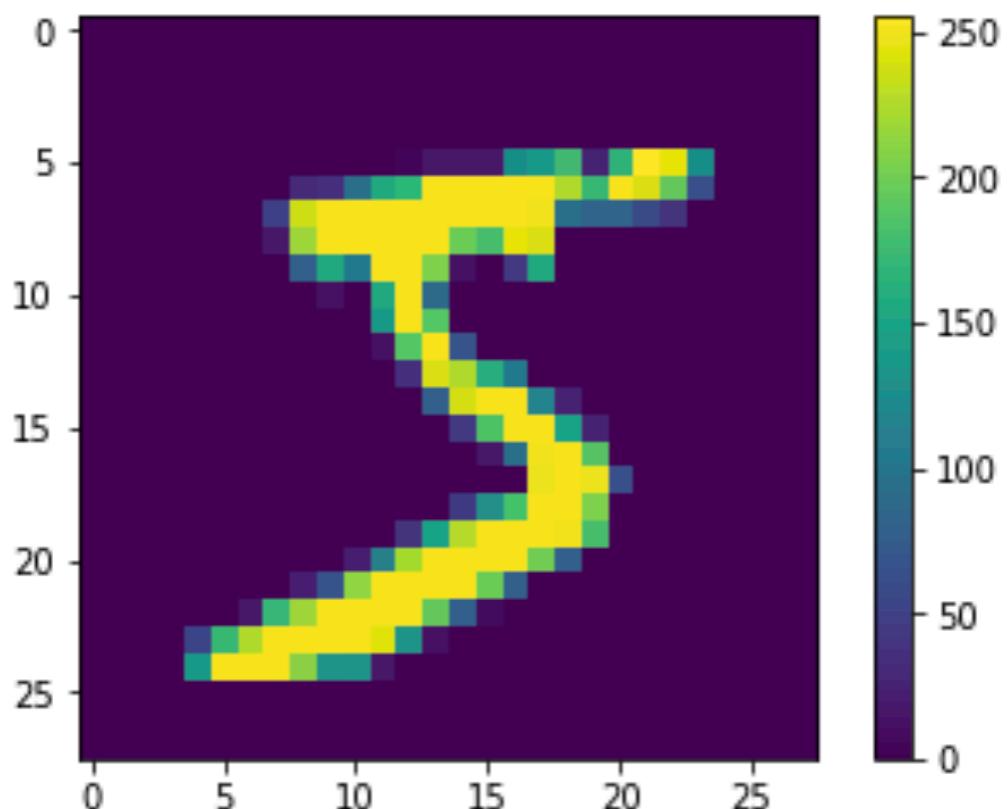
**This subject is the red pill**

## Welcome

|     |        |                                                     |
|-----|--------|-----------------------------------------------------|
| L 1 | Feb. 5 | Machine learning in the computational life sciences |
| L 2 | Feb. 7 | Neural networks and TensorFlow                      |
| R 1 | Feb 7  | Machine Learning Overview and PS 1                  |
| L 3 | Feb 12 | Convolutional and recurrent neural networks         |

Problem Set: Softmax MNIST (PS 1)

