

Homework4

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```
library(ProbBayes)
library(dplyr)
library(ggplot2)
require(gridExtra)
library(reshape)
library(runjags)
library(coda)
library(tidyverse)
library(fastDummies)
crcblue <- "#2905a1"
```

```
CESample <- read.csv("CESample2.csv")
```

I decided that I wanted to use all variables within CESampe logExpenditure, UrbanRural, Race) as estimators for logIncome. Thus, I used a Multilinear regression model in which I scaled Log Income and Log Expenditure by centering at 0 and dividing by standard deviation. I use the following MLR model (where * denotes a standardized continuous variable):

$$\begin{aligned} Y_i^* \mid \beta_0, \beta_1, \dots, \beta_7, \sigma, \mathbf{x}_i^* &\stackrel{ind}{\sim} \text{Normal}(\beta_0 + \beta_1 x_{i,expenditure}^* + \beta_2 x_{i,rural}^* \\ &+ \beta_3 x_{i,race_B} + \beta_4 x_{i,race_N} \\ &+ \beta_5 x_{i,race_A} + \beta_6 x_{i,race_P} \\ &+ \beta_7 x_{i,race_M}, \sigma). \end{aligned} \quad (1)$$

```
CESample <- CESample %>%
  mutate(LogTotalIncome = log(TotalIncomeLastYear))
CESample <- CESample %>%
  mutate(LogTotalExp = log(TotalExpLastQ))
```

```
CESample$Log_TotalExpSTD <- scale(CESample$LogTotalExp)
CESample$Log_TotalIncomeSTD <- scale(CESample$LogTotalIncome)
## create indictor variable for Rural
CESample$Rural = fastDummies::dummy_cols(CESample$UrbanRural)[,names(fastDummies::dummy_cols(CESample$UrbanRural))
== ".data_2"]
```

```
## create indicator variables for Black (2), Native American (3),
## Asian (4), Pacific Islander (5), and Multi-race (6)
CESample$Race_Black = fastDummies::dummy_cols(CESample$Race)[,names(fastDummies::dummy_cols(CESample$Race))
CESample$Race_NA = fastDummies::dummy_cols(CESample$Race)[,names(fastDummies::dummy_cols(CESample$Race))
CESample$Race_Asian = fastDummies::dummy_cols(CESample$Race)[,names(fastDummies::dummy_cols(CESample$Race))
CESample$Race_PI = fastDummies::dummy_cols(CESample$Race)[,names(fastDummies::dummy_cols(CESample$Race))
CESample$Race_M = fastDummies::dummy_cols(CESample$Race)[,names(fastDummies::dummy_cols(CESample$Race))
```

```

modelString <- "
model {
  ## sampling
  for (i in 1:N){
    y[i] ~ dnorm(beta0 + beta1*x_exp[i] + beta2*x_rural[i] +
      beta3*x_race_B[i] + beta4*x_race_N[i] +
      beta5*x_race_A[i] + beta6*x_race_P[i] +
      beta7*x_race_M[i], invsigma2)
  }
  ## priors
  beta0 ~ dnorm(mu0, g0)
  beta1 ~ dnorm(mu1, g1)
  beta2 ~ dnorm(mu2, g2)
  beta3 ~ dnorm(mu3, g3)
  beta4 ~ dnorm(mu4, g4)
  beta5 ~ dnorm(mu5, g5)
  beta6 ~ dnorm(mu6, g6)
  beta7 ~ dnorm(mu7, g7)
  invsigma2 ~ dgamma(a, b)
  sigma <- sqrt(pow(invsigma2, -1))
}
"

```

- Pass the data and hyperparameter values to JAGS:

```

y_income = as.vector(CESample$LogTotalIncome)
x_exp = as.vector(CESample$LogTotalExp)
x_rural = as.vector(CESample$Rural)
x_race_B = as.vector(CESample$Race_Black)
x_race_N = as.vector(CESample$Race_NA)
x_race_A = as.vector(CESample$Race_Asian)
x_race_P = as.vector(CESample$Race_PI)
x_race_M = as.vector(CESample$Race_M)
N = length(y_income) # Compute the number of observations

```

- Pass the data and hyperparameter values to JAGS:

