

Whole-genome sequences of Malawi cichlids reveal multiple radiations interconnected by gene flow

Milan Malinsky^{1,2,10*}, Hannes Svardal^{1,3,4,5,10}, Alexandra M. Tyers^{6,9}, Eric A. Miska^{1,3,7}, Martin J. Genner⁸, George F. Turner⁶ and Richard Durbin^{1,3*}

The hundreds of cichlid fish species in Lake Malawi constitute the most extensive recent vertebrate adaptive radiation. Here we characterize its genomic diversity by sequencing 134 individuals covering 73 species across all major lineages. The average sequence divergence between species pairs is only 0.1–0.25%. These divergence values overlap diversity within species, with 82% of heterozygosity shared between species. Phylogenetic analyses suggest that diversification initially proceeded by serial branching from a generalist *Astatotilapia*-like ancestor. However, no single species tree adequately represents all species relationships, with evidence for substantial gene flow at multiple times. Common signatures of selection on visual and oxygen transport genes shared by distantly related deep-water species point to both adaptive introgression and independent selection. These findings enhance our understanding of genomic processes underlying rapid species diversification, and provide a platform for future genetic analysis of the Malawi radiation.

The formation of every lake or island represents a fresh opportunity for colonization, proliferation and diversification of living forms. In some cases, the ecological opportunities presented by underutilized habitats facilitate adaptive radiation—rapid and extensive diversification of the descendants of the colonizing lineages^{1–3}. Adaptive radiations are thus exquisite examples of the power of natural selection, as seen for example in Darwin's finches in the Galapagos^{4,5}, the Anolis lizards of the Caribbean⁶ and in East African cichlid fishes^{7,8}.

Cichlids are one of the most species-rich and diverse families of vertebrates, and nowhere are their radiations more spectacular than in the Great Lakes of East Africa: lakes Malawi, Tanganyika and Victoria², each of which contains several hundred endemic species, with the largest number in Lake Malawi⁹. Molecular genetic studies have made major contributions to reconstructing the evolutionary histories of these adaptive radiations, especially in terms of the relationships between the lakes^{10,11}, between some major lineages in Lake Tanganyika¹², and in describing the role of hybridization in the origins of the Lake Victoria radiation¹³. However, the task of reconstructing within-lake relationships remains challenging owing both to the retention of large amounts of ancestral genetic polymorphism (that is, incomplete lineage sorting) and the gene flow between taxa^{12,14–18}.

Initial genome assemblies of cichlids from East Africa suggest that an increased rate of gene duplication, together with accelerated evolution of some regulatory elements and protein coding genes, may have contributed to the radiations¹¹. However, our understanding of the genomic mechanisms contributing to adaptive radiations is still in its infancy³.

Here we provide an overview of and insights into the genomic signatures of the haplochromine cichlid radiation of Lake Malawi.

The species that comprise the radiation can be divided into seven groups with differing ecology and morphology (see Supplementary Note): (1) the rock-dwelling 'mbuna'; (2) *Rhamphochromis*—typically midwater pelagic piscivores; (3) *Diplotaxodon*—typically deep-water pelagic zooplanktivores and piscivores; (4) deep-water and twilight-feeding benthic species; (5) 'utaka' feeding on zooplankton in the water column but breeding on or near the lake bottom (here utaka corresponds to the genus *Copadichromis*); (6) a diverse group of benthic species, mainly found in shallow non-rocky habitats; and (7) *Astatotilapia calliptera*, a closely related generalist that inhabits shallow weedy margins of Lake Malawi, and other lakes and rivers in the catchment, as well as river systems to the east and south of the Lake Malawi catchment. This division into seven groups has been partially supported by previous molecular phylogenies based on mitochondrial DNA (mtDNA) and amplified fragment length polymorphism data^{18–20}. However, published phylogenies show numerous inconsistencies and, in particular, the question of whether the groups are genetically separate remained unanswered.

To characterize the genetic diversity, species relationships, and signatures of selection across the whole radiation, we obtained Illumina whole-genome sequence data from 134 individuals of 73 species distributed broadly across the seven groups (Fig. 1a; Supplementary Note). This includes 102 individuals at ~15× coverage and 32 additional individuals at ~6× coverage (Supplementary Table 1).

Results

Low genetic diversity and species divergence. Sequence data were aligned to and variants called against a *Metriaclima zebra* reference genome¹¹. Average divergence from the reference was 0.19% to 0.27% (Supplementary Fig. 1). After filtering and variant refine-

¹Wellcome Sanger Institute, Cambridge, UK. ²Zoological Institute, University of Basel, Basel, Switzerland. ³Department of Genetics, University of Cambridge, Cambridge, UK. ⁴Department of Biology, University of Antwerp, Antwerp, Belgium. ⁵Naturalis Biodiversity Center, Leiden, The Netherlands. ⁶School of Natural Sciences, Bangor University, Bangor, UK. ⁷Gurdon Institute, University of Cambridge, Cambridge, UK. ⁸School of Biological Sciences, University of Bristol, Bristol, UK. ⁹Present address: Max Planck Institute for Biology of Ageing, Cologne, Germany. ¹⁰These authors contributed equally: Milan Malinsky, Hannes Svardal. *e-mail: millanek@gmail.com; rd109@cam.ac.uk

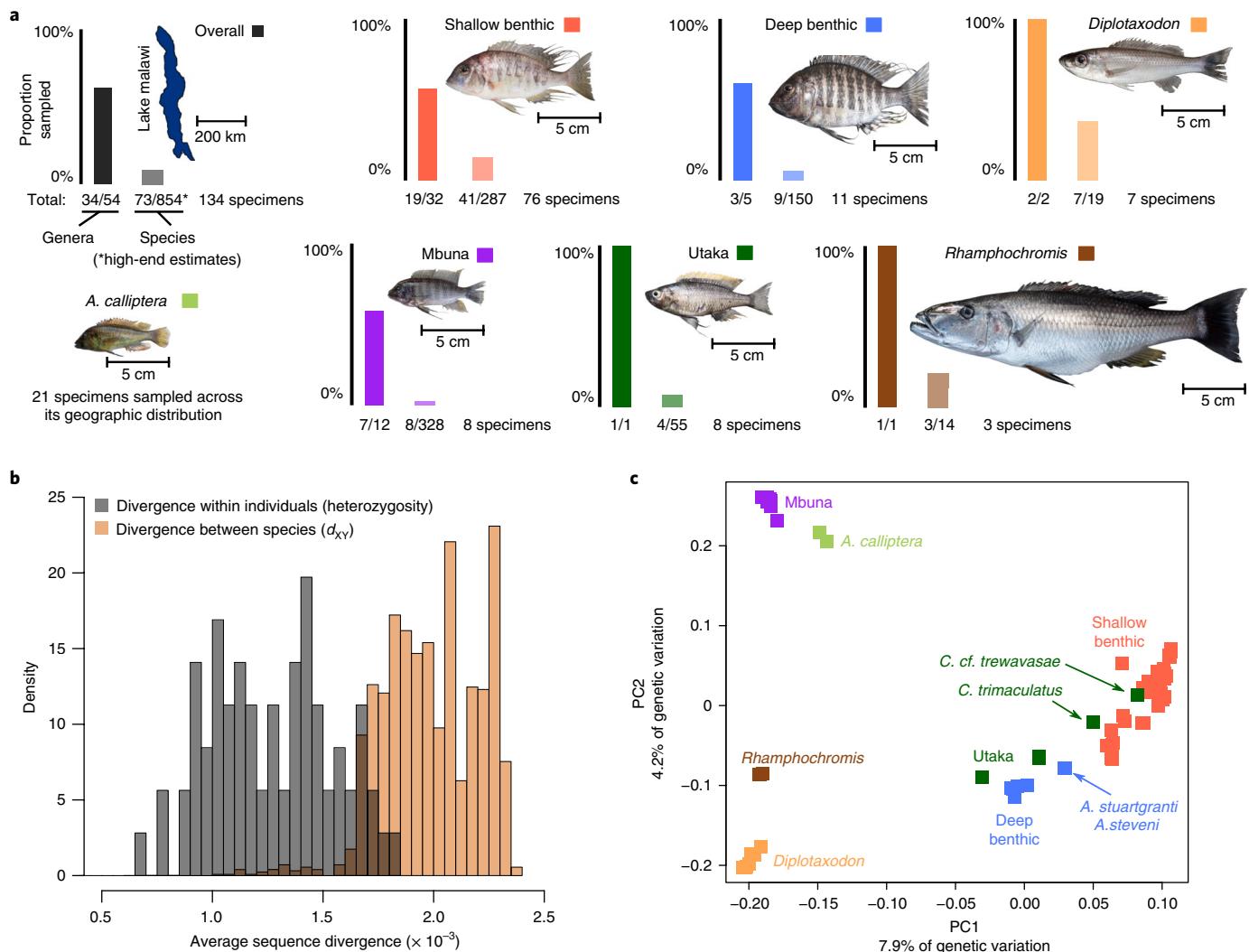


Fig. 1 | The Lake Malawi haplochromine cichlid radiation. **a**, The sampling coverage of this study: overall and for each of the seven main eco-morphological groups within the radiation. A representative specimen is shown for each group (*Diplotaxodon*: *D. limnothrissa*; shallow benthic: *Lethrinops albus*; deep benthic: *Lethrinops gossei*; mbuna: *Metriaclima zebra*; utaka: *Copadichromis virginalis*; *Rhamphochromis*: *R. woodi*). Numbers of species and genera are based on ref. ²⁹. **b**, The distributions of genomic sequence diversity within individuals (heterozygosity; π) and of divergence between species (d_{xy}). **c**, Principal component analysis (PCA) of whole-genome variation data.

ment, we obtained 30.6 million variants, of which 27.1 million were single nucleotide polymorphisms (SNPs) and the rest were short insertions and deletions. All the following analyses are based on biallelic SNPs.

To estimate nucleotide diversity (π) within the species, we measured the frequency of heterozygous sites in each individual. The estimates are distributed within a relatively narrow range between 0.7 and 1.8×10^{-3} per base pair (bp) (Fig. 1b). The mean π estimate of 1.2×10^{-3} per bp is at the low end of values found in other animals²¹. There does not appear to be a relationship between π and the rate of speciation: individuals in the species-rich mbuna and shallow benthic groups show levels of π that are comparable to those of the relatively species-poor utaka, *Diplotaxodon* and *Rhamphochromis* (Supplementary Fig. 1).

Despite their extensive phenotypic differentiation, species within the Lake Malawi radiation are genetically closely related^{22,23}. However, genome-wide genetic divergence has never been quantified. We calculated the average pairwise sequence differences (d_{xy}) between species and compared d_{xy} against heterozygosity, finding that the two distributions partially overlap (Fig. 1b). Thus, the

sequence divergence within a single diploid individual is sometimes higher than the divergence between two distinct species. The average d_{xy} is 2.0×10^{-3} with a range between 1.0 and 2.4×10^{-3} per bp. The maximum d_{xy} is therefore approximately one-fifth of the divergence between human and chimpanzee²⁴. In addition to the low ratio of divergence to diversity, most genetic variation is shared between species. On average both alleles are observed in other species for 82% of heterozygous sites within individuals, consistent with the expected and previously observed high levels of incomplete lineage sorting (ILS)²⁵. Supplementary Fig. 2 shows values of d_{xy} and of the fixation index (F_{ST}) for comparisons between the seven eco-morphological groups and Supplementary Fig. 3 shows patterns of linkage disequilibrium across the radiation, within groups and within individual species.

Low per-generation mutation rate. It has been suggested that the species richness and morphological diversity of teleosts in general and of cichlids in particular might be explained by elevated mutation rates compared to other vertebrates^{25,26}. To obtain a direct estimate of the per-generation mutation rate, we reared offspring of

three species from three different Lake Malawi groups (*A. calliptera*, *Aulonocara stuartgranti* and *Lethrinops lethrinus*). We sequenced both parents and one offspring of each to high coverage (40×), applied stringent quality filtering, and counted variants present in each offspring but absent in both its parents (Supplementary Fig. 4). There was no evidence for significant difference in mutation rates between species. The overall mutation rate (μ) was estimated at 3.5×10^{-9} (95% confidence interval (CI): 1.6×10^{-9} to 4.6×10^{-9}) per bp per generation, approximately three to four times lower than in humans²⁷, although, given much shorter mean generation times, the per-year rate is still expected to be higher in cichlids than in humans. We note that ref. ²⁶ obtained a much higher mutation rate estimate (6.6×10^{-8} per bp per generation) in Midas cichlids, but from relatively low-depth sequencing of restriction-site-associated markers that may have made accurate verification more difficult. We also note that our per-generation rate estimate, although low, is still higher than the lowest μ estimate in vertebrates: 2×10^{-9} per bp per generation recently reported for Atlantic herring²⁸. By combining our mutation rate with nucleotide diversity (π) values, we estimate the long term effective population sizes (N_e) to be in the range of approximately 50,000 to 130,000 breeding individuals (with $N_e = \pi/4\mu$).

Genome data support for eco-morphological groupings. PCA of the whole-genome genotype data generally separates the major eco-morphological groups (Fig. 1c). The most notable exceptions to this are (1) the utaka, for which some species cluster more closely with deep benthics and others with shallow benthics, and (2) two species of the genus *Aulonocara*, *A. stuartgranti* and *A. steveni*, which are located between the shallow and deep benthic groups. Although these have enlarged lateral-line sensory apparatus like many deep benthic species including other *Aulonocara*, they are typically found in shallower water²⁹. Another interesting pattern in the PCA plot is that the utaka and benthic samples are often spread along principal component (PC) axes (Fig. 1c, Supplementary Fig. 5), a pattern typical for admixed populations (for example ref. ³⁰). Along the two main PCs, the deeper-water benthic species extend towards the deep-water *Diplotaxodon*, an observation we will return to in the context of gene flow and shared mechanisms of depth adaptation.

To further verify the consistency of group assignments, we tested whether pairs of species from the same group always share more derived alleles with each other than with any species from other groups. Group assignments were again supported, except for the four species also highlighted in the PCA: the two shallow-living *Aulonocara* are closer to shallow benthics than to deep benthics in 71% and 82% of tests respectively when comparing these alternatives, and *Copadichromis trimaculatus* is closer to shallow benthics than to utaka in 58% of the comparisons. *Copadichromis cf. trewavasae* always clustered with shallow benthics; therefore, we treat it as a member of the shallow benthic group henceforth. With the three intermediate samples removed and *C. cf. trewavasae* reassigned, all other species showed 100% consistency with their group assignment.

Allele sharing inconsistent with tree-like relationships. The above observations suggest that some species may be genetically intermediate between well defined groups, consistent with previous studies that have suggested that hybridization and introgression subsequent to initial separation of species may have played a significant part in cichlid radiations, including in lakes Tanganyika^{12,14–16} and Malawi^{18,20}. Where this happens, there is no single tree relating the species.

To assess the overall extent of violation of tree-like species relationships, we calculated Patterson's D statistic (the ABBA-BABA test)^{31,32} for all possible trios of Lake Malawi species, without assuming any a priori knowledge of their relationships. *N. brichardi* from Lake Tanganyika was always used as the outgroup. The test statistic

D_{\min} is the minimum absolute value of Patterson's D for each trio, across all possible tree topologies. Therefore, a significantly positive D_{\min} score signifies that the sharing of derived alleles between the three species is inconsistent with a single species tree relating them, even in the presence of incomplete lineage sorting.

Overall, 62% of trios (75,616 out of 121,485) have a significantly positive D_{\min} score (Holm–Bonferroni FWER < 0.01). The D_{\min} values are not independent: for example, a single gene-flow event between ancestral lineages can affect multiple contemporary species and thus more trios than would a more recent gene-flow event. However, tree violations are numerous and pervasive throughout the dataset, within all the major groups and also between groups (Fig. 2a), revealing reticulate evolution at multiple levels. Therefore, phylogenetic trees alone cannot fully describe the evolutionary relationships of Lake Malawi cichlids.