# PS 1: Tensor Flow Warm Up



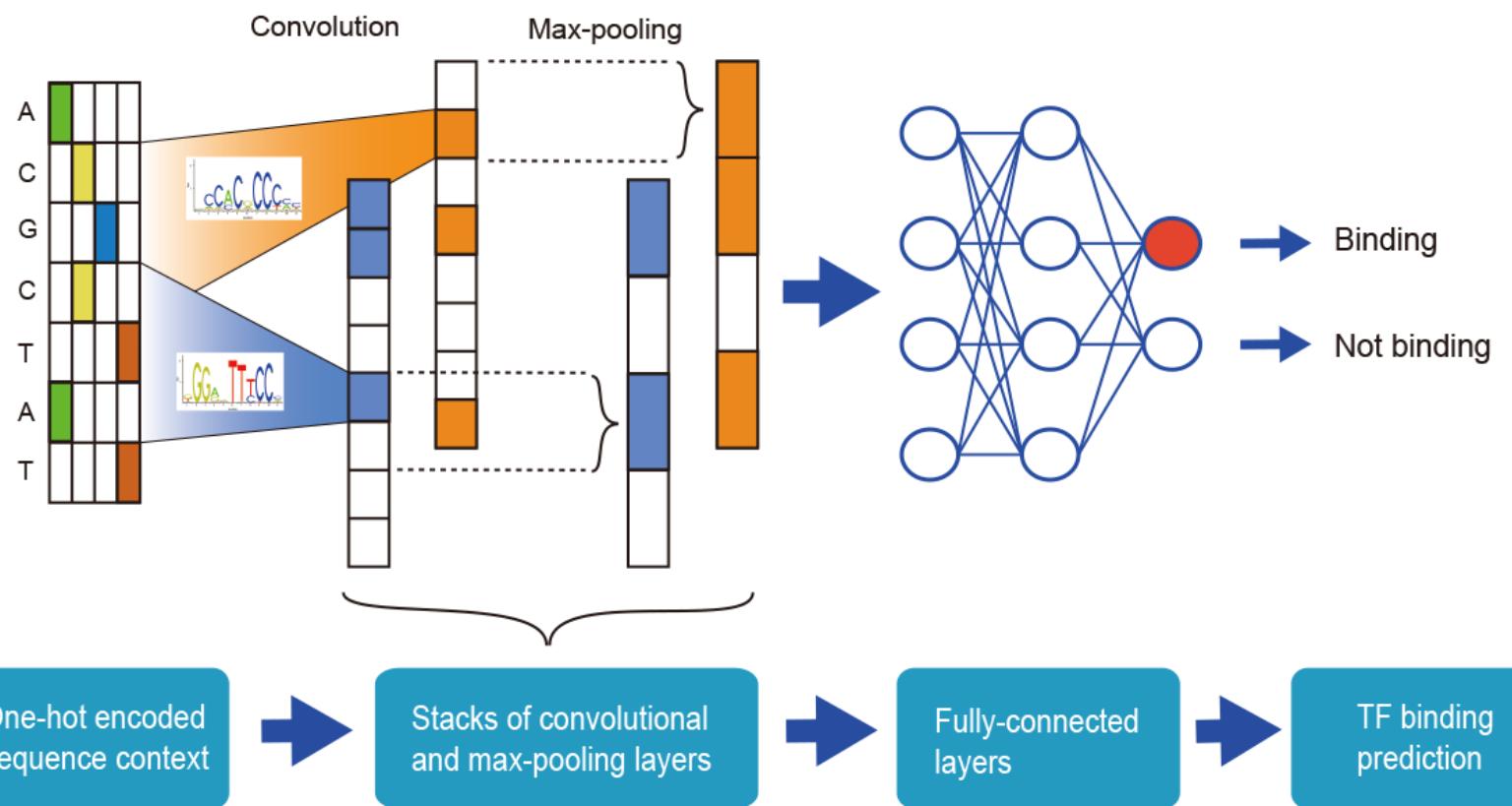
ground truth: 5

## Regulatory Elements / ML models and interpretation

|     |         |                                                    |
|-----|---------|----------------------------------------------------|
| L 4 | Feb 14  | Protein-DNA interactions                           |
| R 2 | Feb. 14 | Neural Networks and TensorFlow                     |
|     | Feb. 19 | (Holiday - President's Day)                        |
| L 5 | Feb. 21 | Models of Protein-DNA Interaction                  |
| R 3 | Feb. 21 | Motifs and models                                  |
| L 6 | Feb. 26 | Model interpretation (Gradient methods, black box) |

Problem Set: Regulatory Grammar

# PS 2: Genomic regulatory codes

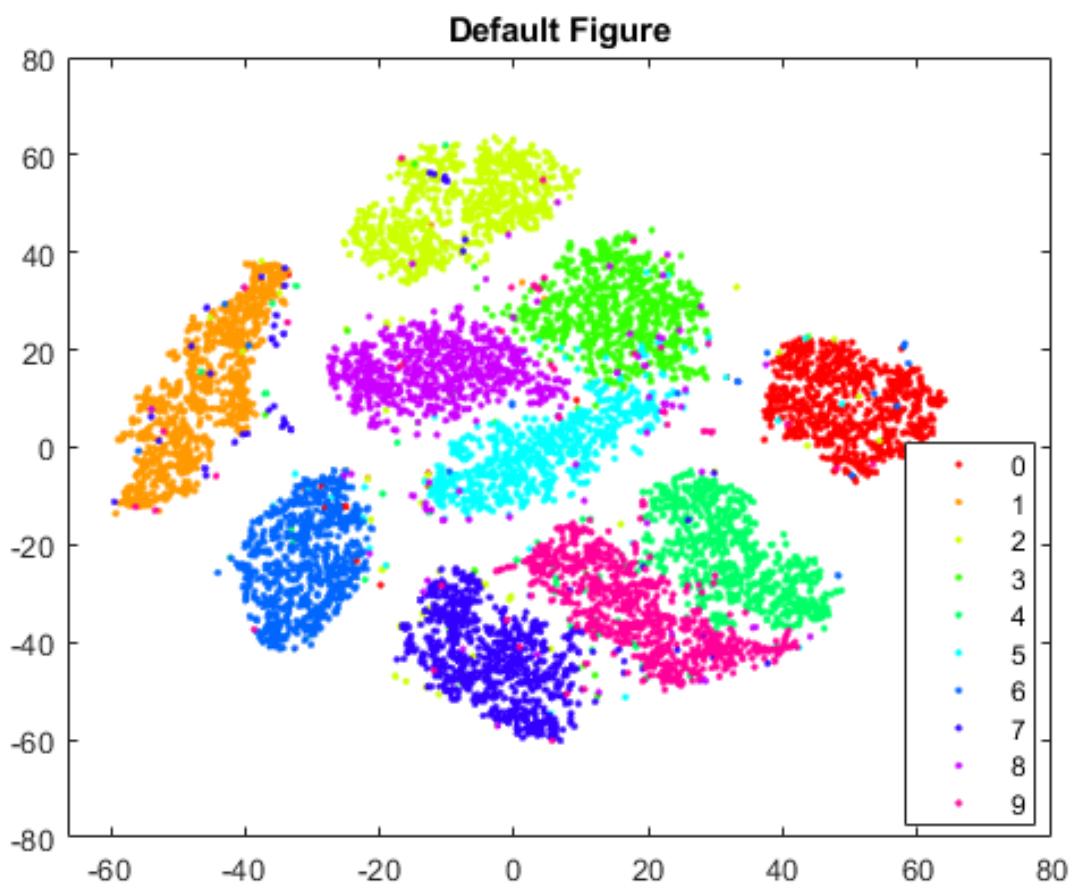


## The Expressed Genome / Dimensionality reduction

|     |         |                                                     |
|-----|---------|-----------------------------------------------------|
| L 7 | Feb. 28 | The expressed genome and RNA splicing               |
| R 4 | Feb 28  | Model interpretation                                |
| L 8 | Mar 5   | PCA, dimensionality reduction (t-SNE), autoencoders |
| L 9 | Mar 7   | scRNA seq and cell labeling                         |
| R 5 | Mar 7   | Compressed state representations                    |

