coexnet: A R package to build CO-EXpression NETworks from Microarray Data

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Contents

1   Abstract

The analysis of biological data through networks has been increasing in the last years due to the capacity of this approach to analyze the high number of information in a simple way and the fact that there is a variety of biological networks to create and analyzed depending on the Data obtained by the researcher (protein-protein interaction, signaling interaction networks, gene regulatory network, among others). Currently, one of the most useful kind of biological network is the co-expression network. Mainly, this kind of network is created from expression measures and considering the existence of particular relations of expression among different genes, proteins or RNA fragments involved in the develpment or arising of a specific phenotype. Here, we present *coexnet*, a new R package to the construction of undirected co-expression networks from microarray data obtained from GEO Datasets database. This package have all the necessary functions to download, normalize and filter expression data to finally create the co-expression network unsing correlation measures as statistic approach.

2   Workflow

2.1   get.info

All microarray raw data associated with the same study is stored in the *CEL* file, this file containe the *GSM* files, each one corresponding to one sample inside the study, you can obtain each *GSM* file in the individual way, but, is preferable to obtain all the samples in the *CEL* file to avoid the work of join each *GSM* and additionally the way to analyze this kind of data is simultainously (in future normalization process for example). Furthermore, all the microarray chips are documented in the *GEO Datasets* database. Each one of them are identified with the letters *GPL* adding a unique number. The information in the *GPL* file is related to the information of each probeset in the microarray chip, including the gene, function, type and other information to take advantage of the analysis of the results.

This function will create in your current pathfile a folder with the *GSE* (unique number) name where the GSM download files are stored and on the other hand the *GPL* (unique number) *.soft* file with the information of the microarray chip.

```
# Downloading the microarray raw data from GSE8216 study
# The accession number of the microarray chip related with this study is GPL2025

get.info(GSE = "GSE8216", GPL = "GPL2025",dir = ".")

# Show the actual pathfile with the folder with the GSE number and the .soft file

dir()
```

Take account

In some cases the information in the *GPL* file are partial, take this in mind in the future analysis and is recomended not store the files in a temporal folder due to in many cases you will need the raw data to re-process the expression values using different methods.

2.2   get.affy

The *AffyBatch object* is one of the most kind of data widely used to process and analyse microarray expression data. The AffyBatch object store information about the date of scan to each one of the samples, the information related with the phenotype, the raw expression values to each probe in the microarray chip, the kind of library to read the expression data among others.

You can use the *AffyBatch object* in many different packages mainly in the affy package, additionally you can modify the *AffyBatch object* if you consider it necessary.

This function search in your current or designaded pathfile the folder with the *GSE* accession number and read the *filelist.txt* file with the name of the every *GSM* samples to recognize them and join in an only *AffyBatch object*.

```
# Reading some GSM samples from GSE4773 study, the folder with the
# GSM files are called GSE1234.

affy <- get.affy(GSE = "GSE1234",dir = system.file("extdata",package = "coexnet"))
```

Take account

In some cases the *AffyBatch object* doesn't have all the information and a warning message is shown when you view the variable with the AffyBatch object, but you can edit the AffyBatch to fill all the requiered information. If you try to process the AffyBatch in some of the packages that use this kind of object you will recieved an error message.

```
# The variable affy doesn't have the CDF (Chip Definition File) information.
# You can include this information modifying the AffyBatch object.

affy@cdfName <- "HG-U133_Plus_2"
```

2.3   gene.symbol

In most cases, the idea of create a co-expression network is to visualize the relations among diferent genes, proteins, specific DNA or RNA fragments or another kind of molecular entity identify with a specific ID. For this reason, is very useful to have the information about the corresponding ID to each one of the probeset in

the microarray. This kind of information will use when you need pass from a matrix of probeset-samples to one of gene(or another ID)-samples before of the construction of the co-expression network.

The *.soft file*, downloaded from *GEO Datasets* database using the *GPL* identifier have the information to create a table with the relationship between probeset and one "molecular ID", in this table one ID can be related with two or more probeset, the process to create only one expression values to one ID from different probeset is called *summarization* (see bellow).

