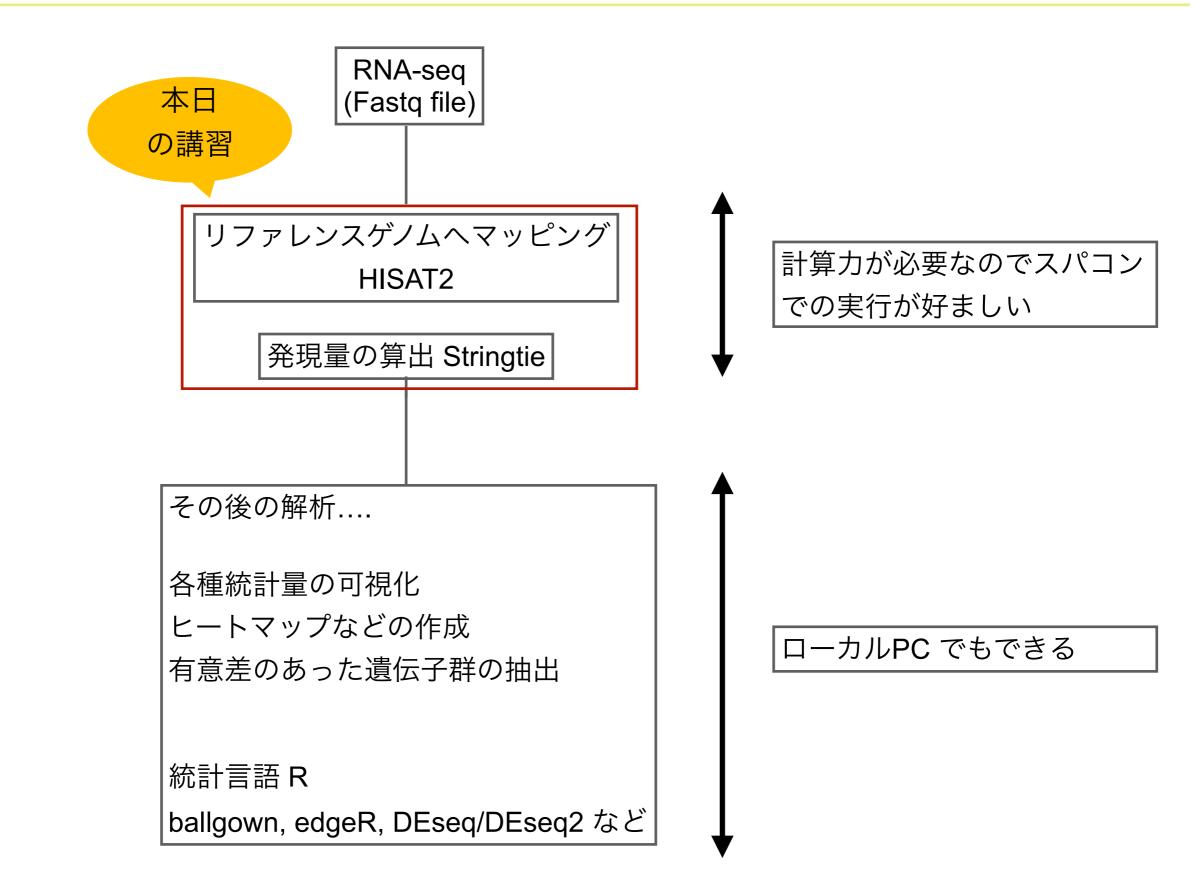
2023/10/30

RNA-seq 解析

国立遺伝学研究所 望月孝子

本日の講習の流れ RNA-seq 発現量解析



配布データのコピー

ホームディレクトリ に移動

\$ cd

ホームディレクトリ に20231030 というファイル/ディレクトリがないかを確認。

\$ ls 20231030

ls: 20231030 にアクセスできません: そのようなファイルやディレクトリはありません

あれば、講習時間だけファイル/ディレクトリ 名前を一時的に変更してください。

\$ mv 20231030 20231030_tmp

講習データをコピー

\$ cp -r /usr/local/shared_data/lecture/20231030 .

本日の講習はこちらのディレクトリで

\$ cd 20231030

配布データの確認 (1)

```
$ ls -al
合計 24

drwxr-xr-x 6 koshu3 koshu 4096 10月 16 15:30 .
drwxr-x--- 13 koshu3 koshu 4096 10月 13 14:50 ..

drwxr-xr-x 5 koshu3 koshu 4096 10月 16 15:28 outputs
drwxr-xr-x 2 koshu3 koshu 4096 10月 16 10:53 reads
drwxr-xr-x 2 koshu3 koshu 4096 10月 16 11:47 reference
drwxr-xr-x 2 koshu3 koshu 4096 10月 17 2022 scripts
```

事前に実行した 結果ファイル

> 講習用リード ファイル

リファレンス ファイル

実行スクリプト

それぞれのディレクトリ の中身は

```
$ ls outputs
$ ls outputs/hisat2_index
$ ls outputs/hisat2
$ ls outputs/stringtie
$ ls reads
$ ls reference
$ ls scripts
```

配布データの確認 (2)

配布データの構成 20231030 outputs #解析結果 hisat2 hisat2 index stringtie |---- reads # リードファイル格納用 SRR453566_1.fastq.gz SRR453566_2.fastq.gz SRR453569 1.fastq.gz SRR453569_2.fastq.gz |---- reference #リファレンスファイル s288c.fa s288c.gff |---- scripts # スクリプト hisat2.sh # condaにて実装 hisat2 index.sh # condaにて実装 stringtie.sh # condaにて実装 _hisat2_singularity.sh # singularityにて実装 _hisat2_index_singularity.sh # singularityにて実装 stringtie singularity.sh # singularityにて実装

Conda activate

\$ conda activate pags_rnaseq

講習用 RNA-seq データ

JOURNAL ARTICLE

A comprehensive comparison of RNA-Seq-based transcriptome analysis from reads to differential gene expression and cross-comparison with microarrays: a case study in *Saccharomyces cerevisiae* 3

Intawat Nookaew, Marta Papini, Natapol Pornputtapong, Gionata Scalcinati, Linn Fagerberg, Matthias Uhlén, Jens Nielsen ➤ Author Notes

Nucleic Acids Research, Volume 40, Issue 20, 1 November 2012, Pages 10084–10097, https://doi.org/10.1093/nar/gks804

Published: 08 September 2012 Article history ▼

Saccharomyces cerevisiae CEN.PK113-7D

バッチ培養 ケモスタット培養

SRR453566	SRR453569
SRR453567	SRR453570
SRR453568	SRR453571

Biological replicates 3回ずつ

\$ ls reads/

SRR453566_1.fastq.gz SRR453566_2.fastq.gz

SRR453569_1.fastq.gz SRR453569_2.fastq.gz

Paired-end データ

Paired-end データ

FASTQ フォーマット

4行で1配列の情報を表す。

```
$ zcat reads/SRR453566_1.fastq.gz | more
```

@SRR453566.1 HWI-ST167:4:1101:1597:1986/1

NAAAACTTTGGATGACTTCAACAACTATTCTTCTGAAATCAACAAAATATCACCAACTTCCGCCAACACAAAGTCTTACAGTGCAACAACAAGTGATGTTG

1行目: @ の後ろにその配列のID

2行目: 配列

3行目: + を記載する。(配列のID を記載してもしなくてもよい)

4行目: その配列のクオリティ値

クオリティ値はアスキーコードで表示 アスキー値 - 33 が クオリティ値

```
.....
    クオリティ値@の場合
                         64 - 33 = 31
!"#$%&'()*+,-./0123456789:;<=>?@ABCDEFGHIJKLMNOPQRSTUVWXYZ[\]^_`abcdefghijklmnopgrstuvwxyz{|}~
33
      59
                 104
                      126
  0.....9......40
        0......40......50
```

```
S - Sanger Phred+33, raw reads typically (0, 40)

X - Solexa Solexa+64, raw reads typically (-5, 40)

I - Illumina 1.3+ Phred+64, raw reads typically (0, 40)

J - Illumina 1.5+ Phred+64, raw reads typically (3, 41)

with 0=unused, 1=unused, 2=Read Segment Quality Control Indicator (bold)

(Note: See discussion above).

L - Illumina 1.8+ Phred+33, raw reads typically (0, 41)

N - Nanopore Phred+33, Duplex reads typically (0, 50)

P - PacBio Phred+33, HiFi reads typically (0, 93)
```