Phylogenetic framework. Despite no tree giving a complete and accurate picture of the relationships between species, standard phylogenetic approaches are useful to provide a framework for discussion. To obtain an initial picture we divided the genome into 2,543 non-overlapping windows, each comprising 8,000 SNPs (average size 274 kb) and constructed a maximum likelihood phylogeny separately for the full sequences within each window, obtaining trees with 2,542 different topologies. We also calculated the maximum clade credibility (MCC) summary tree³³ and a maximum likelihood phylogeny based on the full mtDNA genome (Fig. 2b and Supplementary Fig. 6).

We next applied a range of further phylogenomic methods which are known to be robust to incomplete lineage sorting. These included three multispecies coalescent methods^{34,35}: the Bayesian SNAPP³⁶ (with a subset of 48,922 unlinked SNPs in 12 individuals representing the eco-morphological groups), the algebraic method SVDquartets^{37,38}, which allows for site-specific rate variation and is robust to gene-flow between sister taxa³⁹, and the summary method ASTRAL^{40,41}, using the 2,543 local maximum likelihood trees that were described above as input. We also built a whole-genome neighbour-joining tree using the Dasarathy et al.⁴² algorithm, which has been shown to be a statistically consistent and accurate species tree estimator under ILS^{42,43}. The above methods have also been applied to datasets where the individuals that are genetically intermediate between eco-morphological groups (*C. trimaculatus*, *A. stuartgranti* and *A. steveni*) have been removed, thus probably reducing the extent of violation of the multispecies coalescent model.

Despite extensive variation among the 2,543 individual maximum likelihood trees (at least in part attributable to ILS), and, to a lesser extent, variation between the different genome-wide phylogenetic methods, there is some general consensus (Fig. 2c and Supplementary Figs. 6–10). Except for the three previously identified intermediate species, individuals from within each of the previously identified eco-morphological groups cluster together in all the whole-genome phylogenies, forming well supported reciprocally monophyletic groups. The pelagic *Diplotaxodon* and *Rhamphochromis* together form a sister group to the rest of the radiation, except in the all-sample MCC and SVDquartets phylogenies. Perhaps surprisingly, all the methods place the generalist *A. calliptera* as the sister taxon to the specialized rocky-shore mbuna group in a position that is nested within the Lake Malawi radiation. On a finer scale, many similarities between the resulting phylogenies reflect features of previous taxonomic assignment, but some currently recognized genera are always polyphyletic, including *Placidochromis*, *Lethrinops* and *Mylochromis*.

The mtDNA phylogeny is an outlier, substantially different from all the whole-genome phylogenies and also from the majority of the local maximum likelihood trees (Fig. 2b,c and Supplementary Figs. 6 and 11). Discordances between mtDNA and nuclear phylogenies in Lake Malawi have been reported previously and interpreted as

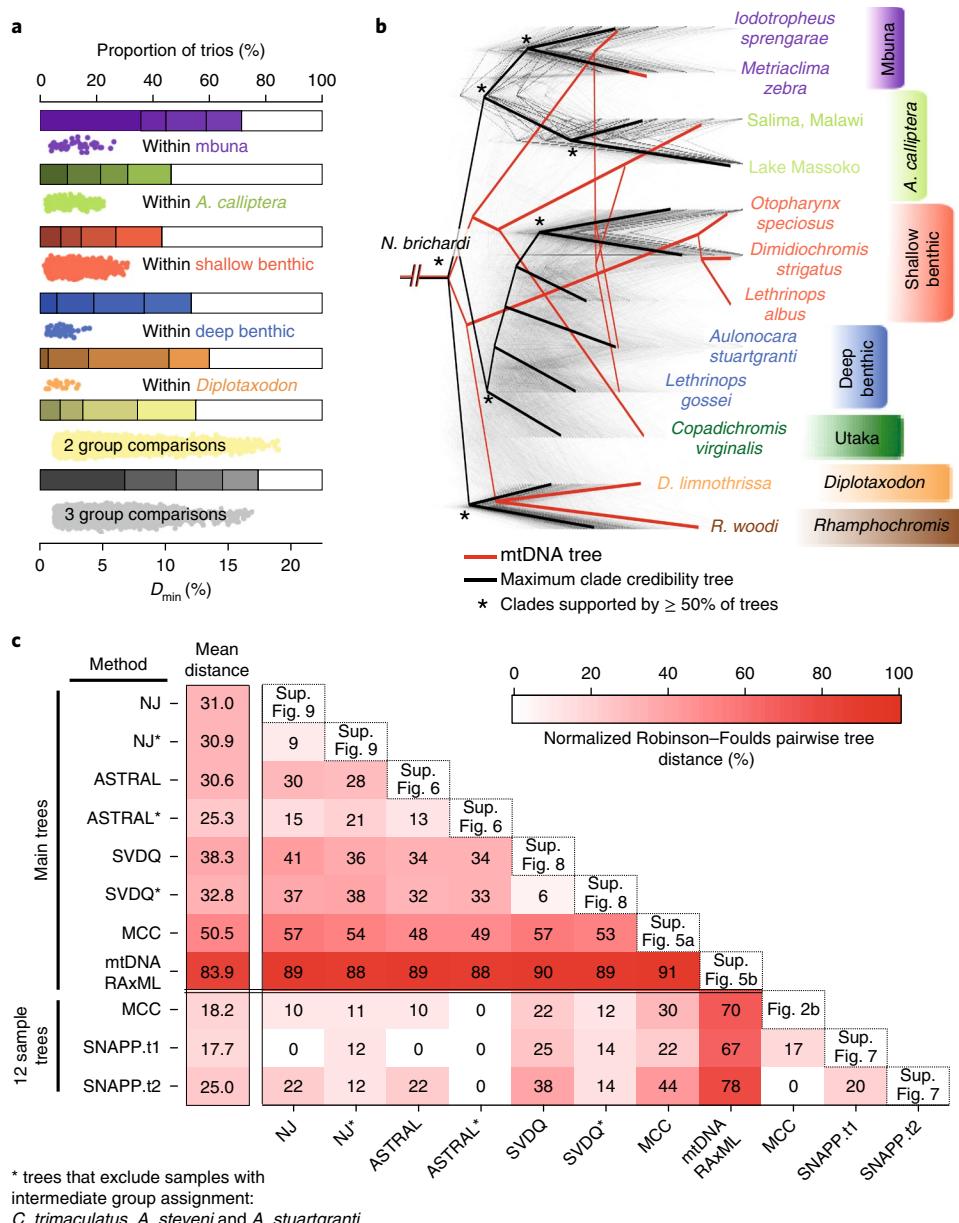


Fig. 2 | Excess allele sharing and patterns of species relatedness. **a**, Derived allele sharing reveals non-tree-like relationships among trios of species. The bars show the proportion of significantly elevated D_{\min} scores (see main text). Shading corresponds to FWER q values of (from light to dark) 10^{-2} , 10^{-4} , 10^{-8} and 10^{-14} . The scatterplots show the D_{\min} scores that were significant with family-wise error rate (FWER) < 0.01 . Results are shown separately for comparisons where all three species in the trio are from the same group, and for cases where the species come from two or three different groups. *Rhamphochromis* and *utaka* within-group comparisons are not shown owing to the low number of data points. **b**, A set of 2,543 maximum likelihood phylogenetic trees for non-overlapping regions along the genome. Branch lengths were scaled for visualization so that the total height of each tree is the same. The local trees were built with 71 species and then subsampled for display to 12 individuals representing the eco-morphological groups. The maximum clade credibility tree shown here was built from the subsampled local trees. A maximum likelihood mitochondrial phylogeny is shown for comparison. **c**, A summary of all phylogenies from this study and the normalized Robinson–Foulds distances between them, reflecting the topological distance between pairs of trees on the scale from zero to 100%. The least-controversial 12-sample tree is SNAPP.t1, with an average distance to other trees of 17.7%, while ASTRAL* is the least controversial among the ‘main trees’ (mean distance of 25.3%). To compare trees with differing sets of taxa, the trees were downsampled so that only matching taxa were present. The position of the outgroup/root was considered in all comparisons. Supplementary figures associated with the phylogenies are indicated for each tree.

a signature of past hybridization events^{18,20}. However, as we discuss below, some of these previously suggested hybridization events are not reflected in the whole-genome data. Indeed, large discrepancies between mitochondrial and nuclear phylogenies have been shown in many other systems, reflecting both that mtDNA as a single locus is not expected to reflect the consensus under ILS, and high incidence of mitochondrial selection^{44–46}. This underlines the

importance of evaluating species relationships in the Lake Malawi radiation from a genome-wide perspective.

Specific signals of introgression. We applied a variety of methods to identify the species and groups whose relationships violate the framework trees described in the previous section. First, we contrasted the pairwise genetic distances used to produce the

neighbour-joining tree against the distances between samples along the tree branches, calculating the residuals (Supplementary Fig. 12). If the tree captured all the genetic relationships in our sample perfectly, the residuals would all be zero. However, as expected in light of the D_{\min} analysis above, we found numerous differences, affecting both groups of species and individual species, with some standout cases. Among the strongest signals on individual species, in addition to the previously discussed *C. trimaculatus*, we can see that (1) *Placidochromis cf. longimanus* is genetically closer to the deep benthic clade and to a subset of the shallow benthic (mainly *Lethrinops* species) than the tree suggests; and (2) our sample of *Otopharynx tetrastigma* (from Lake Ilamba) is much closer to *A. calliptera* (especially to the sample from Lake Kingiri, only 3.2 km away) than is expected from the tree.

Second, the sharing of long haplotypes between otherwise distantly related species is an indication of recent admixture or introgression. To investigate this type of gene flow signature, we used the chromopainter software package⁴⁷ and calculated the ‘co-ancestry matrix’ of all species—a summary of nearest-neighbour (therefore recent) haplotype relationships. The Lake Ilamba *O. tetrastigma* and Lake Kingiri *A. calliptera* also stand out in this analysis, showing a strong signature of recent gene flow between individual species from distinct eco-morphological groups (Supplementary Fig. 13). The other tree-violation signatures described above are also visible on the haplotype sharing level but are less pronounced, consistent with being older events involving the common ancestors of multiple present-day species. However, the chromopainter results indicate additional recent introgression events (for example, the utaka *C. virginalis* with *Diplotaxodon*; more highlighted in Supplementary Fig. 13). Furthermore, the clustering based on recent co-ancestry is different from all phylogenetic trees: in particular a number of shallow benthics, including *P. cf. longimanus*, cluster next to the deep benthics.

Third, we used the f_4 admixture ratio^{31,32,48} (f statistic; closely related to Patterson’s D), computing $f(A,B;C,O)$ for all groups of species that fit the relationships $((A, B), C)$ in the ASTRAL* tree (Supplementary Fig. 7), with the outgroup fixed as *N. brichardi*. When elevated owing to introgression, the f statistic is expected to be linear in relation to the proportion of introgressed material. The ASTRAL* tree has the lowest mean topological distance to all the other trees, and excludes the three species with intermediate group assignment, a choice made here because we were interested in identifying additional signals beyond the admixed status of *A. stuart-granti*, *A. steveni* and *C. trimaculatus*. Out of the 164,320 computed f statistics, 97,889 were significant at $\text{FWER} < 0.001$.

As in the case of D_{\min} , a single gene-flow event can lead to multiple significant f statistics. Noting that the values for different combinations of $((A, B), C)$ groups are not independent as soon as they share branches on the tree, we sought to obtain branch-specific estimates of excess allele sharing that would be less correlated. Building on the logic employed to understand correlated gene flow signals in ref. ⁴⁹, we developed the ‘ f -branch’ metric or $f_b(C)$: a summary of f scores that, on a given tree, captures excess allele sharing between a species C and a branch b compared to the sister branch of b (Methods). Therefore, an $f_b(C)$ score is specific to the branch b (on the y -axis in Fig. 3), but a single introgression event can still lead to significant $f_b(C)$ values across multiple related C values. There were 11,158 $f_b(C)$ scores of which 1,421 were significantly elevated at $\text{FWER} < 0.001$ (Supplementary Fig. 14), and 238 scores were larger than 3% (the value inferred for human–Neanderthal introgression in ref. ³¹). The majority of nodes in the tree are affected: 92 of the 158 branches in the phylogeny show significant excess allele sharing with at least one other species C (Fig. 3).

Overall, the highest $f_b(C)$ (14.2%) is between the ancestor of the two sampled *Ctenopharynx* species from the shallow benthic group and the utaka *Copadichromis virginalis* (Fig. 3). Notably,

Ctenopharynx species, particularly *C. intermedius* and *C. pictus*, have very large numbers of long slender gill rakers, a feature shared with *Copadichromis* species, and believed to be related to a diet of small invertebrates⁵⁰. Several other benthic lineages also share excess alleles with *C. virginalis*, however these signals are less pronounced. Next, the significantly elevated $f_b(C)$ scores between the shallow and the deep benthic lineages suggest that genetic exchanges between these two groups go beyond the clearly admixed shallow-living *Aulonacara* (not included in this analysis). The f -branch signals between *O. tetrastigma* and *A. calliptera* Kingiri are observed in both directions—*A. calliptera* Kingiri with shallow benthics (and most strongly *O. tetrastigma*) and *O. tetrastigma* with *A. calliptera* (most strongly *A. calliptera* Kingiri), suggesting bi-directional introgression.

At the level of the major eco-morphological groups, the strongest signal indicates that the ancestral lineage of benthics and utaka shares excess derived alleles with *Diplotaxodon* and, to a lesser degree, *Rhamphochromis*, as previously suggested by the PCA plot (Fig. 1c). Furthermore, there is evidence for additional ancestry from the pelagic groups in utaka, which could be explained either by an additional, more recent, gene-flow event or by differential fixation of introgressed material, possibly due to selection. Reciprocally, *Diplotaxodon* shares excess derived alleles (relative to *Rhamphochromis*) with utaka and deep benthics, as does *Rhamphochromis* with mbuna and *A. calliptera*. Furthermore, mbuna show excess allele sharing (relative to *A. calliptera*) with *Diplotaxodon* and *Rhamphochromis* (Fig. 3). On the other hand, while ref. ¹⁸ suggested gene flow between the deep benthic and mbuna groups on the basis of a discrepancy between mtDNA and nuclear phylogenies, our genome-wide analysis did not find any signal of substantial genetic exchange between these groups.

The f statistic tests are robust to the occurrence of incomplete lineage sorting, in the sense that ILS alone cannot generate a significant test result³². We note, however, that pronounced population structure within ancestral species, coupled with rapid succession of speciation events, can also substantially violate the assumptions of a strictly bifurcating species tree and lead to significantly elevated f scores^{32,51}. This needs to be taken into account when interpreting non-tree-like relationships, for example among major groups early in the radiation. However, in cases of excess allele sharing between ‘distant’ lineages that are separated by multiple speciation events, ancestral population structure would have needed to segregate through these speciation events without affecting sister lineages, a scenario that is not credible in general. Therefore, we suggest that there is strong evidence for multiple cross-species gene flow events. Additionally, simulations suggest that, compared with treemix⁵², $f_b(C)$ is robust to misspecification of the initial tree (Supplementary Note).

Overall, the neighbour-joining tree residuals, the haplotype sharing patterns and the many elevated $f_b(C)$ scores paint a consistent picture. They confirm the extensive violations of the bifurcating species tree model initially revealed by the D_{\min} analysis, and suggest many independent gene-flow events at different times during the evolutionary history of the adaptive radiation.

