






# Microbial dynamics and soil physicochemical properties explain large-scale variations in soil organic carbon

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## Abstract

First-order organic matter decomposition models are used within most Earth System Models (ESMs) to project future global carbon cycling. These models have been criticized for not accurately representing mechanisms of soil organic carbon (SOC) stabilization and SOC response to climate change. New soil biogeochemical models have been developed, but their evaluation is limited to observations from laboratory incubations or few field experiments. Given the global scope of ESMs, a comprehensive evaluation of such models is essential using in situ observations of a wide range of SOC stocks over large spatial scales before their introduction to ESMs. In this study, we collected a set of in situ observations of SOC, litterfall and soil properties from 206 sites covering different forest and soil types in Europe and China. These data were used to calibrate the model MIMICS (The Microbial-Mineral Carbon Stabilization model), which we compared to the widely used first-order model CENTURY. We show that, compared to CENTURY, MIMICS more accurately estimates forest SOC concentrations and the sensitivities of SOC to variation in soil temperature, clay content and litter input. The ratios of microbial biomass to total SOC predicted by MIMICS agree well with independent observations from globally distributed forest sites. By testing different hypotheses regarding (using alternative process representations) the physicochemical constraints on SOC deprotection and

microbial turnover in MIMICS, the errors of simulated SOC concentrations across sites were further decreased. We show that MIMICS can resolve the dominant mechanisms of SOC decomposition and stabilization and that it can be a reliable tool for predictions of terrestrial SOC dynamics under future climate change. It also allows us to evaluate at large scale the rapidly evolving understanding of SOC formation and stabilization based on laboratory and limited field observation.

#### KEYWORDS

climate change, microbial physiology, soil biogeochemical model, soil carbon classification, soil carbon stabilization, soil organic carbon, soil physicochemical property

## 1 | INTRODUCTION

Soil organic carbon (SOC) is the largest terrestrial carbon (C) pool (Ciais et al., 2013), and it contains more than three times as much C as either the atmosphere or terrestrial vegetation. Therefore, a small change (<1%) in the global SOC pool might drastically alter the land-atmosphere C balance (Heimann & Reichstein, 2008; Shi, Crowell, Luo, Moore, 2018). SOC is also closely related to soil fertility, structure, water holding capacity and ecosystem biogeochemical cycles (Campbell & Paustian, 2015; Six, Bossuyt, Degryze, & Denef, 2004). Dynamics of SOC have received increasing attention in many research areas ranging from small-scale projects for preserving or improving soil health to large-scale climate change mitigation (e.g. the “4per1000” initiative; Lal, 2016). Soil biogeochemical models are the main tools for estimating global land C stock and the interactions between SOC dynamic and changes in climate and land use.

The majority of global soil C models are developed based on first-order kinetics, in which the decomposition rate of organic matter is proportional to the pool size and turnover rate, modified by environmental factors (Manzoni & Porporato, 2009; Parton, Schimel, Cole, & Ojima, 1987). These models are mathematically simple and stable, and have been proven effective for simulating soil organic matter dynamics (e.g. the decreasing trend of remaining organic matter mass during decomposition experiments; Barré et al., 2010; Bonan, Hartman, Parton, & Wieder, 2013). However, these models are unable to mechanically represent the transient SOC dynamics in response to increased fresh litter input (Fontaine et al., 2007; Guenet, Danger, Abbadie, & Lacroix, 2010; Kuzyakov, 2010), likely because they lack explicit representation of microbial decomposition and SOC stabilization (Creamer et al., 2015; Schmidt et al., 2011). Earth System Models (ESMs) which use the first-order soil C models also show poor agreement with global spatial variation of SOC stock (Hararuk & Luo, 2014; Brown et al., 2013; Wu, Piao, Liu, Ciais, & Yao, 2018). Moreover, **conceptual SOC pools used in conventional models are largely not observable** (Abramoff et al., 2018; Elliot, Paustian, & Frey, 1996; Robertson et al., 2019), **making it challenging to validate conventional soil C models using field observations** (Six & Paustian, 2014; Viscarra Rossel et al., 2019).

New theories and soil biogeochemical models have been developed to explicitly represent microbial biomass and physiology

(Abramoff, Torn, Georgiou, Tang, & Riley, 2019; Abramoff et al., 2018; Allison, 2012; Campbell et al., 2016; Cotrufo, Wallenstein, Boot, Denef, & Paul, 2013; Huang et al., 2018; Robertson et al., 2019; Wieder, Grandy, Kallenbach, & Bonan, 2014). These microbial models are valuable for testing specific responses of SOC at small spatial scales, such as the effect of short-term priming observed during litter manipulation experiments or the addition of labile organic matter to the incubated soil samples in the laboratory. However, they introduce parameters determined from short-term experiments or under laboratory conditions. Thus, microbial models add uncertainty to large-scale simulations (Robertson et al., 2019; Shi et al., 2018; Stockmann et al., 2013; Wang et al., 2014) because most of these models are calibrated against observed litter or SOC decomposition rates obtained from limited laboratory or field experiments (Campbell et al., 2016; Georgiou, Abramoff, Harte, Riley, & Torn, 2017; Wieder, Grandy, et al., 2014). Robust datasets which can be used to comprehensively evaluate the simulated quasi-equilibrium SOC pool sizes are still scarce (Wieder, Boehner, & Bonan, 2014). Furthermore, it remains difficult to determine whether microbial explicit models outperform conventional first-order models on predicting large-scale SOC spatial gradients and temporal dynamics (Campbell & Paustian, 2015; Wieder, Grandy, Kallenbach, Taylor, & Bonan, 2015; Wieder et al., 2018). **Microbial models have to be carefully calibrated and evaluated before they are used to replace conventional first-order models in ESMs** (Wang et al., 2014; Wieder, Bonan, & Allison, 2013).

Several studies have calibrated and validated microbial decomposition models (Robertson et al., 2019; Wieder et al., 2013, 2015) using globally gridded soil databases such as the Harmonized World Soils Database (HWSD, FAO/IIASA/ISRIC/ISSCAS/JRC, 2012) and the Northern Circumpolar Soil Carbon Database (NCSDC, Tarnocai et al., 2009). However, these global databases do not contain uncertainty estimates (Dai et al., 2018), and previous studies have identified significant differences between SOC estimates from these databases or between grid-scale estimates from these databases and point-scale in situ observations (Tifafi, Guenet, & Hatté, 2018; Figure S1). In addition, there is still no reliable globally gridded database of plant litter input. Uncertainties in the boundary conditions (e.g. litter inputs simulated by ESMs and soil physical and

chemical properties) used as model forcing data further hamper the use of these global databases for model evaluation. An alternative approach is to calibrate and evaluate the microbial-explicit SOC models using extensive in situ observations of SOC contents, soil properties, litterfall production and climate conditions. Moreover, to ensure that the tested microbial model can capture many key processes related to SOC decomposition and stabilization, rather than only simulate the total SOC contents, it is necessary to evaluate the simulated composition of different C pools to total SOC, the turnover time of each C pool, and the sensitivity of SOC content to litter input and soil properties.

In this study, we compiled a large set of in situ observations of SOC concentrations for northern forests, as well as related soil property measurements (e.g. texture, bulk density and pH), annual litter input and climate from 206 forest sites distributed across different climate zones of Europe and China. Using this database, we calibrated and evaluated the first-order soil biogeochemical model CENTURY (Parton et al., 1987) and the microbial trait-based model MIMICS (Microbial-Mineral Carbon Stabilization, Wieder et al., 2015). To evaluate the simulated SOC composition, we acquired observations of the ratio of microbial biomass to total SOC, and the SOC fractions that represent the different SOC pools in the total SOC stock from sites that are independent from the European and Chinese sites.

The aim of this study is to assess the strength and weakness of microbial implicit and microbial explicit models in simulating the stocks and composition of SOC with the intent of guiding future experiments and model developments. Specifically, we (a) compared CENTURY and MIMICS with observed forest SOC concentrations at the continental scale, and explored the sources of model biases; (b) quantified the sensitivity of CENTURY- and MIMICS-simulated sensitivities of SOC concentration to changing soil conditions and litterfall inputs; (c) evaluated the MIMICS-simulated SOC compositions including ratios of microbial biomass to total SOC and the proportions of different SOC pools using observed values globally and (d) explored the main drivers of the variation in SOC composition. Finally, we discussed the implications of our results for SOC modeling at global scales.

## 2 | MATERIALS AND METHODS

### 2.1 | Observation data on SOC concentration and soil properties

To calibrate and evaluate both soil C models under a wide range of climate conditions and forest types, we compiled observed SOC concentrations and the corresponding plant biomass, litterfall, soil properties (e.g. bulk density, soil texture, pH) and climate conditions (mean annual temperature) from 72 European forest sites and 134 Chinese forest sites (Figure S2). The European sites are part of the International Co-operative Programme on Assessment and Monitoring of Air Pollution Effects on Forests (ICP Forests, <http://icp-forests.net>) operating under the UNECE Air Convention

and featuring consistent methods and harmonized data across the whole network (Fleck, Cools, De Vos, Meesenburg, & Fisher, 2016; Ukonmaanaho, Pitman, Bastrup-Birk, Breda, & Rautio, 2016). The Chinese forest sites belong to a reviewable and consistent nationwide inventory system established by the Chinese Ministry of Forestry (Tang et al., 2018). The forest stand ages at most sites are older than 40 years. In situ observations are mostly conducted during the period from 2000 to 2015, with durations ranging from one to more than 10 years. The observation sites cover four forest types (temperate needle-leaved evergreen forest (TeNE), temperate broad-leaved evergreen forest (TeBE), temperate broad-leaved summer-green forest (TeBS), boreal needle-leaved evergreen forest (BoNE)) and more than 15 soil types (based on the FAO-90 soil classification in HWSD v1.2). Mean annual temperatures of the observation sites span a large range from  $-10^{\circ}\text{C}$  to higher than  $20^{\circ}\text{C}$  (Figure S3a). Values of mean annual total precipitation ranged from less than 300 mm/year to more than 2,000 mm/year (Figure S3b). Annual total litterfall production was between  $100\text{ g C m}^{-2}\text{ year}^{-1}$  and  $2,000\text{ g C m}^{-2}\text{ year}^{-1}$  (Figure S3c). Soil properties at the observation sites vary widely (Figure S3d–i), with soil pH ranges from 4.5 to 8.5, and clay fraction ranges from 1% to 45%. Moreover, observation data at European ICP Forest sites provide measurements of SOC concentrations and soil properties at four different layers (0–10, 10–20, 20–40, 40–80 cm) of the top 80 cm soil, whereas data at Chinese sites provide the mean condition of the top 1 m soil.