講習用 リファレンスファイル ゲノム配列 fasta

\$ ls reference/
s288c.fa s288c.gff

ゲノム配列

ファイルの中身を確認

\$ more reference/s288c.fa

FASTA ヘッダの出力

分の配列

```
$ grep ">" reference/s288c.fa
>NC_001133.9 Saccharomyces cerevisiae S288C chromosome I, complete sequence
>NC_001134.8 Saccharomyces cerevisiae S288C chromosome II, complete sequence
>NC_001135.5 Saccharomyces cerevisiae S288C chromosome III, complete sequence
>NC_001136.10 Saccharomyces cerevisiae S288C chromosome IV, complete sequence
>NC_001137.3 Saccharomyces cerevisiae S288C chromosome V, complete sequence
>NC_001138.5 Saccharomyces cerevisiae S288C chromosome VI, complete sequence
>NC 001139.9 Saccharomyces cerevisiae S288C chromosome VII, complete sequence
>NC_001140.6 Saccharomyces cerevisiae S288C chromosome VIII, complete sequence
>NC_001141.2 Saccharomyces cerevisiae S288C chromosome IX, complete sequence
>NC_001142.9 Saccharomyces cerevisiae S288C chromosome X, complete sequence
>NC_001143.9 Saccharomyces cerevisiae S288C chromosome XI, complete sequence
>NC_001144.5 Saccharomyces cerevisiae S288C chromosome XII, complete sequence
>NC_001145.3 Saccharomyces cerevisiae S288C chromosome XIII, complete sequence
>NC_001146.8 Saccharomyces cerevisiae S288C chromosome XIV, complete sequence
>NC_001147.6 Saccharomyces cerevisiae S288C chromosome XV, complete sequence
>NC 001148.4 Saccharomyces cerevisiae S288C chromosome XVI, complete sequence
```

講習用 リファレンスファイル アノテーションファイル

\$ ls reference/
s288c.fa s288c.gff

アノテー ションファイル

ファイルの中身を確認

```
$ more reference/s288c.gff
##qff-version 3
#!qff-spec-version 1.21
#!processor NCBI annotwriter
#!genome-build R64
#!genome-build-accession NCBI_Assembly:GCF_000146045.2
#!annotation-source SGD R64-3-1
##sequence-region NC_001133.9 1 230218
##species https://www.ncbi.nlm.nih.gov/Taxonomy/Browser/wwwtax.cgi?id=559292
NC_001133.9 RefSeq region 1 230218 . + . ID=NC_001133.9:1..230218;Dbxref=taxor
            RefSeq telomere 1 801 . - .
                                              ID=id-NC_001133.9:1..801;Dbxref=SGD:S0
NC_001133.9
NC_001133.9
                    origin_of_replication 707 776 . + . ID=id-NC_001133.9:707...
            RefSeq
NC_001133.9
            RefSeq
                    gene
                          1807 2169 .
                                              ID=gene-YAL068C;Dbxref=GeneID:851229;N
NC_001133.9
            RefSeq
                    mRNA 1807 2169 .
                                              ID=rna-NM_001180043.1; Parent=gene-YAL@
NC_001133.9
            RefSeq
                          1807 2169 .
                                              ID=exon-NM_001180043.1-1; Parent=rna-NN
                    exon
                                             ID=cds-NP_009332.1; Parent=rna-NM_001186
NC 001133.9
            RefSeq
                    CDS 1807
                               2169 .
                                       - 0
```

GFF フォーマット

遺伝子アノテーションのフォーマット

```
##gff-version 3
#!gff-spec-version 1.21
#!processor NCBI annotwriter
#!genome-build R64
#!genome-build-accession NCBI Assembly:GCF 000146045.2
#!annotation-source SGD R64-2-1
##sequence-region NC 001133.9 1 230218
##species https://www.ncbi.nlm.nih.gov/Taxonomy/Browser/wwwtax.cgi?id=559292
NC 001133.9 RefSeqgene 1807 2169 .
                                                     ID=gene0;Dbxref=GeneID:851229;Name=PAU8;end range=2169,.;gbkey=Gene;gene=PAU8;
NC 001133.9 RefSeqmRNA 1807 2169 .
                                                     ID=rna0;Parent=gene0;Dbxref=GeneID:851229,Genbank:NM 001180043.1;Name=NM 0011
NC 001133.9 RefSeq exon 1807 2169 .
                                                     ID=id3; Parent=rna0; Dbxref=GeneID:851229, Genbank: NM 001180043.1; end range=2169..; gl
NC_001133.9 RefSeqCDS 1807 2169 .
                                                     NC 001133.9 RefSeqgene 2480 2707 .
                                                     ID=gene1;Dbxref=GeneID:1466426;Name=YAL067W-A;end range=2707,.;gbkey=Gene;gene
NC 001133.9 RefSeqmRNA 2480 2707 .
                                                     ID=rna1;Parent=gene1;Dbxref=GeneID:1466426,Genbank:NM 001184582.1;Name=NM 001
NC 001133.9 RefSeq exon 2480 2707 .
                                                     ID=id4;Parent=rna1;Dbxref=GeneID:1466426,Genbank:NM 001184582.1;end range=2707,.;
NC 001133.9 RefSeqCDS 2480 2707 .
                                                     ID=cds1;Parent=rna1;Dbxref=SGD:S000028593,GeneID:1466426,Genbank:NP 878038.1;Nat
NC 001133.9 RefSeqgene 7235 9016 .
                                                     ID=gene2;Dbxref=GeneID:851230;Name=SEO1;end range=9016,.;gbkey=Gene;gene=SEO1;
NC 001133.9 RefSeqmRNA 7235 9016 .
                                                     ID=rna2;Parent=gene2;Dbxref=GeneID:851230,Genbank:NM 001178208.1;Name=NM 0011
NC 001133.9 RefSeq exon 7235 9016 .
                                                     ID=id5;Parent=rna2;Dbxref=GeneID:851230,Genbank:NM 001178208.1;end range=9016,.;gl
                                                     ID=cds2;Parent=rna2;Dbxref=SGD:S000000062,GeneID:851230,Genbank:NP 009333.1;Nam
NC 001133.9 RefSeqCDS 7235 9016 .
```

タブ区切りフォーマット。値がない場合は、"."が設定される。

7. strand: +(forward)、-(reverse)または!!

```
    seqname:染色体 or スキャフォールドの名前
    source:アノテーションを生成したプログラムまたはデータソースの名前
    feature:フィーチャータイプ (mRNA, gene, exon, CDS ....)
    start:スタートポジション (1bp ~)
    end:エンドポジション (1bp ~)
    score:スコア
```

8. frame: 翻訳フレーム (0, 1, 2) 9. attribute: 追加情報。セミコロンで区切られたタグと値のペアのリスト。

リファレンスゲノムへリードをマッピング

ステップ

1. リファレンスゲノムのインデックスを作成

hisat2_index.sh

2. リードをリファレンスゲノムへマッピング

hisat2.sh 2サンプル分をアレイジョブで同時実行

スクリプト

```
$ ls scripts/hisat2*
scripts/hisat2.sh scripts/hisat2_index.sh
scripts/hisat2_index_singularity.sh scripts/hisat2_singularity.sh
```