Origins of the radiation. The generalist *Astatotilapia calliptera* has been referred to as the ‘prototype’ for the endemic Lake Malawi cichlids^{29,53}, and discussions concerning the origin of the radiation often centre on ascertaining its relationship to the Malawi species^{20,54}. Previous phylogenetic analyses, using mtDNA and small numbers of nuclear markers, showed inconsistencies in this respect^{18,20,54}. In contrast, our whole-genome data indicated a clear and consistent position of the Lake Malawi catchment *A. calliptera* as a sister group to the mbuna, in agreement with the nuclear DNA phylogeny in a previous study¹⁸. While it is not certain whether the 320 remaining mbuna species form a monophyletic group with the eight species we used here, the eight species represent the majority of the genera of

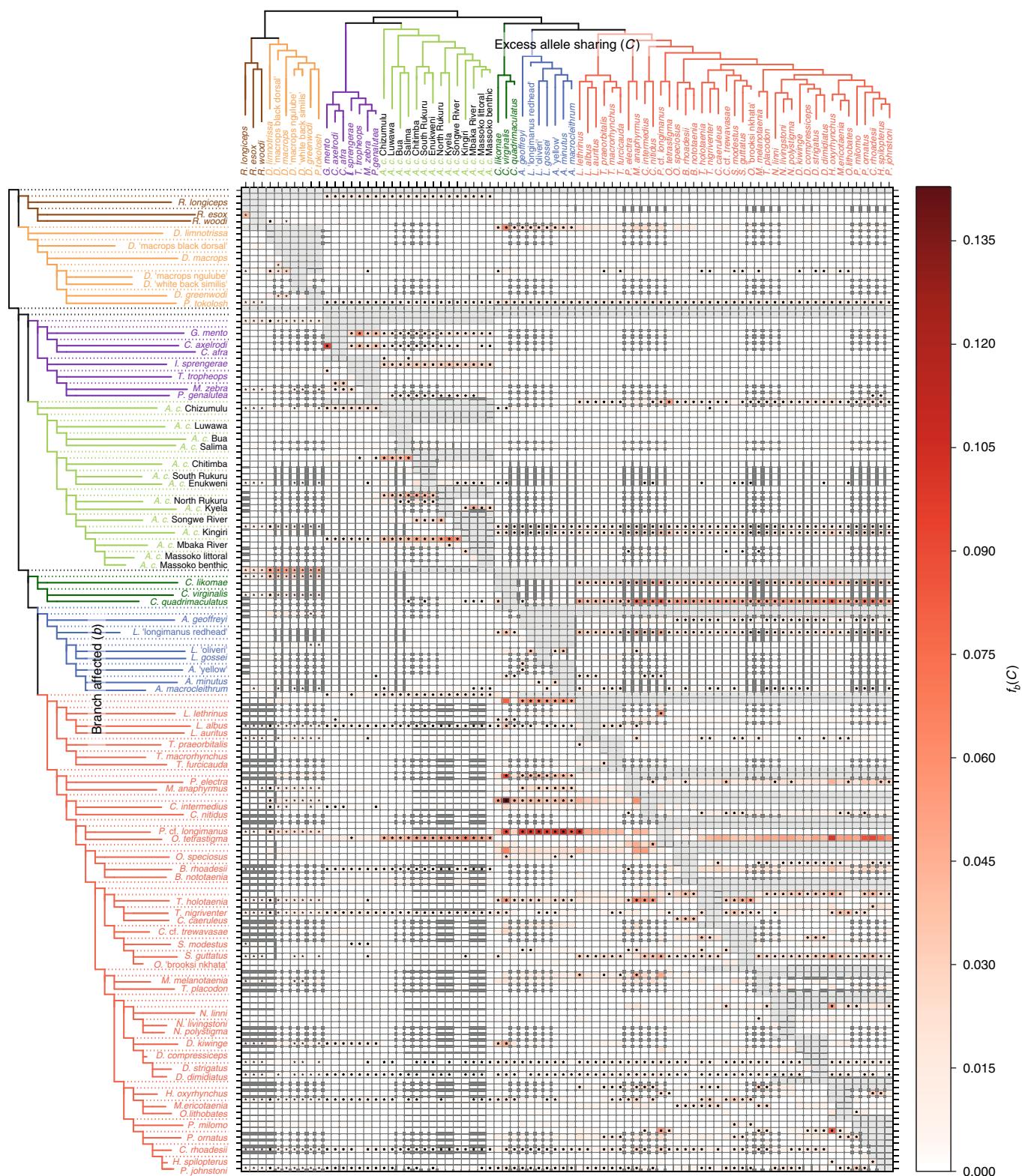


Fig. 3 | Identifying tree violating branches and possible gene-flow events. The branch-specific statistic $f_b(C)$ identifies excess sharing of derived alleles between the branch of the tree on the y axis and the species C on the x axis (see Supplementary Note). The ASTRAL* tree was used as a basis for the branch statistic and grey data points in the matrix correspond to tests that are not consistent with the phylogeny. Colours correspond to eco-morphological groups as in Fig. 1. Asterisks denote block jackknifing significance at $|Z| > 3.17$ (Holm-Bonferroni FWER < 0.001).

mbuna and therefore are likely to be representative of much of the genetic diversity within the group.

To explore the origins of the Lake Malawi radiation in greater detail, we obtained 24 additional *Astatotilapia* whole-genome

sequences from outside Lake Malawi: five *A. calliptera* from Indian Ocean catchments, thus covering most of its geographical distribution, and 19 individuals from seven other *Astatotilapia* species (Supplementary Table 2). We generated new variant calls

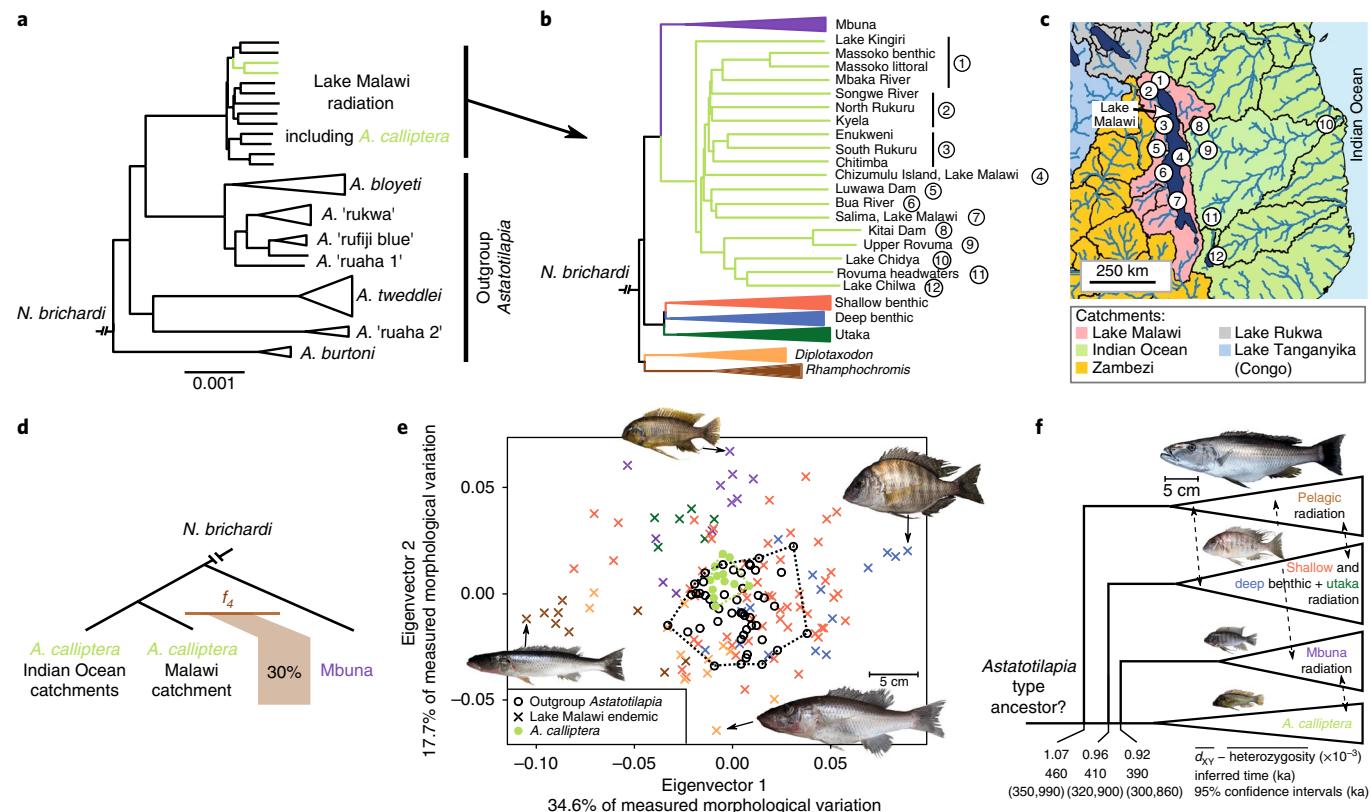


Fig. 4 | Origins of the radiation and the role of *A. calliptera*. **a**, A neighbour-joining phylogeny showing the Lake Malawi radiation in the context of other East African *Astatotilapia* taxa. **b**, A Lake Malawi neighbour-joining phylogeny with expanded view of *A. calliptera*, with all other groups collapsed. **c**, Approximate *A. calliptera* sampling locations shown on a map of the broader Lake Malawi region. Black lines correspond to present-day level 3 catchment boundaries from the US Geological Survey's HYDRO1K dataset (<https://lta.cr.usgs.gov/HYDRO1K>). **d**, Strong f_4 admixture ratio signal showing that Malawi catchment *A. calliptera* are closer to mbuna than their Indian Ocean catchment counterparts. **e**, PCA of body shape variation of Lake Malawi endemics, *A. calliptera* and other *Astatotilapia* taxa, obtained from geometric morphometric analysis. **f**, A phylogeny with the same topology as in **b** but displayed with a straight line between the ancestor and *A. calliptera*. For each branch off this lineage, we show mean sequence divergence (d_{XY}) minus mean heterozygosity, and translation of this value into a mean divergence time estimate with 95% CI reflecting the statistical uncertainty in mutation rate. Dashed lines with arrows indicate likely instances of gene flow between major groups; their true timings are uncertain.

(Supplementary Methods) and first constructed a neighbour-joining tree, finding that all the *A. calliptera* (including Indian Ocean catchments) cluster as a single group nested at the same place within the radiation, whereas the other *Astatotilapia* species branched off well before the lake radiation (Fig. 4a–c). All *A. calliptera* individuals cluster by geography (Fig. 4b,c), except for the specimen from the crater lake Lake Kingiri, whose position in the tree is likely to be a result of admixture with *O. tetrastigma*. Indeed, a neighbour-joining tree built only with *A. calliptera* samples (Supplementary Fig. 17) places the Kingiri individual according to geography with the specimens from the nearby crater lake Lake Massoko and the Mbaka River.

Applying the same logic as above, we tested whether the position of the *A. calliptera* group in the neighbour-joining tree changes when the tree is built without mbuna (as would be expected if *A. calliptera* were affected by hybridization with mbuna). We found that the position of *A. calliptera* is unaffected (Supplementary Fig. 18), suggesting that the nested position is not due to later hybridization. The f statistics in Fig. 3 further support this, because the signals involving the whole mbuna or *A. calliptera* groups are modest and do not suggest erroneous placement of these groups in all phylogenetic analyses. Furthermore, the nested position of *A. calliptera* is also supported by the vast majority of the genome. Searching for the basal branch in a set of 2,638 local maximum likelihood phylogenies, we found results that agree with the whole-genome ASTRAL,

SNAPP and neighbour-joining trees: the most common basal branches are the pelagic groups *Rhamphochromis* and *Diplotaxodon* (in 42.12% of the genomic windows). In comparison, *A. calliptera* (including Indian Ocean catchment samples) were found to be basal only in 5.99% of the windows (Supplementary Fig. 19).

Joyce et al.²⁰ reported that the mtDNA haplogroup of *A. calliptera* from the Indian Ocean catchment clustered with mbuna (as we confirm in Supplementary Fig. 15) and suggested that there had been repeated colonization of Lake Malawi by two independent *Astatotilapia* lineages with different mitochondrial haplogroups: the first founding the entire species flock, and the second, with the Indian Ocean catchment mtDNA haplogroup, introgressing into the Malawi radiation and contributing strongly to the mbuna. This hypothesis predicts that, compared with the Malawi catchment *A. calliptera*, the Indian Ocean catchment *A. calliptera* should be closer to mbuna. However, across the nuclear genome we found a strong signal in the opposite direction, with 30% excess allele sharing between Malawi catchment *A. calliptera* and mbuna (Fig. 4d). Therefore, the Joyce et al.²⁰ hypothesis that the mbuna, the most species-rich group within the radiation, may be a hybrid lineage formed from independent invasions is not supported by genome-wide data.

It has been repeatedly suggested that *A. calliptera* may be the direct descendant of the riverine-generalist lineage that seeded the Lake Malawi radiation^{7,50,53,54}. Our interpretation of this argument

is that the ancestor was probably a riverine generalist that was ecologically and phenotypically similar to *A. calliptera* and other *Astatotilapia*. This hypothesis is lent further support by geometric morphometric analysis. Using 17 homologous body shape landmarks we established that, despite the relatively large genetic divergence, *A. calliptera* is nested within the morphospace of the other more distantly related but ecologically similar *Astatotilapia* species (Fig. 4a,e), and these together have a central position within the morphological space of the Lake Malawi radiation (Fig. 4e and Supplementary Fig. 16).

To reconcile the nested phylogenetic position of *A. calliptera* with its generalist ‘prototype’ phenotype, we propose a model where the Lake Malawi species flock consists of three separate radiations splitting off from the lineage leading to *A. calliptera*. The relationships between the major groups supported by the ASTRAL, SNAPP and neighbour-joining methods suggest that the pelagic radiation was seeded first, then the benthic + utaka, and finally the rock-dwelling mbuna, all in a relatively quick succession, followed by subsequent gene flow as described above (Fig. 4f; the pelagic versus utaka + benthic branching order is swapped in the SVDquartets tree in Supplementary Fig. 9). Applying our per-generation mutation rate to observed genomic divergences we obtained mean divergence time estimates between these lineages between 460 thousand years ago (ka) (95% CI 350–990 ka) and 390 ka (95% CI 300–860 ka) (Fig. 4f), assuming three years per generation as in ref.⁵⁵. The point estimates all fall within the second-most recent prolonged deep-lake phase inferred from the Lake Malawi paleoecological record⁵⁶, while the upper ends of the confidence intervals cover the third deep-lake phase at about 800 ka. Considering that our split time estimates from sequence divergence are likely to be reduced by subsequent gene-flow, leading to underestimates, the data are consistent with a previous report based on fossil time calibration which put the origin of the Lake Malawi radiation at 700–800 ka¹².

The fact that the common ancestor of all the *A. calliptera* appears to be younger than the Malawi radiation suggests that the Lake Malawi *A. calliptera* population has been a reservoir that has repopulated the river systems and more transient lakes following dry–wet transitions in the East African hydroclimate^{56,57}. Our results do not fully resolve whether the lineage leading from the common ancestor to *A. calliptera* retained its riverine generalist phenotype throughout or whether a lacustrine species evolved at some point (for example, the common ancestor of *A. calliptera* and mbuna) and later de-specialized again to recolonize the rivers. However, while it is a possibility, we suggest that it is unlikely that the many strong phenotypic affinities of *A. calliptera* to the basal *Astatotilapia* (Fig. 4e; refs^{58,59}) would be reinvented from a lacustrine species.

Signatures and consequences of selection on coding sequences. To gain insight into the functional basis of diversification and adaptation in Lake Malawi cichlids, we next turned our attention to protein-coding genes. We compared the mean between-species levels of non-synonymous variation \bar{p}_N to synonymous variation \bar{p}_S in 20,664 genes and calculated the difference between these two values ($\delta_{N-S} = \bar{p}_N - \bar{p}_S$). Overall, coding sequence exhibits signatures of purifying selection: the average between-species \bar{p}_N was 54% lower than in a random matching set of non-coding regions. Interestingly, the average between-species synonymous variation \bar{p}_S in genes was 13% higher than in non-coding control regions ($P < 2.2 \times 10^{-16}$, one-tailed Mann–Whitney *U*-test). One possible explanation of this observation would be if intergenic regions were homogenized by gene flow, whereas protein-coding genes were more resistant to this.

To control for statistical effects of variation in gene length and sequence composition we normalized the δ_{N-S} values per gene by taking into account the variance across all pairwise sequence comparisons for each gene, deriving the non-synonymous excess score Δ_{N-S} (see Methods). Values at the upper tail of the distribution of

Δ_{N-S} are substantially over-represented in the actual data when compared to a null model based on random sampling of codons (Fig. 5a). We focus below on the top 5% of the distribution ($\Delta_{N-S} > 40.2$, 1034 candidate genes). Genes with elevated Δ_{N-S} are expected to have been under positive selection at multiple non-synonymous sites, either recently repeatedly within multiple species or ancestrally. Therefore, the statistic reveals only a limited subset of positive selection events from the history of the radiation (for example a selection event on a single amino acid would not be detected). Furthermore, to minimize any effect of gene prediction errors, all the following analyses focus on the 15,980 (77.3% of total) genes for which zebrafish homologues were found in a previous study¹¹; selection scores of genes without homologues are briefly discussed in the Supplementary Note.