This function search in the current or the designaded pathfile the *.soft* file and from this searchand will create a data.frame where the first column have each one of the probeset names and in the second one have the corresponding ID (gene symbol, protein name or another identifier).

```
# Create the table with the relationship between probesets and IDs.

gene_table <- gene.symbol(GPL = "GPL2025",d = system.file("extdata",package = "coexnet"))

head(gene_table)
```

Take account

In some cases, the *.soft* file dosen't have all the IDs to each one of the probeset inside of the microarray, you can ignore this probeset under the assumption that another probeset could have the same ID and this second one have the respective anotation. On the other hand, one ID can have more than two names, this function create a ID with all the related names separed by "-", that is useful in future analysis when the biological information is related at one specific name among the several names for one ID.

```
# The before table have NA and empty information in the IDs.
# We can delete this unuseful information.

# Deletion of IDs with NA information

gene_na <- na.omit(gene_table)

# Deletion of empty IDs

final_table <- gene_na[gene_na$ID != "",]

head(final_table)
```

2.4 exprs.mat

The raw expression data in a microarray experiment mut be processed to transform the original data in the correct way to be analyzed and obtain confidence results. The first step is the normalizaion of the data, that consist in a background correction of the raw data and the follow transformation of the result in a normalize way. The second one is the pass of probeset to gene or any other ID to represent the molecular entity to analyze. Additionally, exists the posibility to make a second kind of background correction based on the batch of samples scaned in a separete way due to the large number of samples and the limitation in the size of the microarray chip used, this correction is knowledge as *Batch Effect Correction*.

Exist different methods to normalize raw expression data from microrray experiments, each one of this methodologies consider a way to generate a background correction, the process of normalization and the pass from probes to probesets. The differencies of this methods consist in the underliying mathematics assumptions used and the range of the noralized results, in some cases the expression data have a range more wide than anothers. In the same way, the process to transform the probeset-samples matrix in a gene(or another ID)-sample matrix consider diferent methodologies to do that including the obtantion of the average

of the expression values of each one of the probeset corresponding at same gene or protein, the selection of the maximum or minimum value, among others.

This function have the possibility to choose among two different methods to normalize the raw expression values, including the process of background correction and the pass from probes to probeset. The first one is *rma* (Robust Multi-Array Average), this method do a background correction and normalization in a separate calculations (Irizarry, *et al.* 2003). The second one is *vsn* (Variance Stabilizing Normalization), this method, contrary to *rma*, generate the background correction and the normalization in the same equation (Huber, *et al.* 2002). On the other hand, the function have the option to do a *Batch Effect Correction* whether you know if the samples were scaned in a separate way using the *Scan Date* information inside the *AffyBatch object* (one of the input in the function).

Additionally, this method considers two ways to calculate the values in the process to pass from probesets to gene / ID. The first one is selecting the representative probeset to each one of the gene, protein or another kind of ID. To do that, it is calculated the average of each one of the probeset associated with the same gene / ID, and the probeset with the most high value in the average is selected. The second one is to obtain the median of each one of the samples to the probesets associated with the same gene / ID, getting only one expression value for sample as the transformation of the normalized data.

```
# Loading AffyBatch object

data("affy")

# Loading table with probeset and gene/ID information

data("info")

# Calculating the expression matrix witj rma

rma <- expr.mat(affy = affy,genes = info,NormalizeMethod = "rma",
SummaryMethod = "median",BatchCorrect = FALSE)
head(rma)
```

Take account

Consider that *rma* is a method whose results have less range of amplitude than *vsn*, take account this to select the method to normalize the raw expresion values. In some cases. The method *vsn* takes account every probes in the normalize process, that could take time. In some cases is a good idea make the normalization using or not the *Batch Effect Correction.*

2.5   cof.var

In some cases, the co-expression network is built from two or more microarrays studies, in this sense, is necessary to define wich of this studies represent the most source of background noise and probably affect in a negative way the future results. One way to determinate the most harmful studies is from variation analysis, the study with more variation among the normalized expression values can be the source of the future background noise and is necessary to consider the use of this studies in the construction of the co-expression network.

To define the variation among the normalized expression values can be determined the coefficient of variation of each one of the genes in each one of the studies and generate the boxplot from the results. So, in a graphical way it is possible to define the studies that will generate noise background watching the atypical information. On the other hand, it is possible to define the number of atypical data and determine the more variant studies using the boxplot and the number of atipic data defining a threshold value, for example the studies with more of 10% of atypical data wont be take account In the construction of the co-expression network.