At the European ICP Forest sites, leaf litterfall (including twig litterfall for some sites) was measured in situ, but not wood and root litterfall. We estimate the wood litterfall based on the ratios of wood litterfall to leaf litterfall, and the root litterfall based on the root turnover rates and the ratios of root biomass to leaf biomass (Table S1). At Chinese sites, there are no in situ observations of litterfall. We calculated the leaf, wood and root litterfall from observed standing biomass (including leaf, wood and root) and the annual leaf and root turnover rates and the ratios of wood litterfall to leaf litterfall (Table S1). The leaf and root turnover rate, the ratios of wood and root litterfall to leaf litterfall and the ratios of root biomass to leaf biomass used in this study were obtained from a statistical analysis of extensive global observations (Holland et al., 2015; Jia, Zhou, & Xu, 2016; Zhang, Yuan, Dong, & Liu, 2014; Figure S4).

C:N ratios of leaf litterfall at both European and Chinese sites were measured in situ. C:N ratios of wood and root litterfall, as well as the litterfall lignin:C ratios for each forest type were obtained from the global Fine-Root Ecology Database (FRED, Iversen et al., 2017), the TRY database (Kattge et al., 2011) and the Long-Term Inter-site Decomposition Experiment Team (LIDET, Harmon et al., 2009).

The soil base saturation (BS, %), Cation Exchange Capacity (CEC, cmol/kg) and soil gravel content (% of volume) at each observation site were obtained from the Global Soil Dataset for Earth System Models (GSDE, Shangguan, Dai, Duan, Liu, & Yuan, 2014). Soil type was determined based on the map from HWSD v1.2. Annual mean soil water content (%) was extracted from the estimation of land surface model ORCHIDEE-trunk (r5504, Krinner et al., 2005). LAI and

NDVI data were extracted from the GLASS (resolution: 0.05°, Liang et al., 2013) and GIMMS NDVI products (resolution: 8 km, Tucker et al., 2005), respectively. Evapotranspiration (ET) and the potential evapotranspiration (PET) were obtained from Jung et al. (2010) and the CRUNCEP v7 database (Viovy, 2018), respectively. More details of the datasets used in this study are shown in Table S1.

## 2.2 | Decomposition models

### 2.2.1 | CENTURY

We selected the CENTURY model (the version presented by Parton et al., 1987) to represent first-order soil biogeochemical models because it has been widely incorporated into ESMs (e.g. Koven et al., 2013; Krinner et al., 2005; Sitch et al., 2003). In CENTURY, organic matter is separated into metabolic litter (high quality,  $LIT_m$ ) and structural litter (low quality,  $LIT_s$ ) and three SOC pools (active pool ( $SOC_{act}$ ),

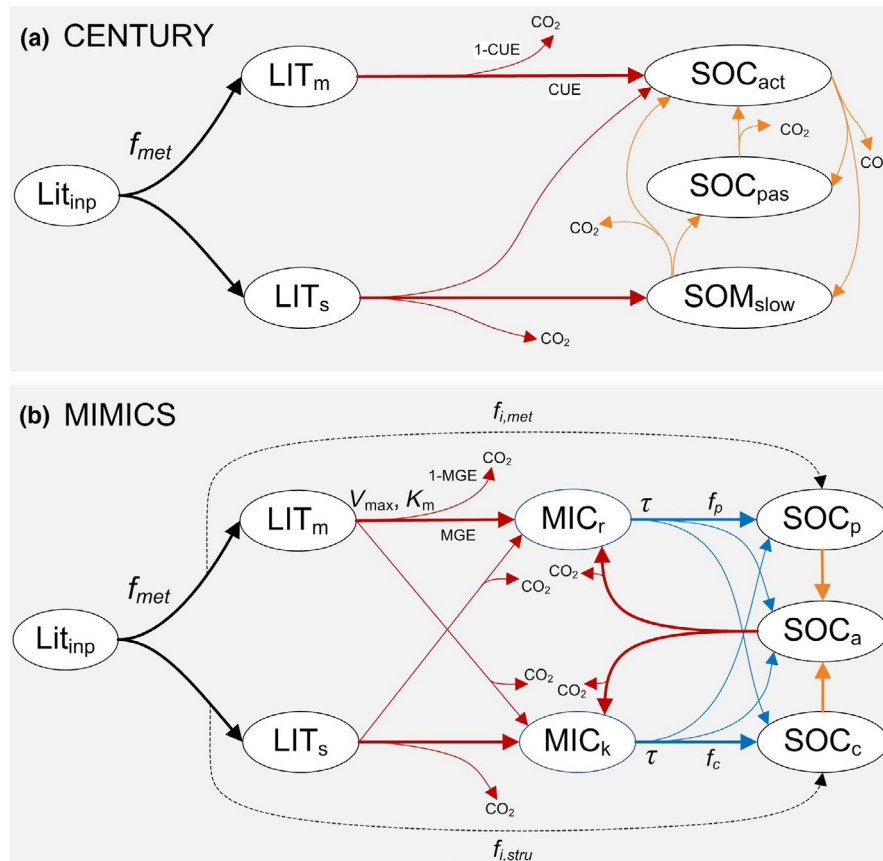
slow pool ( $SOC_{slow}$ ), passive pool ( $SOC_{pas}$ )) with different turnover times (Figure 1a). Fresh litter inputs are partitioned into metabolic and structural litter pools based on a linear function ( $f_{met}$ , dimensionless) of litter lignin to nitrogen (N) ratios (LN; Parton et al., 1987):

$$f_{met} = \max(0.0, 0.85 - 0.013 \times LN). \quad (1)$$

There is no explicit representation of microbial biomass in CENTURY. The decomposition of litter and SOC is described by first-order kinetics. At each daily time step, the decomposition of litter or SOC ( $\text{mg C cm}^{-3} \text{ day}^{-1}$ ) is calculated as following:

$$\frac{dC_s}{dt} = I_c - k_{max} \times C_s \times f(\text{tem}) \times f(\text{swc}) \times f(\text{clay}), \quad (2)$$

where  $C_s$  ( $\text{mg C cm}^{-3}$ ) is an individual litter or SOC pool,  $I_c$  ( $\text{mg C cm}^{-3} \text{ day}^{-1}$ ) is the C input to the pool considered,  $k_{max}$  is the potential maximum turnover rate of  $C_s$  ( $\text{day}^{-1}$ ) and is equal to the reciprocal of maximum turnover time.  $f(\text{tem})$ ,  $f(\text{swc})$  and  $f(\text{clay})$  are the soil



**FIGURE 1** Soil C pools and fluxes represented in CENTURY (a) and MIMICS (b). In both models, litter inputs ( $Lit_{inp}$ ) are partitioned into metabolic and structural litter pools ( $LIT_m$  and  $LIT_s$ ) based on litter quality ( $f_{met}$ ). The soil organic carbon (SOC) in CENTURY are divided into active ( $SOC_{act}$ ), slow ( $SOC_{slow}$ ) and passive ( $SOC_{pas}$ ) pools. CUE is the carbon use efficiency of decomposed litter or SOC. In MIMICS, decomposition of litter and available SOM pools ( $SOC_a$ ) are governed by temperature sensitive Michaelis–Menten kinetics ( $V_{max}$  and  $K_m$ ). Microbial growth efficiency (MGE) determines the partitioning of C fluxes entering microbial biomass pools versus heterotrophic respiration. Turnover of the microbial biomass ( $\tau$ ) depends on microbial functional type ( $MIC_r$  and  $MIC_k$ ), and is partitioned into available, physically and physicochemically protected, and chemically recalcitrant SOC pools ( $SOC_a$ ,  $SOC_p$ , and  $SOC_c$ , respectively).  $f_{i,met}$  and  $f_{i,STRU}$  denote the fraction of decomposed metabolic litter to  $SOC_p$  and the fraction of decomposed structural litter to  $SOC_c$ , respectively.  $f_p$  and  $f_c$  denote the fraction of  $\tau$  partitioned to  $SOC_p$  and the fraction of  $\tau$  partitioned to  $SOC_c$ , respectively [Colour figure can be viewed at [wileyonlinelibrary.com](https://onlinelibrary.wiley.com/doi/10.1111/gcb.14994)]

temperature factor, moisture factor and clay factor modulating decomposition rate, respectively.

## 2.2.2 | MIMICS (default and modified versions)

### The default version of MIMICS (MIMICS-def)

The MIMICS model (Wieder, Grandy, et al., 2014; Wieder et al., 2015) explicitly considers the relationships among litter quality, functional trade-offs in microbial physiology, and the physical and physicochemical protection of microbial byproducts in forming stable soil organic matter. Like CENTURY, MIMICS also has two types of litter pool: metabolic ( $LIT_m$ ) and structural ( $LIT_s$ ) litter (Figure 1b), and the method used to partition fresh litter input into metabolic and structural pools ( $f_{met}$ , Figure 1b) is the same as that used in CENTURY (Equation 1). SOC in MIMICS is divided into three pools: physically and physicochemically protected ( $SOC_p$ ), the chemically recalcitrant ( $SOC_c$ ) and available ( $SOC_a$ ). Two microbial functional types are represented in MIMICS that roughly correspond to microorganisms with copiotrophic (r-strategy,  $MIC_r$ ) and oligotrophic (k-strategy,  $MIC_k$ ) growth strategies (Figure 1b). The  $MIC_r$  is assumed to have higher growth and turnover rates and prefers to consume more labile litter ( $LIT_m$ ), whereas the  $MIC_k$  has relatively lower growth and turnover rates and is more competitive when consuming low-quality litter ( $LIT_s$ ) and chemically recalcitrant SOC ( $SOC_c$ ).