Problem Set: scRNA-seq tSNE

# PS 3: Parametric tSNE

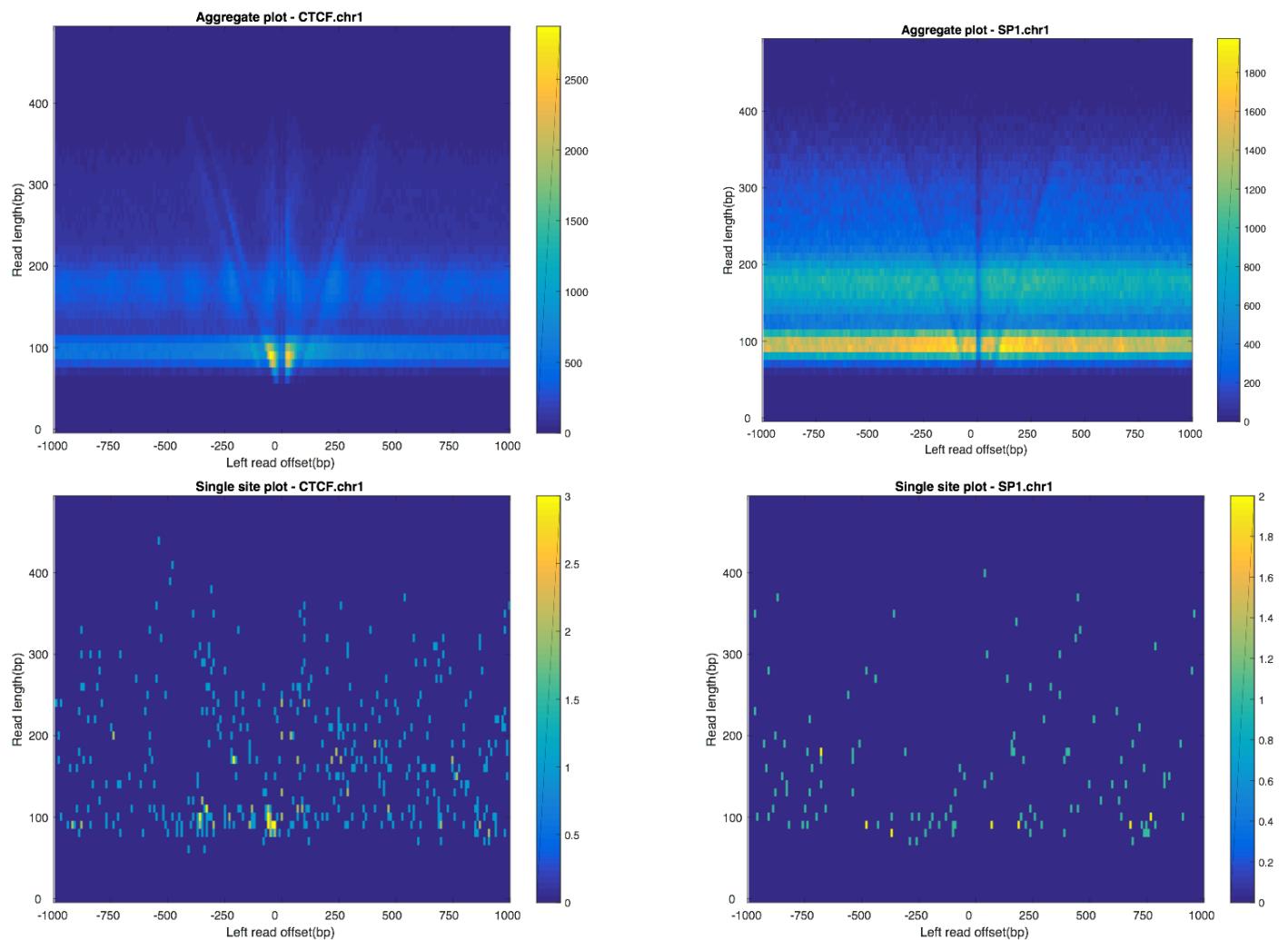


## Gene Regulation / Model selection and uncertainty

|      |        |                                                     |
|------|--------|-----------------------------------------------------|
| L 10 | Mar 12 | Modeling gene expression and regulation             |
| L 11 | Mar 14 | Model uncertainty, significance, hypothesis testing |
| R 6  | Mar 14 | Model selection and L1/L2 regularization            |
| L 12 | Mar 19 | Chromatin accessibility and marks                   |
| L 13 | Mar 21 | Predicting chromatin accessibility                  |
| R 7  | Mar 21 | Chromatin accessibility                             |

Problem Set: CTCF Binding from DNase-seq

# PS 4: Chromatin Accessibility



## Genotype -> Phenotype, Therapeutics

|      |        |                                                |
|------|--------|------------------------------------------------|
| L 14 | Apr 2  | Discovering and predicting genome interactions |
| L 15 | Apr 4  | eQTL prediction and variant prioritization     |
| R 8  | Apr 4  | Lead SNPs to causal SNPs; haplotype structure  |
| L 16 | Apr 9  | Imaging and genotype to phenotype              |
| L 17 | Apr 11 | Generative models: optimization, VAEs, GANs    |
| R 9  | Apr 11 | Generative models                              |
| L 18 | Apr 18 | Deep Learning for eQTLs                        |
| L 19 | Apr 23 | Therapeutic Design                             |
| L 20 | Apr 25 | Exam Review                                    |
| L 21 | Apr 30 | Exam                                           |

Problem Set: Generative models for medical records

# PS 5: Generative Models

Sample 1: discharge instructions: please contact your primary care physician or return to the emergency room if [\*omitted\*] develop any constipation. [\*omitted\*] should be had stop transferred to [\*omitted\*] with dr. [\*omitted\*] or started on a limit your medications. \* [\*omitted\*] see fult dr. [\*omitted\*] office and stop in a 1 mg tablet to tro fever great to your pain in postions, storale. [\*omitted\*] will be taking a cardiac catheterization and take any anti-inflammatory medicines diagness or any other concerning symptoms.