Cichlids have an unexpectedly large number of gene duplicates, which has possibly contributed to their extensive adaptive radiations^{3,11}. To investigate the extent of divergent selection on gene duplicates, we examined how the Δ_{N-S} scores are related to gene copy numbers in the reference genomes. Focusing on homologous genes annotated both in the Malawi reference (*M. zebra*) and in the zebrafish genome, we found that the highest proportion of candidate genes was among genes with two or more copies in both genomes ($N - N$). The relative enrichment in this category is both substantial and highly significant (Fig. 5b). On the other hand, the increase in proportion of candidate genes in the $N - 1$ category (multiple copies in the *M. zebra* genome but only one copy in zebrafish) is much smaller and is not significant (χ^2 test $P = 0.18$), suggesting that selection is occurring more often within ancient multi-copy gene families, rather than on genes with cichlid-specific duplications.

We used GO annotation of zebrafish homologues to test whether candidate genes are enriched for particular functional categories (Methods). We found significant enrichment for 30 GO terms (range: $1.6 \times 10^{-8} < P < 0.01$, weigh algorithm⁶⁰; Supplementary Table 3): 10 in the ‘molecular function’, 4 in the ‘cellular component’ and 16 in the ‘biological process’ category. Combining all the results in a network (connecting terms that share many genes) revealed clear clusters of enriched terms related to (1) haemoglobin function and oxygen transport; (2) phototransduction and visual perception; and (3) the immune system, especially inflammatory response and cytokine activity (Fig. 5c). That evolution of genes in these functional categories has contributed to cichlid radiations has been suggested previously (see below); it is nevertheless interesting that these categories stand out in a genome-wide analysis.

Shared mechanisms of depth adaptation. To gain insight into the distribution of adaptive alleles across the radiation, we built maximum likelihood trees from amino acid sequences of candidate genes, thus summarizing potentially complex haplotype genealogy networks. Focusing on the significantly enriched GO categories, many haplotype trees have features that are unusual in the broader dataset: the haplotypes from the deep benthic group and the deep-water pelagic *Diplotaxodon* tend to group together (despite these two groups being distant in whole-genome phylogenies and monophyletic in only two out of 2,638 local maximum likelihood trees) and also tend to be disproportionately diverse when compared with the rest of the radiation. We quantified both excess similarity and diversity, and found that both measures are elevated for candidate genes in the ‘visual perception’ category (Fig. 6a; Mann–Whitney *U*-tests: $P = 0.007$ for similarity, $P = 0.08$ for shared diversity, and $P = 0.003$ when the scores are added) and also for the ‘haemoglobin complex’ category (P values not significant owing to the small number of genes).

Sharply decreasing levels of dissolved oxygen and low light intensities with narrow short-wavelength spectra are the hallmarks of the habitats below about 50 m to which the deep benthic and *Diplotaxodon* groups have both adapted, either convergently or in

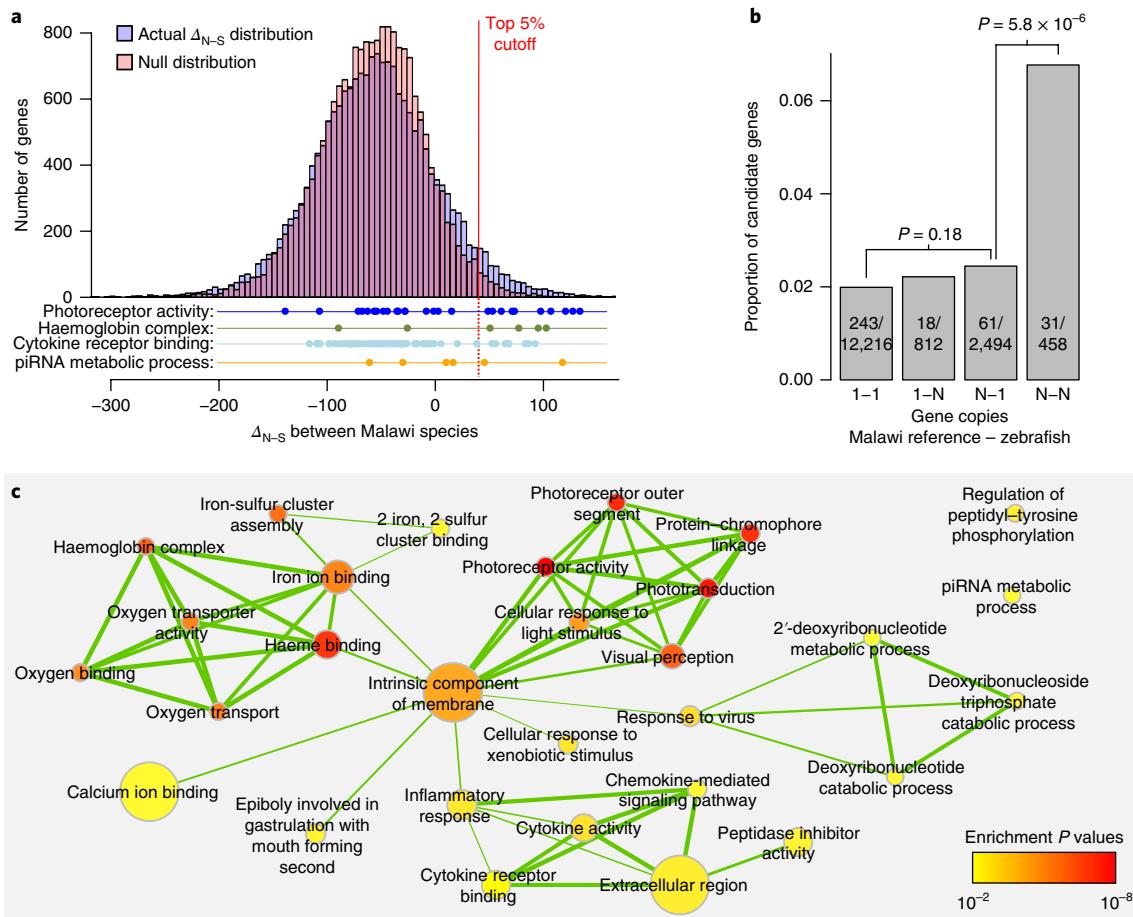


Fig. 5 | Gene selection scores, copy numbers, and ontology enrichment. **a**, The distribution of the non-synonymous variation excess scores (Δ_{N-S}) highlighting the top 5% cutoff, compared against a null model. The null was derived by calculating the statistic on randomly sampled combinations of codons. We also show the distributions of genes in selected gene ontology (GO) categories which are overrepresented in the top 5%. **b**, The relationship between the probability of Δ_{N-S} being in the top 5% and the relative copy numbers of genes in the Lake Malawi reference (*M. zebra*) and zebrafish. The P values are based on χ^2 tests of independence. Genes existing in two or more copies in both zebrafish and Malawi cichlids are disproportionately represented among candidate selected genes. **c**, An enrichment map for significantly enriched GO terms (cutoff at $P \leq 0.01$). The level of overlap between GO enriched terms is indicated by the thickness of the edge between them. The colour of each node indicates the P value for the term and the size of the node is proportional to the number of genes annotated with that GO category.

parallel⁶¹. Shared signatures of selection in genes involved in vision and in oxygen transport therefore point to shared molecular mechanisms underlying this ecological parallelism. Further evidence of shared mechanisms of adaptation is that, for genes annotated with ‘photoreceptor activity’ and ‘haemoglobin complex’ GO terms, the Δ_{N-S} selection score is strongly correlated with the local levels of excess allele sharing between the two depth-adapted groups measured by the f_{dm} statistic, a conservative version of the f statistic more suited to analysing small genomic intervals⁵⁵ (Fig. 6b; $\rho_s = 0.63$ and 0.81, $P = 0.001$ and $P = 0.051$, respectively).

Vision genes with high similarity and diversity scores for the deep benthic and *Diploplotaxodon* groups include three opsins: the green-sensitive *RH2Aβ* and *RH2B*, and rhodopsin (Fig. 6a and Supplementary Fig. 20). The specific residues that distinguish the deep-water-adapted groups from the rest of the radiation differ between the two RH2 copies, with only one shared mutation out of a possible fourteen (Supplementary Fig. 20). *RH2Aβ* and *RH2B* are located less than 40 kilobases (kb) apart on the same chromosome (Fig. 6c); a third parologue, *RH2Aα*, is located between them, but does not show signatures of shared depth adaptation (Supplementary Fig. 21), consistent with reports of functional divergence between *RH2Aα* and *RH2Aβ*^{62,63}. A similar, albeit weaker, signature of shared

depth-related selection is apparent in rhodopsin, which is known to have a role in deep-water adaptation in cichlids⁶⁴. Previously, we discussed the role of coding variants in rhodopsin in the early stages of speciation of *A. calliptera* in the crater lake Lake Massoko⁵⁵. The haplotype tree presented here for the broader radiation shows that the Massoko alleles did not originate by mutation in that lake but were selected out of ancestral variation (Fig. 6a). The remaining opsin genes are less likely to be involved in shared depth adaptation (Supplementary Note).

There have been many studies of selection on opsin genes in fish^{55–67}, including selection associated with depth preference, but having whole-genome coverage allows us to investigate other components of primary visual perception in an unbiased fashion. We found shared patterns of selection between deep benthics and *Diploplotaxodon* in six other vision-associated candidate genes (Fig. 6a). The functions of these genes, together with the fact that *RH2Aβ* and *RH2B* are expressed exclusively in double-cone photoreceptors, suggest a prominent role of cone-cell vision in depth adaptation. The wavelength of maximum absorbance in cone cells expressing a mixture of *RH2Aβ* with *RH2B* ($\lambda_{max} = 498$ nm) corresponds to the part of the visible-light spectrum that best transmits into deep water in Lake Malawi⁶⁷.

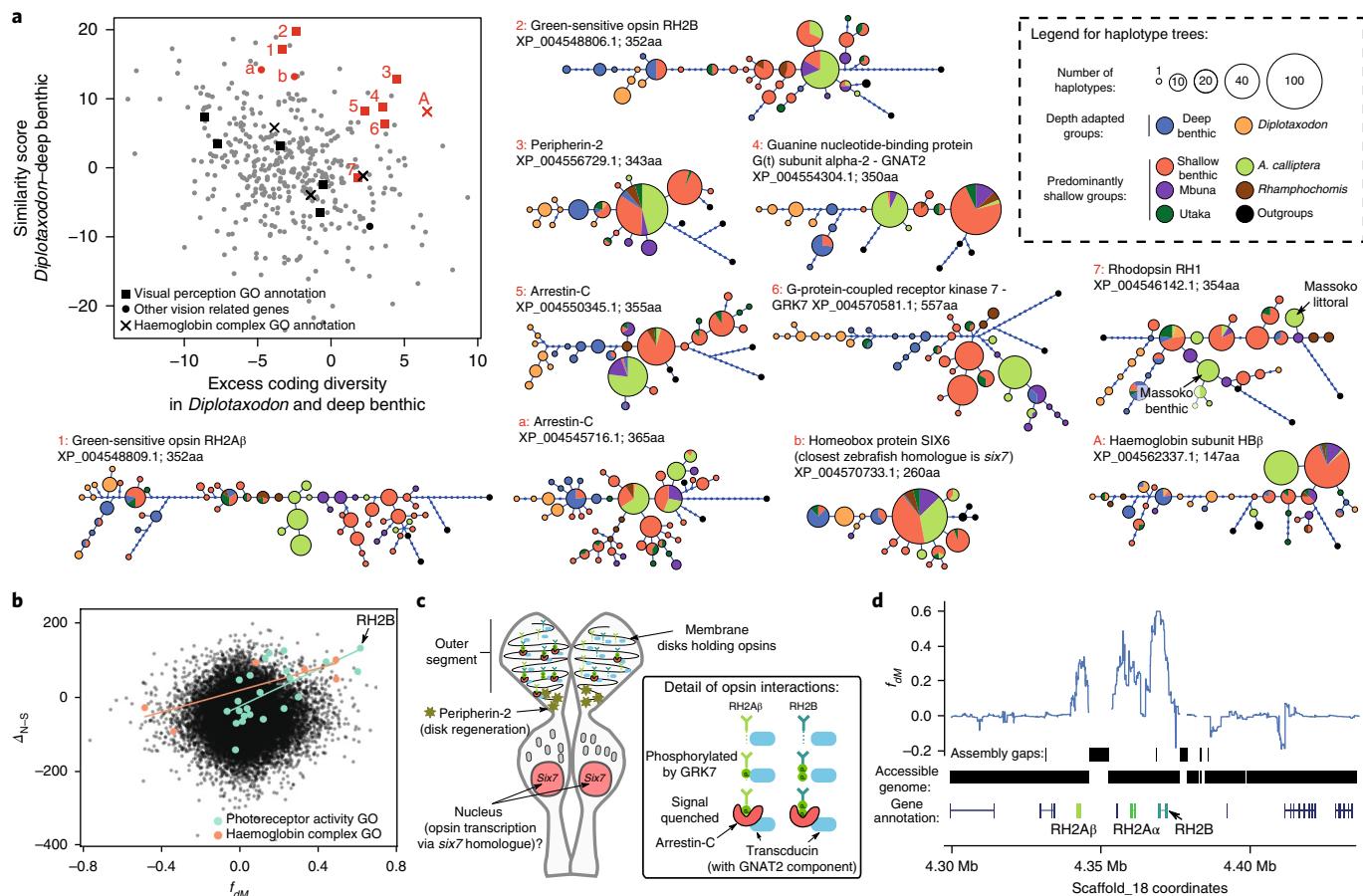


Fig. 6 | Shared selection between the deep-water-adapted groups *Diplotaxodon* and deep benthic. **a**, The scatterplot shows the distribution of genes with high Δ_{N-S} scores (candidates for positive selection) along axes reflecting shared selection signatures. Only genes with zebrafish homologues are shown. Amino acid haplotype trees, shown for genes as indicated by the red symbols and numbers, indicate that *Diplotaxodon* and deep benthic species are often divergent from other taxa, but similar to each other. Outgroups include *Oreochromis niloticus*, *Neolamprologus brichardi*, *Astatotilapia burtoni*, and *Pundamilia nyererei*. **b**, Selection scores plotted against f_{dm} (mbuna, deep benthic; *Diplotaxodon*, *N. brichardi*), a measure of local excess allele sharing between deep benthic and *Diplotaxodon*⁵⁵. Overall there is no correlation between Δ_{N-S} and f_{dm} . However, the strong correlation between Δ_{N-S} and f_{dm} in the highlighted GO categories suggests that positively selected alleles in those categories tend to be subject to introgression or convergent selection between *Diplotaxodon* and the deep benthic group. **c**, A schematic drawing of a double cone photoreceptor expressing the green-sensitive opsins and illustrating the functions of other genes with signatures of shared selection. **d**, f_{dm} calculated in sliding windows of 100 SNPs around the green opsin cluster, revealing that excess allele sharing between deep benthic and *Diplotaxodon* extends far beyond the coding sequences.

Figure 6c illustrates interactions of the vision genes with shared selection patterns in the cichlid double-cone photoreceptor. The homeobox protein *six7* governs the expression of RH2 opsins and is essential for the development of green cones in zebrafish⁶⁸ (specific mutations are highlighted in Supplementary Fig. 20). The kinase GRK7 and the retinal cone arrestin-C have complementary roles in photoresponse recovery: arrestin produces the final shutoff of the cone pigment following phosphorylation by GRK7, thus determining the temporal resolution of motion vision⁶⁹. Bases near to the carboxy terminus in *RH2A β* mutated away from serine (S290Y and S292G), thus reducing the number of residues that can be modified by GRK7 (Supplementary Fig. 20). The transducin subunit GNAT2 is located exclusively in the cone receptors and is a key component of the pathway that converts light stimulus into electrical response in these cells⁷⁰. Finally, peripherin-2 is essential to the development and renewal of the membrane system that holds the opsin pigments in both rod and cone cells⁷¹.

