

# MLEXPRESSO: differential expression and methylation analysis

Case study using RTCGA data

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## 1 Package (Abstract?)

It is considered that the result of increased methylation is decreased gene expression. While, recent studies suggest that the relationship between methylation and expression is more complex than was previously thought. The package **MLEXPRESSO** provides methods to test for differential expression and methylation by use of the negative binomial distribution and t-test. Additionally **MLEXPRESSO** allows to visualize results in a simple way.

## 2 Standard Workflow

In this vignette we will work with the data sets containing information about gene expression and methylation for patients with breast cancer. We will analyze differences in methylation and expression for patients with different subtypes of BRCA cancer.

### 2.1 Methylation

#### 2.1.1 BRCA\_methylation\_chr17 data set

In this section, we will work with the methylation level data from TCGA database. Package contains **BRCA\_methylation\_chr17** dataset. **BRCA\_methylation\_chr17** contains information about methylation of CpG islands for patients with breast cancer. Rows of this data set correspond to patients, more precisely, to samples taken from patients. First column **SUBTYPE** corresponds to a subtype of BRCA cancer, next columns correspond to CpG islands. Values inside the table indicate the methylation level of CpG island for specified sample.

```
library(MLEXPRESSO)
```

```
## Setting options('download.file.method.GEOquery'='auto')
## Setting options('GEOquery.inmemory.gpl'=FALSE)
## Warning: namespace 'bit64' is not available and has been replaced
## by .GlobalEnv when processing object 'silent'

## Warning: namespace 'bit64' is not available and has been replaced
## by .GlobalEnv when processing object 'silent'

## No methods found in "genoset" for requests: toGenomeOrder
##
```

```
library(MLEXPRESSodata)
```

```
head(BRCA_methylation_chr17)[1:5,1:4]
```

```
##                               SUBTYPE cg00021527 cg00031162  cg00032227
## TCGA-A1-A0SD-01A-11D-A112-05      LumA 0.03781858  0.7910348 0.006391233
## TCGA-A2-A04N-01A-11D-A112-05      LumA 0.01437552  0.7359370 0.008752293
## TCGA-A2-A04P-01A-31D-A032-05      Basal 0.01360124  0.6967802 0.009442039
## TCGA-A2-A04Q-01A-21D-A032-05      Basal 0.01525656  0.5341244 0.014674247
## TCGA-A2-A04T-01A-21D-A032-05      Basal 0.01167384  0.7378100 0.012251559
```

In this analysis we would like to find genes with different methylation and expression. At first we need to use function `map_to_gene`, which generates new data frame with CpG islands mapped to genes.

```
BRCA_methylation_gen <- aggregate_probes(BRCA_methylation_chr17[, -1])
head(BRCA_methylation_gen)[1:5, 1:4]
```

```
##                               AANAT  AARSD1  AATF  AATK
## TCGA-A1-A0SD-01A-11D-A112-05 0.7148533 0.8625816 0.24294092 0.7835302
## TCGA-A2-A04N-01A-11D-A112-05 0.5850106 0.8355825 0.21367129 0.8466190
## TCGA-A2-A04P-01A-31D-A032-05 0.4495537 0.8786166 0.03277413 0.3417919
## TCGA-A2-A04Q-01A-21D-A032-05 0.7120650 0.8819490 0.03460160 0.7264985
## TCGA-A2-A04T-01A-21D-A032-05 0.6010397 0.7739978 0.02501599 0.6276399
```

In this case we have two conditions, connected with subtypes of breast cancer.

