Annotation and filtering



Learning outcomes

At the end of this lecture, you should be able to:

- 1. Describe and evaluate available annotations
- 2. Justify filtering strategies using genetic knowledge
- 3. Contrast variant-led and phenotype-led approaches

Analysis workflow



Raw reads **Processed BAM** Visualise data raw sequence data **FASTQ** Quality control of Assess quality and Call variants Variant calling process reads and QC **Processed reads** Variant and sample **FASTQ** quality control Map to reference **Annotate** Alignment> Public databases genome **Annotation** Post process overlaps **Annotate** and duplicates **Pathogenicity** Post alignment processing Post process Filter and prioritise and assessment InDel realignment variants strategies Filtering Post process Integrate with clinical Visualise data BaseQ recalibration information Assess depth and **Functional Shortlist of disease** breadth of coverage follow-up related variants



Data to annotate

- Variant functional effect
- Allele frequencies
- Conservation
- Functional effect prediction scores
- Disease databases for disease of interest
 - Genes
 - Variants



Functional effect

- Is variant synonymous, stop-gain, frameshift...
 - Different transcripts may be affected differently
 - Worst affected transcript may not be clinically relevant
 - Strict guidelines (http://varnomen.hgvs.org/)
- Choice of transcript database (RefSeq or ensembl)
- Reduces complex biology to an absolute, sometimes imperfectly

NOD2:NM_001293557:exon3:c.T953C:p.V318A

Variant type	p. description	Remarks
	p.(Arg490Ser)	The protein change is predicted (no experimental proof)
Substitution	p.Arg490Ser/p.R490S p.Trp87Ter / p.Trp78*/p.W87*	Both three- (preferred) and one-letter amino acid code may be used; * accepted for one- and three-letter code
Deletion	p.Asp388_Gln393del	No specification of deleted amino acid(s)
Duplication	p.Asp388_Gln393dup	No specification of duplicated amino acid(s)
Insertion	p.Ala228_Val229insTrpPro p.Ala228_Val229insLys*	Mandatory specification of inserted amino acids
Inversions		Not possible
Frame shift	p.(Arg97fs) p.(Arg97Profs*23)	Short and long form accepted; long form contains "fsTer" or "fs*"



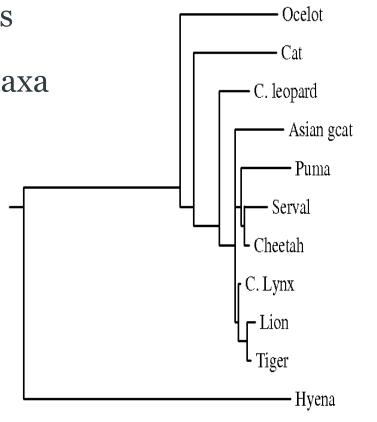
Allele frequencies

- Require baseline of human variance
 - Rare/novel variant ≠ pathogenic
 - Massive collaborations (and infrastructure) required
- 1000 Genomes 2,504 WGS individuals globally ('healthy')
 - 843 authors; 168 affiliations
- Exome Sequencing Project 6,515 Americans (EA and AA)
 - 396 authors;
- ExAC 60,706 WES globally (common disease cohorts)
 - 74 authors + consortium; 52 affiliations



Conservation

- Measure how much a base varies
- Based on sequence data across taxa
- Requires genome alignments
 - Non-trivial
- PhyloP is common score
- GERP++ more complex



Highly conserved → more damaging if changed?