Haemoglobin genes in teleost fish are found in two separate chromosomal locations: the minor 'LA' cluster and the major 'MN' cluster⁷². The region around the LA cluster has been highlighted by selection scans among four *Diplotaxodon* species by ref. ⁷³, who

also noted the similarity of the haemoglobin subunit beta (*HB β*) haplotypes between *Diplotaxodon* and deep benthic species. We confirmed signatures of selection in the two annotated LA cluster haemoglobins. In addition, we found that four haemoglobin subunits (*HB β 1*, *HB β 2*, *HB α 2* and *HB α 3*) from the MN cluster are also among the genes with high selection scores (Supplementary Fig. 22). The shared patterns of depth selection may be particular to the β -globin genes (Supplementary Fig. 22), although this hypothesis remains tentative, because the repetitive nature of the MN cluster precludes us from confidently examining all haemoglobin genes.

A key question concerns the mechanism leading to the similarity of haplotypes in *Diplotaxodon* and deep benthics. Possibilities include parallel selection on variation segregating in both groups owing to common ancestry, selection on the gene flow that we described in a previous section, or independent selection on new mutations. From considering the haplotype trees and local patterns of excess allele sharing (using f_{dm} statistics⁵⁵), there is evidence for each of these processes acting on different genes. The haplotype trees for rhodopsin and *HB β* have outgroup taxa (and also *A. caliptera*) appearing at multiple locations on their haplotype networks (Fig. 6a), suggesting that the haplotype diversity of these genes

may reflect ancestral variation. In contrast, trees for the green cone genes show the Malawi radiation all being derived with respect to outgroups and we found substantially elevated f_{dm} scores extending for around 40 kb around the RH2 cluster (Fig. 6d), consistent with adaptive introgression in a pattern reminiscent of mimicry loci in *Heliconius* butterflies⁷⁴. Finally, the peaks in f_{dm} around peripherin-2 and one of the arrestin-C genes are narrow, ending at the gene boundaries, and f_{dm} scores are elevated only for non-synonymous variants; synonymous variants do not show excess allele sharing (Supplementary Fig. 23). Owing to the close proximity of non-synonymous and synonymous sites within the same gene, this suggests that for these two genes there may have been independent selection on the same de novo mutations.

Discussion

Variation in genome sequences forms the substrate for evolution. Here we described genome variation at the full sequence level across the Lake Malawi haplochromine cichlid radiation. We focused on ecomorphological diversity, representing more than half the genera from each major group, rather than obtaining deep coverage of species within any particular group. Therefore, we have more samples from the morphologically highly diverse benthic lineages than, for example, from the mbuna where there are relatively fewer genera and many species are largely recognized by colour differences.

The observation that cichlids within an African Great Lake radiation are genetically very similar is not new⁷⁵, but we now quantify the relationship of this to within-species variation, and the consequences for variation in local phylogeny across the genome. The fact that between-species divergence is generally only slightly higher than within species diversity, is probably the result of the young age of the radiation, the relatively low mutation rate and of gene flow between taxa. Within-species diversity itself is relatively low for vertebrates, at around 0.1%, suggesting that low genome-wide nucleotide diversity levels do not necessarily limit rapid adaptation and speciation, results that are in contrast to a recent report that found that high diversity levels may have been important for rapid adaptation in Atlantic killifish⁷⁶. One possibility is that in cichlids repeated selection has maintained diversity in adaptive alleles for a range of traits that support ecological diversification, as we have concluded for rhodopsin and HB β and as appears to be the case for some adaptive variants in sticklebacks⁷⁷.

We provide evidence that gene flow during the radiation, although not ubiquitous, has certainly been extensive. Overall, the numerous violations of the bifurcating species tree model suggest that full resolution of interspecies relationships in this system will require network approaches (see for example section 6.2 of ref. ³⁵) and population genomic analyses within the structured coalescent framework with gene flow. The majority of the signals affect groups of species, suggesting events involving their common ancestors, or are between closely related species within the major ecological groups. The only strong and clear example of recent gene flow between individual distantly related species is not within Lake Malawi itself, but between *Otopharynx tetrastigma* from crater Lake Ilamba and local *A. calliptera*. Lake Ilamba is very turbid and the scenario is reminiscent of cichlid admixture in low-visibility conditions in Lake Victoria⁷⁸. It is possible that some of the earlier signals of gene flow between lineages we observed in Lake Malawi may have happened during periods of low lake level when the water is known to have been more turbid⁵⁶.

Our model of the early stages of radiation in Lake Malawi (Fig. 4f) is broadly consistent with the model of initial separation by major habitat divergence²³, although we propose a refinement in which there were three relatively closely spaced separations from a generalist *Astatotilapia* type lineage, initially of pelagic genera *Rhamphochromis* and *Diplotaxodon*, then of shallow- and deep-water benthics and utaka (this includes Kocher's sand dwellers^{23,29}),

and finally of mbuna. Thus, we suggest that Lake Malawi contains three separate haplochromine cichlid radiations stemming from the generalist lineage, interconnected by subsequent gene flow.

The finding that cichlid-specific gene duplicates do not tend to diverge particularly strongly in coding sequences (Fig. 5b) suggests that other mechanisms of diversification following gene duplications may be more important. Divergence via changes in expression patterns has previously been illustrated and discussed¹¹, and future studies addressing structural variation between cichlid genomes will assess the contribution of differential retention of duplicated genes.

The evidence concerning shared adaptation of the visual and oxygen-transport systems to deep-water environments between deep benthics and *Diplotaxodon* suggests different evolutionary mechanisms acting on different genes, even within the same cellular system. It will be interesting to see whether the same genes or even specific mutations underlie depth adaptation in Lake Tanganyika, which harbours specialist deep-water species in least two different tribes⁷⁹ and has a similar light attenuation profile but a steeper oxygen gradient than Lake Malawi⁶¹.

Over the last few decades, East African cichlids have emerged as a model for studying rapid vertebrate evolution^{11,23}. Taking advantage of recently assembled reference genomes¹¹, our data and results provide insight into patterns of sequence sharing and adaptation across the Lake Malawi radiation, and into mechanisms of rapid phenotypic diversification. The datasets are publicly available (see 'Data availability') and will underpin further studies on specific taxa and molecular systems. For example, we envisage that our results, clarifying the relationships between all the main lineages and many individual species, will facilitate speciation studies, which require investigation of taxon pairs at varying stages on the speciation continuum^{80,81}, and studies on the role of adaptive gene flow in speciation.

Methods

Samples. Ethanol-preserved fin clips were collected by M. J. Genner and G. F. Turner between 2004 and 2014 from Tanzania and Malawi, in collaboration with the Tanzania Fisheries Research Institute (the MolEcoFish Project) and with the Fisheries Research Unit of the Government of Malawi (various collaborative projects). Samples were collected and exported with the permission of the Tanzania Commission for Science and Technology, the Tanzania Fisheries Research Institute, and the Fisheries Research Unit of the Government of Malawi.

From sequencing to a variant callset. The analyses presented above are based on SNPs obtained from Illumina short (100–125 bp) reads, aligned to the *M. zebra* reference assembly version 1.1¹¹ with bwa-mem⁸², followed by GATK haplotype caller⁸³ and samtools/bcftools⁸⁴ variant calling restricted to 653 Mb of 'accessible genome' where variants can be determined confidently with short reads, filtering, genotype refinement, imputation and phasing in BEAGLE⁸⁵ and further haplotype phasing with shapeit v2⁸⁶, including the use of phase-informative reads⁸⁷. For details please see Supplementary Methods.

Linkage disequilibrium calculations. The haplotype disequilibrium coefficient⁸⁸ r^2 between pairs of SNPs was calculated along the phased scaffolds 0 to 201 (scaffolds are assembled fragments of the reference genome and scaffolds 0–201 are longer than 1 Mb), using vcftools v0.1.12b⁸⁹ with the options --hap-r2 --ld-window-bp 50000. To reduce the computational burden, we used a random subsample of 10% of SNPs. We binned the r^2 values according to the distance between SNPs into 1-kb or 100-bp windows and plotted the average values in each bin.

To estimate background linkage disequilibrium, we calculated haplotype r^2 between variants mapping to different linkage groups in the *Oreochromis niloticus* genome assembly. First, we used the chain files generated by the whole genome alignment pipeline⁹⁰ (see Supplementary Methods) and the UCSC liftOver tool (http://hgdownload.soe.ucsc.edu/downloads.html#source_downloads) to translate the genomic coordinates of all SNPs to the *O. niloticus* coordinates. Then we calculated linkage disequilibrium between variants mapping to linkage groups LG1 and LG2.

De novo mutation rate estimation. In each trio we looked for mutations in the child that were not present in either of its parents. Because the results of this analysis are very sensitive to false positives and false negative rates, we used higher coverage sequencing (about 40× average) and applied more stringent genome masks than in the population genomic work. Increased coverage supports clean separation of sequencing errors and somatic mutations from true heterozygous

calls in the offspring, and improved ability to distinguish single copy versus multi-copy sequence on a per-individual basis.

First we determined the ‘accessible genome’ (that is the regions of the genome in which the mutations can be confidently called (de novo mutations) for each trio by excluding:

1. Genomic regions where mapped read depth in any member of a trio is $\leq 25 \times$ or $> 50 \times$
2. Bases where either of the parents has a mapped read that does not match the reference (the specific bases where any read has non-reference alleles in the parents were masked)
3. Sequences where indels (base insertion or deletion) were called in any sample (we also excluded ± 3 bp of sequence surrounding the indel)
4. Sites that were called as multiallelic among the nine samples in the overall trios dataset
5. Known segregating variable sites—that is, sites with alternative alleles found in four and more copies in the overall Lake Malawi dataset
6. Sites in the reference where less than 90% of overlapping 50-mers (sub-sequences of length 50) could be matched back uniquely and without 1-difference. For this we used Heng Li’s SNPable tool (<http://lh3lh3.users.sourceforge.net/snitable.shtml>), dividing the reference genome into overlapping k-mers (sequences of length k ; we used $k=50$), and then aligning the extracted k-mers back to the genome (we used bwa aln -R 1000000 -O 3 -E 3).

After excluding sites in the categories above, we were left with an ‘accessible genome’ of 516.6 Mb in the *A. calliptera* trio, 459 Mb in the *A. stuartgranti* trio and 404 Mb in the *L. lethrinus* trio. Because any observed de novo mutation could have occurred either on the chromosome inherited from the mother or on the chromosome inherited from the father, the point estimate of the per-generation per-base-pair mutation rate is: $\mu = n_{\text{mutations}} / (2 \times \text{the size of the accessible genome})$.

Next we set out to search for de novo mutations: that is, heterozygous sites in the offspring within the accessible genome. Under random sampling there is an equal probability of seeing a read with either of the two alleles at a heterozygous site. Therefore, N_a (the number of reads supporting the alternative allele) is distributed as approximately Binomial(read depth, 0.5). We filtered out variants with observed N_a values below the 2.5th or above the 97.5th percentiles of this distribution, thus accepting a false-negative rate of 5%. We also filtered out sites where the offspring call had Read Position or Base Quality rank-sum test Z-score exceeding the 99.5th percentile of the standard normal distribution or where the strand-bias phred-scaled P value ($-\log_{10}(\text{error probability})$) was ≥ 20 or where the phred-scaled genotype quality in either mother, father or offspring was ≤ 30 . For simplicity, assuming these filters are independent, they are expected to introduce a false-negative rate of 7.17%. The mutation rate estimate was adjusted to account for this.

After filtering, we found nine de novo mutations across the three offspring. For each mutation we double-checked the alignment in the IGV genome browser and found all of them were single base mutations supported by high number of reads (> 8) in the offspring. The 95% confidence intervals for the number of observed mutations were calculated using the ‘exact’ method relating χ^2 and Poisson distributions^{91,92}. If N is the number of observed mutations, the lower (ci N_L) and upper (ci N_U) limits are:

$$\text{ci}N_L = \frac{P(\chi^2_{2N} \leq 0.025)}{2} \quad \text{ci}N_U = \frac{P(\chi^2_{2(N+1)} \geq 0.975)}{2}$$

where $2N$ and $2(N+1)$ are the degrees of freedom of the corresponding χ^2 distributions.

PCA. SNPs with minor allele frequency ≥ 0.05 were selected using the bcftools (v1.2) view option --min-af 0.05:minor. The program vcftools v0.1.12b was then used to export that data into PLINK format⁹³. Next, the variants were linkage-disequilibrium-pruned to obtain a set of variants in approximate linkage equilibrium (unlinked sites) using the --indep-pairwise 50 5 0.2 option in PLINK v1.0.7. PCA on the resulting set of variants was performed using the smartpca program from the eigensoft v5.0.2 software package⁹⁴ with default parameters.

Genome-wide F_{ST} calculations. In addition to performing PCA, the smartpca program from the eigensoft v5.0.2 software package also calculates genome-wide F_{ST} for all pairs of populations specified by the sixth column in the .pedind file. For the calculation, it uses the Hudson estimator, as defined previously⁹⁵ in their equation (10), and the ‘ratio of averages’ is used to combine estimates of F_{ST} across multiple variants, as they recommended. We used all SNPs (no minor allele frequency filtering).

Allele sharing test for group assignment. We tested whether two individuals who come from the same group always share more derived alleles with each other than with any individuals from other groups. Technically, we implemented this using the D statistic (ABBA-BABA tests) framework^{31,32}, by calculating $D(A, G_1; G_2, O)$ for all permutations of individuals, where G_1 and G_2 come from the same

eco-morphological group and A from a different group. The outgroup O was always *N. brichardi* from Lake Tanganyika. Note that this is an unusual use of the D statistics and our aim here was not to look for gene flow but to test whether allele sharing is greater within eco-morphological groups (G_1 with G_2) compared to across groups (A with G_2), in which case $D(A, G_1; G_2, O) > 0$. All results were statistically significant, which was assessed using block jackknife³¹ on windows of 60,000 SNPs.

D_{\min} statistic. Here we calculated the D statistic for each trio of species (A, B, C) and for all possible tree topologies (the outgroup again fixed as *N. brichardi*). Therefore, $D_{\min} = \min(|D(A, B; C, O)|, |D(A, C; B, O)|, |D(C, B; A, O)|)$. If this is significantly elevated, then allele sharing within the trio of species is inconsistent with any simple tree topology. Note that this approach is conservative in the sense that the D_{\min} score for each trio is considered in isolation and we ignore ‘higher-order’ inconsistencies where different D_{\min} trio topologies are inconsistent with each other. Statistical significance was assessed using block jackknife³¹ on windows of 60,000 SNPs and family wise error rate (FWER) was calculated following the Holm–Bonferroni method.

Sample selection for demographic analyses. To prevent potential confounding effects of uneven sequencing depth, we limited these analyses to one high-coverage (15 \times) individual per species. Species without a high-coverage sample (*P. subocularis*, *F. rostratus* and *L. trewavasae*) were not included.

Outgroup sequences/alleles. Outgroup (Supplementary Table 5) sequences in *M. zebra* genomic coordinates were obtained based on pairwise whole-genome alignments (Supplementary Methods). Insertions in the outgroup were ignored and deletions filled by ‘N’ characters.

Local phylogenetic trees and maximum clade credibility. To generate a multiple alignment input in fasta format we used the getWGSeq subprogram of *evo*. We set the window size in terms of the numbers of variants rather than physical length (8,000 variants; the --split 8,000 option) aiming for the local regions to have similar strengths of phylogenetic signal. Small windows at the ends of scaffolds were discarded. We limited the sequence output to the accessible genome using the --accessibleGenomeBED option. The *N. brichardi* outgroup sequence in *M. zebra* genomic coordinates was added via the --incl-Pn option.

Maximum likelihood phylogenies were inferred using RAxML version 7.7.8⁹⁶ under the GTRGAMMA model. The best tree for each region was selected out of twenty alternative runs on distinct starting maximum parsimony trees (the -N 20 option).