Before we go to the testing, we need to define condition values for each sample. We would like to test for differences between LumA subtype and other subtypes of breast cancer, so we create a vector, which each element corresponds to a sample. Our division into this two groups relies on numbers of occurrences of each subtype. The LumA subtype is the most common, in case of breast cancer.

```
condition_met <- ifelse(BRCA_methylation_chr17$SUBTYPE=="LumA", "LumA", "other")
head(condition_met, 8)
```

```
## [1] "LumA" "LumA" "other" "other" "other" "other" "LumA" "other"
```

## 2.1.2 Testing

```
res_met <- calculate_test(BRCA_methylation_chr17[, -1], condition_met, test="ttest")
head(res_met)
```

```
##           id      mean log2.fold      pval      padj
## cg10275770 cg10275770 0.3330801 -0.1515132 3.795188e-17 5.556155e-14
```

```
## cg06144905 cg06144905 0.4111616 0.1738623 9.908386e-14 7.252939e-11
## cg04049033 cg04049033 0.6462602 -0.1027159 1.513031e-13 7.383589e-11
## cg12045829 cg12045829 0.2485025 -0.1341285 5.910592e-12 2.163277e-09
## cg02473123 cg02473123 0.8127112 0.0982269 1.653082e-11 4.840224e-09
## cg05246522 cg05246522 0.5549808 0.1997340 2.069380e-11 5.049286e-09
```

## 2.2 Expression

### 2.2.1 BRCA\_mRNAseq\_chr17 data set

Data set `BRCA_mRNAseq_chr17` contains information about gene expression. This data set contains per-gene read counts computed for genes for 736 patients with breast cancer. Rows of this data set correspond to samples taken from patients. First column `SUBTYPE` corresponds to a subtype of BRCA cancer, next columns correspond to genes.

```
BRCA_mRNAseq_chr17[1:5,1:5]
```

```
##                SUBTYPE AANAT AARSD1 AATF AATK
## TCGA-A1-AOSB-01A-11R-A144-07 Normal    9  2354 2870 317
## TCGA-A1-AOSD-01A-11R-A115-07 LumA      2  1846 5656 312
## TCGA-A1-AOSE-01A-11R-A084-07 LumA     11  3391 9522 736
## TCGA-A1-AOSF-01A-11R-A144-07 LumA      0  2169 4625 169
## TCGA-A1-AOSG-01A-11R-A144-07 LumA      1  2273 3473  92
```

In our example we will test for differential expression between groups with LumA breast cancer subtype and other subtypes of that cancer. Again we will use vector `conditions`, which consist of two values corresponds to subtype of breast cancer: LumA and other.

```
condition_exp <- ifelse(BRCA_mRNAseq_chr17$SUBTYPE=="LumA", "LumA", "other")
head(condition_exp, 8)
```

```
## [1] "other" "LumA" "LumA" "LumA" "LumA" "LumA" "other" "LumA"
```

### 2.2.2 Testing

```
res_exp <- calculate_test(BRCA_mRNAseq_chr17[, -1], condition_exp, test="ttest")
head(res_exp)
```

```
##      id      mean log2.fold      pval      padj
## AURKB AURKB 1485.01  1784.844 3.890262e-10 2.839891e-07
## BIRC5 BIRC5 4448.76  4701.273 3.134210e-09 1.143987e-06
## MAPT  MAPT 13092.44 -17598.929 7.177688e-09 1.746571e-06
## SPAG5 SPAG5 3938.81  3174.993 2.597013e-08 4.739548e-06
## CBX2  CBX2 2574.48  3664.093 3.357756e-08 4.902324e-06
## KPNA2 KPNA2 19433.77 14880.016 5.242496e-08 6.378370e-06
```

## 2.3 Comparing test results

```
genes_comparison <- calculate_comparison_table(BRCA_mRNAseq_chr17[, -1], BRCA_methylation_gen,
                                              condition_exp, condition_met, test1="nbinom2", test2="ttest")
head(genes_comparison)
```

```
##      id nbinom2.log2.fold nbinom2.pval ttest.log2.fold  ttest.pval
## 1  AANAT      0.36342624  0.22109578    0.07237890 1.026904e-05
## 2  AARSD1     -0.07435954  0.45027777    0.01114861 5.052981e-01
## 3   AATF      0.13308688  0.18597163    0.01360521 2.136847e-01
## 4   AATK      0.02308606  0.92313513    0.06619897 1.427292e-05
## 5  ABCA5     -0.07034249  0.73314806    0.00510806 6.353608e-01
## 6  ABCA6     -0.44685708  0.08266088   -0.03402781 4.393248e-02
##      genes_rank
## 1 0.001506798
## 2 0.476995295
## 3 0.199347171
## 4 0.003629853
## 5 0.682505353
## 6 0.060261904
```