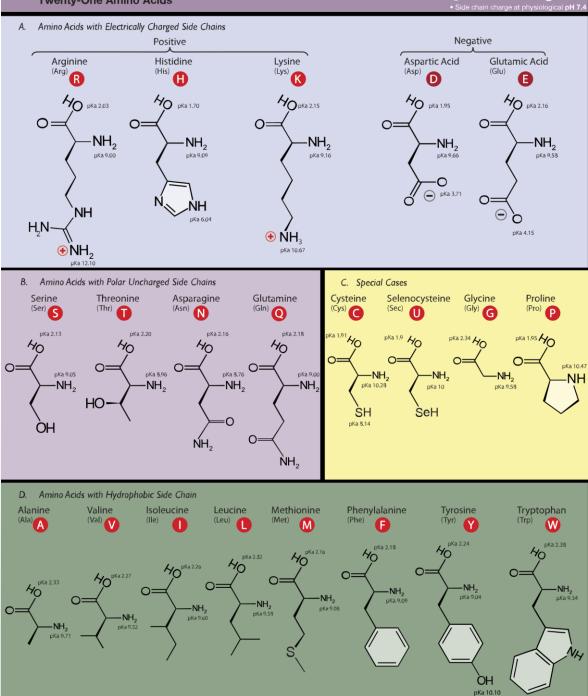
Functional effect scores

- Based on the difference a mutation may make to protein function
- Amino acid changes Grantham scores
- Plus conservation information SIFT scores
- Structure based PolyPhen-2
 - Machine learning based on structural/biochemical features
- Consensus CADD

Twenty-One Amino Acids

Negativ

Medicine

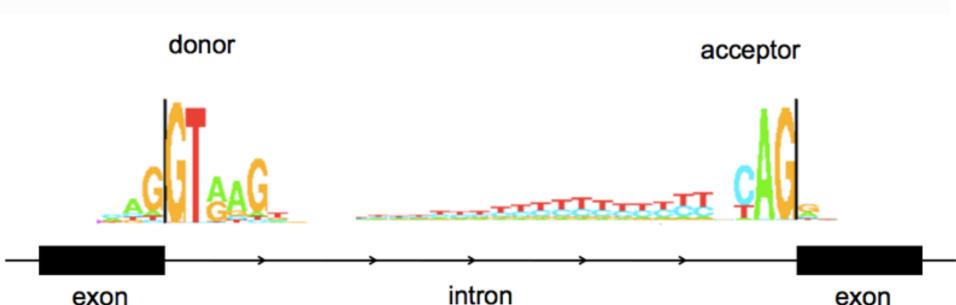






Splicing

- Splicing mutations can have similar impact to frameshifts
- Complex process which is hard to predict
- Conservation of motifs at splice sites





Disease databases

- Can use for gene and variant prioritisation
- Human Gene Mutation Database (HGMD)
 - Team of dedicated curators, industry funded

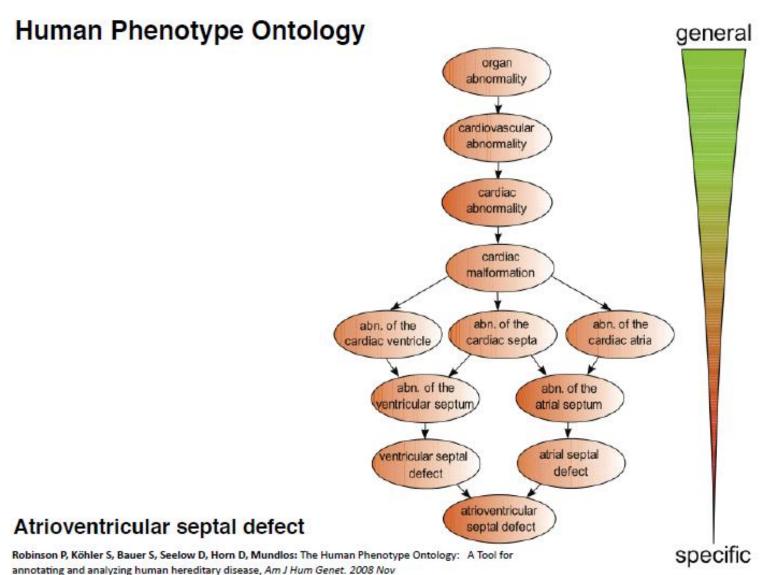
- Online Mendelian Inheritance in Man (OMIM)
 - Dedicated curators, public grant funded
- Leiden Open Variation Database (LOVD)
 - Infrastructure only, community implemented