マッピング用インデックスの作成

スクリプトの確認

```
$ more scripts/hisat2_index.sh
#$ -S /bin/bash
#$ -pe def_slot 2
#$ -cwd
#$ -l mem_req=10G,s_vmem=10G
```

conda activate pags_rnaseq

GENOME=./reference/s288c.fa
INDEX=./reference/s288c.fa

hisat2-build \$GENOME \$INDEX

conda 仮想環境 の指定

リファレンスゲノムの インデックス化

qsub コマンドのオプション

- -S 使用するインタプリタのパス
- -pe def_slot 1 ジョブスロット数
- -cwd ホームディレクトリではなく、qsubコマンド実行時のディレクトリでジョブ を実行。標準出力 / 標準エラー出力ファイルは、qsubコマンド実行時のディレクトリに出力。
- -l 主にキューの選択、メモリ利用上限の変更に使う

mem_req: 使用するメモリの量を宣言する。(ジョブ管理システムUGEのジョブリソース管理に対する宣言)

s_vmem: ジョブが使用可能な仮想メモリの上限値。 (OS に対する宣言)

キューの指定: Thin ノードへの投入は、キューの指定は不要。

```
$ hisat2-build -h
HISAT2 version 2.2.1 by Daehwan Kim (infphilo@gmail.com, http://www.ccb.jhu.edu/people/infphilo)
Usage: hisat2-build [options]* <reference_in> <ht2_index_base>
    reference in
                            comma-separated list of files with ref sequences
                            write ht2 data to files with this dir/basename
   hisat2 index base
Options:
                            reference sequences given on cmd line (as
    -c
                            <reference in>)
                            force generated index to be 'large', even if ref
    --large-index
                            has fewer than 4 billion nucleotides
    -a/--noauto
                            disable automatic -p/--bmax/--dcv memory-fitting
                            number of threads
    -p <int>
    --bmax <int>
                            max bucket sz for blockwise suffix-array builder
                            max bucket sz as divisor of ref len (default: 4)
    --bmaxdivn <int>
    --dcv <int>
                            diff-cover period for blockwise (default: 1024)
                            disable diff-cover (algorithm becomes quadratic)
    --nodc
                            don't build .3/.4.ht2 (packed reference) portion
    -r/--noref
    -3/--justref
                            just build .3/.4.ht2 (packed reference) portion
    -o/--offrate <int>
                            SA is sampled every 2^offRate BWT chars (default: 5)
    -t/--ftabchars <int>
                            # of chars consumed in initial lookup (default: 10)
                            SA (local) is sampled every 2^offRate BWT chars (default: 3)
    --localoffrate <int>
                           # of chars consumed in initial lookup in a local index (default: 6)
    --localftabchars <int>
                            SNP file name
    --snp <path>
                            haplotype file name
    --haplotype <path>
                            Splice site file name
    --ss <path>
    --exon <path>
                            Exon file name
    --repeat-ref <path>
                            Repeat reference file name
                            Repeat information file name
    --repeat-info <path>
    --repeat-snp <path>
                            Repeat snp file name
    --repeat-haplotype <path>
                                Repeat haplotype file name
    --seed <int>
                            seed for random number generator
    -q/--quiet
                            disable verbose output (for debugging)
    -h/--help
                            print detailed description of tool and its options
                            print this usage message
    --usage
    --version
                            print version information and quit
```

実行

講習中はqsub を実行しないでください。

\$ qsub scripts/hisat2_index.sh

Your job 24706258 ("hisat2_index.sh") has been submitted



ステータスの確認

<pre>\$ qstat job-ID</pre>	prior	name	user	state	submit/start at	queue	jclass slots ja-task-ID
24704406 24706258		QLOGIN hisat2_ind	koshu3 koshu3		10/16/2023 10:42:12 10/16/2023 11:47:09	login.q@at138	1 2

実行 待機



<pre>\$ qstat job-ID</pre>	prior	name	user	state	submit/start at	queue	jclass slots ja-task-ID
24704406 24706258		QLOGIN hisat2_ind	koshu3 koshu3		10/16/2023 10:42:12 10/16/2023 11:47:12	5	1 2

実行 中



<pre>\$ qstat job-ID</pre>	prior	name	user	state	submit/start at	queue	jclass slots ja-task-ID
24704406	0.25026	QLOGIN	koshu3	r	10/16/2023 10:42:12	login.q@at138	1

ジョブが終了すると該当ジョブID が表示されなくなる。

```
$ ls -al
合計 32
drwxr-xr-x 6 koshu3 koshu 4096 10月 16 11:58 .
drwxr-x--- 13 koshu3 koshu 4096 10月 13 14:50 ..
-rw-r--r-- 1 koshu3 koshu 2262 10月 16 11:47 hisat2_index.sh.e24706258
-rw-r--r-- 1 koshu3 koshu 3919 10月 16 11:47 hisat2_index.sh.o24706258
-rw-r--r-- 1 koshu3 koshu 0 10月 16 11:47 hisat2_index.sh.pe24706258
-rw-r--r-- 1 koshu3 koshu 0 10月 16 11:47 hisat2_index.sh.pe24706258
drwxr-xr-x 2 koshu3 koshu 4096 10月 16 11:58 outputs
drwxr-xr-x 2 koshu3 koshu 4096 10月 16 10:53 reads
drwxr-xr-x 2 koshu3 koshu 4096 10月 16 11:47 reference
drwxr-xr-x 2 koshu3 koshu 4096 10月 16 11:47 reference
drwxr-xr-x 2 koshu3 koshu 4096 10月 17 2022 scripts
```

インデックスが作成された

```
$ ls -al reference/
合計 46228
drwxr-xr-x 2 koshu3 koshu
                             4096 10月 16 11:47 .
drwxr-xr-x 6 koshu3 koshu
                             4096 10月 16 11:58 ...
-rw-r--r-- 1 koshu3 koshu 12245035 10月 17 2022 s288c.fa
-rw-r--r-- 1 koshu3 koshu 8219756 10月 16 11:47 s288c.fa.1.ht2
-rw-r--r-- 1 koshu3 koshu 3017836 10月 16 11:47 s288c.fa.2.ht2
-rw-r--r-- 1 koshu3 koshu
                              152 10月 16 11:47 s288c.fa.3.ht2
-rw-r--r-- 1 koshu3 koshu
                         3017832 10月 16 11:47 s288c.fa.4.ht2
                         5357645 10月 16 11:47 s288c.fa.5.ht2
-rw-r--r-- 1 koshu3 koshu
-rw-r--r-- 1 koshu3 koshu 3071004 10月 16 11:47 s288c.fa.6.ht2
                               12 10月 16 11:47 s288c.fa.7.ht2
-rw-r--r-- 1 koshu3 koshu
                                8 10月 16 11:47 s288c.fa.8.ht2
-rw-r--r-- 1 koshu3 koshu
-rw-r--r-- 1 koshu3 koshu 12377219 10月 17 2022 s288c.gff
```

ログの確認

```
Settings:
    Output files: "./reference/s288c.fa.*.ht2"
    Line rate: 6 (line is 64 bytes)
    Lines per side: 1 (side is 64 bytes)
    Offset rate: 4 (one in 16)
    FTable chars: 10
    .
    .
    Total time for call to driver() for forward index: 00:00
```

```
$ more hisat2_index.sh.o24706258
Building DifferenceCoverSample
Building sPrime
Building sPrimeOrder
V-Sorting samples
...
Returning block of 1908813 for bucket 7
```

\$ more hisat2_index.sh.e24706258

事前実行した結果で確認

```
$ ls outputs/hisat2_index/
$ more outputs/hisat2_index/hisat2_index.sh.e24706258
$ more outputs/hisat2_index/hisat2_index.sh.o24706258
```

```
$ more scripts/hisat2.sh
#$ -S /bin/bash
                               アレイジョブを
#$ -pe def_slot 4
#$ -cwd
                                    指定
#$ -t 1-2:1
#$ -l mem_req=8G,s_vmem=8G
conda activate pags_rnaseq
# Batch culture: SRR453566
# chemostat: SRR453569
ACESSIONS=(453566 453569)
                                   アレイジョブ
no=`expr ${SGE TASK ID} - 1`
                                    のタスクID
NUM=${ACESSIONS[${no}]}
PREFIX=SRR${NUM}
# read file
DIR=./reads/
QUERY1_1=${DIR}${PREFIX}"_1.fastq.gz"
                                             —dta: reports alignments tailored
QUERY1 2=${DIR}${PREFIX}" 2.fastq.qz"
                                            for transcript assemblers
hisat2 -p ${NSLOTS} -x reference/s288c.fa --dta \
       -1 ${QUERY1 1} -2 ${QUERY1 2} \
       -S ${PREFIX}.sam
# convert sam to bam
# sort by position
samtools sort -@ ${NSLOTS} ${PREFIX}.sam -o ${PREFIX}.sorted.bam
```