```

the_data <- list("y" = y_income, "x_exp" = x_exp,
  "x_rural" = x_rural, "x_race_B" = x_race_B,
  "x_race_N" = x_race_N, "x_race_A" = x_race_A,
  "x_race_P" = x_race_P, "x_race_M" = x_race_M,
  "N" = N,
  "mu0" = 0, "g0" = 0.0001, "mu1" = 0, "g1" = 0.0001,
  "mu2" = 0, "g2" = 1, "mu3" = 0, "g3" = 1,
  "mu4" = 0, "g4" = 1, "mu5" = 0, "g5" = 1,
  "mu6" = 0, "g6" = 1, "mu7" = 0, "g7" = 1,
  "a" = 1, "b" = 1)

```

- Pass the data and hyperparameter values to JAGS:

```

initsfunction <- function(chain){
  .RNG.seed <- c(1,2)[chain]
  .RNG.name <- c("base::Super-Duper",
                 "base::Wichmann-Hill")[chain]
  return(list(.RNG.seed=.RNG.seed,
              .RNG.name=.RNG.name))
}

```

- Run the JAGS code for this model:

```

posterior_MLR <- run.jags(modelString,
  n.chains = 1,
  data = the_data,
  monitor = c("beta0", "beta1", "beta2",
              "beta3", "beta4", "beta5",
              "beta6", "beta7", "sigma"),
  adapt = 1000,
  burnin = 5000,
  sample = 5000,
  thin = 50,
  inits = initsfunction)

## Calling the simulation...
## Welcome to JAGS 4.3.0 on Tue Feb 25 14:14:51 2020
## JAGS is free software and comes with ABSOLUTELY NO WARRANTY
## Loading module: basemod: ok
## Loading module: bugs: ok
## . . Reading data file data.txt
## . Compiling model graph
##   Resolving undeclared variables
##   Allocating nodes
## Graph information:
##   Observed stochastic nodes: 994
##   Unobserved stochastic nodes: 9
##   Total graph size: 9984
## . Reading parameter file inits1.txt
## . Initializing model
## . Adaptation skipped: model is not in adaptive mode.
## . Updating 5000
## -----| 5000
## ***** 100%
## . . . . . Updating 250000
## -----| 250000
## ***** 100%
## . . . . Updating 0
## . Deleting model
## .
## Note: the model did not require adaptation
## Simulation complete. Reading coda files...
## Coda files loaded successfully
## Calculating summary statistics...

## Warning: Convergence cannot be assessed with only 1 chain

```

```
## Finished running the simulation
```

JAGS output for the MLR model

```
summary(posterior_MLR)
```

##	Lower95	Median	Upper95	Mean	SD	Mode
## beta0	3.669740	4.28610000	4.9266900	4.2946630	0.32414868	NA
## beta1	0.647560	0.72123550	0.7888190	0.7206641	0.03640298	NA
## beta2	-0.336150	-0.05379105	0.2074630	-0.0543172	0.13912170	NA
## beta3	-0.348372	-0.16199450	0.0359885	-0.1631919	0.09822631	NA
## beta4	-1.264630	-0.58281600	0.0728679	-0.5837983	0.34284402	NA
## beta5	-0.149501	0.13428050	0.4534340	0.1331292	0.15589175	NA
## beta6	-1.039080	-0.33076200	0.4318230	-0.3323508	0.37281544	NA
## beta7	-0.941018	-0.49236050	-0.0524113	-0.4891195	0.22801659	NA
## sigma	0.913835	0.95479350	0.9986350	0.9551042	0.02162080	NA
##	MCerr	MC%ofSD	SSeff	AC.500	psrf	
## beta0	0.0097435585	3.0	1107	1.657507e-02	NA	
## beta1	0.0010948661	3.0	1105	1.915412e-02	NA	
## beta2	0.0019674779	1.4	5000	3.204546e-03	NA	
## beta3	0.0013819565	1.4	5052	1.890181e-03	NA	
## beta4	0.0048485466	1.4	5000	5.349766e-03	NA	
## beta5	0.0022896118	1.5	4636	3.068741e-02	NA	
## beta6	0.0052724065	1.4	5000	9.152724e-03	NA	
## beta7	0.0032246415	1.4	5000	1.546961e-02	NA	
## sigma	0.0003057642	1.4	5000	-2.900956e-05	NA	

```
post_MLR <- as.mcmc(posterior_MLR)
```

```
synthesize <- function(X, index, n){  
  mean_Y <- post_MLR[index, "beta0"] + X$y_income * post_MLR[index, "beta1"] + X$x_rural * post_MLR[index, "beta2"]  
  synthetic_Y <- rnorm(n, mean_Y, post_MLR[index, "sigma"])  
  data.frame(X$y_income, synthetic_Y)  
}
```

