

# qbs121\_hw7\_gibran

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## Data Analyses

### 2.1 PBC (50 points)

Use the dataset pbc that comes with the library survival in R. The time and status indicators compose the endpoint.

```
library(survival)
head(pbc)
```

```
##   id time status trt      age sex ascites hepato spiders edema bili chol
## 1  1  400      2   1 58.76523  f      1      1      1  1.0 14.5 261
## 2  2 4500      0   1 56.44627  f      0      1      1  0.0  1.1 302
## 3  3 1012      2   1 70.07255  m      0      0      0  0.5  1.4 176
## 4  4 1925      2   1 54.74059  f      0      1      1  0.5  1.8 244
## 5  5 1504      1   2 38.10541  f      0      1      1  0.0  3.4 279
## 6  6 2503      2   2 66.25873  f      0      1      0  0.0  0.8 248
##   albumin copper alk.phos   ast trig platelet protime stage
## 1    2.60    156  1718.0 137.95  172     190    12.2     4
## 2    4.14     54  7394.8 113.52   88     221    10.6     3
## 3    3.48    210   516.0  96.10   55     151    12.0     4
## 4    2.54     64  6121.8  60.63   92     183    10.3     4
## 5    3.53    143   671.0 113.15   72     136    10.9     3
## 6    3.98     50   944.0  93.00   63      NA    11.0     3
```

1. Competing Risks: The variable status has 3 levels, 0 for censoring, 1 for transplant, 2 for death.

(a) (2pts) If someone had a transplant and died sometime later what would this data record?

The status should show 2 for someone who had transplant and died when the study is running.

(b) Download the library, cmprsk

```
library(cmprsk)
```

```
## Warning: package 'cmprsk' was built under R version 4.1.1
```

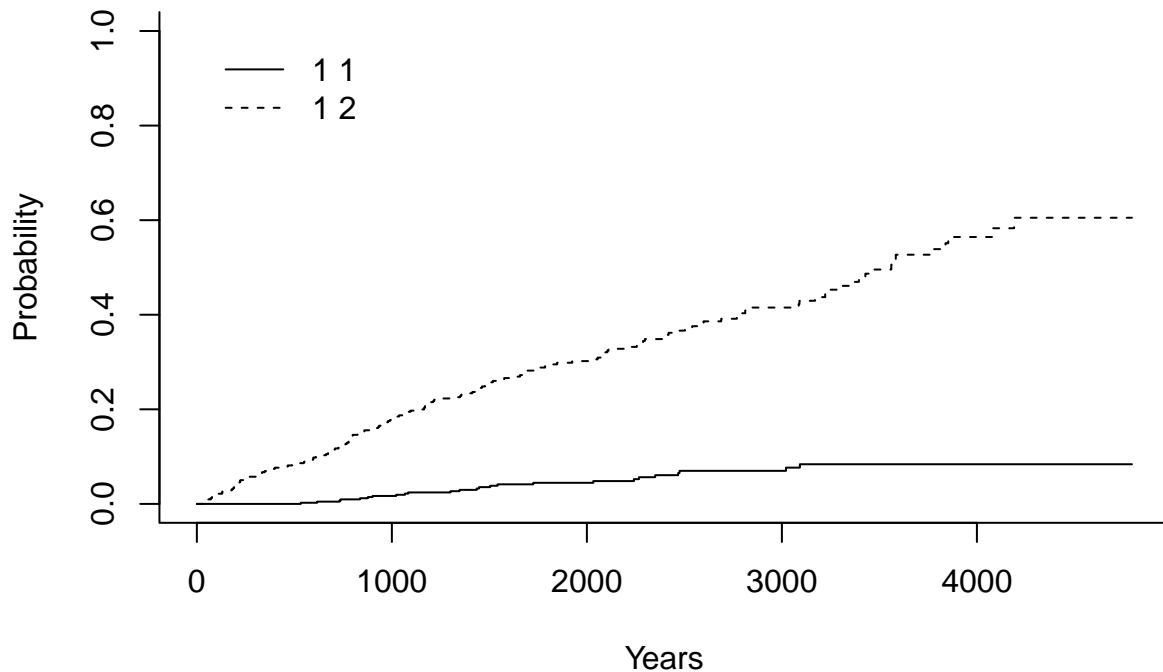
(c) (3) Run the command to get “cumulative incidence” o=with(pbc, cuminc(time,status))

```
o <- with(pbc, cuminc(time, status))
o
```

```
## Estimates and Variances:
## $est
##           1000           2000           3000           4000
## 1 1 0.01681293 0.04457393 0.0698984 0.08382977
## 1 2 0.18239707 0.30198210 0.4152135 0.56435900
##
## $var
##           1000           2000           3000           4000
## 1 1 3.981527e-05 0.0001134119 0.0002123444 0.0003017893
## 1 2 3.590633e-04 0.0005594984 0.0008438443 0.0015692409
```