C fluxes in MIMICS are simulated at an hourly (h) time step. Decomposition of litter and SOC pools ( $mg\ C\ cm^{-3}\ hr^{-1}$ ) is based on temperature-sensitive Michaelis–Menten kinetics (Allison, Wallenstein, & Bradford, 2010; Schimel & Weintraub, 2003) through the equation:

$$\frac{dC_s}{dt} = I_c - MIC \times \frac{V_{max} \times C_s}{K_m + C_s}, \quad (3)$$

where  $C_s$  ( $mg\ C/cm^3$ ) is a substrate pool ( $LIT$  or  $SOC$ ) and  $MIC$  ( $mg\ C/cm^3$ ) corresponds to the biomass of each microbial pool ( $MIC_r$  or  $MIC_k$ ).  $I_c$  is the C input to the pool considered ( $mg\ C\ cm^{-3}\ hr^{-1}$ ).  $V_{max}$  and  $K_m$  are the microbial maximum reaction velocity ( $mg\ C\ (mg\ MIC)^{-1}\ hr^{-1}$ ) and half-saturation constant ( $mg\ C/cm^3$ ), respectively. They are calculated as follows:

$$V_{max} = e^{V_{slope} \times T + V_{int}} \times av \times V_{mod}, \quad (4)$$

$$K_m = e^{K_{slope} \times T + K_{int}} \times ak \times K_{mod}, \quad (5)$$

where  $T$  is the soil temperature ( $^{\circ}C$ ),  $V_{mod}$  and  $K_{mod}$  represent the modifications of  $V_{max}$  and  $K_m$  based on assumptions regarding to microbial functional types, litter chemical quality and soil texture effects,  $av$  and  $ak$  are the tuning coefficient of  $V_{max}$  and  $K_m$ , respectively.  $V_{slope}$  and  $K_{slope}$  are two regression coefficients.  $V_{int}$  and  $K_{int}$  are the regression intercepts.

Decomposition rate of substrates and the microbial growth efficiency (MGE, Figure 1b) determine the growth rate of microbes.

The turnover of  $MIC_r$  and  $MIC_k$  ( $MIC_r, mg\ C\ cm^{-3}\ hr^{-1}$ ) at each time step is calculated based on their specific turnover rate ( $k_{mic}, hr^{-1}$ ), annual total litterfall input ( $LIT_{tot}, g\ C\ m^{-2}\ year^{-1}$ ) and  $f_{met}$  by following:

$$MIC_r = a_r \times k_{mic} \times e^{c \times f_{met}} \times \max\left(\min\left(\sqrt{LIT_{tot}}, 1.2\right), 0.8\right) \times MIC, \quad (6)$$

where  $a_r$  ( $=1.0$ , dimensionless) is a tuning coefficient of  $k_{mic}$ .  $c$  is the regression coefficients, and its value is 0.3 for  $MIC_r$  and 0.1 for  $MIC_k$ . Turnover of microbial biomass provides C inputs to SOC pools (Figure 1b). The fractions of microbial residues to different SOC pools are determined by soil clay content ( $f_{clay}$ ) and the quality of litter inputs (lignin:N), and can be specifically calculated by following:

$$f_{rp} = \min\left(1.0, a_1 \times e^{1.3 \times f_{clay}}\right), \quad (7)$$

$$f_{kp} = \min\left(1.0, a_2 \times e^{0.8 \times f_{clay}}\right), \quad (8)$$

$$f_{rc} = \min\left(1.0 - f_{rp}, a_4 \times e^{a_3 \times f_{met}}\right), \quad (9)$$

$$f_{kc} = \min\left(1.0 - f_{kp}, a_5 \times e^{a_3 \times f_{met}}\right), \quad (10)$$

$$f_{ra} = 1.0 - f_{rp} - f_{rc}, \quad (11)$$

$$f_{ka} = 1.0 - f_{kp} - f_{kc}, \quad (12)$$

where  $f_{rp}$ ,  $f_{kp}$ ,  $f_{rc}$ ,  $f_{kc}$ ,  $f_{ra}$  and  $f_{ka}$  represent the fractions of  $MIC_r$  and  $MIC_k$  residues to  $SOC_p$ ,  $SOC_c$  and  $SOC_a$ , respectively. LN is the lignin:N ratio.  $a_{1-5}$  are coefficients and their values in default MIMICS are found in Table S1. In addition to microbial residues, a fraction of inputs ( $f_{i,met}$  and  $f_{i,STRU}$ ) which bypasses litter and microbial biomass pools is transferred directly to corresponding SOC pools (Figure 1b).

The transfer of  $SOC_p$  to  $SOC_a$  ( $D, mg\ C\ cm^{-3}\ hr^{-1}$ ), which is intended to represent the deprotection of SOC, that is, desorption of physicochemically protected SOC from mineral surfaces and/or the breakdown of aggregates deprotecting physically protected SOC, is calculated as a function of soil clay content ( $f_{clay}$ ) by following:

$$D = 1.5 \times 10^{-5} \times k_d \times e^{-1.5 \times f_{clay}}, \quad (13)$$

where  $k_d$  ( $=1.0$ , dimensionless) is a tuning coefficient of the deprotection rate. Some parameter values of the default MIMICS are provided in Table S1. Please see Wieder, Grandy, et al. (2014), Wieder et al. (2015) for more details of the structure, algorithms, parameters and underlying assumptions of MIMICS.

### MIMICS with revised SOC deprotection rate (MIMICS-D)

In addition to the default version of MIMICS (MIMICS-def), we also developed and tested a new version of MIMICS (MIMICS-D) that considers the saturation of SOC protected by the mineral



matrix ( $\text{SOC}_p$ ). In the MIMICS-def, the deprotection rate of  $\text{SOC}_p$  in a specific soil was a fixed value determined by the abundance of the soil clay fraction (Equation 13). However, field and laboratory research suggests that there might be an upper limit, or 'saturation level', in the amount of physicochemically and physically protected SOC that can be held in soil (Robertson et al., 2019; Six et al., 2002; Stewart, Paustian, Conant, Plante, & Six, 2007). Deprotection rate of the SOC protected by the mineral matrix is closely related to this saturation degree (defined as the ratio of existing  $\text{SOC}_p$  to the soil maximum adsorption capacity; Kothawala, Moore, & Hendershot, 2008; Wang, Post, & Mayes, 2013). In this study, we did not calculate the maximum adsorption capacity directly, as it is determined by soil physical and chemical characteristics, and there is still no widely recognized method to calculate it (Campbell & Paustian, 2015; Huang et al., 2018; Lützow et al., 2006). The upper limit of  $\text{SOC}_p$  was represented by assuming that the deprotection rate increases exponentially with the pool size of  $\text{SOC}_p$ :

$$D = 1.5 \times 10^{-5} \times k_d \times e^{-1.5 \times f_{\text{clay}}} \times e^{k_{\text{dp}} \times \text{SOC}_p}, \quad (14)$$

where  $k_{\text{dp}}$  is a coefficient for tuning the relationship between the deprotection rate ( $D$ ) and the pool size of  $\text{SOC}_p$ .

#### MIMICS considering the impact of BS on deprotection rate (MIMICS-DB)

We tested several new modifications of MIMICS to see whether the inclusion of soil chemical properties (BS and pH) could further decrease the uncertainties in simulated SOC concentrations. We modified the microbial maximum reaction velocity ( $V_{\text{max}}$ , Equation 4), the C input rates to  $\text{SOC}_p$  ( $f_p$  and  $f_{i,\text{met}}$  in Figure 1b) and the deprotection rate of  $\text{SOC}_p$  with some simple linear or exponential functions of soil BS and pH, separately. In this study, we only present the results from the modification called MIMICS-DB, where the modified deprotection rate of  $\text{SOC}_p$  is calculated as:

$$D = 1.5 \times 10^{-5} \times k_d \times e^{-1.5 \times f_{\text{clay}}} \times e^{k_{\text{dp}} \times \text{SOC}_p} \times e^{k_{\text{bs}} \times \text{BS}}, \quad (15)$$

where  $k_{\text{bs}}$  is a coefficient modifying the impacts of BS on the deprotection rate.

#### MIMICS considering density-dependent microbial turnover rate (MIMICS-DBT)

Following the method of Georgiou et al. (2017), we also incorporated a density-dependent microbial turnover rate into MIMICS. In this version (MIMICS-DBT), microbial turnover rate increases with growing microbial biomass density (MIC, mg C/cm<sup>3</sup>) by modifying Equation (6):

$$\text{MIC}_r = a_r \times k_{\text{mic}} \times e^{c \times f_{\text{met}}} \times \max\left(\min\left(\sqrt{\text{LIT}_{\text{tot}}}, 1.2\right), 0.8\right) \times (\text{MIC})^\beta, \quad (16)$$

where  $\beta$  is the density-dependence exponent.

## 2.3 | Model parameterization and validation against SOC concentrations

We assumed that all the forest sites included in this study are at steady state (i.e. no interannual variation of SOC, litterfall and stand biomass). CENTURY and the four versions of MIMICS introduced above (Table 1) were then calibrated and evaluated against the 'equilibrium' SOC concentrations using observation data of soil texture, annual total litterfall and mean annual temperature. We also ignored the interannual and seasonal dynamics of climate and vegetation. Historical climate, litterfall input and soil properties were all assumed to be similar to the average condition during the observation period. Vertical discretization in SOC and soil properties are not considered in CENTURY and MIMICS. We focus only on the spatial variation of average SOC concentrations in the upper soil horizons (0–80 cm for European sites and 0–1 m for Chinese sites). The semi-analytic approach was used to calculate the steady state microbial and soil C pool sizes (Xia, Luo, Wang, Weng, & Hararuk, 2012) based on annual total litterfall production (evenly distributed to each time step of simulation), annual mean soil temperature and model conditions and observed soil properties at each forest site. Parameters of CENTURY and MIMICS were optimized against the observed SOC concentrations (Table 1). Although many parameters (e.g. carbon use efficiency and parameters related to the constraints of temperature and soil clay on C decomposition rate) of CENTURY and MIMICS can impact the simulated SOC concentrations, we only optimized the parameters which directly control the organic matter decomposition rates. Because these parameters generally contain large uncertainties and the simulated SOC stocks are generally more sensitive to these parameters than to other model parameters (Shi et al., 2018; Wieder, Grandy, et al., 2014; Wieder et al., 2015). Specifically, we added two scaling parameters  $k_{\text{litt}}$  and  $k_{\text{soc}}$  (dimensionless) in CENTURY to tune the turnover rates of litter and SOC pools, respectively.