# Your programming environment

## Problem 2

In this problem, we wish to use CNN to learn the motif of CTCF from sequences with similar di-nucleotide frequency. The positive samples are 101bp sequences centered at CTCF ChIP-seq peaks from GM12878 cell line. The negative sequences are generated by permuting the nucleotides in the positive sequences while keeping the di-nucleotide frequency.

We will provide functions for loading data, training and testing. You will:

- implement a CNN model with given specifications
- specify the initialization of parameters in the model
- train the model and evaluate on the test set

All the places where you need to fill in begins with "TODO" and ends with "END OF YOUR CODE".

```
In [1]: import tensorflow as tf, sys, numpy as np, h5py
from os.path import join, dirname, basename, exists, realpath
from os import makedirs
from tensorflow.examples.tutorials.mnist import input_data
from sklearn.metrics import roc_auc_score
```

```
In [2]: data_folder = '../data/motif_disc'
batch_size = 128
valid_size = 2000
epochs = 20
best_model_file = join('../output', basename(data_folder), 'best_model.ckpt')
if not exists(dirname(best_model_file)):
    makedirs(dirname(best_model_file))
```

```
In [3]: # Function to load the data embedded in the previous problem and their labels
def load_data(mydir):
    train = h5py.File(join(mydir, 'train.h5'), 'r')
```



Your computing resource



# Cloud Platform Education Grants

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Thank you for your interest in Google Cloud Platform Education Grants. Please fill out the form below to receive a coupon code for credit to use on Google Cloud Platform.

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If you do not see your domain listed, please contact your course instructor: [gifford@mit.edu](mailto:gifford@mit.edu)

By clicking "Submit" below, you agree that we may share the following information with your educational institution and course instructor ([gifford@mit.edu](mailto:gifford@mit.edu)): (1) personal information that you provide to us on this form and (2) information regarding your use of the coupon and Google Cloud Platform products.

**Submit**

Your grade is based on 5 problem sets,  
an exam, and a final project

- Five Problem Sets (40%)
  - Individual contribution
  - Done using Google Cloud, Jupyter Notebook
- In class exam (1.5 hours), one sheet of notes (30%)
- Final Project (30%)
  - Done individually or in teams (6.874 by permission)
  - Substantial question

# Amgen could not reproduce the findings of 47/53 (89%) landmark preclinical cancer papers

## REPRODUCIBILITY OF RESEARCH FINDINGS

Preclinical research generates many secondary publications, even when results cannot be reproduced.

| Journal impact factor | Number of articles | Mean number of citations of non-reproduced articles* | Mean number of citations of reproduced articles |
|-----------------------|--------------------|------------------------------------------------------|-------------------------------------------------|
| >20                   | 21                 | 248 (range 3–800)                                    | 231 (range 82–519)                              |
| 5–19                  | 32                 | 169 (range 6–1,909)                                  | 13 (range 3–24)                                 |

Results from ten-year retrospective analysis of experiments performed prospectively. The term 'non-reproduced' was assigned on the basis of findings not being sufficiently robust to drive a drug-development programme.

\*Source of citations: Google Scholar, May 2011.

<http://www.nature.com/nature/journal/v483/n7391/pdf/483531a.pdf>

# Direct and conceptual replication is important

- **Direct replication** is defined as attempting to reproduce a previously observed result with a procedure that provides no a priori reason to expect a different outcome
- **Conceptual replication** uses a different methodology (such as a different experimental technique or a different model of a disease) to test the same hypothesis; tries to avoid confounders

<https://elifesciences.org/content/6/e23383>

# Reproducibility Project: Cancer Biology Registered Report/Replication Study Structure

- A **Registered Report** details the experimental designs and protocols that will be used for the replications, and experiments cannot begin until this report has been peer reviewed and accepted for publication.
- The results of the experiments are then published as a **Replication Study**, irrespective of outcome but subject to peer review to check that the experimental designs and protocols were followed.

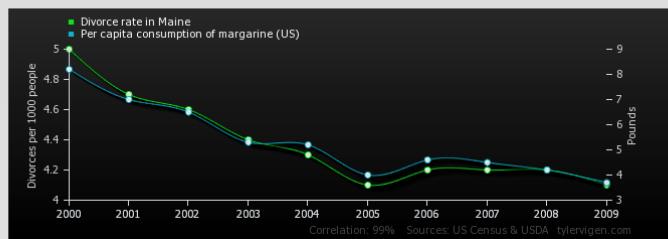
<https://elifesciences.org/content/6/e23383>