(d) (5) Plot the output, `plot(o)`, and interpret the resulting curves

```
plot(o)
```



2. (2) What proportion of patients are censored?

```
library(tidyverse)
```

```
## -- Attaching packages ----- tidyverse 1.3.1 --
```

```
## v ggplot2 3.3.5          v purrr 0.3.4
## v tibble 3.1.5          v dplyr 1.0.7.9000
## v tidyr 1.1.4           v stringr 1.4.0
## v readr 2.0.2           v forcats 0.5.1

## Warning: package 'ggplot2' was built under R version 4.1.1

## Warning: package 'tibble' was built under R version 4.1.1

## Warning: package 'tidyr' was built under R version 4.1.1

## Warning: package 'readr' was built under R version 4.1.1

## Warning: package 'stringr' was built under R version 4.1.1

## Warning: package 'forcats' was built under R version 4.1.1

## -- Conflicts ----- tidyverse_conflicts() --
## x dplyr::filter() masks stats::filter()
## x dplyr::lag() masks stats::lag()
```

```

pbc %>%
  group_by(pbc$status == 0) %>%
  summarise(n=n()) %>%
  mutate(freq = n/sum(n))

```

```
## # A tibble: 2 x 3
##   'pbc$status == 0'      n freq
##   <lgl>              <int> <dbl>
## 1 FALSE              186 0.445
## 2 TRUE               232 0.555
```

3. (3) What is the total follow-up time in this study?

```
sum(pbc$time)
```

```
## [1] 801633
```

4. (5) Calculate the incidence rate of transplant and death (e.g. number of events divided by total follow-up time)

```
sum(pbc$status == 1)/sum(pbc$time)
```

```
## [1] 3.118634e-05
```

5. Evaluate if trt arm impacts the composite endpoint, transplant or death by

(a) (2) first coding the composite endpoint, tr.death=status> 0

```
tr.death <- pbc$status > 0
```

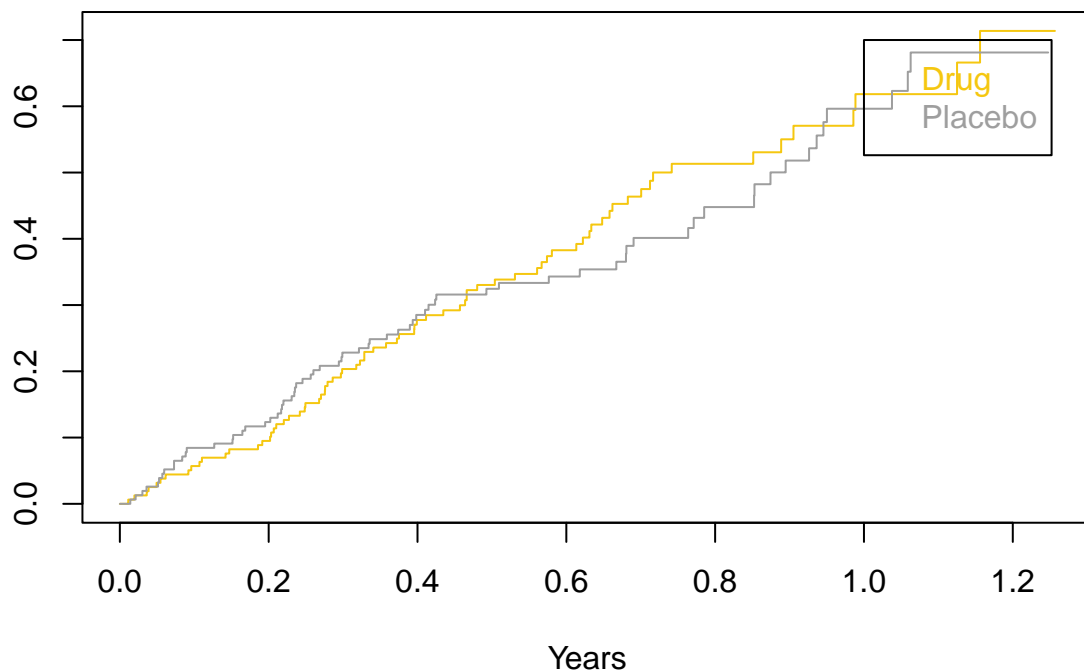
(b) (3) reporting the number of events and rate in each arm

```
tapply(tr.death, pbc$trt, sum)
```

```
## 1 2  
## 75 69
```