$$k_{\text{max\_litt\_opt}} = k_{\text{litt}} \times k_{\text{max\_litt}}, \quad (17)$$

$$k_{\text{max\_soc\_opt}} = k_{\text{soc}} \times k_{\text{max\_soc}}, \quad (18)$$

**TABLE 1** Tested models in this study and parameters subject to optimization of each model

Model	Optimized parameters
CENTURY	$k_{\text{litt}}, k_{\text{soc}}$
MIMICS-def	$a_v, a_k, k_d$
MIMICS-D	$a_v, a_k, k_d, k_{\text{dp}}$
MIMICS-DB	$a_v, a_k, k_d, k_{\text{dp}}, k_{\text{bs}}$
MIMICS-DBT	$a_v, a_k, k_d, k_{\text{dp}}, k_{\text{bs}}, \beta$

Note:  $k_{\text{litt}}$  and  $k_{\text{soc}}$  tune the turnover rate of litter and SOC pools in CENTURY, respectively.  $a_v$  and  $a_k$  are parameters tune microbial maximum reaction velocity (Equation 4) and half-saturation constant (Equation 5).  $k_d, k_{\text{dp}}$  and  $k_{\text{bs}}$  tune the deprotection rate of  $\text{SOC}_p$  (Equations 13–15).  $\beta$  tunes the density-dependent microbial turnover rate (Equation 16).

where  $k_{\max\_litt}$  and  $k_{\max\_litt\_opt}$  are the default and optimized litter turnover rates, respectively.  $k_{\max\_soc}$  and  $k_{\max\_soc\_opt}$  are the default and optimized SOC turnover rates, respectively. The default litter and SOC turnover rates (see Table S2) were obtained from Parton et al. (1987). Optimization of only  $k_{litt}$  and  $k_{soc}$  may be not enough to minimize the uncertainties in the turnover rates of litter and SOC pools and the simulated SOC concentrations. We therefore also tested the effectiveness of CENTURY on capturing observed SOC concentrations when five free parameters were introduced to tune the turnover rates of metabolic litter, structural litter, active SOC, slow SOC and passive SOC, respectively (Figure S5).

For the MIMICS models, we optimized the scaling parameters ( $av$ ,  $ak$  and  $k_d$ ) of the microbial maximum reaction velocity ( $V_{\max}$ , Equation 4), half-saturation constant ( $K_m$ , Equation 5) and of the deprotection rate of  $SOC_p$  (Equations 13–15), as they are all closely related to the decomposition and the physical stabilization of organic matter (Wieder, Grandy, et al., 2014; Wieder et al., 2015). Parameters in the newly introduced equations (Equations 14–16) for modifying deprotection rates and microbial turnover rate were also optimized (Table 1).

Parameter optimization was performed using the shuffled complex evolution (SCE) algorithm developed by Duan, Gupta, and Sorooshian (1993), Duan, Sorooshian, and Gupta (1994), which has proven to be effective for global optimization by many previous studies (e.g. Franchini, Galeati, & Berra, 2009; Muttill & Jayawardena, 2008). Prior value and the range of each parameter used for the SCE algorithm are listed in Table S3. Root mean square error (RMSE, Equation 19) between simulated ( $SOC_{sim,i}$ ) and observed ( $SOC_{obs,i}$ ) SOC concentrations (g C/kg soil) was used as the objective function, and parameters that minimized the RMSE were regarded as optimal.

$$RMSE = \sqrt{\left( \frac{\sum_{i=1}^n (SOC_{obs,i} - SOC_{sim,i})^2}{n} \right)}, \quad (19)$$

where  $n$  is the number of observation sites. In addition to RMSE, the Akaike information criterion (AIC, Equation 20, Akaike, 1974), which considers both the goodness of fit and the number of free model parameters ( $n_{param}$ ), were also used to evaluate the optimized models (Table 1).

$$AIC = n \times \ln \left( \frac{\sum_{i=1}^n (SOC_{obs,i} - SOC_{sim,i})^2}{n} \right) + 2n_{param}. \quad (20)$$

Our preliminary analyses indicated that parameter optimizations of MIMICS based solely on observed SOC concentration might result in unrealistic estimates of SOC composition (e.g. the  $SOC_p$  pool approaching to zero at all sites) if turnover rates (e.g. the  $SOC_p$  turnover rates being significantly larger than  $SOC_a$ ), although the simulated concentrations of total SOC agreed well with the observations. To mitigate this problem, some additional constraints on simulated SOC composition and turnover rates were

incorporated into our optimization scheme (see below). Parameter sets that did not meet the imposed constraints on SOC composition and turnover rates were excluded. Note that the simulated turnover rates of different SOC pools from CENTURY are always consistent with the definition of SOC pools (i.e. the active pool has the largest turnover rate, followed by the slow pool, and the passive pool has the lowest turnover rate), and the simulated SOC composition (mainly determined by the turnover rate of each pool, see Section 3.2) did not show any 'abnormalities' (i.e. no simulated SOC pool declined to very small values approaching zero), so we did not incorporate additional constraints when optimizing the parameters of CENTURY.

Previous studies suggest that the organic C associated with soil minerals or stored within soil aggregates, corresponding to the  $SOC_p$  pool of MIMICS, is the most stable fraction of SOC with turnover times approaching hundreds to thousands of years. Furthermore, the recalcitrant SOC fractions composed by structurally complex compounds corresponding to the  $SOC_c$  pool of MIMICS generally have longer turnover time than the labile SOC fraction (Benbi, Boparai, & Brar, 2019; Robertson et al., 2019; Sokol, Sanderman, & Bradford, 2019). Therefore, we set a constraint that the simulated mean  $SOC_p$  turnover time for all of the 206 observation sites must be longer than that of  $SOC_c$ , and that the mean  $SOC_c$  turnover time must be longer than  $SOC_a$ .

Observations found that a large fraction (e.g. 10%–50%) of SOC is in stable pool (Barré et al., 2010; Benbi et al., 2014; Lützw et al., 2007; Viscarra Rossel et al., 2019). To avoid the optimized parameters giving a very low (approaching to zero) estimate of the fraction of  $SOC_p$ , we also added as a constraint of model results with optimized parameters that the simulated average proportion of  $SOC_p$  at the 206 observation sites (not for every individual site) must be larger than 5%, that average proportion of  $SOC_c$  cannot exceed 70%, and that the total amount of  $SOC_p$  and  $SOC_c$  should be higher than  $SOC_a$ .

Note that the parameters ( $a_{1-5}$  in Equations 7–10) controlling the partition of microbial residues to different SOC pools were modified before the parameters listed in Table 1 are optimized because MIMICS did not give reasonable estimates of the SOC concentrations, compositions and the turnover rates simultaneously when only the parameters listed in Table 1 were calibrated. The modified values of  $a_{1-5}$  are provided in Table S2.

To explore the sources of simulation errors (i.e. the difference between simulated and observed SOC concentrations), we first calculated the partial correlation coefficient between the errors of the simulated SOC concentration and different soil (e.g. texture, pH, BS and CEC), plant (NDVI and LAI) and climate (temperature, precipitation, ET) variables (see Section 2.1 and Table S1 for the source of each variable). Then, we fitted a linear mixed-effects (LME) model to quantify the combined contribution of the fixed effects (soil, plant and climate variables listed above) and site-specific random effects (e.g. soil type, forest type, stand age and micro-topography) on explaining the simulation errors. All the important variables that might potentially affect SOC dynamics, for example soil texture,

temperature, pH, moisture, BS, CEC, bulk density, litterfall inputs, precipitation and ET, were included as fixed effects in the LME. Observation site was used as a random effect. We also fitted a multiple linear regression (MLR) with all of the fixed effects of the LME as the predictor variables to quantify the relative contributions of fixed effects and random effects to the simulation errors. Then, the relative contributions of fixed effects and random effects were quantified based on the coefficient of determination of the LME ( $R^2_{\text{LME}}$ ) and MLR ( $R^2_{\text{MLR}}$ ). The contributions of model choice ( $f_{\text{model}}$ ), fixed effects ( $f_{\text{fixed}}$ ) and random effects ( $f_{\text{random}}$ ) to explaining the variation of SOC concentrations can be quantified by:

$$f_{\text{model}} = R^2_{\text{model}} \quad (21)$$

$$f_{\text{fixed}} = R^2_{\text{MLR}} \times (1 - R^2_{\text{model}}), \quad (22)$$

$$f_{\text{model}} = (R^2_{\text{LME}} - R^2_{\text{MLR}}) \times (1 - R^2_{\text{model}}), \quad (23)$$

where  $R^2_{\text{model}}$  is the determining coefficient of the regression equation between simulated and observed SOC concentrations.

## 2.4 | Model evaluation against sensitivities of SOC concentrations to key model drivers

To assess whether each model simulated the variations of SOC concentrations for the right reasons, we first identified the key drivers of the spatial variations of SOC concentration, and then compared modelled sensitivities of SOC concentration to these drivers to the values derived from the observations. **The potential key drivers we evaluated include soil temperature, moisture, clay content, litterfall input, the mean C:N ratio and the lignin:C ratio of litterfall.** The sensitivities of organic matter decomposition rate to manipulated soil temperature, moisture and litter inputs have been widely investigated via laboratory and field experiments (Bonan et al., 2013; Parton et al., 2007; Sierra, Trumbore, Davidson, Vicca, & Janssens, 2015). However, no experiments have measured the sensitivity of equilibrium SOC stock to changing soil properties and litter inputs, as it would take decades to hundreds of years for the SOC pool to reach equilibrium after manipulating litter. Here we estimated the sensitivities by making use of observed spatial variation of SOC with different drivers, including soil temperature, water content, clay fraction, annual total litter input and the C:N ratio and lignin:C ratio of litter input. We assumed the soil-litter system is in steady state, and the sensitivities of equilibrium SOC to different drivers were quantified by multiple linear regression. The regression coefficient of each driver was regarded as the observed sensitivity.