# Claim precision is key to science

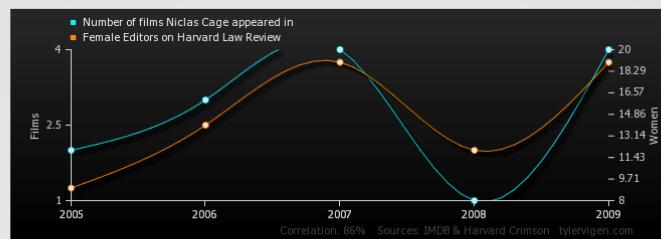
- “We have discovered the regulatory elements”
- “We have predicted the regulatory elements”
- “The variant causes a difference in gene expression”
- “The variant is associated with a difference in gene expression”

# Correlation ≠ Causation

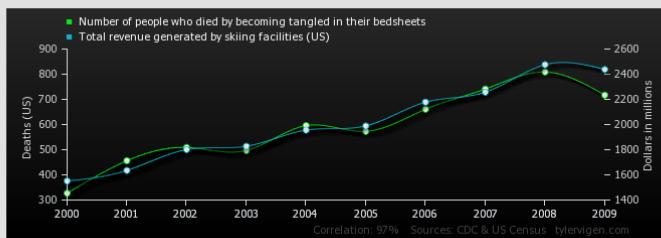
Divorce rate in Maine  
correlates with  
Per capita consumption of margarine (US)



Number of films Niclas Cage appeared in  
correlates with  
Female Editors on Harvard Law Review

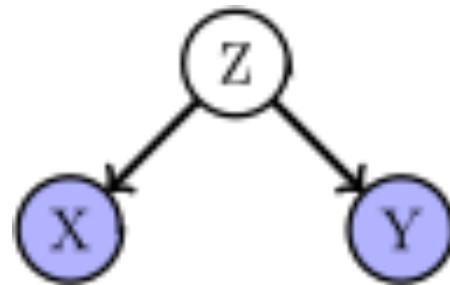


Number of people who died by becoming tangled in their bedsheets  
correlates with  
Total revenue generated by skiing facilities (US)

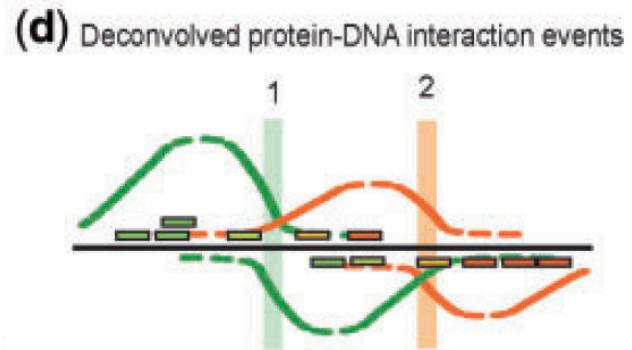
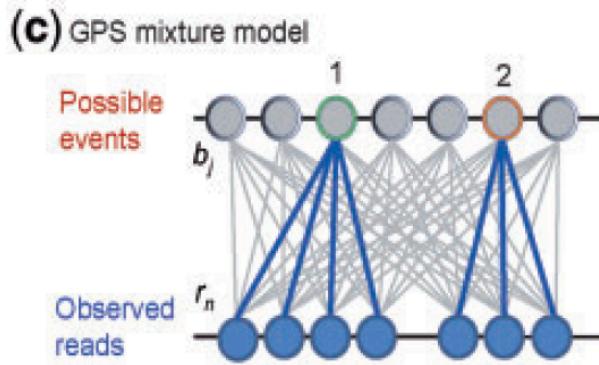
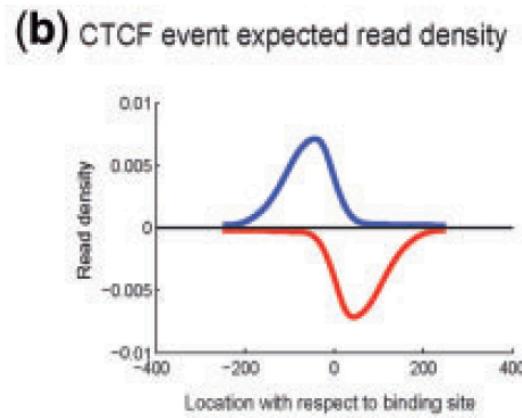
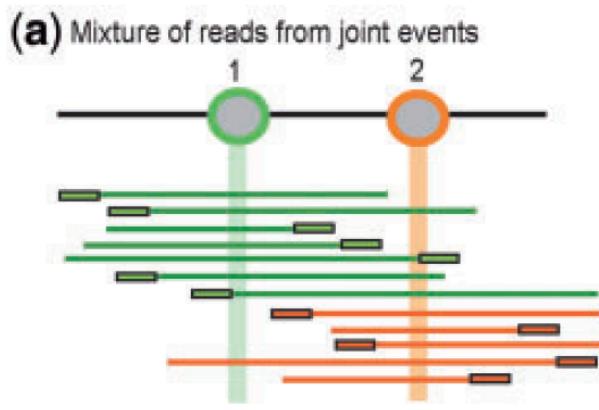


# Interventions enable causal statements

- Observation only data can be influenced by confounders
- A confounder is an unobserved variable that explains an observed effect
- Interventions on a variable allow for the detection of its direct and indirect effects