(c) (5) plotting survival curves and reporting median time to transplant or death in each treatment group

```
plot(survfit(Surv(time/3625.5, tr.death) ~ trt, data = pbc),  
     fun = "event",  
     col=7:8,  
     xlab="Years")  
  
legend(1,0.7,  
       text.col=7:8,  
       legend=c("Drug", "Placebo"))
```



(d) (5) conducting a log-rank test

```
survdif(Surv(time, tr.death) ~ trt, data = pbc)
```

```
## Call:
## survdiff(formula = Surv(time, tr.death) ~ trt, data = pbc)
##
## n=312, 106 observations deleted due to missingness.
##
##           N Observed Expected (O-E)^2/E (O-E)^2/V
## trt=1 158      75      72.9   0.0595   0.121
## trt=2 154      69      71.1   0.0611   0.121
##
## Chisq= 0.1  on 1 degrees of freedom, p= 0.7
```

(e) (5) conducting a Peto-Peto-Prentice test (generalization of the Wilcoxon rank test)

```
survdif(Surv(time, tr.death) ~ trt, rho=1, data=pbc)
```

```
## Call:
## survdiff(formula = Surv(time, tr.death) ~ trt, data = pbc, rho = 1)
##
## n=312, 106 observations deleted due to missingness.
##
##           N Observed Expected (O-E)^2/E (O-E)^2/V
## trt=1 158      55.6      54.6   0.0183   0.0476
## trt=2 154      51.8      52.8   0.0190   0.0476
##
## Chisq= 0  on 1 degrees of freedom, p= 0.8
```

(f) (5) using a Weibull proportional hazards model model to estimate the hazard ratio with a 95%CI

```
summary(survreg(Surv(time, tr.death) ~ trt, dist="weibull", data=pbc))
```

```
##
## Call:
## survreg(formula = Surv(time, tr.death) ~ trt, data = pbc, dist = "weibull")
##           Value Std. Error      z      p
## (Intercept)  8.2291      0.2258 36.45 <2e-16
## trt          0.0438      0.1431  0.31  0.759
## Log(scale) -0.1532      0.0729 -2.10  0.036
##
## Scale= 0.858
##
## Weibull distribution
## Loglik(model)= -1348.2  Loglik(intercept only)= -1348.3
## Chisq= 0.09 on 1 degrees of freedom, p= 0.76
## Number of Newton-Raphson Iterations: 5
## n=312 (106 observations deleted due to missingness)
```

(g) (5) using a Cox's proportional hazards model to estimate the hazard ratio with a 95%CI

```
summary(coxph(Surv(time, tr.death) ~ trt, data = pbc))
```

```
## Call:
## coxph(formula = Surv(time, tr.death) ~ trt, data = pbc)
##
##      n= 312, number of events= 144
##      (106 observations deleted due to missingness)
##
##              coef exp(coef) se(coef)      z Pr(>|z|)
## trt -0.0581      0.9436   0.1669 -0.348   0.728
##
##      exp(coef) exp(-coef) lower .95 upper .95
## trt    0.9436      1.06    0.6803    1.309
##
## Concordance= 0.502  (se = 0.023 )
## Likelihood ratio test= 0.12  on 1 df,  p=0.7
## Wald test               = 0.12  on 1 df,  p=0.7
## Score (logrank) test = 0.12  on 1 df,  p=0.7
```

## 2.2 Lung

Use the dataset lung that comes with the library survival in R. The time and status indicators compose the endpoint.

1. (2pts) What are the levels of the status variable?

```
unique(lung$status)
```

```
## [1] 2 1
```

1=censored, 2=dead

2. (2) What proportion of patients are censored?

```
perct_censored <- lung %>%
  group_by(lung$status == 1) %>%
  summarise(n=n()) %>%
  mutate(freq = n/sum(n))

as.numeric(perct_censored[2,3]*100)
```

```
## [1] 27.63158
```

3. (3) What is the total follow-up time in this study?

```
sum(lung$time)
```

```
## [1] 69593
```

4. (3) Calculate the mortality rate (e.g. number of deaths divided by total follow-up time)