The sensitivities of simulated SOC concentration to soil and litter properties from optimized CENTURY and MIMICS were obtained using Monte Carlo simulations. We sampled 1,000 sets of unique soil and litter input condition within the observed space of each variable using Latin Hypercube technique (Tang & Zhuang,

2009). All soil and litter variables were assumed to be uniformly distributed and the range of each variable was set based on the maximum and minimum observed values at the European and Chinese sites. For each combination of soil and litter input condition, the sensitivity ( $S_i$ ) of SOC concentration to each variable ( $d_i$ ) was calculated as

$$S_i = \frac{f(d_1, d_2, \dots, d_i + \delta, \dots, d_n) - f(d_1, d_2, \dots, d_i, \dots, d_n)}{\delta}, \quad (24)$$

where  $\delta$  is the step size of a change in variable  $d_i$  assumed to be 1% of the difference between maximum and minimum  $d_i$  (i.e.  $\delta = (d_{i_{\text{max}}} - d_{i_{\text{min}}})/100$ ).

## 2.5 | Model evaluation against SOC composition

We evaluated the simulated proportions of the different SOC pools using observations from sites that are independent of the European and Chinese forest sites, for which the model parameters were calibrated. The simulated ratios of microbial biomass to total SOC were validated against 655 observations from forest sites around the world (Xu, Thornton, & Post, 2013). The simulated SOC composition from CENTURY and MIMICS was compared to measurements of SOC composition from 505 sites under native forests and grasslands in Australia (Viscarra Rossel & Hicks, 2015; Viscarra Rossel et al., 2019). These data were partitioned into three fractions, the particulate organic C (POC), humic organic C (HOC) and resistant organic C (ROC, which is the mineral-associated organic carbon) based on the particle size and chemical compositions of organic matter. We acknowledge the fact that the observed pools are not modelled conceptual pools and we propose a correspondence between both in Table S4. We compared the simulated SOC pools to the observed SOC fractions to assess their correspondence in terms of their expected/assumed turnover rates.

## 2.6 | Model evaluation against the key drivers of variations in SOC composition

To determine whether the key drivers of variations in SOC composition in MIMICS and CENTURY models are consistent with the observations, we calculated the partial correlation coefficient between fraction of each SOC pool and different model drivers using the simulated proportions of different SOC pools by optimized MIMICS and CENTURY models at all of the 206 forest sites in Europe and China (Figure S2), and using the observed proportions of different SOC pools at the 505 Australia sites (Viscarra Rossel et al., 2019). The key drivers we considered in this analysis include soil temperature, moisture, clay fraction, BS, annual litterfall input, litter C:N and lignin:C ratios and the total SOC pool size. For each model driver, all of the other drivers described above were used as the controlling factor for calculating the partial correlation coefficient.



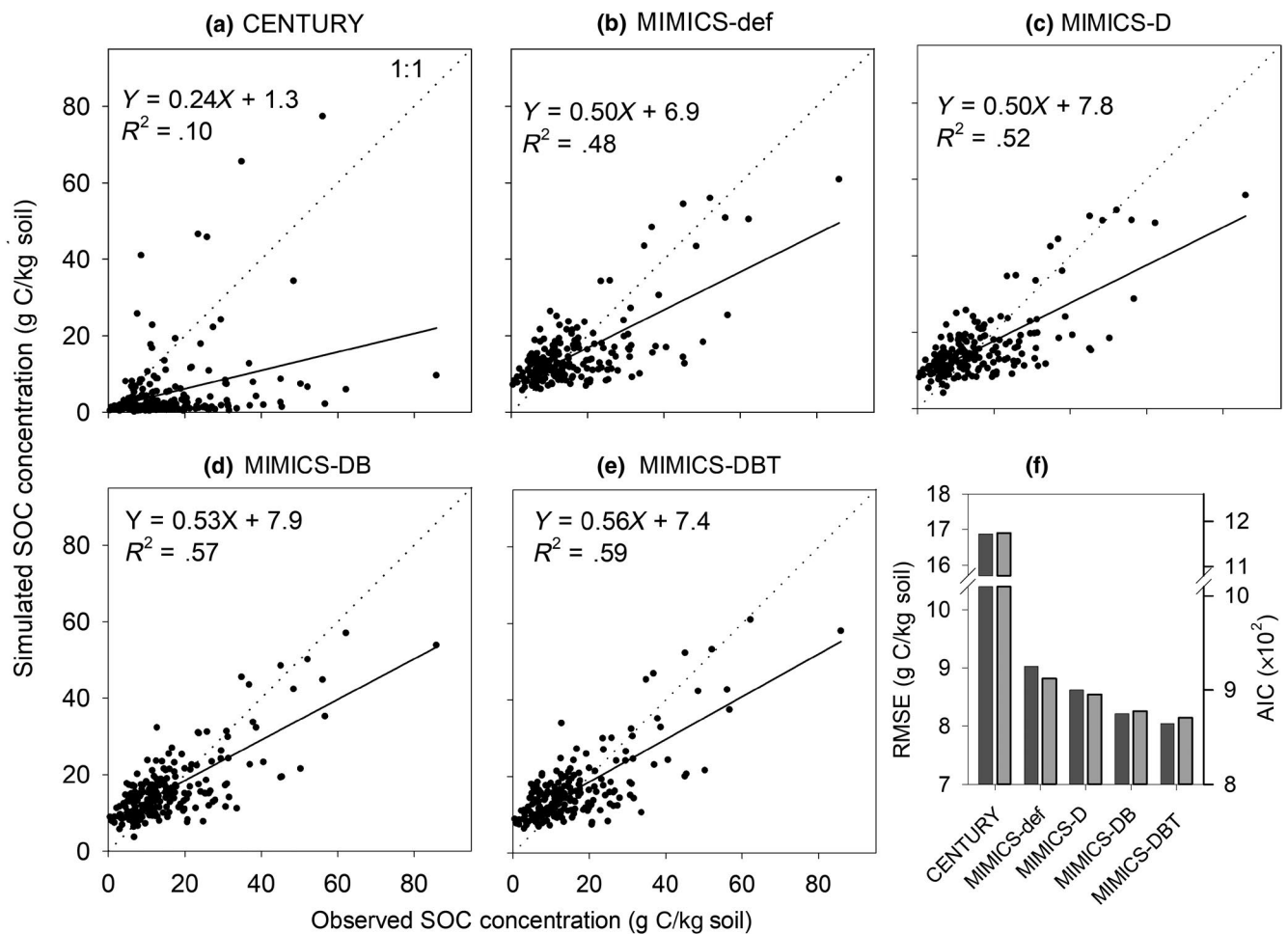
### 3 | RESULTS

#### 3.1 | Evaluation of simulated SOC concentrations

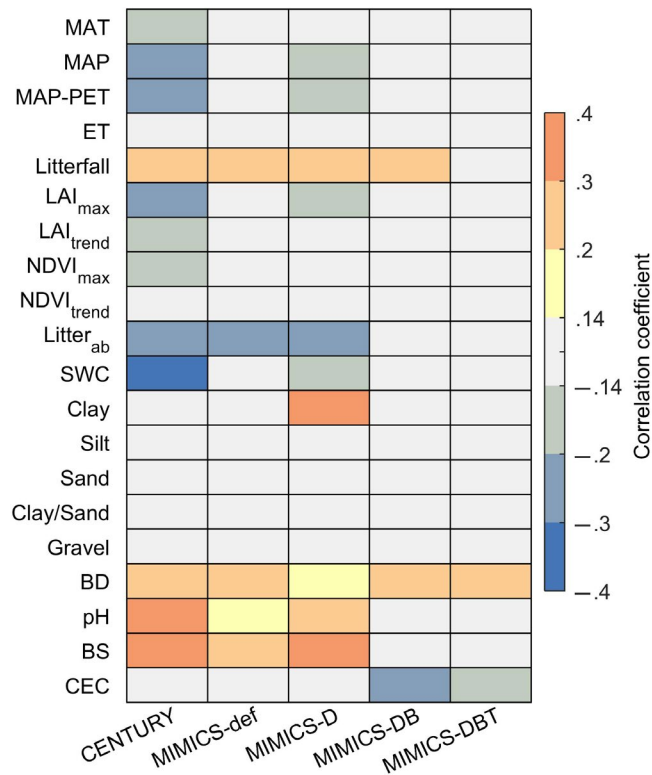
Our evaluation indicates that MIMICS can better capture the observed spatial variation of SOC concentrations than CENTURY across European and Chinese forest sites. The default version MIMICS-def explains 48% observed SOC spatial variation, as compared to only 10% by CENTURY model (Figure 2). MIMICS-D, MIMICS-DB and MIMICS-DBT explain 52%, 57% and 59% SOC spatial variation, respectively (Figure 2). The RMSE and AIC indicate that all MIMICS versions estimate the spatial variation of SOC concentration more accurately than CENTURY, with MIMICS-DBT having the best performance overall (Figure 2f). We also note that the CENTURY model with five free parameters for tuning turnover rates of litter and SOC pools (Figure S5a) does not estimate SOC concentrations more accurately than the CENTURY with two free parameters (Table 1). CENTURY with five free parameters has a slightly smaller RMSE (16.89) but a higher AIC (1,174.7) than the

RMSE (16.97) and AIC (1,170.5), respectively, from CENTURY with two parameters (Figure S5a).