# ML resolves Protein-DNA binding events

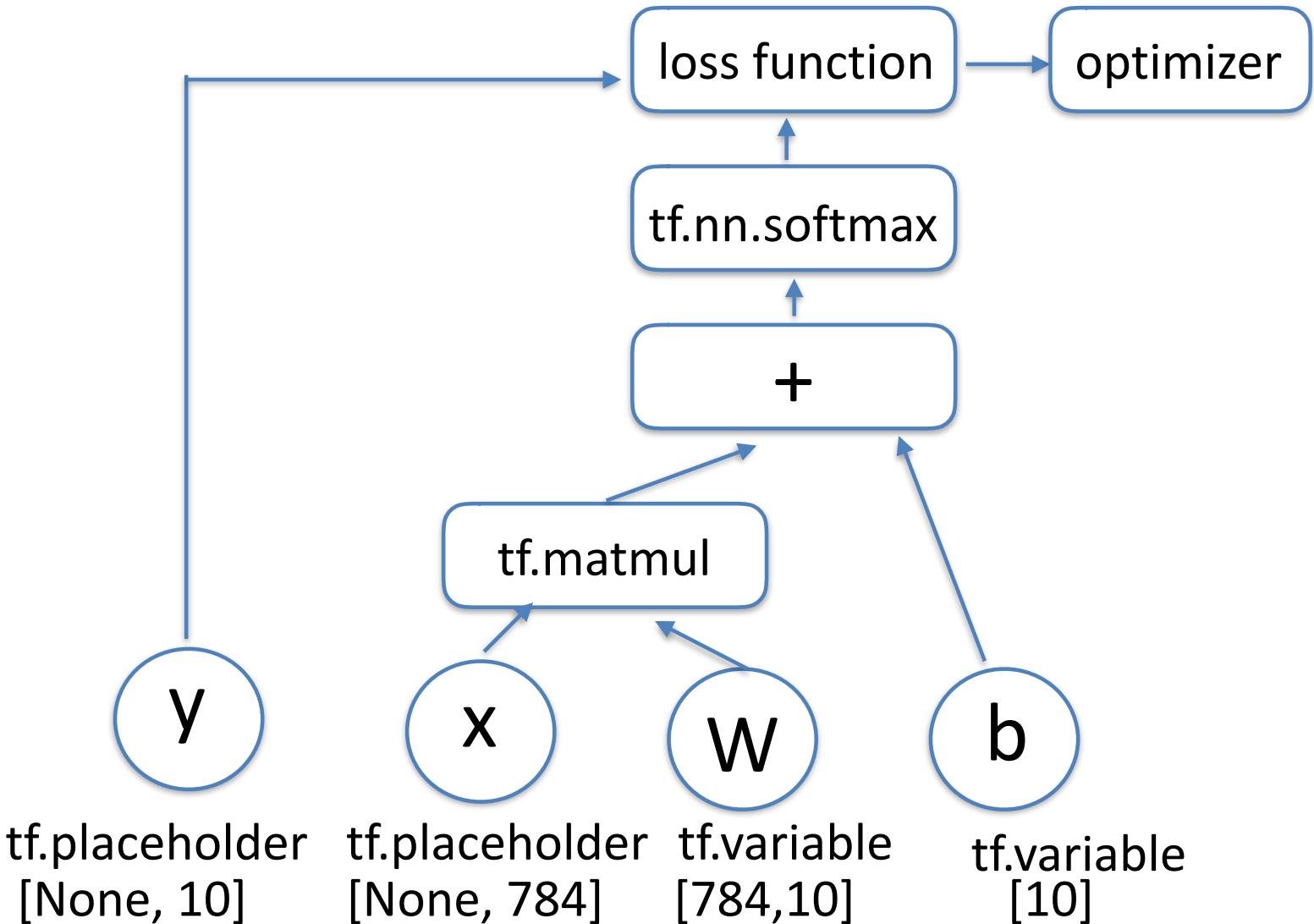


- Who - what protein(s) are binding?
- Where - where are they binding?
- Why - what chromatin state and sequence motif causes their binding?
- When - what differential binding is observed in different cell states or genotypes?
- How - are accessory factors or modifications of the factor involved?

# How can we establish ground truth?

- Replicate experiments should have consistent observations
- Independent tests for same hypothesis (different antibody, different assay)
- Statistical test against a null hypothesis - what is the probably of seeing the reads at random? We need a *null model* for this test.

## Problem Set 1 Structure



# Programming model

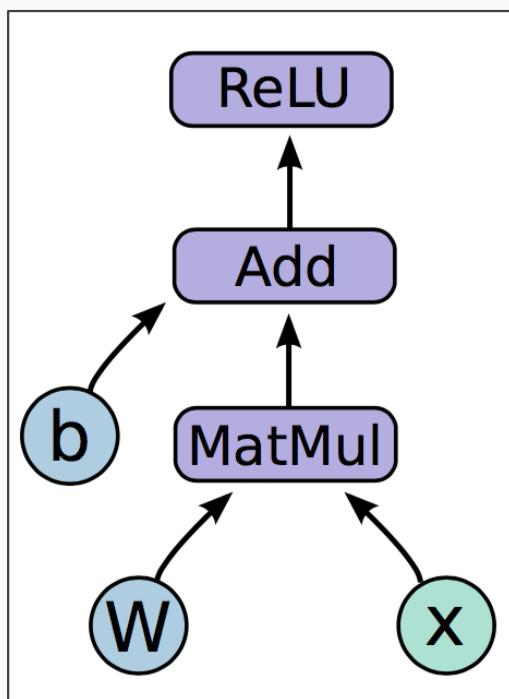
Big idea: Express a numeric computation as a **graph**.