HISAT2 オプションの確認

```
$ hisat2 --help
HISAT2 version 2.2.1 by Daehwan Kim (infphilo@gmail.com, www.ccb.jhu.edu/people/infphilo)
Usage:
  hisat2 [options]* -x <ht2-idx> \{-1 <m1> -2 <m2> | -U <r>} [-S <sam>]
  <ht2-idx> Index filename prefix (minus trailing .X.ht2).
             Files with #1 mates, paired with files in <m2>.
  <m1>
             Could be gzip'ed (extension: .gz) or bzip2'ed (extension: .bz2).
             Files with #2 mates, paired with files in <m1>.
  <m2>
             Could be gzip'ed (extension: .gz) or bzip2'ed (extension: .bz2).
             Files with unpaired reads.
  <r>
             Could be gzip'ed (extension: .gz) or bzip2'ed (extension: .bz2).
             File for SAM output (default: stdout)
  <sam>
  <m1>, <m2>, <r> can be comma-separated lists (no whitespace) and can be
  specified many times. E.g. '-U file1.fq,file2.fq -U file3.fq'.
Options (defaults in parentheses):
 Input:
                     query input files are FASTQ .fq/.fastq (default)
  -q
                     query input files are in Illumina's gseg format
  --qseq
                     query input files are (multi-)FASTA .fa/.mfa
  -f
                     query input files are raw one-sequence-per-line
  -r
                     <m1>, <m2>, <r> are sequences themselves, not files
  -c
                     skip the first <int> reads/pairs in the input (none)
  -s/--skip <int>
                     stop after first <int> reads/pairs (no limit)
  -u/--upto <int>
                     trim <int> bases from 5'/left end of reads (0)
  -5/--trim5 <int>
                     trim <int> bases from 3'/right end of reads (0)
  -3/--trim3 <int>
                     qualities are Phred+33 (default)
  --phred33
                     qualities are Phred+64
  --phred64
  --int-quals
                     qualities encoded as space-delimited integers
 Presets:
                          Same as:
   --fast
                          --no-repeat-index
   --sensitive
                          --bowtie2-dp 1 -k 30 --score-min L,0,-0.5
   --very-sensitive
                          --bowtie2-dp 2 -k 50 --score-min L,0,-1
 Alignment:
  --bowtie2-dp <int> use Bowtie2's dynamic programming alignment algorithm (0) - 0: no dynamic programming, 1: conditional
dynamic programming, and 2: unconditional dynamic programming (slowest)
                     func for max # non-A/C/G/Ts permitted in aln (L,0,0.15)
  --n-ceil <func>
  --ignore-quals
                     treat all quality values as 30 on Phred scale (off)
                     do not align forward (original) version of read (off)
  --nofw
                     do not align reverse-complement version of read (off)
  --norc
  --no-reneat-index do not use reneat index
```

```
$ samtools sort --help
sort: unrecognized option '--help'
Usage: samtools sort [options...] [in.bam]
Options:
  -l INT
             Set compression level, from 0 (uncompressed) to 9 (best)
             Output uncompressed data (equivalent to -l 0)
  -u
  -m INT
             Set maximum memory per thread; suffix K/M/G recognized [768M]
             Use minimiser for clustering unaligned/unplaced reads
  -M
  -K INT
             Kmer size to use for minimiser [20]
             Sort by read name (not compatible with samtools index command)
  -n
             Sort by value of TAG. Uses position as secondary index (or read name if -n is set)
  -t TAG
            Write final output to FILE rather than standard output
  -o FILE
  -T PREFIX Write temporary files to PREFIX.nnnn.bam
      --no-PG
               Do not add a PG line
      --template-coordinate
               Sort by template-coordinate
      --input-fmt-option OPT[=VAL]
               Specify a single input file format option in the form
               of OPTION or OPTION=VALUE
  -0, --output-fmt FORMAT[,OPT[=VAL]]...
               Specify output format (SAM, BAM, CRAM)
      --output-fmt-option OPT[=VAL]
               Specify a single output file format option in the form
               of OPTION or OPTION=VALUE
      --reference FILE
               Reference sequence FASTA FILE [null]
  -@, --threads INT
               Number of additional threads to use [0]
      --write-index
               Automatically index the output files [off]
      --verbosity INT
               Set level of verbosity
```

SAM フォーマット/ BAM フォーマット

```
VN:1.0 S0:unsorted
@HD
      SN:NC 001133.9
@SQ
                         LN:230218
      SN:NC_001134.8
@SQ
                         LN:813184
                                            @ヘッダ行
      SN:NC 001135.5
@SQ
                         LN:316620
@SQ
      SN:NC 001136.10
                         LN:1531933
                                            HD: ヘッダ行 SAMフォーマットのバージョンなど
      SN:NC_001137.3
                         LN:576874
@SQ
      SN:NC 001138.5
                         LN:270161
@SQ
                                            SQ: リァレンスの情報
@SQ
      SN:NC 001139.9
                         LN:1090940
@SQ
      SN:NC 001140.6
                         LN:562643
                                            PG ツールの実行情報
@SQ
      SN:NC_001141.2
                         LN:439888
@SQ
      SN:NC 001142.9
                         LN:745751
      SN:NC 001143.9
@SQ
                         LN:666816
@SQ
      SN:NC 001144.5
                         LN:1078177
@SQ
      SN:NC_001145.3
                         LN:924431
@SQ
      SN:NC_001146.8
                         LN:784333
@SQ
      SN:NC 001147.6
                         LN:1091291
      SN:NC 001148.4
@SQ
                         LN:948066
      SN:NC_001224.1
@SQ
                         LN:85779
@PG ID:hisat2 PN:hisat2
                                      CL:"/home/koshu3/miniconda3/envs/pags_rnaseq/bin/hisat2-align-s --wrapper basic-0 -p 4 -x referen
                         VN:2.2.1
SRR453566.24 163
                  NC 001139.9 72751860
                                            69M
                                                         727620 203
                                                                     TTAATCAAG... =DFFFFHHHH... AS:i:0 XN:i:0 XM:i:0 XO:i:0 XG:i:0 NM:i:0 MD
SRR453566.22 99
                                                                      CAAAGCGTA... CCCFFFFFHG... AS:i:-6
                  NC 001142.9 50970560
                                            101M =
                                                         509740 136
                                                                                                            XN:i:0 XM:i:1 X0:i:0 XG:i:0 NM
SRR453566.22 147
                  NC 001142.9 50974060
                                                                     GGTATATTT.... @DA@:>>>@C.... AS:i:-4
                                                                                                            ZS:i:-7
                                            101M =
                                                         509705 –136
                                                                                                                        XN:i:0 XM:i:1 X0
SRR453566.23 99
                                                                     TTTTCTTCA.... @BCFFFFFHH.... AS:i:-6
                                                                                                            XN:i:0 XM:i:1 X0:i:0 XG:i:0 NM
                  NC 001134.8 67424060
                                            101M =
                                                         674286 131
                                                                                  ??>C>@5(@>.... AS:i:-4
                                                                     AACAAAAGC....
SRR453566.23 147
                  NC 001134.8 67428660
                                            85M
                                                         674240 -131
                                                                                                            ZS:i:-10
                                                                                                                        XN:i:0 XM:i:1 X0
QNAME
                                                                       SEQ
                                                                                   QUAL
            FLG
                    RNAME
                               POS MAPQ CIGAR RNEXT PNEXT TLEN
                                                                                                 optional fields
```