Phenotype-led interrogation

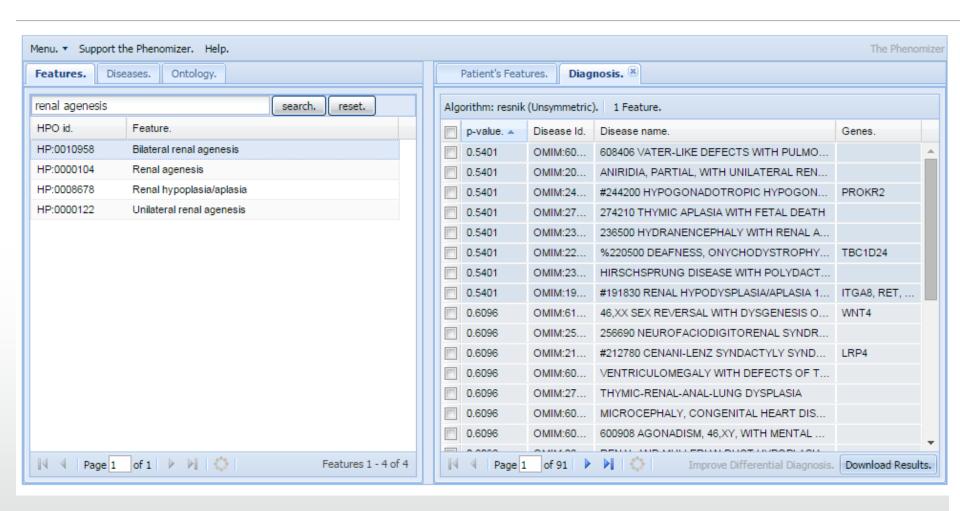
- Can use systematic phenotyping for prioritisation
- Human phenotype ontology (HPO) provides a platform
- Link phenotypes to known causal genotypes in a network
- Create gene lists (Phenomizer, Phenotips)
- Prioritise variants (Exomiser, PhenIX)





MediciPhenomizer





MediciPhenotype defined target



Narrowing diagnostic scope to phenotype-defined target

Unmasking exome data using phenotypically compatible genes

Advantages

Individually defined for each patient, in accordance to presenting phenotype

Does not depend on the existence of diagnostic hypothesis

Is robust to diagnostic re-classification

Based on a **continuously updated** resource (HPO)