The MCC trees were calculated in TreeAnnotator version 2.4.2, a part of the BEAST2 platform⁹⁷. Clade credibility is the frequency with which a clade appears in the tree set; the MCC tree is the tree (from among the trees in the set) that maximizes the product of the frequencies of all its clades³³. The node heights for the MCC trees are derived as a summary from the heights of each clade in the whole tree set via the ‘common ancestor’ heights option.

Mitochondrial DNA phylogenies. The mtDNA sequence corresponds to scaffolds 747 and 2,036 in the *M. zebra* reference. Variants from these scaffolds were subjected to the same filtering as in the rest of the genome except for the depth filter because the mapped read depth was much higher (approximately 300–400 \times per sample). Because of the greater sequence diversity in the mtDNA genome, we found that more than 10% of variants were multiallelic. Therefore, we separated SNPs from indels at multiallelic sites using bcftools norm with the --multiallelics - option, then removed indels and the merged multiallelic SNPs back together with the --multiallelics + option. Sequences in the fasta format were generated using the bcftools consensus command, and missing genotypes in the VCF replaced by the ‘N’ character with the --mask option. The *N. brichardi* outgroup sequence in *M. zebra* genomic coordinates was added to the fasta files.

A maximum likelihood tree was inferred using RAxML version 7.7.8⁹⁶ under the GTRGAMMA model. The best tree was selected out of twenty alternative runs on distinct starting maximum parsimony trees (using the -N 20 option) and two hundred bootstrap replicates were obtained using RAxML’s rapid bootstrapping algorithm⁹⁸ satisfying the -N autoFC frequency-based bootstrap stopping criterion. Bipartition bootstrap support was drawn on the maximum likelihood tree using the RAxML -f b option.

Neighbour-joining trees and the residuals. For the neighbour-joining⁹⁹ trees we calculated the average numbers of single-nucleotide differences between haplotypes for each pair of species. This simple pairwise difference matrix was divided by the accessible genome size to obtain pairwise differences per base pair, which are equivalent to the \hat{p}_{AB} variable of Dasarathy et al.⁴². Then we followed equation (8) from Dasarathy et al.⁴² and calculated their corrected measure of dissimilarity:

$$\hat{d}_{AB} = -\frac{3}{4} \log \left(1 - \frac{4}{3} \hat{p}_{AB} \right)$$

The \hat{d}_{AB} values were then used as input into the nj() tree-building function implemented in the APE package¹⁰⁰ in the R language.

We measured the distances between all pairs of species in the reconstructed neighbour-joining tree (that is the lengths of branches) using the `get_distance()` method implemented in the ETE3 toolkit for phylogenetic trees¹⁰. Our first measure of ‘tree violation’ is the difference between these distances and the distances between samples in the original matrix that was used to build the neighbour-joining tree.

Multispecies coalescent methods. We applied three different methods that attempt to reconstruct the species tree under the multispecies coalescent model. For a brief discussion of these approaches see Supplementary Methods.

For SNAPP³⁶ we used a random subset of about 0.5% of genome-wide SNPs (48,922 SNPs) for 12 individuals representing the eco-morphological groups and the Lake Victoria outgroup *P. nyererei*, whose alleles were filled in based on the whole-genome alignment. The *P. nyererei* alleles were assigned as ‘ancestral’ (0 in the nexus input file). The ‘forward’ and ‘backward’ mutation rate parameters u and v were calculated directly from the data by SNAPP (the ‘Calc mutation’ rates option). The default value 10 was used for the ‘Coalescent rate’ parameter and the value of the parameter was sampled (estimated in the Markov chain Monte Carlo (MCMC) chain). We used uninformative priors as we do not assume strong a priori knowledge about the parameters. The prior for ancestral population sizes was chosen to be a relatively broad gamma distribution with parameters $\alpha = 4$ and $\beta = 20$. The tree height prior λ was set to the initial value of 100 but sampled in the MCMC chain with an uninformative uniform hyperprior on the interval [0, 50,000]. We ran three independent MCMC chains with the same starting parameters, each on 30 threads with a total runtime of over 10 central processing unit (CPU) years. The first one million steps from each MCMC chain was discarded as burn-in. In total, more than 30 million MCMC steps were sampled in the three runs. For the MCMC traces for each run, see Supplementary Fig. 24.

Next we used SVDquartets^{37,38} as implemented in PAUP* (v4.0a, build 159)¹⁰². We prepared the data into the NEXUS ‘dn’ format, using `evo` with the `getWGSeq --whole-genome --makeSVDinput -r` options. This command outputs for each individual the DNA base at each variable site, randomly sampling one of the two alleles at heterozygous sites, and ignoring sites that become monomorphic owing to this random sampling of alleles. The final dataset contained 17,833,187 SNPs. Then we ran SVDquartets in PAUP* setting outgroup to *N. brichardi* and then executing `svdq evalq=all`; specifying that all quartets should be evaluated (not just a random subset). In the final step, PAUP* version of the QFM algorithm¹⁰³ is used to search for the overall tree that minimizes the number of quartets that are inconsistent with it.

Finally we used ASTRAL⁴⁰ (v.5.6.1) with default parameters and the full set of 2,543 local trees generated by RAxML (see above) as input.

Tree comparisons. To summarize the degree of (dis)agreement between the topologies of trees produced by different phylogenetic methods (Fig. 2c), we calculated the normalized Robinson–Foulds distances between pairs of trees¹⁰⁴ using the `RF.dist` function from the `phangorn`¹⁰⁵ package in R with the option `normalize=TRUE`.

Chromopainter and fineSTRUCTURE. Singleton SNPs were excluded using the `bcftools v1.1 -c 2:minor` option, before exporting the remaining variants in the PLINK format⁹³. The chromopainter v0.0.4 software⁴⁷ was then run for the 201 largest genomic scaffolds on shapeit-phased SNPs. Briefly, we created a uniform recombination map using the `makeuniformrefile.pl` script, then estimated the effective population size (N_e) for a subsample of 20 individuals using the chromopainter inbuilt expectation-maximization procedure⁴⁷, averaged over the 20 N_e values using the provided `neverage.pl` script. The chromopainter program was then run for each scaffold independently, with the `-a 0 0` option to run all individuals against all others. Results for individual scaffolds were combined using the `chromocombine` tool before running fineSTRUCTURE v0.0.5 with 1,000,000 burn-in iterations, and 200,000 sample iterations, recording a sample every 1,000 iterations (options `-x 1000000 -y 200000 -z 1000`). Finally, the sample relationship tree was built with fineSTRUCTURE using the `-m T` option and 20,000 iterations.

The f-branch statistic. The f -admixture ratio (f statistic) statistic was developed to estimate the proportion of introgressed material in an admixed population (see SOM18 in ref. ³¹, and f_G in ref. ⁴⁸). However, when calculated for different subsets of samples within the same phylogeny, there are a very large number of highly correlated f values that are hard to interpret. To make the interpretation easier, we developed the ‘ f -branch’ metric or $f_b(C)$: $f_b(C) = \text{median}_A[\min_B[f(A, B; C, O)]]$, where B are samples descending from branch b , and A are samples descending from the sister branch of b . The outgroup O was always *N. brichardi*. The $f_b(C)$ score provides for each branch b of a given phylogeny and each sample C a summary of excess allele sharing of branch b with sample C (Fig. 3, Supplementary Fig. 26). Each $f_b(C)$ score was also assigned an associated z -score to assess statistical significance $Z_b(C) = \text{median}_A[\min_B[Z(A, B; C, O)]]$. Additional information on the f and $f_b(C)$ statistics, including detailed reasoning behind the design of $f_b(C)$, are in Supplementary Methods.

Geometric morphometric analyses. A total of 168 photographs were used to compare the gross body morphology of *Astatotilapia calliptera* to that of endemic Lake Malawi species and other East African *Astatotilapia* lineages (Supplementary Table 7). Coordinates for 17 homologous landmarks (following ref. ¹⁰⁶) were collected using tpsDig2 v2.26¹⁰⁷. After landmark digitization, analysis of shape variation was carried out in R (v3.3.2) using the package GeoMorph v3.0.2¹⁰⁸. First a General Procrustes Analysis was applied to remove non-shape variation and shape data were corrected for allometric size effects by performing a regression of Procrustes coordinates (10,000 iterations). The resulting allometry-corrected residuals were used in PCA.

Maps. Present-day catchment boundary maps are based on ‘level 3’ detail of the Hydro1K dataset from the US Geological Survey. We downloaded the watershed boundary data from the United Nations Environment Programme website (<http://ede.grid.unep.ch>) and processed it using the QGIS geographic information system software (<http://www.qgis.org/en/site/>).

Protein-coding gene annotations. We used the BROADMZ2 annotation generated by the cichlid genome project¹¹ and removed overlapping transcripts using Jim Kent’s genePredSingleCover program. Genes whose annotated length in nucleotides was not divisible by three were discarded, as they typically had inaccuracies in annotation that would require manual curation (2,495 out of 23,698 genes). We also used the cichlid genome project¹¹ assignment of homologues between the *M. zebra* genome reference and zebrafish (*Danio rerio*).

Coding sequence positive selection scan. We used `evo` with the `getCodingSeq -H b --no-stats` options to obtain the coding sequences for each allele and each gene. The excess of non-synonymous variation (δ_{N-S}) and the non-synonymous variation excess score (Δ_{N-S}) were calculated on a per-gene basis as follows. Let N_{TS} be the number of possible non-synonymous transitions and N_{TV} the number of possible non-synonymous transversion between two sequences; analogously S_{TS} and S_{TV} represent possible synonymous differences. We do not specify the ancestral allele, and therefore consider it equally likely that allele i mutated into allele j or that allele j mutated to allele i . Then let N be the number of observed non-synonymous mutations and S the number of observed synonymous mutations. If there is more than one difference within a codon, all ‘mutation pathways’ (that is, the different orders in which mutations could have happened) have equal probabilities. When a particular allele contained a premature stop codon, the remainder of the sequence after the stop was excluded from the calculations.

Because the transition:transversion ratio in the Lake Malawi dataset was 1.73, and hence (because there are two possible transversions for each possible transition) the prior probability of each transition is 3.46 times that of each transversion, we account for the unequal probabilities of transitions and transversions in calculating the proportions of non-synonymous (p_N) and of synonymous (p_S) as follows:

$$p_N = \frac{N}{3.46 \times N_{TS} + N_{TV}} \quad p_S = \frac{S}{3.46 \times S_{TS} + S_{TV}}$$

The excess of non-synonymous variation (δ_{N-S}) is the average of $p_N - p_S$ over pairwise sequence comparisons. Only between-species sequence comparisons are considered for the Lake Malawi dataset. We normalized the δ_{N-S} values in order to take into account the effect on the variance of this statistic introduced by differences in gene length and by sequence composition. To achieve this, we used the leave-one-out jackknife procedure across different pairwise comparisons for each gene, estimating the standard error. The non-synonymous variation excess score (Δ_{N-S}) is then:

$$\Delta_{N-S} = \frac{\delta_{N-S}}{\text{jackknife_se}(\delta_{N-S})}$$

Note that because the sequences are related by a genealogy, there is a correlation structure between the pairwise comparisons. Therefore, the jackknife approach substantially underestimates the true standard error of δ_{N-S} and is used here simply as a normalization factor.

The null model shown in Fig. 5a was derived by splitting all the coding sequence into its constituent codons, and then randomly sampling these codons with replacement to build new sequences that matched the actual coding genes in their numbers and the length distribution. Then we calculated the Δ_{N-S} scores, as we did for the actual genes and compared the two distributions. High positive values at the upper tail of the distribution are substantially over-represented in the actual data when compared to a null model.

We also calculated the above statistics for random non-coding regions, matching the gene sequences in length. We used the bedtools v2.26.0¹⁰⁹ ‘shuffle’ command to permute the locations of exons along the chromosomes. Of the total length of all the permuted sequences, 98.4% were within the ‘accessible genome’ and outside coding sequences (we required at least 95% in any of the

permuted locations). The specific command was bedtools shuffle -chrom -I exons.bed -excl InaccessibleGenome_andExons.bed -f 0.05 -g chrom.sizes.

GO enrichment. Zebrafish has the most extensive functional gene annotation of any fish species, providing a basis for GO¹¹⁰ term enrichment analysis. GO enrichment for the genes that were candidates for being under positive selection (the top 5% of Δ_{N-S} values) was calculated in R using the topGO v2.26.0 package¹¹¹ from the Bioconductor project¹¹². The GO hierarchical structure was obtained from the GOdb v3.4.0 annotation and linking zebrafish gene identifiers to GO terms was accomplished using the org.Dr.gd.v3.4.0 annotation package. Genome-wide, between 9,024 and 9,353 genes had a GO annotation that could be used by topGO, the exact number depending on the GO category being assessed. The nodeSize parameter was set to 5 to remove GO terms which have fewer than five annotated genes, as suggested in the topGO manual.

There is often an overlap between gene sets annotated with different GO terms, in part because the terms are related to each other in a hierarchical structure¹¹⁰. This is partly accounted for by our use in topGO of the weight algorithm that accounts for the GO graph structure by down-weighing genes in the GO terms that are neighbours of the locally most significant terms in the GO graph⁶⁰. All the *P* values we report are from the weight algorithm, which the authors suggest should be reported without multiple testing correction¹¹¹.

Some interdependency between significant GO terms remains after using the weight algorithm. Therefore, we used the Enrichment Map¹¹³ app for Cytoscape (<http://www.cytoscape.org>) to organize all the significantly enriched terms into networks where terms are connected if they have a high overlap, that is if they share many genes.

Diplotaxodon and deep benthic convergence. To obtain a quantitative measure of the similarity between and the extent of excess diversity in the *Diplotaxodon* and deep benthic amino acid sequences, we calculated simple statistics based on the proportions of non-synonymous differences (p_N scores). Intuitively, the similarity score is high if *Diplotaxodon* and deep benthic jointly have higher p_N than all the others, but are not very different from each other relative to how much diversity there is within *Diplotaxodon* and deep benthic.

Specifically, the similarity score s is calculated as follows:

$$s_{\text{raw}} = \bar{p}_N^O - (\bar{p}_N^B - \bar{p}_N^W)$$

and

$$s = \frac{s_{\text{raw}}}{\text{jackknife_se}(p_N)} - \text{mean} \left(\frac{s_{\text{raw}}}{\text{jackknife_se}(p_N)} \right)$$

where \bar{p}_N^O is the mean p_N between *Diplotaxodon* jointly with deep benthic and all the other Lake Malawi species, \bar{p}_N^B is the mean p_N between *Diplotaxodon* and deep benthic, and \bar{p}_N^W is the mean p_N within *Diplotaxodon* and deep benthic. The jackknife normalization is analogous to the one used for Δ_{N-S} and the mean (\bar{s}_{raw}) is subtracted to centre the statistic at zero.

The excess diversity score is high when the mean p_N scores within *Diplotaxodon* and within deep benthic are high relative to the mean p_N in the rest of the radiation. Specifically, the excess score ex is defined as:

$$ex = \frac{[(\bar{p}_N^D + \bar{p}_N^{DB})/2] - \bar{p}_N^R}{\text{jackknife_se}(p_N)}$$

where \bar{p}_N^D is the mean p_N within *Diplotaxodon*, \bar{p}_N^{DB} is the mean p_N within deep benthic, and \bar{p}_N^R is the mean p_N within the rest of the radiation.

Haplotype trees. To view the relationship between haplotypes for genes of interest, we translated nucleotide sequences to amino acid sequences and loaded these into Haplotype Viewer (<http://www.cibiv.at/~greg/haplerviewer>). This software requires that a tree is loaded together with the sequences. Therefore, we inferred gene trees using RAxML v7.7.8⁹⁶ with the PROTGAMMADAYHOFF model of substitution.

Local excess allele sharing between *Diplotaxodon* and deep benthic. We used an extension of the f_d statistic⁴⁸; this extension⁵⁵ is referred to as f_{dm} . f_{dm} is a conservative version of the f statistic that is particularly suited for analysis of small genomic windows^{48,55}. For the gene scores shown in Fig. 6b, we calculated f_{dm} (mbuna, deep benthic, *Diplotaxodon*, *N. brichardi*) for each gene in window from the transcription start site (TSS) to 10 kb into the gene. For the 'along the genome' plots, as shown in Fig. 6d and Supplementary Fig. 23, we used a product of two f_{dm} statistics (f_{dm} (shallow benthic; deep benthic, *Diplotaxodon*, *N. brichardi*) $\times f_{dm}$ (*Rhamphochromis*, *Diplotaxodon*; deep benthic, *N. brichardi*)), an approach which we found to increase the local resolution. This score was calculated in sliding windows of 100 SNPs across a region of ± 100 kb around the genes. Finally,

we also calculated f_{dm} (mbuna, deep benthic; *Diplotaxodon*, *N. brichardi*) separately for synonymous and non-synonymous mutations in each gene.