QNAME	リード名								
FLG	アラインメント情報。参考 <u>https://broadinstitute.github.io/picard/explain-flags.html</u>								
RNAME	マップされたリファレンス名								
POS	マップポジション	マップポジション							
MAPQ	マッピングスコア	マッピングスコア							
CIGAR	マッピングの状況 ex) M アライメントマッチ I リファレンスにインサーションあり など								
RNEXT	ペアエンドの場合、ペアのリード名(=: QNAME)。								
PNEXT	ペアエンドの場合、ペアのマップされた開始位置。								
TLEN	ペアエンドのリード間の距離。								
SEQ	FASTQ の塩基配列データ								
QUAL	FASTQ のクオリティデータ。	BAM は SAM をバイナリ形式にしたファイ川							

実行

```
$ qsub scripts/hisat2.sh
                                                                  講習中はqsub を実行しないでください。
Your job-array 24707186.1-2:1 ("hisat2.sh") has been submitted
$ qstat
iob-ID
                                    state submit/start at
                                                                           jclass slots ja-task-ID
          prior
                             user
                  name
                                                              queue
 24704406 0.25081 OLOGIN
                             koshu3 r
                                          10/16/2023 10:42:12 login.g@at138
 24707186 0.25068 hisat2.sh
                             koshu3 r
                                          10/16/2023 14:16:24 intel.g@it008
                                                                                      4 1
                                          10/16/2023 14:15:32
                                                                                      4 2
  24707186 0.25068 hisat2.sh
                             koshu3 qw
```

```
$ ls -al
合計 8310040
drwxr-xr-x 6 koshu3 koshu
                                4096 10月 16 14:19 .
                                4096 10月 13 14:50 ...
drwxr-x--- 13 koshu3 koshu
-rw-r--r-- 1 koshu3 koshu 4173908395 10月 16 14:17 SRR453566.sam
          1 koshu3 koshu 821523448 10月 16 14:18 SRR453566.sorted.bam
          1 koshu3 koshu 2922686962 10月 16 14:19 SRR453569.sam
          1 koshu3 koshu 591294623 10月 16 14:19 SRR453569.sorted.bam
                                 693 10月 16 14:18 hisat2.sh.e24707186.1
           1 koshu3 koshu
-rw-r--r--
                                 688 10月 16 14:19 hisat2.sh.e24707186.2
           1 koshu3 koshu
-rw-r--r--
                                   0 10月 16 14:16 hisat2.sh.o24707186.1
           1 koshu3 koshu
-rw-r--r--
-rw-r--r-- 1 koshu3 koshu
                                   0 10月 16 14:18 hisat2.sh.o24707186.2
          1 koshu3 koshu
                                   0 10月 16 14:16 hisat2.sh.pe24707186.1
-rw-r--r--
           1 koshu3 koshu
                                   0 10月 16 14:18 hisat2.sh.pe24707186.2
-rw-r--r--
           1 koshu3 koshu
                                   0 10月 16 14:16 hisat2.sh.po24707186.1
-rw-r--r--
          1 koshu3 koshu
                                   0 10月 16 14:18 hisat2.sh.po24707186.2
                                2262 10月 16 11:47 hisat2_index.sh.e24706258
          1 koshu3 koshu
                                3919 10月 16 11:47 hisat2_index.sh.o24706258
           1 koshu3 koshu
                                   0 10月 16 11:47 hisat2_index.sh.pe24706258
           1 koshu3 koshu
-rw-r--r--
-rw-r--r-- 1 koshu3 koshu
                                   0 10月 16 11:47 hisat2 index.sh.po24706258
drwxr-xr-x 2 koshu3 koshu
                                4096 10月 16 11:58 outputs
drwxr-xr-x 2 koshu3 koshu
                                4096 10月 16 10:53 reads
drwxr-xr-x 2 koshu3 koshu
                                4096 10月 16 11:47 reference
                                4096 10月 17 2022 scripts
drwxr-xr-x 2 koshu3 koshu
```

事前実行した結果で確認

\$ Is -al outputs/hisat2/

マッピング 実行結果 ログの確認

```
$ more hisat2.sh.e24707186.1
5725730 reads; of these:
                                                                  事前実行した結果で確認
 5725730 (100.00%) were paired; of these:
   1222756 (21.36%) aligned concordantly 0 times
                                                                  $ more outputs/hisat2/hisat2.sh.e16626062.1
   4258780 (74.38%) aligned concordantly exactly 1 time
   244194 (4.26%) aligned concordantly >1 times
    1222756 pairs aligned concordantly 0 times; of these:
      128491 (10.51%) aligned discordantly 1 time
   1094265 pairs aligned 0 times concordantly or discordantly; of these:
      2188530 mates make up the pairs; of these:
        1470694 (67.20%) aligned 0 times
        662896 (30.29%) aligned exactly 1 time
        54940 (2.51%) aligned >1 times
87.16% overall alignment rate
[bam_sort_core] merging from 1 files and 4 in-memory blocks...
```

```
$ more hisat2.sh.e24707186.2
4032514 reads; of these:
 4032514 (100.00%) were paired; of these:
   975045 (24.18%) aligned concordantly 0 times
                                                                  事前実行した結果で確認
   2882289 (71.48%) aligned concordantly exactly 1 time
                                                                  $ more outputs/hisat2/hisat2.sh.e16626062.2
    175180 (4.34%) aligned concordantly >1 times
   975045 pairs aligned concordantly 0 times; of these:
      89479 (9.18%) aligned discordantly 1 time
   885566 pairs aligned 0 times concordantly or discordantly; of these:
      1771132 mates make up the pairs; of these:
        1274482 (71.96%) aligned 0 times
        459285 (25.93%) aligned exactly 1 time
        37365 (2.11%) aligned >1 times
84.20% overall alignment rate
[bam_sort_core] merging from 0 files and 4 in-memory blocks...
```

マッピング 実行結果 アライメントファイル (sam)の確認

```
$ more SRR453566.sam
@HD VN:1.0 SO:unsorted
@SQ SN:NC_001133.9 LN:230218
                                                     事前実行した結果で確認
@SQ SN:NC_001134.8 LN:813184
@SQ SN:NC_001135.5 LN:316620
                                                     $ more outputs/hisat2/SRR453566.sam
@SQ SN:NC_001136.10
                  LN:1531933
@SQ SN:NC_001137.3 LN:576874
@SQ SN:NC_001138.5 LN:270161
@SQ SN:NC_001139.9LN:1090940
@SQ SN:NC_001140.6 LN:562643
@SQ SN:NC_001141.2 LN:439888
@SQ SN:NC_001142.9 LN:745751
@SQ SN:NC_001143.9 LN:666816
@SQ SN:NC_001144.5 LN:1078177
@SQ SN:NC_001145.3 LN:924431
@SQ SN:NC_001146.8 LN:784333
@SQ SN:NC_001147.6 LN:1091291
@SQ SN:NC_001148.4 LN:948066
@PG ID:hisat2 PN:hisat2 VN:2.2.1 CL:"/home/koshu3/tools/miniforge3/envs/pags_rnaseq/bin/hisat2-align-s --wrapper basic-0 -p
             73 NC 001136.10 107190060 101M=10719000NTTCGCTAGCTTCCGCTTTGATCTCTTCCTGTAGCTTGGCGGCGTCTTTATTGTCCAAAGGAACAGTTTCC
SRR453566.2
             133 NC_001136.10
                                       SRR453566.2
                             10719000
             73 NC 001145.3
SRR453566.1
                              887941 60 101M=8879410NAAAACTTTGGATGACTTCAACAACTATTCTTCTGAAATCAACAAAATATCACCAACTTCCGCCAACACAAAA
s more SRR453569.sam
@HD VN:1.0 SO:unsorted
                                                     事前実行した結果で確認
@SQ SN:NC_001133.9 LN:230218
@SQ SN:NC_001134.8 LN:813184
                                                     $ more outputs/hisat2/SRR453569.sam
@SQ SN:NC 001135.5 LN:316620
@SQ SN:NC_001136.10 LN:1531933
```