Can be used a **standardized approach** for panel generation

Disadvantages

Depend on the completeness of genephenotype mappings

Depend on the completeness of describing patient phenotype presentation

Generally result in larger gene target for analysis

Moderate increase in the possibility of finding pertinent findings

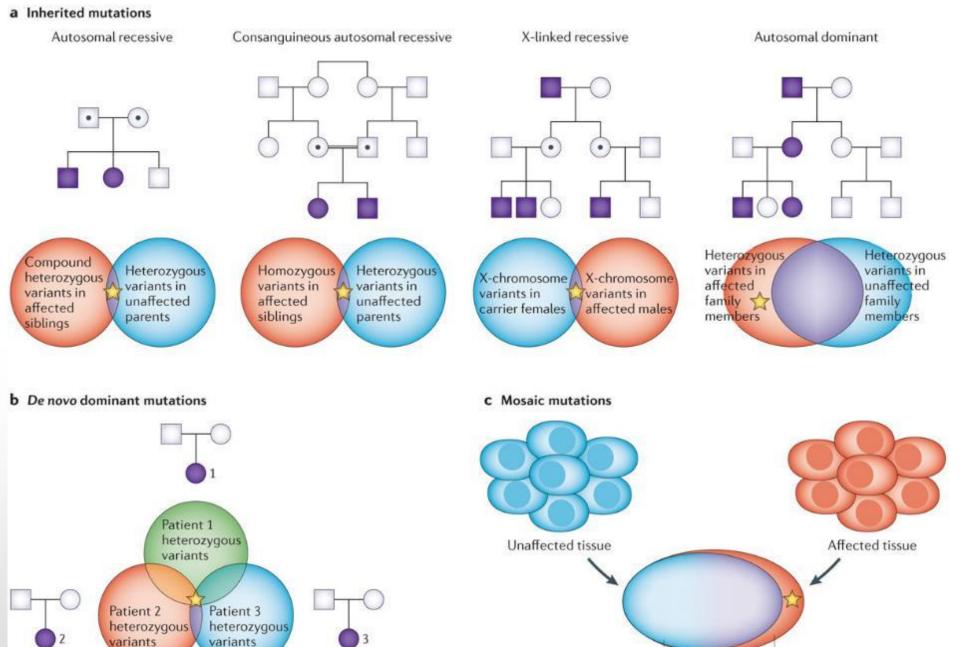
Variant shortlist



- Use Phenotype to restrict the candidates
- However, different resources give different phenotype to gene interactions

Renal Agenesis	
OMIM	RET, ITGA8, PAX2
Orphanet	FGF20, ITGA8
Congenital anomalies of the kidney and urinary tract (CAKUT) panel	BICC1, BMP4, CHD1L, EYA1, FOXC1, GATA3, GDNF, RET, ROBO2, SIX1, SIX5, SOX17, TFAP2A, TRAP1, UPK3A, WT1
Phenomizer	PROKR2, RET, PAX2, TBC1D24, LRP4,

						Rank	
Gene	Diagnosis	PhenIX	Exomiser	Exomiser with CADD	OVA	eXtasy (order statistics)	eXtasy (combined max)
ARID1B	COFFIN-SIRIS SYNDROME; CSS;;FIFTH DIGIT SYNDROME	2	95	132	1037	6013	6184
KCNQ2	EPILEPTIC ENCEPHALOPATHY, EARLY INFANTILE, 7; EIEE7	1	85	104	Not listed	1458	8508
SGCE	MYOCLONIC DYSTONIA	7	Not listed	Not listed	Not listed	239	9304
MED13L	MENTAL RETARDATION, AUTOSOMAL RECESSIVE 15; MRT15	106	14	10	1004	2230	4511
RYR1	CONGENITAL FIBER-TYPE DISPROPORTION MYOPATHY	1	68	85	74	422	8624
SACS	SPASTIC ATAXIA, CHARLEVOIX-SAGUENAY TYPE	3	89	77	308	3264	5032
UBE3A	ANGELMAN SYNDROME	12	74	77	Not listed	178	8728
PTEN	PTEN HAMARTOMA TUMOR SYNDROME	1	1	1	Not listed	126	8822
DYNC1H1	SPINAL MUSCULAR ATROPHY, LOWER EXTREMITY, AUTOSOMAL DOMINANT; SMALED	10	85	86	20	1759	4687
SCN1A	DRAVET SYNDROME	2	27	53	72	250	8188
TCOF1	TREACHER COLLINS SYNDROME 3; TCS3;;MANDIBULOFACIAL DYSOSTOSIS, TREACHER COLLINS TYPE, AUTOSOMAL RECESSIVE	9	99	92	45	259	8858
OTX2	MICROPHTHALMIA, ISOLATED 1	5	60	70	73	Not listed	Not listed
EHMT1	KLEEFSTRA SYNDROME	10	88	95	Not listed	Not listed	Not listed
EFNB1	CRANIOFRONTONASAL SYNDROME; CFNS;;CRANIOFRONTONASAL DYSPLASIA; CFND;;CRANIOFRONTONASAL DYSOSTOSIS	1	1	1	Not listed	254	8997
HRAS	COSTELLO SYNDROME	7	1	1	52	1	9328
PTPN11	NOONAN SYNDROME 6; NS6	1	82	83	Not listed	1	9328
EIF2B1	LEUKOENCEPHALOPATHY WITH VANISHING WHITE MATTER; VWM	11	Not listed	144	Not listed	30	9216
FGFR3	MUENKE SYNDROME; MNKES	1	1	1	50	7	9281
POLG	ALPERS SYNDROME	1	89	98	402	14	8876
СОМР	PSEUDOACHONDROPLASIA	1	78	90	53	10	9310