Graph nodes are **operations** which have any number of inputs and outputs

Graph edges are **tensors** which flow between nodes

## Programming model: NN feedforward

$$h_i = \text{ReLU}(Wx + b)$$

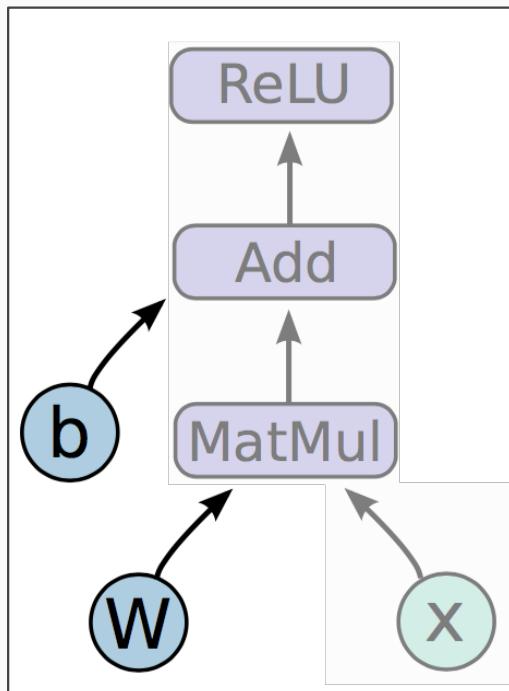


## Programming model: NN feedforward

$$h_i = \text{ReLU}(Wx + b)$$

**Variables** are 0-ary stateful nodes which output their current value.  
(State is retained across multiple executions of a graph.)

(parameters, gradient stores, eligibility traces, ...)

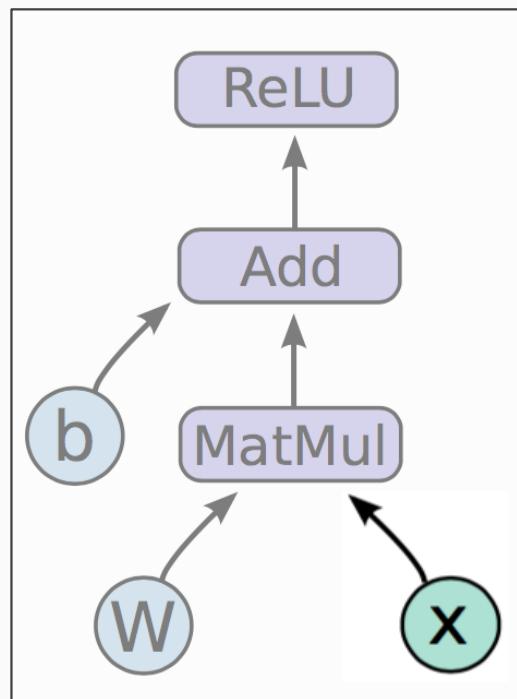


## Programming model: NN feedforward

$$h_i = \text{ReLU}(Wx + b)$$

**Placeholders** are 0-ary nodes whose value is fed in at execution time.

(inputs, variable learning rates, ...)



## Programming model: NN feedforward

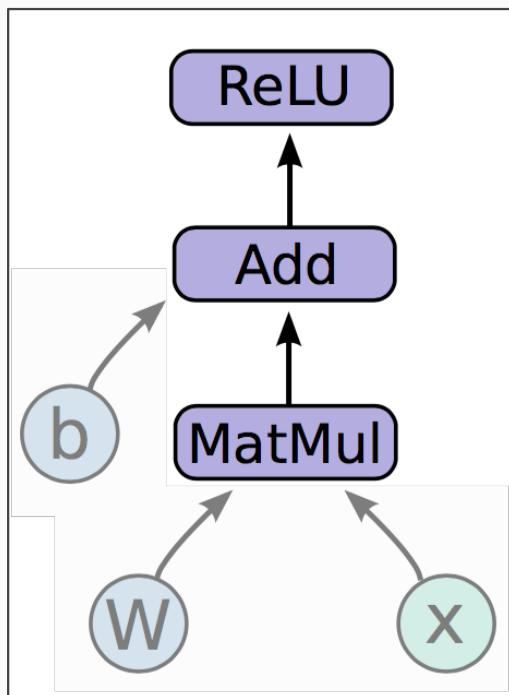
$$h_i = \text{ReLU}(Wx + b)$$

### Mathematical operations:

**MatMul**: Multiply two matrix values.

**Add**: Add elementwise (with broadcasting).

**ReLU**: Activate with elementwise rectified linear function.



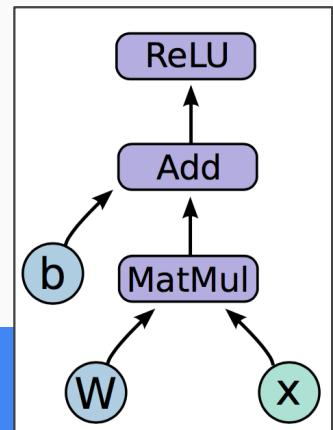
# In code, please!

1. Create model weights, including initialization
  - a.  $W \sim Uniform(-1, 1)$ ;  $b = 0$
2. Create input placeholder  $x$ 
  - a.  $m * 784$  input matrix
3. Create computation graph

```
import tensorflow as tf

1 b = tf.Variable(tf.zeros((100,)))
W = tf.Variable(tf.random_uniform((784,
100), -1, 1))
2 x = tf.placeholder(tf.float32, (None,
784))
3 h_i = tf.nn.relu(tf.matmul(x, W) + b)
```

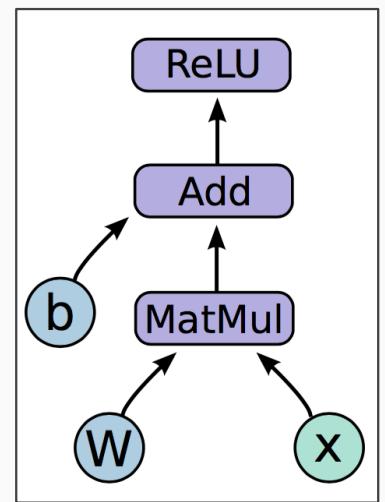
$$h_i = \text{ReLU}(Wx + b)$$



# How do we run it?

So far we have defined a **graph**.