NTGGACTTTAGTGCGTTGAACGACATATTAAATGAACATGGTATTAGTATACTCCCAGCTAACGCATCACAATATGTCAAAAGA

24039 60 101M=240390TTCGAACGTCTCAAAGACATATGAGGAGCATATTGAGACCGTTTAATCACTTTTCATTGCTCTCTAGGGCGTG

77 * 0

141 *

0

89 NC 001143.9

0 0

0 0

SRR453569.2

SRR453569.2

SRR453569.4

発現量の算出

```
$ more scripts/stringtie.sh
#$ -S /bin/bash
#$ -pe def_slot 4
#$ -cwd
#$ -l mem_req=8G,s_vmem=8G
conda activate pags rnaseg
ACESSIONS=(453566 453569)
for NUM in ${ACESSIONS[@]}
do
 PREFIX=SRR${NUM}
  BAM=${PREFIX}".sorted.bam"
  stringtie -e -B -p ${NSLOTS} \
         -G reference/s288c.gff \
         -o ballgown/$PREFIX/${PREFIX}.out.gtf \
         -A ${PREFIX}.gene_abund.tab \
         $BAM
done
```

```
    -e only estimate the abundance of given reference transcripts (requires -G)
    -B enable output of Ballgown table files which will be created in the same directory as the output GTF (requires -G, -o recommended)
    -p number of threads (CPUs) to use (default: 1)
    -G reference annotation to use for guiding the assembly process (GTF/GFF)
    -o output path/file name for the assembled transcripts GTF (default: stdout)
    -A gene abundance estimation output file
```

発現量の算出

```
$ stringtie -h
StringTie v2.2.1 usage:
stringtie <in.bam ..> [-G <guide_gff>] [-l <prefix>] [-o <out.gtf>] [-p <cpus>]
[-v] [-a <min_anchor_len>] [-m <min_len>] [-j <min_anchor_cov>] [-f <min_iso>]
[-c <min_bundle_cov>] [-g <bdist>] [-u] [-L] [-e] [--viral] [-E <err_margin>]
[--ptf <f_tab>] [-x <seqid,..>] [-A <gene_abund.out>] [-h] {-B|-b <dir_path>}
 [--mix] [--conservative] [--rf] [--fr]
Assemble RNA-Seq alignments into potential transcripts.
Options:
--version: print just the version at stdout and exit
--conservative: conservative transcript assembly, same as -t -c 1.5 -f 0.05
--mix: both short and long read data alignments are provided
        (long read alignments must be the 2nd BAM/CRAM input file)
--rf: assume stranded library fr-firststrand
--fr : assume stranded library fr-secondstrand
-G reference annotation to use for guiding the assembly process (GTF/GFF)
--ptf : load point-features from a given 4 column feature file <f_tab>
-o output path/file name for the assembled transcripts GTF (default: stdout)
-l name prefix for output transcripts (default: STRG)
-f minimum isoform fraction (default: 0.01)
-L long reads processing; also enforces -s 1.5 -g 0 (default:false)
-R if long reads are provided, just clean and collapse the reads but
    do not assemble
-m minimum assembled transcript length (default: 200)
-a minimum anchor length for junctions (default: 10)
-j minimum junction coverage (default: 1)
-t disable trimming of predicted transcripts based on coverage
    (default: coverage trimming is enabled)
-c minimum reads per bp coverage to consider for multi-exon transcript
    (default: 1)
-s minimum reads per bp coverage to consider for single-exon transcript
    (default: 4.75)
-v verbose (log bundle processing details)
-g maximum gap allowed between read mappings (default: 50)
-M fraction of bundle allowed to be covered by multi-hit reads (default:1)
-p number of threads (CPUs) to use (default: 1)
-A gene abundance estimation output file
-E define window around possibly erroneous splice sites from long reads to
    look out for correct splice sites (default: 25)
-B enable output of Ballgown table files which will be created in the
    same directory as the output GTF (requires -G, -o recommended)
-b enable output of Ballgown table files but these files will be
   created under the directory path given as <dir_path>
-e only estimate the abundance of given reference transcripts (requires -G)
--viral: only relevant for long reads from viral data where splice sites
```

発現量の算出

講習中はqsub を実行しないでください。

\$ qsub scripts/stringtie.sh

Your job 24707314 ("stringtie.sh") has been submitted

```
$ 合計 8311004
drwxr-xr-x 7 koshu3 koshu
                                4096 10月 16 14:47 .
drwxr-x--- 13 koshu3 koshu
                                4096 10月 13 14:50 ...
-rw-r--r-- 1 koshu3 koshu
                              490409 10月 16 14:46 SRR453566.gene_abund.tab
-rw-r--r-- 1 koshu3 koshu 4173908395 10月 16 14:17 SRR453566.sam
-rw-r--r-- 1 koshu3 koshu 821523448 10月 16 14:18 SRR453566.sorted.bam
-rw-r--r 1 koshu3 koshu
                              491170 10月 16 14:47 SRR453569.gene_abund.tab
-rw-r--r-- 1 koshu3 koshu 2922686962 10月 16 14:19 SRR453569.sam
-rw-r--r-- 1 koshu3 koshu 591294623 10月 16 14:19 SRR453569.sorted.bam
drwxr-xr-x 4 koshu3 koshu
                                4096 10月 16 14:46 ballgown
-rw-r--r-- 1 koshu3 koshu
                                 693 10月 16 14:18 hisat2.sh.e24707186.1
-rw-r--r-- 1 koshu3 koshu
                                 688 10月 16 14:19 hisat2.sh.e24707186.2
-rw-r--r-- 1 koshu3 koshu
                                   0 10月 16 14:16 hisat2.sh.o24707186.1
-rw-r--r 1 koshu3 koshu
                                   0 10月 16 14:18 hisat2.sh.o24707186.2
-rw-r--r 1 koshu3 koshu
                                   0 10月 16 14:16 hisat2.sh.pe24707186.1
-rw-r--r-- 1 koshu3 koshu
                                   0 10月 16 14:18 hisat2.sh.pe24707186.2
-rw-r--r-- 1 koshu3 koshu
                                   0 10月 16 14:16 hisat2.sh.po24707186.1
-rw-r--r-- 1 koshu3 koshu
                                   0 10月 16 14:18 hisat2.sh.po24707186.2
-rw-r--r-- 1 koshu3 koshu
                                2262 10月 16 11:47 hisat2_index.sh.e24706258
                                3919 10月 16 11:47 hisat2 index.sh.o24706258
-rw-r--r-- 1 koshu3 koshu
                                   0 10月 16 11:47 hisat2_index.sh.pe24706258
-rw-r--r-- 1 koshu3 koshu
                                   0 10月 16 11:47 hisat2_index.sh.po24706258
-rw-r--r-- 1 koshu3 koshu
drwxr-xr-x 2 koshu3 koshu
                                4096 10月 16 11:58 outputs
drwxr-xr-x 2 koshu3 koshu
                                4096 10月 16 10:53 reads
drwxr-xr-x 2 koshu3 koshu
                                4096 10月 16 11:47 reference
drwxr-xr-x 2 koshu3 koshu
                                4096 10月 17 2022 scripts
-rw-r--r 1 koshu3 koshu
                                   0 10月 16 14:46 stringtie.sh.e24707314
-rw-r--r-- 1 koshu3 koshu
                                   0 10月 16 14:46 stringtie.sh.o24707314
                                   0 10月 16 14:46 stringtie.sh.pe24707314
-rw-r--r-- 1 koshu3 koshu
                                   0 10月 16 14:46 stringtie.sh.po24707314
-rw-r--r 1 koshu3 koshu
```