This function take the normalized ID-sample matrix and claculate the median and the coefficient of variation to each one of the ID, this process is neccesary to doing study-by-study. Additionally, this function allow to calculate the mean and the coefficient of variation to cases and controls samples separately using a vector of 0s and 1s to identify the cases and control samples. This vector is possible be defined in the description of each sample in the GEO Datasets database and is necessary in the process of identify the genes (or another ID as proteins) differentially expressed (see bellow).

```r
# Simulated expression data

n <- 200
m <- 20

# The vector with treatment samples and control samples

t <- c(rep(0,10),rep(1,10))

# Calculating the expression values normalized

mat <- as.matrix(rexp(n, rate = 1))
norm <- t(apply(mat, 1, function(nm) rnorm(m, mean=nm, sd=1)))

# Calculating the coefficient of variation to case samples

case <- cof.var(data = norm,complete = FALSE,treatment = t,type = "case")
head(case)

# Creating the boxplot to coefficient of variation results

boxplot(case$cv)

# Extracting the number of atipic data

length(boxplot.stats(case$cv)$out)
```

Take account

The desition of delete a microarray study from the result of coefficient of variation result depend of the data and the criteria of the researcher to filter the studies (the selection of a threshold value), don't exist a gold rule to discard a study, is advisable calculate the coefficient of variation of all samples at time to compare and determinate the most variant.

```r
# Calculating the coefficient of variation to whole matrix

complete <- cof.var(norm)
head(complete)

# Creating the boxplot to coefficient of variation results

boxplot(complete$cv)

# Extracting the number of atipic data

length(boxplot.stats(complete$cv)$out)
```

2.6 dif.exprs

When expression data of gene, proteins, or another kind of ID are used to build a co-expression network, in the mostly cases is interested carry on this process to genes/IDs that present interest behavior, one of them is about the differences in the expression value with respect to the rest of gene/ID data. To know wich of them have this kind of behavior, a differential expression analysis must be do, this process allow identify the genes/IDs over-expressed or under-expressed and these kind of disturbance in the expression are the factor to establish the molecular components associated directly or indirectly with the develop of a certain phenotype and finally use this genes/IDs to create the co-expression network.

To do that, is necessary to know wich of the samples are the control and wich of them are the cases (treatment), to compare them in a mathematical way and consider which of the genes/IDs with the most high variation to be consider as differentially expressed compared with the differences in case/control in the remain genes/IDs. Exist several methodologies to identify the genes/IDs differentially expressed, some of them are more predictive than another one, it means, depending of the method, is possible obtain genes/IDs crearly differentialized in their expression values respecting another one or obtain genes/IDs whose expression value are lightly differentiated from the another one, but by the criteria of the method are considered differentialy expressed. In both cases is posible to have genes/IDs selected by error and is necessary to consider a measure of this error. One of the most used measures or error is the *False Discover Rate* or *FDR*, this describe the probability of one gene/ID has been selected as differentially expressed by error and depending of the method used the *FDR* value is calculated.

This function considers two ways to calculate the differentially expressed genes / IDs. In this case, it is possible to carry out a predictive methodology to try to obtain the most possible amount of genes, or IDs considered differentially expressed, in this case the * sam * method is used. This method basically uses a difference of means to calculate the genes / IDs over-expressed or under-expressed using the permutation process to test the results and prove that the genes / IDs selected where not randomly selected, throught of aforementioned permutations the * FDR * value is calculated (Tusher, * et al * 2001). On the other hand, this function uses the method * acde * to calculate and obtain the genes / IDs differentially expressed in a less predictive way. This method consists essentially of the application of the main components to characterize the genes differentially expressed by calculating the FDR using multiple hypothesis tests according to Benjamini and Hochberg (1995) (Acosta & López-Kleine).

```r
# Creating a matrix with 200 genes and 20 samples

n <- 200
m <- 20

# The vector with treatment samples and control samples

t <- c(rep(0,10),rep(1,10))

# Calculating the expression values normalized

mat <- as.matrix(rexp(n, rate = 1))
norm <- t(apply(mat, 1, function(nm) rnorm(m, mean=nm, sd=1)))

# Running the function using the two approaches

sam <- dif.exprs(eset = norm,treatment = t,fdr = 0.2,DifferentialMethod = "sam")
head(sam)
```

Take account

This function identify the genes/IDs differentially expressed taking account the expected *FDR*, it means, independientely of the method used (*sam* or *acde*), the number of genes/IDs identify as differentially expressed

will be guided by the *FDR* expected by the user increasing the predictive power in the final results.