We can deploy this graph with a **session**: a binding to a particular execution context (e.g. CPU, GPU)



# Getting output

```
sess.run(fetches, feeds)
```

**Fetches:** List of graph nodes.  
Return the outputs of these nodes.

**Feeds:** Dictionary mapping from graph nodes to concrete values. Specifies the value of each graph node given in the dictionary.

```
import numpy as np
import tensorflow as tf

1 b = tf.Variable(tf.zeros((100,)))
W = tf.Variable(tf.random_uniform((784,
2 100) -1, 1))
x = tf.placeholder(tf.float32, (None, 784))
3 h_i = tf.nn.relu(tf.matmul(x, W) + b)
```

```
sess = tf.Session()
sess.run(tf.global_variables_initializer())
sess.run(h_i, {x: np.random.random(64,
784)})
```

# Basic flow

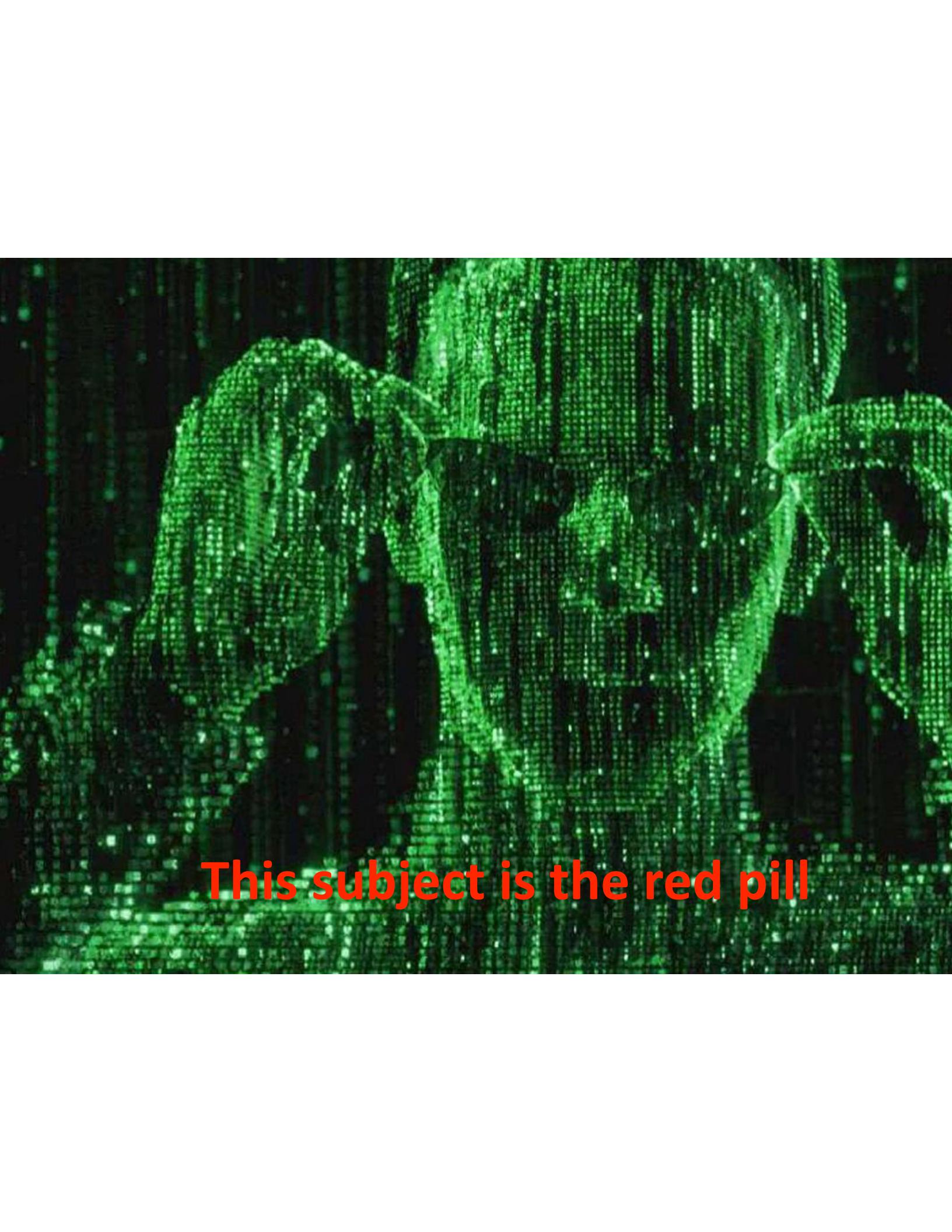
## 1. Build a graph

- a. Graph contains parameter specifications, model architecture, optimization process, ...
- b. Somewhere between 5 and 5000 lines

## 2. Initialize a session

## 3. Fetch and feed data with Session.run

- a. Compilation, optimization, etc. happens at this step — you probably won't notice

A close-up photograph of a person's face, where the skin texture is replaced by a dense grid of green digital code or binary digits (0s and 1s). The person has dark hair and is looking slightly to the right.

**This subject is the red pill**