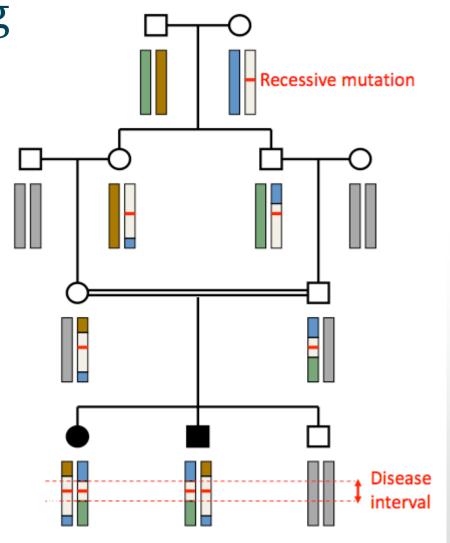
Variants in unaffected tissue

Variants in affected tissue

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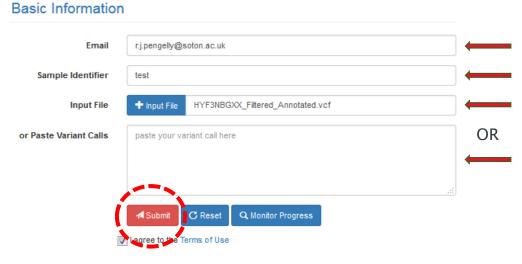
Autozygosity mapping

- Useful in consanguineous scenarios
- Looking for regions of genome from the same recent ancestor (i.e. autozygous)
- Does not require trios
 - Identify tracts of homozygosity in proband



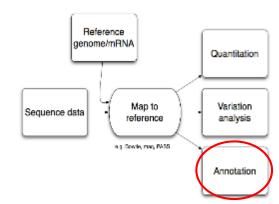


Medicine



please enter your focused disease/phenotype terms

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Disease/Phenotype

Enter Disease or

Phenotype Terms

	Please use semicolon or enter as so Try to use multiple terms instead of OMIM IDs are also accepted, like 11- Better Combined with wANNOVAR's	a super long term 4480 for 'Breast cancer'	
Parameter Settin	gs		
Result duration	1 day	· Q	
Reference Genome	hg19	• Q	
Input Fomat	VCF	a	
Gene Definition	RefSeq Gene	• Q	
Individual analysis	Individual analysis	→	
Disease Model	none	Q	

http://wannovar.usc.edu/



WANNOVAR Home Tutorial Example Related projects → WŁob

Submission ID: 71432

Sample identifier = test

File_name=HYF3NBGXX_Filtered_Annotated.vcf

File_format=vcf4

Reference_genome=hg19

Disease_model=no filtering

Processed variants=170

Basic Information

exome summary results	view	CSV file	TXT file
genome summary results	view	CSV file	TXT file