There are systematic biases in the simulated SOC concentrations along the gradients of SOC pool size, soil properties, and climate and plant variables (Figure 3 and Figure S6). Both CENTURY and MIMICS overestimate the low SOC concentrations but underestimate the high concentrations (Figure 2 and Figure S6). The simulation biases of CENTURY are significantly correlated with soil (e.g. moisture, BS, pH and bulk density), plant (e.g. litterfall, LAI) and climate (e.g. mean annual temperature and annual total precipitation) variables (Figure 3), suggesting that CENTURY has structural biases in the processes depending upon those factors. Similar to CENTURY, the simulation bias of MIMICS is also significantly correlated with some soil and litterfall-related variables. By including the effect of BS on deprotection rate into MIMICS (MIMICS-DB), the significant relationships between simulation biases and soil, plant and climate variables are largely eliminated, but a significant negative relationship between simulation biases and soil CEC appears. The significant relationship between simulation biases and



**FIGURE 2** Comparison of CENTURY (a) and MIMICS (b–e) for simulating large-scale variation of soil organic carbon (SOC) concentrations across the 206 forest sites in Europe and China. RMSE (f) is the root mean square error, and AIC (f) is the Akaike information criterion. MIMICS versions include the default model (MIMICS-def), revised SOC deprotection rate (MIMICS-D), using base saturation to modify deprotection rates (MIMICS-DB); and density-dependent microbial turnover rate (MIMICS-DBT; see Section 2.2.2)



**FIGURE 3** Partial correlation coefficients between the biases of simulated SOC concentrations and the climate condition, amount and quality of litter input, and soil physical and chemical properties. MAT: mean annual temperature ( $^{\circ}\text{C}$ ), MAP: mean annual total precipitation (mm), MAP-PET: the difference between annual total precipitation and potential evapotranspiration (mm), ET: evapotranspiration (mm),  $\text{LAI}_{\text{max}}$ : mean of the annual maximum leaf area index at the observation site during the period from 1982 to 2000,  $\text{LAI}_{\text{trend}}$ : change trend of the  $\text{LAI}_{\text{max}}$  during the period from 1982 to 2000 ( $\text{year}^{-1}$ ),  $\text{NDVI}_{\text{max}}$ : mean of the annual maximum normalized difference vegetation index at the observation site during the period from 1982 to 2000,  $\text{LAI}_{\text{trend}}$ : change trend of the  $\text{NDVI}_{\text{max}}$  during the period from 1982 to 2000 ( $\text{year}^{-1}$ ),  $\text{litter}_{\text{ab}}$ : aboveground litter-C stock ( $\text{g C m}^{-2}$ ), SWC: soil water content, BD: bulk density ( $\text{g cm}^{-3}$ ), BS: base saturation (0–1, dimensionless), CEC: Cation of exchange capacity ( $\text{cmol kg}^{-1}$ ). Partial correlation coefficients between  $-0.14$  and  $0.14$  were not significant ( $p > .05$ ) [Colour figure can be viewed at [wileyonlinelibrary.com](https://onlinelibrary.wiley.com)]

annual litterfall input can be eliminated only when the density-dependence of microbial turnover rate in MIMICS-DBT is represented. Moreover, the simulation biases of all models are positively related to soil bulk density (Figure 3).

Soil properties, litter input rate and the plant and climate conditions together can only explain a small portion of the simulation biases in SOC concentrations, especially for MIMICS (Figures S7 and S8). The linear mixed-effects (LME) models which consider both fixed factors (i.e. the soil, litter and climate variables) and site-specific random factor (e.g. soil type, forest type, stand age and micro-topography) explain most of the variations in the simulation biases (Figure S7). Further statistics indicated that the SOC variation explained by CENTURY, fixed factors and random factors

are 10%, 27% and 54%, respectively (Figure S8). But for MIMICS, the model itself explained the largest part (48%–59%) of SOC variation, followed by the random factor (24%–32%), with fixed factors explaining 5%–9% of SOC variation (Figure S8). Our further analysis on the potential contributors to random factors indicated that CENTURY estimations of SOC are consistently biased regardless of soil type, plant type and stand age (Figure S9). But the estimations of SOC made by MIMICS are, with few exceptions, unbiased across sites with different soil types, plant types and stand ages. Overall, the constraints of soil, litter and climate factors on SOC stocks are significantly better represented in MIMICS than in CENTURY.

### 3.2 | Evaluation of simulated sensitivities of SOC concentration to key model drivers

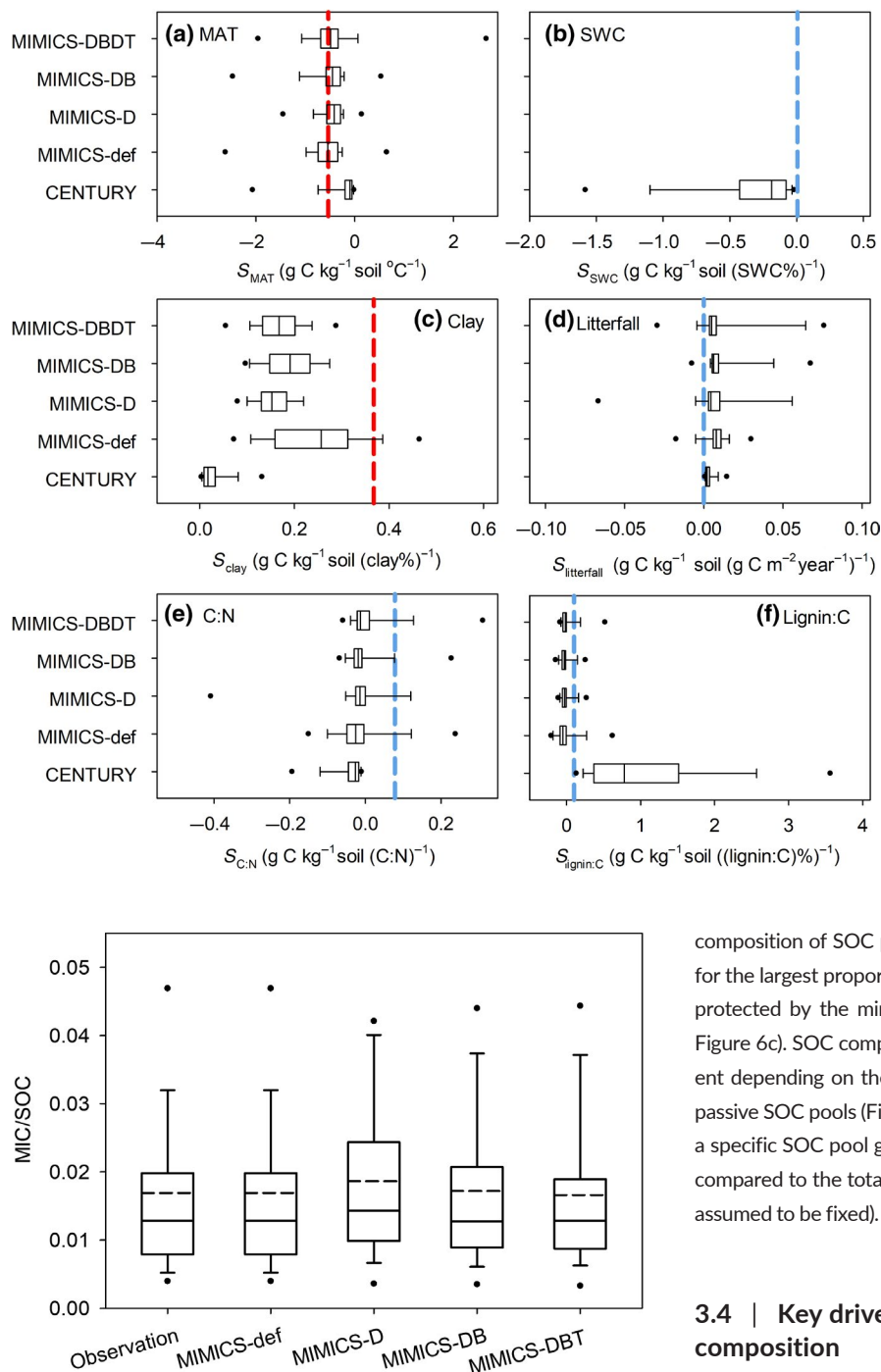
Based on observations, SOC concentrations are sensitive to local soil temperature and soil clay content (Figure 4a,c), but are not sensitive to local soil moisture or litter quantity and quality (Figure 4b,d,e,f). On average, SOC concentration declines by  $0.53 \text{ g C/kg soil}$  with a  $1^{\circ}\text{C}$  increase in soil temperature, and increases by  $0.37 \text{ g C/kg soil}$  with a 1% increase in soil clay fraction.

MIMICS models provide more accurate estimates of the observation-based partial sensitivity of SOC concentration to changes in soil temperature, compared to CENTURY (Figure 4a). With a  $1^{\circ}\text{C}$  increase in soil temperature, the simulated SOC concentration declines by  $0.4$ – $0.55 \text{ g C/kg soil}$  (median value) depending on the version of MIMICS. The sensitivity is comparable to the value calculated based on observation data, but significantly lower than the value simulated by CENTURY ( $-0.92 \pm 4.1 \text{ g C kg}^{-1} \text{ soil } ^{\circ}\text{C}^{-1}$ ). Both CENTURY and MIMICS underestimate the observed sensitivity of SOC to soil clay fraction. Despite this, the sensitivities estimated by MIMICS ( $0.17$ – $0.26 \text{ g C kg}^{-1} \text{ soil (clay\%)}^{-1}$ ) are closer to the observed value than CENTURY ( $0.02 \text{ g C kg}^{-1} \text{ soil (clay\%)}^{-1}$ , Figure 4c). In CENTURY or MIMICS, the sensitivities of SOC concentration to these variables generally show large variations. Overall, SOC simulated by CENTURY is more sensitive to the changes in soil condition and litter input than MIMICS.

### 3.3 | Evaluation of simulated SOC composition

The simulated ratios of microbial biomass (MIC) to total SOC stock (MIC/SOC) from the MIMICS models are broadly consistent with the observations collected from global forest sites (Xu et al., 2013), both in terms of mean (or median) value and the range of variation (Figure 5). Overall, both observed and simulated MIC/SOC ranged from 0.005 to approximately 0.05, with a mean value of approximately 0.017 (0.015–0.019) and a median value of 0.013 (0.012–0.014).