事前実行した結果で確認

\$ Is outputs/stringtie

```
$ ls ballgown/*
ballgown/SRR453566:
SRR453566.out.gtf e2t.ctab e_data.ctab
i2t.ctab i_data.ctab t_data.ctab

ballgown/SRR453569:
SRR453569.out.gtf e2t.ctab e_data.ctab
i2t.ctab i_data.ctab t_data.ctab
```

次のステップとして、R などを用いることで可視化などができる。

ballgown を使う場合は、このballgown ディレクトリを そのまま入力データとして使用できる。

事前実行した結果で確認

\$ Is outputs/stringtie/ballgown

\$ more SRR453566.gene_abund.tab

Gene ID Gene Name Reference Strand Start End Coverage FPKM 1.058091 1.01341 1.058091 1.01341 1.058091 1.01341 1.058091 1.01341 1.058091 1.01341 1.058091 1.01341 1.058091 1.01341 1.058091 1.01341 1.058091 1.01341 1.058091 1.01341 1.058091 1.01341 1.058091 1.01341 1.058091 1.01341 1.058091 1.01341 1.058091 1.01341 1.058091 1.0013341 1.0									
gene-YAL030W SNC1 NC_001133.9 + 87286 87752 70.827682 74.101746 80.072098 gene-YAL029C MY04 NC_001133.9 - 87855 92270 35.674591 37.323677 40.330833 gene-YAL028W FRT2 NC_001133.9 + 92900 94486 5.316950 5.562730 6.010918 gene-YAL027W SAW1 NC_001133.9 + 94687 95472 24.430025 25.559322 27.618629 gene-YAL026C DRS2 NC_001133.9 - 95630 99697 27.811796 29.097420 31.441788 gene-YNCA0001W HRA1 NC_001133.9 - 99305 99868 3.410652 3.568312 3.855810 gene-YAL025C MAK16 NC_001133.9 - 100225 101145 146.168289 152.925034 165.246155 gene-YAL023C PMT2 NC_001133.9 - 106272 108551 185.183334 193.743591 209.353424 gene-YAL021C CCR4	Gene ID	Gene Name	Reference	Strand	Start	End	Coverage	FPKM	TPM
gene-YAL029C MY04 NC_001133.9 - 87855 92270 35.674591 37.323677 40.330833 gene-YAL028W FRT2 NC_001133.9 + 92900 94486 5.316950 5.562730 6.010918 gene-YAL027W SAW1 NC_001133.9 + 94687 95472 24.430025 25.559322 27.618629 gene-YAL026C DRS2 NC_001133.9 - 95630 99697 27.811796 29.097420 31.441788 gene-YAL025C MAK16 NC_001133.9 - 99305 99868 3.410652 3.568312 3.855810 gene-YAL024C LTE1 NC_001133.9 - 100225 101145 146.168289 152.925034 165.246155 gene-YAL023C PMT2 NC_001133.9 - 101565 105872 9.433844 9.869931 10.665148 gene-YAL022C FUN26 NC_001133.9 - 108877 110430 36.694981 38.391239 41.484402 gene-YAL020C ATS1 NC_001133.9 - 113614 114615 31.161676 32.602150 35.	gene-YAL068C	PAU8	NC_001133.9	_	1807	2169	1.011341	1.058091	1.143341
gene-YAL028W FRT2 NC_001133.9 + 92900 94486 5.316950 5.562730 6.010918 gene-YAL027W SAW1 NC_001133.9 + 94687 95472 24.430025 25.559322 27.618629 gene-YAL026C DRS2 NC_001133.9 - 95630 99697 27.811796 29.097420 31.441788 gene-YAL025C MAK16 NC_001133.9 - 100225 101145 146.168289 152.925034 165.246155 gene-YAL024C LTE1 NC_001133.9 - 101565 105872 9.433844 9.869931 10.665148 gene-YAL023C PMT2 NC_001133.9 - 106272 108551 185.183334 193.743591 209.353424 gene-YAL022C FUN26 NC_001133.9 - 108877 110430 36.694981 38.391239 41.484402 gene-YAL021C CCR4 NC_001133.9 - 113614 114615 31.161676 32.602150 35.228893 gene-YAL019W FUN30 NC_001133.9 - 113614 114919 118314 30.554476	gene-YAL030W	SNC1	NC_001133.9	+	87286	87752	70.827682	74.101746	80.072098
gene-YAL027W SAW1 NC_001133.9 + 94687 95472 24.430025 25.559322 27.618629 gene-YAL026C DRS2 NC_001133.9 - 95630 99697 27.811796 29.097420 31.441788 gene-YAL020C MRK16 NC_001133.9 - 99868 3.410652 3.568312 3.855810 gene-YAL024C LTE1 NC_001133.9 - 100225 101145 146.168289 152.925034 165.246155 gene-YAL023C PMT2 NC_001133.9 - 106272 108551 185.183334 193.743591 209.353424 gene-YAL022C FUN26 NC_001133.9 - 108877 110430 36.694981 38.391239 41.484402 gene-YAL021C CCR4 NC_001133.9 - 110846 113359 35.354412 36.988697 39.968864 gene-YAL020C ATS1 NC_001133.9 - 113614 114615 31.161676 32.602150 35.228893 gene-YAL018C LDS1 NC_001133.9 - 118564 119541 1.130879 1.183155 1.278481	gene-YAL029C	MY04	NC_001133.9	_	87855	92270	35.674591	37.323677	40.330833
gene-YAL026C DRS2 NC_001133.9 - 95630 99697 27.811796 29.097420 31.441788 gene-YNCA0001W HRA1 NC_001133.9 + 99305 99868 3.410652 3.568312 3.855810 gene-YAL025C MAK16 NC_001133.9 - 100225 101145 146.168289 152.925034 165.246155 gene-YAL024C LTE1 NC_001133.9 - 101565 105872 9.433844 9.869931 10.665148 gene-YAL023C PMT2 NC_001133.9 - 106272 108551 185.183334 193.743591 209.353424 gene-YAL022C FUN26 NC_001133.9 - 108877 110430 36.694981 38.391239 41.484402 gene-YAL021C CCR4 NC_001133.9 - 110846 113359 35.354412 36.988697 39.968864 gene-YAL020C ATS1 NC_001133.9 - 113614 114615 31.161676 32.602150 35.228893 gene-YAL018C LDS1 NC_001133.9 - 118564 119541 1.130879 1.183155 1.278481	gene-YAL028W	FRT2	NC_001133.9	+	92900	94486	5.316950	5.562730	6.010918
gene-YNCA0001W HRA1 NC_001133.9 + 99305 99868 3.410652 3.568312 3.855810 gene-YAL025C MAK16 NC_001133.9 - 100225 101145 146.168289 152.925034 165.246155 gene-YAL024C LTE1 NC_001133.9 - 101565 105872 9.433844 9.869931 10.665148 gene-YAL023C PMT2 NC_001133.9 - 106272 108551 185.183334 193.743591 209.353424 gene-YAL022C FUN26 NC_001133.9 - 108877 110430 36.694981 38.391239 41.484402 gene-YAL021C CCR4 NC_001133.9 - 110846 113359 35.354412 36.988697 39.968864 gene-YAL019W FUN30 NC_001133.9 + 113614 114615 31.161676 32.602150 35.228893 gene-YAL018C LDS1 NC_001133.9 - 118564 119541 1.130879 1.183155 1.278481	gene-YAL027W	SAW1	NC_001133.9	+	94687	95472	24.430025	25.559322	27.618629
gene-YAL025C MAK16 NC_001133.9 - 100225 101145 146.168289 152.925034 165.246155 gene-YAL024C LTE1 NC_001133.9 - 101565 105872 9.433844 9.869931 10.665148 gene-YAL023C PMT2 NC_001133.9 - 106272 108551 185.183334 193.743591 209.353424 gene-YAL022C FUN26 NC_001133.9 - 108877 110430 36.694981 38.391239 41.484402 gene-YAL021C CCR4 NC_001133.9 - 110846 113359 35.354412 36.988697 39.968864 gene-YAL019W FUN30 NC_001133.9 - 113614 114615 31.161676 32.602150 35.228893 gene-YAL018C LDS1 NC_001133.9 - 118564 119541 1.130879 1.183155 1.278481	gene-YAL026C	DRS2	NC_001133.9	_	95630	99697	27.811796	29.097420	31.441788
gene-YAL024C LTE1 NC_001133.9 - 101565 105872 9.433844 9.869931 10.665148 gene-YAL023C PMT2 NC_001133.9 - 106272 108551 185.183334 193.743591 209.353424 gene-YAL022C FUN26 NC_001133.9 - 108877 110430 36.694981 38.391239 41.484402 gene-YAL021C CCR4 NC_001133.9 - 110846 113359 35.354412 36.988697 39.968864 gene-YAL020C ATS1 NC_001133.9 - 113614 114615 31.161676 32.602150 35.228893 gene-YAL019W FUN30 NC_001133.9 + 114919 118314 30.554476 31.966883 34.542442 gene-YAL018C LDS1 NC_001133.9 - 118564 119541 1.130879 1.183155 1.278481	gene-YNCA0001W	HRA1	NC_001133.9	+	99305	99868	3.410652	3.568312	3.855810
gene-YAL023C PMT2 NC_001133.9 - 106272 108551 185.183334 193.743591 209.353424 gene-YAL022C FUN26 NC_001133.9 - 108877 110430 36.694981 38.391239 41.484402 gene-YAL021C CCR4 NC_001133.9 - 110846 113359 35.354412 36.988697 39.968864 gene-YAL020C ATS1 NC_001133.9 - 113614 114615 31.161676 32.602150 35.228893 gene-YAL019W FUN30 NC_001133.9 + 114919 118314 30.554476 31.966883 34.542442 gene-YAL018C LDS1 NC_001133.9 - 118564 119541 1.130879 1.183155 1.278481	gene-YAL025C	MAK16	NC_001133.9	_	100225	101145	146.168289	152.925034	165.246155
gene-YAL022C FUN26 NC_001133.9 - 108877 110430 36.694981 38.391239 36.988697 39.968864 gene-YAL021C CCR4 NC_001133.9 - 110846 113359 35.354412 36.988697 39.968864 gene-YAL020C ATS1 NC_001133.9 - 113614 114615 31.161676 32.602150 35.228893 gene-YAL019W FUN30 NC_001133.9 + 114919 118314 30.554476 31.966883 34.542442 gene-YAL018C LDS1 NC_001133.9 - 118564 119541 1.130879 1.183155 1.278481	gene-YAL024C	LTE1	NC_001133.9	_	101565	105872	9.433844	9.869931	10.665148
gene-YAL021C CCR4 NC_001133.9 - 110846 113359 35.354412 36.988697 39.968864 gene-YAL020C ATS1 NC_001133.9 - 113614 114615 31.161676 32.602150 35.228893 gene-YAL019W FUN30 NC_001133.9 + 114919 118314 30.554476 31.966883 34.542442 gene-YAL018C LDS1 NC_001133.9 - 118564 119541 1.130879 1.183155 1.278481	gene-YAL023C	PMT2	NC_001133.9	_	106272	108551	185.183334	193.743591	209.353424
gene-YAL020C ATS1 NC_001133.9 - 113614 114615 31.161676 32.602150 35.228893 gene-YAL019W FUN30 NC_001133.9 + 114919 118314 30.554476 31.966883 34.542442 gene-YAL018C LDS1 NC_001133.9 - 118564 119541 1.130879 1.183155 1.278481	gene-YAL022C	FUN26	NC_001133.9	_	108877	110430	36.694981	38.391239	41.484402
gene-YAL019W FUN30 NC_001133.9 + 114919 118314 30.554476 31.966883 34.542442 gene-YAL018C LDS1 NC_001133.9 - 118564 119541 1.130879 1.183155 1.278481	gene-YAL021C	CCR4	NC_001133.9	_	110846	113359	35.354412	36.988697	39.968864
gene-YAL018C LDS1 NC_001133.9 - 118564 119541 1.130879 1.183155 1.278481	gene-YAL020C	ATS1	NC_001133.9	_	113614	114615	31.161676	32.602150	35.228893
_	gene-YAL019W	FUN30	NC_001133.9	+	114919	118314	30.554476	31.966883	34.542442
gene-YAL017W PSK1 NC_001133.9 + 120225 124295 18.278311 19.123240 20.663992	gene-YAL018C	LDS1	NC_001133.9	_	118564	119541	1.130879	1.183155	1.278481
	gene-YAL017W	PSK1	NC_001133.9	+	120225	124295	18.278311	19.123240	20.663992