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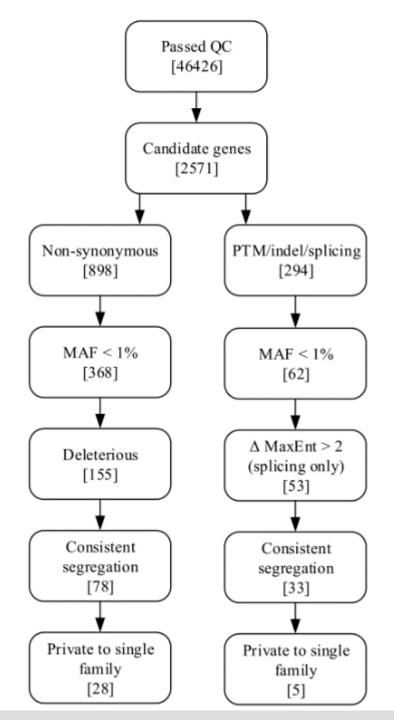
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Func	Gene	ExonicFunc	AAChange	Conserved	SegDup	ESP5400 ALL	1000g2012feb ALL	dbSNP135	AVSIFT	LJB PhyloP	LJB PhyloP Pred	LJB SIFT	LJ SI Pr
exonic	SLC16A1	nonsynonymous SNV	NM_001166496:c.T1470A:p.D490E	13		0.666	0.66	rs1049434	1	0.862	N	0.22	Т
exonic	LMNA	synonymous SNV	NM_005572:c.C51T:p.S17S	1075		0.00939	0.01	rs11549668					
exonic	LMNA	synonymous SNV	NM_001257374:c.G276A:p.L92L	369		0.00595	0.0037	rs12117552					
exonic	LMNA	synonymous SNV	NM_001257374:c.T525C:p.A175A			0.203	0.16	rs538089					
exonic	LMNA	synonymous SNV	NM_001257374:c.C631T:p.L211L	124									
exonic	LMNA	synonymous SNV	NM_001257374:c.T1002C:p.D334D	344		0.263	0.21	rs505058					
exonic	LMNA	synonymous SNV	NM_001257374:c.C1230T:p.C410C	142		0.00149	0.0014	rs149339264					
exonic;splicing	LMNA; LMNA	synonymous SNV	NM_001257374:c.C1362T:p.H454H	216		0.194	0.21	rs4641					
exonic	SLC19A2	synonymous SNV	NM_006996:c.G639A:p.K213K	260		0.00093	0.0023	rs137970656					
exonic	ENAH	synonymous SNV	NM_001008493:c.T1062C:p.P354P	55									
exonic	ENAH	synonymous SNV	NM_001008493:c.G759A:p.R253R	218		0.0354	0.03	rs1340868					
exonic	ENAH	nonsynonymous SNV	NM_001008493:c.G651T:p.E217D	82						0.5	N	0.89	Т
exonic	ENAH	synonymous SNV	NM_001008493:c.G615A:p.E205E	82									
exonic	KLF11	nonsynonymous SNV	NM_001177716:c.A134G:p.Q45R	209		0.0941	0.06	rs35927125	0.11	0.245	N	0.64	Т
exonic	KLF11	synonymous SNV	NM_001177716:c.A1134T:p.V378V	426		0.808	0.84	rs11687357					
exonic	ALMS1	synonymous SNV	NM 015120:c.G57A:p.E19E										
exonic	ALMS1	synonymous SNV	NM 015120:c.G60A:p.E20E			0.000892	0.01	rs183407241					
exonic	ALMS1	synonymous SNV	NM_015120:c.A75G:p.E25E					rs13009043					
exonic	ALMS1	synonymous SNV	NM_015120:c.G975A:p.S325S			0.000103							
exonic	ALMS1	nonsynonymous SNV	NM_015120:c.C1174T:p.R392C			0.389	0.34	rs3813227	0.18	0.143	N	1.0	D
exonic	ALMS1	nonsynonymous SNV	NM_015120:c.G1267A:p.V423I			0.00385	0.0005	rs45630557	0.98	0.805	N	1.0	D
exonic	ALMS1	nonsynonymous SNV	NM_015120:c.A1868G:p.H623R			0.0194	0.01	rs41291187	0.12	0.138	N	0.96	D
exonic	ALMS1	nonsynonymous SNV	NM 015120:c.T2012G:p.V671G			0.881	0.86	rs2037814	0.14	0.772	N	1.0	D
exonic	ALMS1	synonymous SNV	NM 015120:c.C2187T:p.F729F			0.51	0.54	rs7598901					
exonic	ALMS1	synonymous SNV	NM 015120:c.C2532T:p.D844D			0.0331	0.04	rs77517267					
exonic	ALMS1	nonsynonymous SNV	NM_015120:c.C3304G:p.P1102A			0.000519			0.1	0.853	N	0.94	Т
exonic	ALMS1	synonymous SNV	NM 015120:c.A3891G:p.Q1297Q			0.0592	0.05	rs112034360					
exonic	ALMS1	synonymous SNV	NM_015120:c.A4176G:p.Q1392Q			0.365	0.32	rs6546836					
exonic	ALMS1	nonsynonymous SNV	NM_015120:c.G4241C:p.G1414A			0.389	0.34	rs6546837	1	0.036	N	0.92	Т
exonic	ALMS1	synonymous SNV	NM 015120:c.G4956A:p.Q1652Q										
exonic	ALMS1	nonsynonymous SNV	NM_015120:c.A5356G:p.N1786D			0.0115	0.01	rs45608038	0.35	0.000859	N	0.94	Т
exonic	ALMS1	nonsynonymous SNV	NM 015120:c.A5623G:p.I1875V			0.386	0.33	rs6546838	1	0.0262	N	0.76	Т
exonic	ALMS1	nonsynonymous SNV	NM_015120:c.C6122T:p.T2041I						0	0.984	С	0.99	D
exonic	ALMS1	nonsynonymous SNV	NM_015120:c.T6209C:p.I2070T			0.137	0.10	rs10496192	0	0.0271	N	0.9	Т
exonic	ALMS1	nonsynonymous SNV	NM 015120:c.C6299T:p.S2100L			0.0232	0.02	rs28730854	0.02	0.981	С	0.99	D
exonic	ALMS1	nonsynonymous SNV	NM 015120:c.T6333A:p.S2111R			0.39	0.34	rs6724782	1	0.0136	N	0.99	D