## 2.4 Choosing most different genes

Sorting ...

```
genes_comparison_sorted <- genes_comparison[order(genes_comparison$ttest.pval), ]
head(genes_comparison_sorted)
```

```
##      id nbinom2.log2.fold nbinom2.pval ttest.log2.fold  ttest.pval
## 302  ICAM2      0.2077055 2.102511e-01   -0.15151320 3.754116e-17
## 519   RILP      0.3139026 5.252448e-02   -0.05073691 2.575168e-13
## 466  PIPOX      0.3589979 4.371416e-02    0.11505558 5.360053e-12
## 652 TNFSF12    -0.4357410 3.486764e-04   -0.13412855 5.867083e-12
## 133   CD7      1.3728093 9.099866e-07    0.09822690 1.641919e-11
## 341   KSR1      0.4261993 1.658465e-02    0.19973400 2.054467e-11
##      genes_rank
## 302 2.809461e-09
## 519 1.163011e-07
## 466 4.840560e-07
## 652 4.522956e-08
## 133 3.865391e-09
## 341 5.837174e-07
```

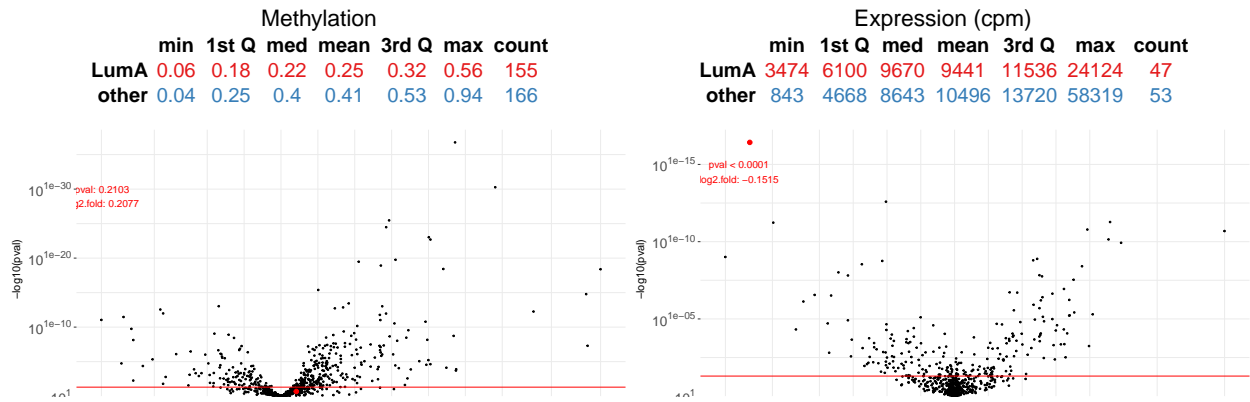
ICAM2!

## 2.5 Visualization

Visualizing chosen gene - IGFALS.

```
test_exp <- genes_comparison[,c(1,2,3)]
test_met <- genes_comparison[,c(1,4,5)]
```

```
plot_volcanoes(condition_exp, condition_met,
               BRCA_mRNAseq_chr17[, -1], BRCA_methylation_chr17[, -1],
               "ICAM2",
               test_exp, test_met,
               values=TRUE)
```



Note that `plot_gene` methylation require data frame with cpg islands, not genes.

```
plot_gene(condition_exp, condition_met, BRCA_mRNAseq_chr17, BRCA_methylation_chr17, "ICAM2")
```