MIMICS-simulated fractions of SOC pools are consistent with measurements of the Australian soil samples based on the particle size and chemical compositions of organic matter (Table S4), but CENTURY did not (Figure 6). Observations at 505 Australian sites indicate that HOC



**FIGURE 4** Sensitivity of simulated soil organic carbon (SOC) concentration to mean annual temperature ( $S_{MAT}$ , a), soil water content ( $S_{SWC}$ , b), soil clay fraction ( $S_{clay}$ , c), annual litterfall input ( $S_{litterfall}$ , d), the C:N ratio of litterfall ( $S_{C:N}$ , e) and the lignin:C ratio of litterfall ( $S_{lignin:C}$ , f). The blue and red dashed lines denote insignificant and significant ( $p < .05$ ) sensitivity calculated based on observation data, respectively. The solid line in each box denotes the median value. Box boundaries show the 25th and 75th percentiles, whiskers denote the 10th and 90th percentiles, and the black dots denote the 5th and 95th percentiles [Colour figure can be viewed at [wileyonlinelibrary.com](http://wileyonlinelibrary.com)]

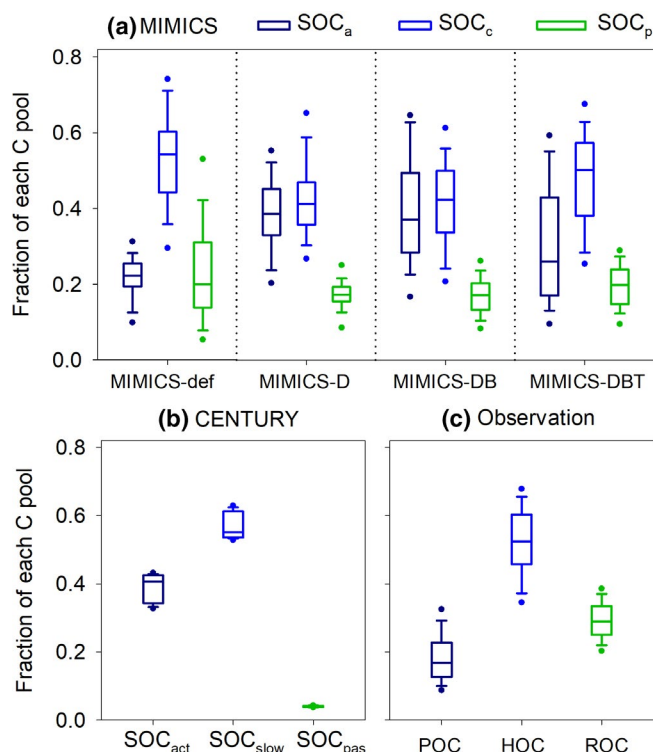
**FIGURE 5** Comparison between the simulated ratio of microbial C (MIC) to total soil organic carbon (SOC) from different versions of MIMICS and the observed values at globally distributed forest sites. The dashed and solid lines in each box are the mean and median value, respectively. Box boundaries show the 25th and 75th percentiles, whiskers denote the 10th and 90th percentiles, the dots below and above each box denote the 5th and 95th percentiles, respectively. The 655 samples of observed MIC/SOC at globally distributed forest sites are collected by Xu et al. (2013)

(46%–60%) accounts for the largest proportion of SOC, followed by the most stable pool ROC (25%–33%). The labile pool POC makes up a small fraction (12%–23%) of total SOC (Figure 6a). MIMICS predicts a similar

composition of SOC pools. The moderately stable pool ( $SOC_m$ ) accounts for the largest proportion of total SOC, followed by the most stable pool protected by the mineral matrix ( $SOC_p$ ) and the available pool ( $SOC_a$ , Figure 6c). SOC composition simulated by CENTURY can be very different depending on the optimized turnover rates of the active, slow and passive SOC pools (Figure 6b and Figure S5b). Increasing turnover rate of a specific SOC pool generally results in a smaller proportion of this pool compared to the total SOC (if the turnover rates of other SOC pools are assumed to be fixed).

### 3.4 | Key drivers of the variation in SOC composition

The key factors controlling the simulated SOC composition in CENTURY and MIMICS are different from the observations (Figure 7). Based on observation data, soil moisture, clay fraction, BS and litter input show significant empirical correlations with SOC composition, whereas soil temperature shows no significant correlation. In both CENTURY and MIMICS, soil temperature strongly affects SOC composition. Higher temperature however decreases the 'stable' SOC fraction ( $SOC_{pas}$ ) in CENTURY, but increases the stable fraction ( $SOC_p$ ) in MIMICS. MIMICS can represent the impacts of litter input on SOC composition, but CENTURY does not. Similar to the observations, higher litter input rate increases the proportion of the stable SOC pools (ROC and  $SOC_p$ ) but decreases the proportion of moderately

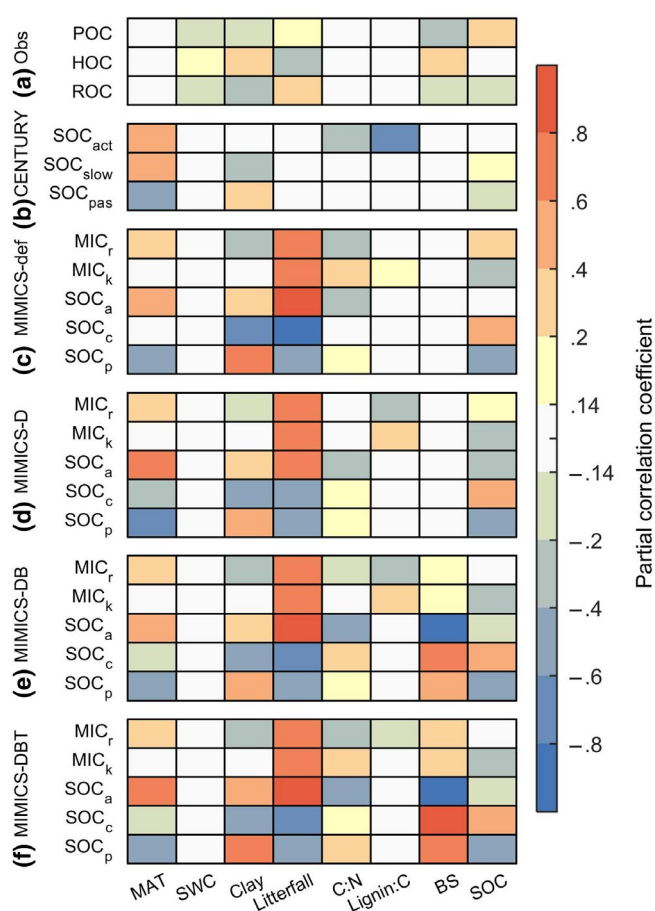


**FIGURE 6** Comparison between the simulated soil organic carbon (SOC) compositions from optimized MIMICS (a) and CENTURY (b) model and the observed SOC compositions at 505 sites in Australia (c). The observation data in Australia are obtained from Viscarra Rossel et al. (2019). Viscarra-Rossel et al. partitioned total SOC into three fractions with different particle sizes: the particulate organic carbon (POC), the humic organic carbon (HOC) and the resistant organic carbon (ROC, which is the mineral-associated organic carbon). The line in each box denotes median value. Box boundaries show the 25th and 75th percentiles, whiskers denote the 10th and 90th percentiles, and the dots below and above each box denote the 5th and 95th percentiles [Colour figure can be viewed at [wileyonlinelibrary.com](https://onlinelibrary.wiley.com/doi/10.1111/gcb.14994)]

stable pools (HOC and SOC<sub>c</sub>). The simulated decreasing trend of labile SOC (SOC<sub>a</sub>) with increasing litter input is contrary to the observation (POC). In MIMICS-DB and MIMICS-DBT, soil chemical properties represented by BS also show strong impact on SOC composition. Moreover, SOC composition also changes with the pool size of total SOC. It is necessary to note that the partial correlation coefficients might not be able to fully represent the relationships between SOC composition and soil and litter variables (Figure 7), as SOC composition might not be linearly related to these variables (Figure S10).

## 4 | DISCUSSION

Using in situ observations of SOC, litterfall and soil properties from 206 forest sites in Europe and China, we compared the performance of a first-order soil biogeochemical model (CENTURY) and four different versions of the microbial trait-based model (MIMICS) for simulating the large-scale spatial variation of SOC concentrations, the sensitivity



**FIGURE 7** Partial correlation coefficients between fraction of each SOC pool and model drivers, including mean annual temperature (MAT, °C), soil water content (SWC, dimensionless), soil clay content (clay, dimensionless), annual total litterfall production (Litterfall, g C m<sup>-2</sup> year<sup>-1</sup>), litter C:N ratio (C:N), litter lignin:C ratio (Lignin:C), base saturation (BS, 0–1, dimensionless) and total SOC concentration (SOC), Figure (a) Obs show the results based on observation data from Australia. Figure (b)–(f) showed the results based on optimized CENTURY and MIMICS models. Partial correlation coefficients between –0.14 and 0.14 were not significant ( $p > .05$ ) [Colour figure can be viewed at [wileyonlinelibrary.com](https://onlinelibrary.wiley.com/doi/10.1111/gcb.14994)]

of SOC concentration to key model drivers and the SOC composition. Our evaluation provides strong evidence that soil biogeochemical models with explicit microbial processes can be applied to simulate the large-scale SOC dynamics across different soil, vegetation and climate conditions. Below, we discuss in detail the implications of these results, uncertainties associated with the analysis, and an outlook for future data and model needs.

## 4.1 | Implications of simulation results

### 4.1.1 | Decomposition model should be calibrated and evaluated comprehensively

This study reveals the necessity to calibrate and evaluate MIMICS comprehensively. Preliminary parameter estimates for this study



showed that although parameters optimized based solely on observed SOC concentrations can accurately estimate total SOC stocks; they may not be able to estimate SOC composition and turnover time. To avoid unreasonable estimates of SOC composition (e.g. SOC<sub>p</sub> of MIMICS calibrated only against the SOC concentrations at European and China forest sites always approaches to zero) and C turnover times, we imposed additional constraints to restrict the ranges of proportions and turnover times of MIMICS SOC pools (see Section 2.3). Our results highlight the need for comparing model results with total SOC and microbial biomass, SOC composition and turnover time, as well as the response of SOC to changed climate, litter input and soil properties with a wide range of observations. Moreover, the optimized parameter values of both CENTURY and MIMICS in this study (Table S3) are different from the default values calibrated against manipulated decomposition experiments (Parton et al., 1987; Wieder et al., 2015), suggesting that **model parameters obtained based on local decomposition experiments might not work well at large spatial scales.**

#### 4.1.2 | Importance of explicitly representing microbial dynamics in decomposition model

Explicit representation of microbial biomass and substrate-limited growth rates is important for soil biogeochemical models to accurately capture the observed SOC concentration variations and the responses of SOC to climate changes (Campbell & Paustian, 2015; Wieder, Grandy, et al., 2014). In our research, simulations of SOC concentration at forest sites using MIMICS were more accurate and parsimonious compared to using CENTURY (Figure 2), and MIMICS better capture the observed sensitivities of SOC concentrations to temperature and soil clay than CENTURY. Conventional first-order models do not explicitly simulate microbial activity, but instead strongly emphasizes the relationship between litter chemical recalcitrance and soil C stock (Jenkinson & Rayner, 1977; Parton et al., 1987; Wieder, Grandy, et al., 2014). Recent analytical and experimental advances have demonstrated that molecular structure alone does not control SOC stability. Rather, microbial products of decomposition are the main precursors of stable SOC (Cotrufo et al., 2013; Kallenbach, Frey, & Grandy, 2016), suggesting that, in fact, environmental and biological controls predominate (Lehmann & Kleber, 2015; Lützow et al., 2006; Schmidt et al., 2011).