2.7 find.threshold

When you have the final expression matrix, is necessary analyze it in a way to obtain the co-expression network. Exist two methods widely used to do this process, both of them are related with the definition of correlation value between all the genes/IDs creating a square matrix. On one hand you can calculate the *Pearson Correlation Coefficient*, this method calculate the correlation between every genes/IDs from the expression values of each one, as result, the square matrix will have values between zero and one, due to in the future construction of co-expression network is necessary to use the absolute value of the results. On the other hand, the *Mutual Information* is based on entropy of the data and in a same way in the aforementioned method a square matrix is created, but, in this case, is necessary an additional transformation of the results to obtain a square matrix with values between zero and one.

The obtaintion of a square matrix with a range of values from zero to one is necessary to the future transformation of this correlation matrix to the adjacency matrix (see bellow). Additionally, is necessary this range of values because a threshold value must be defined to establish the final relations between the genes/IDs, to do this, a value among zero to one is defined and the values of correlation less than threshold value will indicate the no-existance of a real correlation among them, getting finally the relations among the genes/IDs as co-expression network.

This function computes a threshold value using a novel method based on two * Biological Systems * approaches. First, each possible threshold value, from 0.01 to 0.99 with an increase of 0.01 is examined. Each one of this possible threshold values is then analyzed in the first approche aforementioned mentioned. To each value, it is calculated the * Clustering Coefficient * to the network created using the current threshold value under test. After, it is calculated an artificial * Clustering Coefficient * to simulate a random network created using the same threshold value. Then, it is calculated the difference between the two * Clustering Coefficient * values, and the result that meets the criteria of Elo, * et al * (2008), will be take account to the next analysis. Finally, the remaining threshold values will be analyzed using the * Degree Distribution * under normal distribution using * Kolmogorov-Smirnov * test, expecting the resulting p-value rejecting the distribution, as a result of the assumption that the biological networks don't Have Normal distribution when the * Degree Distribution * is analyzed. Finally, the minimum threshold value that satisfies this two criteria will be selected as the final threshold value for the construction of the co-expression network.

```
# Loading data

pathfile <- system.file("extdata","expression_example.txt",package = "coexnet")
data <- read.table(pathfile,stringsAsFactors = FALSE)

# Find threshold value

cor_pearson <- find.threshold(difexp = data,method = "correlation")
cor_pearson
```

Take account

Mutual Information is used, in most cases when you need to analyze a lot of amount of expression data, for example, when the study was designed to use a lot of samples and you must be process all information simultaneously. In most cases, the threshold value is selected by the researcher without any biological or biology systems assumption. Here, we present this novel methodology to select a threshold value under biology netwrok assumptions.

2.8 create.net

The last step in the construction of a co-expression network is the creation of a kind of structure that will be read and store the information ncessary to create a network. Once you had defined a threshold value, the last step is transform the expression matrix in a adjacency matrix using the correlation or the mutual information method to obtain the values of relationship between the diferent genes or proteins (or another kind of ID).

The process to pass from expression matrix to network consist of two steps. The first one is the build of a correlation matrix, to do that is necessary to apply some of the two method to calculate the relation among the genes/ID (Pearson correlation or mutual information, see above) (López-Kleine, *et al.* 2013). The second one is the pass from expression matrix to adjacency matrix, to create this kind of matrix is necessary the threshold value, every correlation value inside the matrix less than the threshold will be replace by zero, while all remainded values will be replaced by one. Aditionally, the product of related one gene/ID with itself will be always zero.

Finally, from the adjacency matrix, is created a list where one gene/ID and another one conected to the first are related separated by a space. It means, if *gene A* and the *gene B*, in the adjacency matrix have a value of one, then, in the final edge list they appear as:

gene A – gene B

and so on to the every genes/IDs connected in the final co-expression network.

This function takes the expression matrix and create the correlation matrix using *Pearson Correlation Coefficient* or *Mutual Informtion*. After that, creates the adjacency matrix taking account the threshold value given by the user and finally create the network from the adjacency matrix as an igraph object to be analyze using the igraph R package or another one that recognize this kind of object.

```
# Loading data

pathfile <- system.file("extdata","expression_example.txt",package = "coexnet")
data <- read.table(pathfile,stringsAsFactors = FALSE)

# Building the network

cor_pearson <- create.net(difexp = data,threshold = 0.7,method = "correlation")
head(cor_pearson)
```

Take account

In the process of construction of adjacency matrix, some gene/ID could not be related with another one and in the process to pass from matrix to edge list, this gene/ID will be deleted. Additionally, the network in the igraph object can be export as edge list using the igraph package to viualize the network in another program as *Cytoscape* or *Gephi.*

3   References

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