Reporting Summary. Further information on experimental design is available in the Nature Research Reporting Summary linked to this article.

Code availability. The majority of the custom code used in this project is available on Github as a part of the evo package (<https://github.com/millanek/evo>). All other custom codes are available from the authors upon request.

Data availability

All raw sequencing reads have been deposited to the NCBI Short Read Archive: (BioProjects PRJEB1254 and PRJEB15289). Sample accessions are listed in Supplementary Table 4. In addition, we are making whole-genome variant calls in the Variant Call Format (VCF), phylogenetic trees and protein coding sequence alignments, and tables with f4 statistics available through the Dryad Digital Repository (<https://doi.org/10.5061/dryad.7rj8k6c>).

Received: 4 December 2017; Accepted: 10 October 2018;

Published online: 19 November 2018

References

1. Losos, J. B. & Ricklefs, R. E. Adaptation and diversification on islands. *Nature* **457**, 830–836 (2009).
2. Wagner, C. E., Harmon, L. J. & Seehausen, O. Ecological opportunity and sexual selection together predict adaptive radiation. *Nature* **487**, 366–369 (2012).
3. Berner, D. & Salzburger, W. The genomics of organismal diversification illuminated by adaptive radiations. *Trends Genet.* **31**, 491–499 (2015).
4. Darwin, C. *On the Origin of Species* (Oxford Univ. Press, Oxford, 2008).
5. Lamichhaney, S. et al. Evolution of Darwin's finches and their beaks revealed by genome sequencing. *Nature* **518**, 371–375 (2015).
6. Losos, J., Jackman, T., Larson, A., Queiroz, K. & Rodriguez-Schettino, L. Contingency and determinism in replicated adaptive radiations of island lizards. *Science* **279**, 2115–2118 (1998).
7. Fryer, G. & Iles, T. D. *The Cichlid Fishes of the Great Lakes of Africa: their Biology and Evolution* (Oliver and Boyd, Edinburgh, 1972).
8. Salzburger, W., Van Boeckelaer, B. & Cohen, A. S. Ecology and evolution of the African Great Lakes and their faunas. *Annu. Rev. Ecol. Evol. Syst.* **45**, 519–545 (2014).
9. Gennar, M. J. et al. How does the taxonomic status of allopatric populations influence species richness within African cichlid fish assemblages? *J. Biogeogr.* **31**, 93–102 (2004).
10. Meyer, A. Phylogenetic relationships and evolutionary processes in East African cichlid fishes. *Trends. Ecol. Evol.* **8**, 279–284 (1993).
11. Brawand, D. et al. The genomic substrate for adaptive radiation in African cichlid fish. *Nature* **513**, 375–381 (2014).
12. Meyer, B. S., Matschiner, M. & Salzburger, W. Disentangling incomplete lineage sorting and introgression to refine species-tree estimates for Lake Tanganyika cichlid fishes. *Syst. Biol.* **66**, 531–550 (2016).
13. Meier, J. I. et al. Ancient hybridization fuels rapid cichlid fish adaptive radiations. *Nat. Commun.* **8**, 14363 (2017).
14. Koblmüller, S., Egger, B., Sturmbauer, C. & Sevc, K. M. Rapid radiation, ancient incomplete lineage sorting and ancient hybridization in the endemic Lake Tanganyika cichlid tribe Tropheini. *Mol. Phylogenet. Evol.* **55**, 318–334 (2010).
15. Weiss, J. D., Cotterill, F. P. D. & Schliwesen, U. K. Lake Tanganyika—a ‘melting pot’ of ancient and young cichlid lineages (Teleostei: Cichlidae)? *PLoS ONE* **10**, e0125043 (2015).
16. Gante, H. F. et al. Genomics of speciation and introgression in Princess cichlid fishes from Lake Tanganyika. *Mol. Ecol.* **25**, 6143–6161 (2016).
17. Wagner, C. E. et al. Genome-wide RAD sequence data provide unprecedented resolution of species boundaries and relationships in the Lake Victoria cichlid adaptive radiation. *Mol. Ecol.* **22**, 787–798 (2012).
18. Gennar, M. J. & Turner, G. F. Ancient hybridization and phenotypic novelty within Lake Malawi's cichlid fish radiation. *Mol. Biol. Evol.* **29**, 195–206 (2012).
19. Moran, P., Kornfield, I. & Reinhart, P. N. Molecular systematics and radiation of the haplochromine cichlids (Teleostei: Perciformes) of Lake Malawi. *Copeia* **1994**, 274 (1994).
20. Joyce, D. A. et al. Repeated colonization and hybridization in Lake Malawi cichlids. *Curr. Biol.* **21**, R108–R109 (2011).
21. Lefler, E. M. et al. Revisiting an old riddle: what determines genetic diversity levels within species? *PLoS Biol.* **10**, e1001388 (2012).
22. Albertson, R. C., Markert, J. A., Danley, P. D. & Kocher, T. D. Phylogeny of a rapidly evolving clade: the cichlid fishes of Lake Malawi, East Africa. *Proc. Natl. Acad. Sci. USA* **96**, 5107–5110 (1999).

23. Kocher, T. D. Adaptive evolution and explosive speciation: the cichlid fish model. *Nat. Rev. Genet.* **5**, 288–298 (2004).
24. Chimpanzee Sequencing and Analysis Consortium Initial sequence of the chimpanzee genome and comparison with the human genome. *Nature* **437**, 69–87 (2005).
25. Ravi, V. & Venkatesh, B. Rapidly evolving fish genomes and teleost diversity. *Curr. Opin. Genet. Dev.* **18**, 544–550 (2008).
26. Recknagel, H., Elmer, K. R. & Meyer, A. A hybrid genetic linkage map of two ecologically and morphologically divergent Midas cichlid fishes (*Amphilophus* spp.) obtained by massively parallel DNA sequencing (ddRADSeq). *Genes Genomes Genet.* **3**, 65–74 (2013).
27. Ségurel, L., Wyman, M. J. & Przeworski, M. Determinants of mutation rate variation in the human germline. *Annu. Rev. Genom. Hum. Genet.* **15**, 47–70 (2014).
28. Feng, C. et al. Moderate nucleotide diversity in the Atlantic herring is associated with a low mutation rate. *eLife* **6**, e23907 (2017).
29. Konings, A. *Malawi Cichlids in Their Natural Habitat* (Cichlid Press, El Paso, 2007).
30. 1000 Genomes Project Consortium An integrated map of genetic variation from 1,092 human genomes. *Nature* **491**, 56–65 (2012).
31. Green, R. E. et al. A draft sequence of the Neandertal genome. *Science* **328**, 710–722 (2010).
32. Durand, E. Y., Patterson, N., Reich, D. & Slatkin, M. Testing for ancient admixture between closely related populations. *Mol. Biol. Evol.* **28**, 2239–2252 (2011).
33. Heled, J. & Bouckaert, R. R. Looking for trees in the forest: summary tree from posterior samples. *BMC Evol. Biol.* **13**, 221 (2013).
34. Edwards, S. V. Is a new and general theory of molecular systematics emerging? *Evolution* **63**, 1–19 (2009).
35. Edwards, S. V. et al. Implementing and testing the multispecies coalescent model: a valuable paradigm for phylogenomics. *Mol. Phylogenet. Evol.* **94**, 447–462 (2016).
36. Bryant, D., Bouckaert, R., Felsenstein, J., Rosenberg, N. A. & RoyChoudhury, A. Inferring species trees directly from biallelic genetic markers: bypassing gene trees in a full coalescent analysis. *Mol. Biol. Evol.* **29**, 1917–1932 (2012).
37. Chifman, J. & Kubatko, L. Quartet inference from SNP data under the coalescent model. *Bioinformatics* **30**, 3317–3324 (2014).
38. Chifman, J. & Kubatko, L. Identifiability of the unrooted species tree topology under the coalescent model with time-reversible substitution processes, site-specific rate variation, and invariable sites. *J. Theor. Biol.* **374**, 35–47 (2015).
39. Long, C. & Kubatko, L. The effect of gene flow on coalescent-based species-tree inference. *Syst. Biol.* **67**, 770–785 (2018).
40. Mirarab, S. et al. ASTRAL: genome-scale coalescent-based species tree estimation. *Bioinformatics* **30**, i541–i548 (2014).
41. Zhang, C., Rabiee, M., Sayyari, E. & Mirarab, S. ASTRAL-III: polynomial time species tree reconstruction from partially resolved gene trees. *BMC Bioinformatics* **19**, 153 (2018).
42. Dasarathy, G., Nowak, R. & Roch, S. Data requirement for phylogenetic inference from multiple loci: a new distance method. *IEEE/ACM Trans. Comput. Biol. Bioinformatics* **12**, 422–432 (2015).
43. Rusinko, J. & McPartlon, M. Species tree estimation using neighbor joining. *J. Theor. Biol.* **414**, 5–7 (2017).
44. Ballard, J. W. O. & Whitlock, M. C. The incomplete natural history of mitochondria. *Mol. Ecol.* **13**, 729–744 (2004).
45. Toews, D. P. L. & Brelsford, A. The biogeography of mitochondrial and nuclear discordance in animals. *Mol. Ecol.* **21**, 3907–3930 (2012).
46. Consuegra, S., John, E., Verspoor, E. & de Leaniz, C. G. Patterns of natural selection acting on the mitochondrial genome of a locally adapted fish species. *Genet. Sel. Evol.* **47**, 58 (2015).
47. Lawson, D. J., Hellenthal, G., Myers, S. & Falush, D. Inference of population structure using dense haplotype data. *PLoS Genet.* **8**, e1002453 (2012).
48. Martin, S. H., Davey, J. W. & Jiggins, C. D. Evaluating the use of ABBA-BABA statistics to locate introgressed loci. *Mol. Biol. Evol.* **32**, 244–257 (2015).
49. Martin, S. H. et al. Genome-wide evidence for speciation with gene flow in *Heliconius* butterflies. *Genome Res.* **23**, 1817–1828 (2013).
50. Eccles, D. H. & Trewavas, E. *Malawian Cichlid Fishes* (Lake Fish Movies, Herten, 1989).
51. Eriksson, A. & Manica, A. Effect of ancient population structure on the degree of polymorphism shared between modern human populations and ancient hominins. *Proc. Natl Acad. Sci. USA* **109**, 13956–13960 (2012).
52. Pickrell, J. K. & Pritchard, J. K. Inference of population splits and mixtures from genome-wide allele frequency data. *PLoS Genet.* **8**, e1002967 (2012).
53. Peterson, E. N., Cline, M. E., Moore, E. C., Roberts, N. B. & Roberts, R. B. Genetic sex determination in *Astatotilapia calliptera*, a prototype species for the Lake Malawi cichlid radiation. *Naturwissenschaften* **104**, 41 (2017).
54. Genner, M. J., Ngatunga, B. P., Mzighani, S., Smith, A. & Turner, G. F. Geographical ancestry of Lake Malawi's cichlid fish diversity. *Biol. Lett.* **11**, 20150232 (2015).
55. Malinsky, M. et al. Genomic islands of speciation separate cichlid ecomorphs in an East African crater lake. *Science* **350**, 1493–1498 (2015).
56. Ivory, S. J. et al. Environmental change explains cichlid adaptive radiation at Lake Malawi over the past 1.2 million years. *Proc. Natl Acad. Sci. USA* **113**, 11895–11900 (2016).
57. Lyons, R. P. et al. Continuous 1.3-million-year record of East African hydroclimate, and implications for patterns of evolution and biodiversity. *Proc. Natl Acad. Sci. USA* **112**, 15568–15573 (2015).
58. Greenwood, P. H. Towards a phyletic classification of the 'genus' *Haplochromis* (Pisces, Cichlidae) and related taxa. Part 1. *Bull. Brit. Mus. Nat. Hist.* **35**, 265–322 (1979).
59. Lippitsch, E. A phyletic study on lacustrine haplochromine fishes (Perciformes, Cichlidae) of East Africa, based on scale and squamation characters. *J. Fish. Biol.* **42**, 903–946 (1993).
60. Alexa, A., Rahnenführer, J. & Lengauer, T. Improved scoring of functional groups from gene expression data by decorrelating GO graph structure. *Bioinformatics* **22**, 1600–1607 (2006).
61. Van Bochlaer, B., Schultheiß, R., Plisnier, P.-D. & Albrecht, C. Does the decline of gastropods in deep water herald ecosystem change in Lakes Malawi and Tanganyika? *Freshwat. Biol.* **57**, 1733–1744 (2012).
62. Spady, T. C. et al. Evolution of the cichlid visual palette through ontogenetic subfunctionalization of the opsin gene arrays. *Mol. Biol. Evol.* **23**, 1538–1547 (2006).
63. Weadick, C. J. & Chang, B. S. W. Complex patterns of divergence among green-sensitive (RH2a) African cichlid opsins revealed by clade model analyses. *BMC Evol. Biol.* **12**, 206 (2012).
64. Sugawara, T. et al. Parallelism of amino acid changes at the RH1 affecting spectral sensitivity among deep-water cichlids from lakes Tanganyika and Malawi. *Proc. Natl Acad. Sci. USA* **102**, 5448–5453 (2005).
65. Bowmaker, J. K. & Hunt, D. M. Evolution of vertebrate visual pigments. *Curr. Biol.* **16**, R484–R489 (2006).
66. Davies, W. I. L., Collin, S. P. & Hunt, D. M. Molecular ecology and adaptation of visual photopigments in craniates. *Mol. Ecol.* **21**, 3121–3158 (2012).
67. Carleton, K. L., Dalton, B. E., Escobar-Camacho, D. & Nandamuri, S. P. Proximate and ultimate causes of variable visual sensitivities: insights from cichlid fish radiations. *Genesis* **54**, 299–325 (2016).
68. Ogawa, Y., Shiraki, T., Kojima, D. & Fukada, Y. Homeobox transcription factor Six7 governs expression of green opsin genes in zebrafish. *Proc. Biol. Sci.* **282**, 20150659 (2015).
69. Renninger, S. L., Gesemann, M. & Neuhauss, S. C. F. Cone arrestin confers cone vision of high temporal resolution in zebrafish larvae. *Eur. J. Neurosci.* **33**, 658–667 (2011).
70. Brockerhoff, S. E. et al. Light stimulates a transducin-independent increase of cytoplasmic Ca^{2+} and suppression of current in cones from the zebrafish mutant *nof*. *J. Neurosci.* **23**, 470–480 (2003).
71. Boesel-Battaglia, K. & Goldberg, A. F. X. Photoreceptor renewal: a role for peripherin/rds. *Int. Rev. Cytol.* **217**, 183–225 (2002).
72. Opazo, J. C., Butts, G. T., Nery, M. F., Storz, J. F. & Hoffmann, F. G. Whole-genome duplication and the functional diversification of teleost fish hemoglobins. *Mol. Biol. Evol.* **30**, 140–153 (2013).
73. Hahn, C., Genner, M. J., Turner, G. F. & Joyce, D. A. The genomic basis of cichlid fish adaptation within the deepwater 'twilight zone' of Lake Malawi. *Evol. Lett.* **1**, 184–198 (2017).
74. Heliconius Genome Consortium Butterfly genome reveals promiscuous exchange of mimicry adaptations among species. *Nature* **487**, 94–98 (2012).
75. Meyer, A., Kocher, T. D., Basasibwaki, P. & Wilson, A. C. Monophyletic origin of Lake Victoria cichlid fishes suggested by mitochondrial DNA sequences. *Nature* **347**, 550–553 (1990).
76. Reid, N. M. et al. The genomic landscape of rapid repeated evolutionary adaptation to toxic pollution in wild fish. *Science* **354**, 1305–1308 (2016).
77. Jones, F. C. et al. The genomic basis of adaptive evolution in threespine sticklebacks. *Nature* **484**, 55–61 (2012).
78. Seehausen, O. Cichlid fish diversity threatened by eutrophication that curbs sexual selection. *Science* **277**, 1808–1811 (1997).
79. Konings, A. *Tanganyika Cichlids in their Natural Habitat* (Cichlid Press, El Paso, 2015).
80. Coyne, J. A. & Orr, H. A. *Speciation* (Sinauer Associates, Sunderland, 2004).
81. Feder, J. L., Egan, S. P. & Nosil, P. The genomics of speciation-with-gene-flow. *Trends Genet.* **28**, 342–350 (2012).
82. Li, H. Aligning sequence reads, clone sequences and assembly contigs with BWA-MEM. Preprint at <https://arxiv.org/abs/1303.3997v2> (2013).
83. McKenna, A. et al. The Genome Analysis Toolkit: a MapReduce framework for analyzing next-generation DNA sequencing data. *Genome Res.* **20**, 1297–1303 (2010).