#### 4.1.3 | Impacts of soil physiochemical properties on SOC decomposition and stabilization

Besides microbial dynamics, it is also necessary to accurately represent the effects of soil physiochemical properties on SOC dynamics in soil biogeochemical models, especially for the formation and release of SOC protected by the mineral matrix. It has been widely recognized that soil clay fractions can influence SOC

stock and stabilization by promoting the sorption of organic C to mineral surfaces and entrapment into micropores (Schimel et al., 1994; Wagner, Cattle, & Scholten, 2007). CENTURY uses the soil clay fraction to modify the decomposition rate of the active SOC pool and the C transfer from active to slow pool (Parton et al., 1987). As the active pool generally accounts for only a small fraction (c. 3.5%) of total SOC (Figure 6b), this might explain why the sensitivity of SOC concentration to soil clay content in CENTURY is drastically underestimated compared to the observation-based sensitivity (Figure 4c). In MIMICS, soil clay influences both the decomposition rate of available SOC pool and the deprotection rate of protected by the mineral matrix. MIMICS thus better represents current understanding of SOC stabilization processes and appears to more accurately estimate the sensitivity of SOC to soil clay fraction than CENTURY (Figure 4c).

Numerous experimental studies also reported the significant impacts of soil chemical properties such as pH, exchangeable cations (e.g. Ca<sup>2+</sup>) and extractable metals (e.g. iron- and aluminium-oxyhydroxides) on SOC dynamics (Doetterl et al., 2015; Rasmussen et al., 2018; Six et al., 2004; Viscarra Rossel et al., 2019), and the relative importance of these factors likely varies across scales and ecosystems (Jobbágy & Jackson, 2000; Schmidt et al., 2011; Viscarra Rossel et al., 2019). Indeed, representing the diversity of mechanisms by which the soil physicochemical environment influences the persistence of soil organic matter in numerically tractable ways remains an outstanding challenge in models (Bailey et al., 2018). Our work suggests one opportunity to use BS as a proxy variable that can modify C deprotection rates from the SOC<sub>p</sub> pool in MIMICS (MIMICS-DB). This modification significantly decreased the biases in simulated SOC concentrations (Figure 2) and eliminated the systematic estimation biases along gradients of soil pH, clay content and annual precipitation at the observation sites (Figure 3). Moreover, our analysis on the relative contributions of model choice, fixed effects and site-specific random effects to explaining the SOC variation (Figure S8) reveals that the constraints of soil physical (e.g. temperature and clay content) and chemical (e.g. BS) properties on SOC dynamics has been better represented in MIMICS than in CENTURY, as the fixed effects including all potentially important soil variables can only explain a small part of the simulation errors of MIMICS, but a considerable part (~30%) of the simulation errors of CENTURY (Figures S7 and S8).

#### 4.1.4 | Impacts of litter inputs on SOC decomposition and stabilization

First-order models like CENTURY assume a linear relationship with productivity and soil C stocks (Todd-Brown et al., 2013), and the same is true for default parameterizations of MIMICS. Our analysis shows that the simulated SOC concentrations from CENTURY and MIMICS models are systematically biased from observations along the gradients of local litterfall production, except for the MIMICS-DBT which considers the density-dependent turnover of microbes



(Figure 3). This suggests that at the community level, regulatory mechanisms like competition, space constraints and other controls that depend on the density of individuals (such as disease and production of toxins) may limit microbial population sizes (Hibbing, Fuqua, Parsek, & Peterson, 2010; Kaiser, Franklin, Dieckmann, & Richter, 2014; Kaiser, Franklin, Richter, & Dieckmann, 2015). Indeed, a recent study from Georgiou et al. (2017) indicated that the density-dependent microbial processes can play an essential, but often overlooked role in regulating SOC dynamics. We recognize that the parameterization of density-dependent turnover implemented in MIMICS-DBT simplifies the complex community interactions that occur in soils, but they represent a tractable means for capturing the emergent dynamics in models that are intended for global-scale application and projections.

Litter input is not as important as soil physicochemical properties for predicting total SOC stock (Figure 4d), but it nevertheless strongly affects SOC composition (Figure 7), which determines the vulnerability of SOC (i.e. risk of C loss) to perturbations such as climate change and human disturbances. Litter quality can impact microbial C use efficiency and short-term SOC dynamics (Manzoni et al., 2017; Zhang et al., 2018), but evidence is inclusive on the significant role of litter quality in long-term SOC dynamics (Gentile, Vanlauwe, & Six, 2011; Helfrich, Ludwig, Potthoff, & Flessa, 2008). The effect of litter quality on SOC stabilization is mostly modulated by the extent of soil C saturation, and it may alter SOC stocks only when there is a saturation deficit (Castellano, Mueller, Olk, Sawyer, & Six, 2015). Consistent with our results (Figure 7), previous studies also reported that litter quantity rather than quality is one of the main determinants of SOC stability (Carrington, Hernes, Dyda, Plante, & Six, 2012; Dungait, Hopkins, Gregory, & Whitmore, 2012). Experiments by Wang, He, and Liu (2016) suggested that the ratio between different SOC fractions is related to microbial biomass and community composition (which depends on the amounts of litter inputs), but not to litter chemical composition.

## 4.2 | Uncertainties in this study

Some uncertainties in our simulation results may be caused by biases of forcing and validation data. In this study, we assumed the forest and soil C at all observation sites are at equilibrium. However, even though most observation sites have a stand age older than 40 years and have not been strongly disturbed by fire or human activities (e.g. reforestation and deforestation can induce a 30% change in soil C stock, Don, Schumacher, & Freibauer, 2011), the forest systems at some sites may not be at equilibrium, especially under the background of global climate change. Some uncertainties also arise due to lack of observations. Specifically, the wood and root litterfall at European sites have not been measured and Chinese observation data only provide measurements of plant biomass but not litterfall, so we have adopted the leaf turnover rates and ratios of wood litter and root litter to leaf litter from databases of plant traits and litterfall production to

calculate the total litterfall production at each observation site (see Section 2.1). Moreover, most of the litter C:N ratios and the lignin:C ratios were obtained from previously compiled litterfall databases and publications and not from site-level observations. Thus, biases and uncertainties that exist in the litter input data are poorly quantified.

Additional uncertainties are related to model structural assumptions and parameterizations. Specifically, soil moisture has been widely regarded as one of the primary physical factors that control microbial activity (Arnold, Ghezzehei, & Berhe, 2015; Ghezzehei, Sulman, Arnold, Bogie, & Berhe, 2019; Manzoni, Moyano, Kätterer, & Schimel, 2016); however, the soil moisture control over microbial dynamics is not used in the current parameterization of MIMICS. Soil structure (characterized by porosity or bulk density) determines soil O<sub>2</sub> availability and the accessibility of C particles to microbes (Davidson, Samanta, Caramori, & Savage, 2012; Lützow et al., 2006). Soil nutrient availability (e.g. mineral nitrogen and phosphorus) strongly affects microbial C use efficiency and growth rate (Manzoni et al., 2017). Again, soil moisture, structure and nutrient availability have not been considered in this implement MIMICS. Finally, neither of the models considered here implement vertically resolved soil biogeochemistry, which are clearly important to capture soils with strong vertical profiles or vertical perturbations such as in permafrost C (Koven, Lawrence, & Riley, 2015; McGuire et al., 2018). The insufficient representation of interactions between soil physicochemical properties, nutrient availability, microbial dynamics and SOC stabilization therefore may induce additional uncertainties in our results. We appreciate that these additional complexities in model form also generates greater data demands to appropriately parameterize and evaluate models, but may be necessary to build confidence in soil carbon projections (Bradford et al., 2016).

## 4.3 | Outlooks and challenges

A study by Wieder, Grandy, et al. (2014) demonstrated that MIMICS could capture the observed temporal decreasing trends of litter and SOC stocks in field decomposition experiments. Our evaluation further demonstrates that MIMICS can simulate SOC stock and composition across ecosystems with different climate, and soil and forest types. MIMICS also represents the SOC decomposition and stabilization processes more realistically (e.g. explicitly represents microbial dynamics) than conventional first-order models. Therefore, MIMICS can be used to replace the conventional decomposition models used in existing ESMs.

The parameters, structure and algorithms of MIMICS can still be improved. We encourage future studies to assess the global applicability of MIMICS or similar models based on more integrated in situ observations on plant biomass, litterfall (both aboveground and belowground), SOC stock and composition, soil physicochemical properties and local climate from more ecosystems, in particular observations from grasslands and tropical forests. We also

encourage more studies to quantify the interactions between soil physicochemical properties, microbial dynamics and the stabilization of SOC. In this study, the MIMICS model considering the physicochemical constraints of soil properties on SOC deprotection rate and microbe turnover more accurately estimated SOC concentration than the default model (Figure 2). But the empirical functions (Equations 13 and 14) used to represent physicochemical constraints were built empirically based on analysis of the biases of simulated SOC concentration from the default version of MIMICS (Figure 3). More experiments investigating influences of soil physicochemical properties on microbial activity and the C adsorption/desorption rate of mineral soil are needed to improve these empirical functions. Furthermore, many soil properties are significantly correlated (e.g. Figure S12) and the changes in litter inputs and SOC contents can in return dramatically alter soil physical, chemical and biological properties (Murphy, 2015; Schmidt et al., 2011). Thus, research focusing on the interactions between litter, SOC and different soil properties is also essential.

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## DATA AVAILABILITY STATEMENT


The European ICP forest data can always be requested from the Programme Co-ordinating Centre (<http://icp-forests.net/page/data-requests>) of ICP Forests in Eberswalde, Germany. The Chinese forest data can be obtained by contacting the Prof. Tang X (xltang@scib.ac.cn) in South China Botanical Garden, Chinese Academy of Sciences, Guangzhou, China. All of the other databases of soil, climate, litterfall and vegetation are publicly accessible, and the specific references and links to these databases are provided in Section 2.1.

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## SUPPORTING INFORMATION

Additional supporting information may be found online in the Supporting Information section.

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# Microbial dynamics and soil physicochemical properties explain large-scale variations in soil organic carbon

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