Sort by:	•			
Filter by:				
1000G_ALL: ▼	1000G_AFR:	▼	1000G_EUR:	▼.
ExAC_Freq:	ExAC_AMR:	▼	ExAC_NFE:	▼
ESP6500si_ALL:	CG46:	▼	COSMIC_ID:	▼
ClinVar_DIS:	ClinVar_ID:	▼	ClinVar_DBID:	•
GWAS_DIS:	GWAS_OR:	•		
Chr:				
Ctarte				
Start: 🔻				
End: ▼				
Gene:				
1000G_ALL:				
1000G_EAS: ▼				
1000G_AFR: ▼				
Func:	ExonicFunc:			
exonic	frameshift insertion	on		
exonic;splicing	frameshift deletion	n		
splicing	nonframeshift dele	etion		
UTR3	nonframeshift inse	ertion		
UTR5	nonsynonymous SN	IV		
intronic intronic	synonymous SNV			
intergenic intergenic	stopgain SNV			
upstream	stoploss SNV			
downstream	unknown			
upstream;downstream	ı			
ncRNA_exonic				
ncRNA_intronic				
ncRNA_UTR3				
ncRNA_UTR5				

Filtering

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Variant assessment

- Does variant look real?
- Is variant in gene associated with phenotype?
 - Predictive phenotyping
- Is it a known pathogenic variant?
- Does it alter the protein in a way reported to be damaging?
- Is the variant rare (not just in patient's population)?
- Is the variant predicted to be deleterious (multiple scores)?
- Is the variant being pathogenic biologically plausible?

Analysis workflow



Raw reads **Processed BAM** Visualise data raw sequence data **FASTQ** Quality control of Assess quality and Call variants Variant calling process reads and QC Processed reads Variant and sample **FASTQ** quality control Map to reference **Annotate** Alignment> Public databases genome **Annotation** Post process overlaps **Annotate** and duplicates **Pathogenicity** Post alignment processing Post process Filter and prioritise and assessment InDel realignment variants strategies Filtering Post process Integrate with clinical Visualise data BaseQ recalibration information Assess depth and **Functional Shortlist of disease** breadth of coverage follow-up related variants

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Follow up

Literature review Animal models

Biochemical assays

Cell based assays

Patient cohorts

In silico modelling



Summary

- Filtering is required to produce usable variant shortlist
- Allele frequencies are an effective filter
- Predictive scores are informative for prioritisation
- Segregation is a powerful tool
- Phenotype led approaches can be helpful, but a work in progress

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