84. Li, H. A statistical framework for SNP calling, mutation discovery, association mapping and population genetic parameter estimation from sequencing data. *Bioinformatics* **27**, 2987–2993 (2011).
85. Browning, S. R. & Browning, B. L. Rapid and accurate haplotype phasing and missing-data inference for whole-genome association studies by use of localized haplotype clustering. *Am. J. Hum. Genet.* **81**, 1084–1097 (2007).
86. Delaneau, O., Marchini, J. & Zagury, J.-F. A linear complexity phasing method for thousands of genomes. *Nat. Methods* **9**, 179–181 (2012).
87. Delaneau, O., Howie, B., Cox, A. J., Zagury, J.-F. & Marchini, J. Haplotype estimation using sequencing reads. *Am. J. Hum. Genet.* **93**, 687–696 (2013).
88. Pritchard, J. K. & Przeworski, M. Linkage disequilibrium in humans: models and data. *Am. J. Hum. Genet.* **69**, 1–14 (2001).
89. Danecek, P. et al. The variant call format and VCFtools. *Bioinformatics* **27**, 2156–2158 (2011).
90. Miller, W. et al. 28-way vertebrate alignment and conservation track in the UCSC Genome Browser. *Genome Res.* **17**, 1797–1808 (2007).
91. Ulm, K. A simple method to calculate the confidence interval of a standardized mortality ratio (SMR). *Am. J. Epidemiol.* **131**, 373–375 (1990).
92. Dobson, A. J., Kuulasmaa, K., Eberle, E. & Scherer, J. Confidence intervals for weighted sums of Poisson parameters. *Stat. Med.* **10**, 457–462 (1991).
93. Purcell, S. et al. PLINK: a tool set for whole-genome association and population-based linkage analyses. *Am. J. Hum. Genet.* **81**, 559–575 (2007).
94. Patterson, N., Price, A. L. & Reich, D. Population structure and eigenanalysis. *PLoS Genet.* **2**, e190 (2006).
95. Bhatia, G., Patterson, N., Sankararaman, S. & Price, A. L. Estimating and interpreting FST: the impact of rare variants. *Genome Res.* **23**, 1514–1521 (2013).
96. Stamatakis, A. RAxML-VI-HPC: maximum likelihood-based phylogenetic analyses with thousands of taxa and mixed models. *Bioinformatics* **22**, 2688–2690 (2006).
97. Bouckaert, R. et al. BEAST 2: a software platform for Bayesian evolutionary analysis. *PLoS Comput. Biol.* **10**, e1003537 (2014).
98. Stamatakis, A., Hoover, P. & Rougemand, J. A rapid bootstrap algorithm for the RAxML Web servers. *Syst. Biol.* **57**, 758–771 (2008).
99. Saitou, N. & Nei, M. The neighbor-joining method: a new method for reconstructing phylogenetic trees. *Mol. Biol. Evol.* **4**, 406–425 (1987).
100. Paradis, E., Claude, J. & Strimmer, K. APE: analyses of phylogenetics and evolution in R language. *Bioinformatics* **20**, 289–290 (2004).
101. Huerta-Cepas, J., Serra, F. & Bork, P. ETE 3: reconstruction, analysis, and visualization of phylogenomic data. *Mol. Biol. Evol.* **33**, 1635–1638 (2016).
102. Swofford, D. L. *PAUP*: Phylogenetic Analysis using Parsimony (and Other Methods)* (Sinauer Associates, Sunderland, MA, 2002).
103. Reaz, R., Bayzid, M. S. & Rahman, M. S. Accurate phylogenetic tree reconstruction from quartets: a heuristic approach. *PLoS ONE* **9**, e104008 (2014).
104. Robinson, D. F. & Foulds, L. R. Comparison of phylogenetic trees. *Math. Biosci.* **53**, 131–147 (1981).
105. Schliep, K. P. phangorn: phylogenetic analysis in R. *Bioinformatics* **27**, 592–593 (2011).
106. Theis, A., Ronco, F., Indermauer, A., Salzburger, W. & Egger, B. Adaptive divergence between lake and stream populations of an East African cichlid fish. *Mol. Ecol.* **23**, 5304–5322 (2014).
107. Rohlf, F. J. *tpsDig*, digitize landmarks and outlines. Version 2.0 (Department of Ecology and Evolution, State Univ. New York, 2004); <https://www.swmath.org/software/18407>
108. Adams, D. C. & Castillo, E. O. geomorph: an R package for the collection and analysis of geometric morphometric shape data. *Methods Ecol. Evol.* **4**, 393–399 (2013).
109. Quinlan, A. R. & Hall, I. M. BEDTools: a flexible suite of utilities for comparing genomic features. *Bioinformatics* **26**, 841–842 (2010).
110. Ashburner, M. et al. Gene ontology: tool for the unification of biology. The Gene Ontology Consortium. *Nat. Genet.* **25**, 25–29 (2000).
111. Alexa, A. & Rahnenfuhrer, J. topGO: enrichment analysis for gene ontology. R package version 2.28.0 (R Foundation for Statistical Computing, 2010).
112. Huber, W. et al. Orchestrating high-throughput genomic analysis with Bioconductor. *Nat. Methods* **12**, 115–121 (2015).
113. Merico, D., Isserlin, R., Stueker, O., Emili, A. & Bader, G. D. Enrichment map: a network-based method for gene-set enrichment visualization and interpretation. *PLoS ONE* **5**, e13984 (2010).

Acknowledgements

This work was supported by the Wellcome Trust (grant number 097677/Z/11/Z to M.M., grant numbers WT206194 and WT207492 to R.D. and H.S.), the Royal Society-Leverhulme Trust Africa Awards (grant numbers AA100023 and AA130107 to M.J.G. and G.F.T.) and the European Molecular Biology Organization (grant number ALTF 456-2016 to M.M.). We thank the Sanger Institute sequencing core for DNA sequencing, M. Du for DNA extractions, D. Swofford and M. Matschiner for advice on phylogenomic analyses, and W. Salzburger and I. Wilson for comments on the manuscript. We also thank the Tanzania Fisheries Research Institute, and the Fisheries Research Unit of the Government of Malawi for their assistance and support.

Author contributions

E.A.M., G.F.T., M.J.G., M.M. and R.D. devised the study. G.F.T. and M.J.G. collected the samples. A.M.T. bred parent-offspring trios and performed geometric morphometric analyses. M.M. performed the DNA extractions. H.S. and M.M. analysed the genomic data. All authors participated in the interpretation of the results. M.M., H.S. and R.D. drafted the manuscript, and all others commented.

Competing interests

R.D. declares that he owns stock in Illumina from previous consulting. The authors declare no other competing interests.

Additional information

Supplementary information is available for this paper at <https://doi.org/10.1038/s41559-018-0717-x>.

Reprints and permissions information is available at www.nature.com/reprints.

Correspondence and requests for materials should be addressed to M.M. or R.D.

Publisher's note: Springer Nature remains neutral with regard to jurisdictional claims in published maps and institutional affiliations.



Open Access This article is licensed under a Creative Commons Attribution 4.0 International License, which permits use, sharing, adaptation, distribution and reproduction in any medium or format, as long as you give appropriate credit to the original author(s) and the source, provide a link to the Creative Commons license, and indicate if changes were made. The images or other third party material in this article are included in the article's Creative Commons license, unless indicated otherwise in a credit line to the material. If material is not included in the article's Creative Commons license and your intended use is not permitted by statutory regulation or exceeds the permitted use, you will need to obtain permission directly from the copyright holder. To view a copy of this license, visit <http://creativecommons.org/licenses/by/4.0/>.

© The Author(s) 2018

Reporting Summary

Nature Research wishes to improve the reproducibility of the work that we publish. This form provides structure for consistency and transparency in reporting. For further information on Nature Research policies, see [Authors & Referees](#) and the [Editorial Policy Checklist](#).

Statistical parameters

When statistical analyses are reported, confirm that the following items are present in the relevant location (e.g. figure legend, table legend, main text, or Methods section).

n/a Confirmed

- The exact sample size (*n*) for each experimental group/condition, given as a discrete number and unit of measurement
- An indication of whether measurements were taken from distinct samples or whether the same sample was measured repeatedly
- The statistical test(s) used AND whether they are one- or two-sided
Only common tests should be described solely by name; describe more complex techniques in the Methods section.
- A description of all covariates tested
- A description of any assumptions or corrections, such as tests of normality and adjustment for multiple comparisons
- A full description of the statistics including central tendency (e.g. means) or other basic estimates (e.g. regression coefficient) AND variation (e.g. standard deviation) or associated estimates of uncertainty (e.g. confidence intervals)
- For null hypothesis testing, the test statistic (e.g. *F*, *t*, *r*) with confidence intervals, effect sizes, degrees of freedom and *P* value noted
Give P values as exact values whenever suitable.
- For Bayesian analysis, information on the choice of priors and Markov chain Monte Carlo settings
- For hierarchical and complex designs, identification of the appropriate level for tests and full reporting of outcomes
- Estimates of effect sizes (e.g. Cohen's *d*, Pearson's *r*), indicating how they were calculated
- Clearly defined error bars
State explicitly what error bars represent (e.g. SD, SE, CI)

Our web collection on [statistics for biologists](#) may be useful.

Software and code

Policy information about [availability of computer code](#)

Data collection

tpsDig2 v2.26

Data analysis

bwa-mem v.0.7.10, samtools v.1.2, bcftools v.1.2, picard-tools v.1.124, GATK v3.3.0 , vcftools v0.1.12b, BEAGLE v.4.0 (r1398), shapeit v2 (r790), SNPable tool (<http://lh3lh3.users.sourceforge.net/snnable.shtml>), lastz v1.0, Kent UCSC Tools (<git://genome-source.cse.ucsc.edu/kent.git>; v293_base-36-g9e7d0af), PLINK v1.0.7, eigensoft v5.0.2, RAxML v7.7.8, TreeAnnotator v.2.4.2, BEAST 2.4.2, SNAPP 1.3.0, ETE3 toolkit, PAUP* v4.0a (build 159), ASTRAL v.5.6.1, chromopainter v0.0.4, fineSTRUCTURE v0.0.5, R (and pin particular packages: GeoMorph v3.0.2, topGO v2.26.0, org.Dr.eg.db v3.4.0), bedtools v2.26.0, Haplotype Viewer (<http://www.cibiv.at/~greg/haploviewer>), evo (<https://github.com/millanek/evo>), modules dstat and tensorstats of pypopgen (<https://github.com/feilchenfeldt/pypopgen>)

For manuscripts utilizing custom algorithms or software that are central to the research but not yet described in published literature, software must be made available to editors/reviewers upon request. We strongly encourage code deposition in a community repository (e.g. GitHub). See the Nature Research [guidelines for submitting code & software](#) for further information.

Data

Policy information about [availability of data](#)

All manuscripts must include a [data availability statement](#). This statement should provide the following information, where applicable:

- Accession codes, unique identifiers, or web links for publicly available datasets
- A list of figures that have associated raw data
- A description of any restrictions on data availability

All raw sequencing reads have been deposited to the NCBI Short Read Archive: (BioProjects PRJEB1254 and PRJEB15289). Sample accessions are listed in Supplementary Table 4. In addition, we are making whole-genome variant calls in the Variant Call Format (VCF), phylogenetic trees and protein coding sequence alignments, and tables with f4 statistics available through the Dryad Digital Repository (see the doi in the "Data availability" section of the manuscript)

Field-specific reporting

Please select the best fit for your research. If you are not sure, read the appropriate sections before making your selection.

Life sciences Behavioural & social sciences Ecological, evolutionary & environmental sciences

For a reference copy of the document with all sections, see nature.com/authors/policies/ReportingSummary-flat.pdf

Ecological, evolutionary & environmental sciences study design

All studies must disclose on these points even when the disclosure is negative.

Study description	The goal of our study was to provide primary whole-genome based analysis of the evolutionary diversity of Lake Malawi cichlid fishes.
Research sample	Our sample selection provides broad coverage of all the major lineages of the Lake Malawi cichlid radiation: rapidly radiating cichlid tribe Haplochromini.
Sampling strategy	We focused on ecomorphological diversity, representing more than half the genera from each major group, rather than obtaining deep coverage of any particular group.
Data collection	DNA sequencing was performed by the Wellcome Sanger Institute sequencing core facility
Timing and spatial scale	In this study, we sequenced DNA from ethanol preserved finclips that were collected by M.J.Genner and G.F.Turner between 2004 and 2014 from Tanzania and Malawi, in collaboration with the Tanzania Fisheries Research Institute (the MolEcoFish Project) and with the Fisheries Research Unit of the Government of Malawi (various collaborative projects). All of the DNA was extracted and sequenced at the Wellcome Sanger Institute between 2012 and 2015.
Data exclusions	The analyses used only single nucleotide polymorphism (SNP) data. Larger insertions, deletions, and structural variation were excluded from this study.
Reproducibility	All our attempts at replication of analyses were successful, although this was not done in a systematic manner. To assist independent researchers in reproducing and verifying our findings, we provide the SNPs ready to use in a Variant Call Format (VCF) file.
Randomization	A number of the reported analyses rely on comparisons between major groups of species, which were determined based on published literature, and correspond to the clustering of the whole genome data. Random assignment was not appropriate.
Blinding	During the genomic data analysis, the investigators were aware of the species identity of the genomic sequences analysed (no blinding).

Did the study involve field work? Yes No

Reporting for specific materials, systems and methods

Materials & experimental systems

n/a	Involved in the study
<input checked="" type="checkbox"/>	Unique biological materials
<input checked="" type="checkbox"/>	Antibodies
<input checked="" type="checkbox"/>	Eukaryotic cell lines
<input checked="" type="checkbox"/>	Palaeontology
<input type="checkbox"/>	<input checked="" type="checkbox"/> Animals and other organisms
<input checked="" type="checkbox"/>	Human research participants

Methods

n/a	Involved in the study
<input checked="" type="checkbox"/>	ChIP-seq
<input checked="" type="checkbox"/>	Flow cytometry
<input checked="" type="checkbox"/>	MRI-based neuroimaging

Animals and other organisms

Policy information about [studies involving animals; ARRIVE guidelines](#) recommended for reporting animal research

Laboratory animals

Trios were obtained by mating wild-caught fish obtained from Lake Malawi by commercial aquarium fish exporters (Stuart M. Grant Ltd) and exported to UK ornamental fish importers. Adults were quarantined at Bangor University's research aquarium, maintained at ca 25C in filtered aquaria with a mix of salts added to mimic Lake Malawi water conditions, and fed on standard aquarium fish flake food. Multiple females (5-10) were kept with a single adult male, until a female was seen to be carrying eggs (indicated by her bulging throat and loss of interest in food). Mouthbrooding females were removed to brooding tanks containing shelter for mother in the form of a PVC tube and shelter for the offspring in the form of rocks and artificial plants. When free-swimming fry were observed in the tank (usually after about 3 weeks), the mother was removed and, along with the father, euthanised by anaesthetic (MS-222) overdose. Offspring were reared for 3-6 months, before euthanasia.

Wild animals

The study did not involve wild animals.

Field-collected samples

Finclips were preserved in ethanol for DNA extraction.