```
n <- dim(CESample)[1]  
params <- data.frame(y_income, x_rural, x_race_B, x_race_N, x_race_A, x_race_P, x_race_M)  
synthetic_one <- synthesize(params, 1, n)  
names(synthetic_one) <- c("LogIncome_org", "LogIncome_syn")
```

```
m <- 20  
synthetic_m <- vector("list", m)  
for (l in 1:m){  
  params <- data.frame(y_income, x_rural, x_race_B, x_race_N, x_race_A, x_race_P, x_race_M)  
  synthetic_i <- synthesize(params, 4980+l, n)  
  names(synthetic_i) <- c("LogIncome_org", "LogIncome_syn")  
  synthetic_m[[l]] <- synthetic_i  
}
```

Here I write a function to calculate analysis specific utility measures, which I will run on each synthetic data set.

```

utilitymeasure<- function(list_i){
  exp<-mean(list_i$LogIncome_syn)
  med<-median(list_i$LogIncome_syn)
  stand<-sd(list_i$LogIncome_syn)
  pointEstAnal<- lm(CESample$LogTotalExp ~ list_i$LogIncome_syn)
  pointEst<-pointEstAnal$coefficients[1]
  unitInc<-pointEstAnal$coefficients[2]
  data.frame(exp,med,stand,pointEst,unitInc)
}

```

ASUM stands for Analysis Specific Utility Measures.

```

asum_m<- data.frame(utilitymeasure(synthetic_m[[1]]))
names(asum_m)<-c("mean", "median", "standard_dev", "point_estimate", "unit_increase")
if(m>1){
  for (j in 2:m){
    asum_i<-utilitymeasure(synthetic_m[[j]])
    names(asum_i)<-c("mean", "median", "standard_dev", "point_estimate", "unit_increase")
    asum_m<-bind_rows(asum_m,asum_i)
  }
}
asum_m

```

##	mean	median	standard_dev	point_estimate	unit_increase
## 1	11.84910	11.83972	1.305444	5.932063	0.2406901
## 2	11.88153	11.93562	1.280932	5.976175	0.2363202
## 3	11.94784	11.99617	1.324023	5.753981	0.2536058
## 4	11.97089	12.01812	1.320689	5.583272	0.2673778
## 5	12.00438	12.11602	1.297229	5.666384	0.2597083
## 6	11.89179	11.91895	1.269411	5.825632	0.2487758
## 7	11.86799	11.85585	1.268448	5.498784	0.2768150
## 8	11.94576	12.01081	1.271905	5.722889	0.2562526
## 9	11.90847	11.92977	1.318916	5.764561	0.2535558
## 10	11.84804	11.91408	1.232910	5.726666	0.2580473
## 11	11.85281	11.90632	1.221803	5.454562	0.2809006
## 12	11.93360	12.02291	1.257350	5.574590	0.2689408
## 13	11.84667	11.88360	1.301429	6.108460	0.2258493
## 14	11.97612	12.02908	1.288296	5.392621	0.2831803
## 15	11.89510	11.92999	1.331687	5.611213	0.2667325
## 16	11.85142	11.87861	1.281479	5.749847	0.2560180
## 17	11.96110	11.97159	1.255948	5.818081	0.2479657
## 18	11.92738	11.91579	1.283672	5.616151	0.2655965
## 19	11.93186	11.97143	1.260838	5.540059	0.2718740
## 20	12.00961	12.10996	1.316495	5.795065	0.2488804

Here I create the calcQ function which calculates the approximation for mean and variance of all 20 synthesized data sets.

```

calcQ<-function(list_q, list_u, m){
  qm_bar<-sum(list_q)/m
  bm<-0
  for(i in 1:m){

```

```

    bm = bm + (list_q[i]-qm_bar)^2/(m-1)
  }
  um_bar<-sum(list_u)/m
  Tp<-bm/m+um_bar
  return(c(qm_bar, Tp))
}
inferences<- calcQ(asum_m$mean, (asum_m$standard_dev)^2,m)
inferences<-data.frame(inferences[1], inferences[2])
names(inferences)<-c("mean", "variance")
inferences

```

```

##          mean variance
## 1 11.91507 1.650822

```