発現量のノーマライズ

FPKM: Fragments Per Kilobase of exon per Million reads mapped TPM: Transcripts Per kilobase Milion

FPKMもTPMも以下の二つで補正するが、補正する順番が異なる。

- (1) 総リード数での補正 (総リード数 100万)
- (2) 遺伝子長での補正 (遺伝子長 1000b)

FPKM (1) -> (2) TPM (2) -> (1)

Singularity を利用したスクリプト

```
$ more scripts/hisat2_singularity.sh
#$ -S /bin/bash
#$ -pe def slot 4
#$ -cwd
#$ -t 1-2:1
#$ -1 mem req=8G,s vmem=8G
conda activate pags_rnaseq
# Batch culture: SRR453566
# chemostat: SRR453569
ACESSIONS=(453566 453569)
no=`expr ${SGE TASK ID} - 1`
NUM=${ACESSIONS[${no}]}
PREFIX=SRR${NUM}
# read file
DIR=./reads/
QUERY1_1=${DIR}${PREFIX}"_1.fastq.qz"
QUERY1_2=${DIR}${PREFIX}"_2.fastq.gz"
singularity exec /usr/local/biotools/h/hisat2:2.2.1--h87f3376_4 \
hisat2 -p ${NSLOTS} -x reference/s288c.fa --dta \
        -1 ${QUERY1_1} -2 ${QUERY1_2} \
        -S ${PREFIX}.sam
# convert sam to bam
# sort by position
singularity exec /usr/local/biotools/s/samtools:1.17--hd87286a_2 \
 samtools sort -@ ${NSLOTS} ${PREFIX}.sam -o ${PREFIX}.sorted.bam
```

おわり

GTF format とは

stringtie -e -B -p 4 -G reference/s288c.gff -o ballgown/SRR453566/SRR453566.out.gtf -A SRR453566.gene_abund.tab SRR453566.sorted.bam # StringTie version 2.2.1 NC 001133.9 StringTie 21691000 - . gene_id "gene-YAL068C"; transcript_id "rna-NM_001180043.1"; ref_gene_name transcript 1807 21691000 - . gene id "gene-YAL068C"; transcript id "rna-NM 001180043.1"; exon number ' NC 001133.9 StringTie 1807 exon 87286 877521000 + . gene id "gene-YAL030W"; transcript id "rna-NM 001178175.1"; ref gene nam NC 001133.9 StringTie transcript NC 001133.9 87286 873871000 + . gene id "gene-YAL030W"; transcript id "rna-NM 001178175.1"; exon number StringTie exon 87501 877521000 + . gene id "gene-YAL030W"; transcript id "rna-NM 001178175.1"; exon number NC 001133.9 StringTie exon

タブ区切りフォーマット。値がない場合は、"." が設定される。

1. seqname: 染色体 or スキャフォールドの名前

2. source:アノテーションを生成したプログラムまたはデータソースの名前

3. feature: フィーチャータイプ (mRNA, gene, exon, CDS)

4. start: スタートポジション (1bp ∼)

5. end: エンドポジション (1bp ~)

6. score:スコア

7. strand: +(forward)、-(reverse)または!.'

8. frame:翻訳フレーム(0, 1, 2)

9. attribute: 追加情報。セミコロンで区切られたタグと値のペアのリスト。

GFF では、gene_id=XXXXX; の形式に対して、GTF では、gene_id "XXXXXX"; と記載していく。