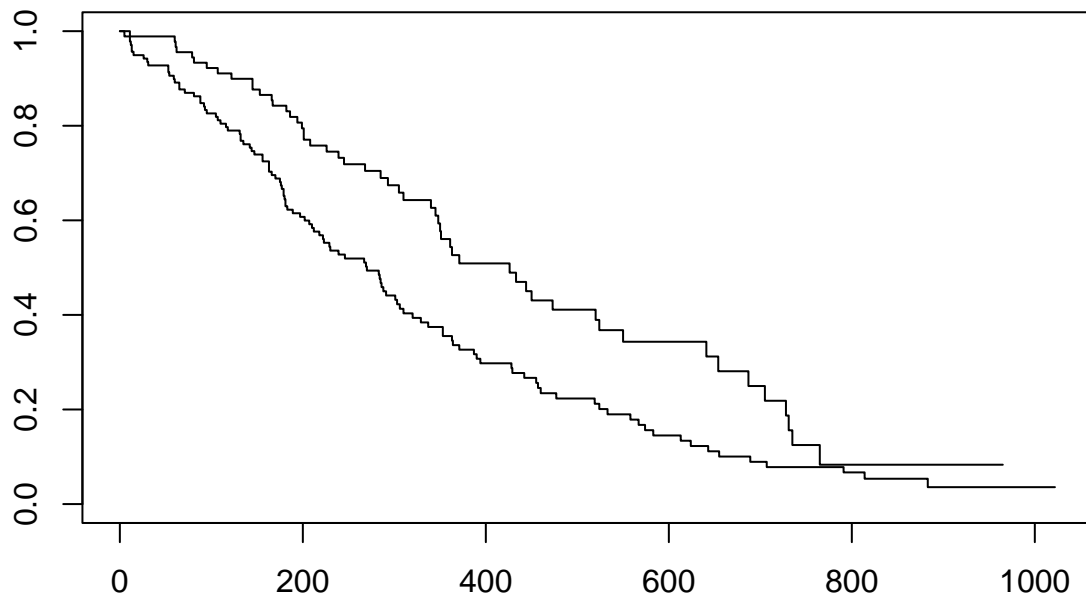
```
sum(lung$status == 2) / sum(lung$time)
```

```
## [1] 0.002370928
```

5. Examine the association of biological sex with survival as follows

(a) (3) plotting male and female survival curves

```
surv_gender <- survfit(Surv(time, status) ~ sex, data = lung)  
plot(surv_gender)
```



(b) (3) reporting median survival time for each biological sex

```
male <- lung %>%  
  filter(lung$sex == 1) #male  
  
female <- lung %>%  
  filter(lung$sex == 2) #female  
  
print(median(male$time))
```

```
## [1] 224
```

```
print(median(female$time))
```

```
## [1] 292.5
```

(c) (3) conducting a log-rank test

```
survdif(Surv(time, status) ~ sex, data = lung)
```

```
## Call:
## survdif(formula = Surv(time, status) ~ sex, data = lung)
##
##           N Observed Expected (O-E)^2/E (O-E)^2/V
## sex=1 138      112     91.6      4.55      10.3
## sex=2  90       53     73.4      5.68      10.3
##
##  Chisq= 10.3  on 1 degrees of freedom, p= 0.001
```

(d) (3) conducting a Peto-Peto-Prentice test (generalization of the Wilcoxon rank test)

```
survdif(Surv(time, status) ~ sex, rho=1, data = lung)
```

```
## Call:
## survdif(formula = Surv(time, status) ~ sex, data = lung, rho = 1)
##
##           N Observed Expected (O-E)^2/E (O-E)^2/V
## sex=1 138      70.4     55.6      3.95      12.7
## sex=2  90      28.7     43.5      5.04      12.7
##
##  Chisq= 12.7  on 1 degrees of freedom, p= 4e-04
```

(e) (5) using a Weibull proportional hazards model model to estimate the hazard ratio with a 95% CI

```
summary(survreg(Surv(time, status) ~ sex, dist="weibull", data = lung))
```

```
##
## Call:
## survreg(formula = Surv(time, status) ~ sex, data = lung, dist = "weibull")
##           Value Std. Error      z      p
## (Intercept)  5.4886      0.1790 30.66 < 2e-16
## sex          0.3956      0.1276  3.10  0.0019
## Log(scale)  -0.2809      0.0619 -4.54 5.7e-06
##
## Scale= 0.755
##
## Weibull distribution
## Loglik(model)= -1148.7  Loglik(intercept only)= -1153.9
##  Chisq= 10.4 on 1 degrees of freedom, p= 0.0013
## Number of Newton-Raphson Iterations: 5
## n= 228
```

(f) (5) using a Cox's proportional hazards model to estimate the hazard ratio with a 95% CI



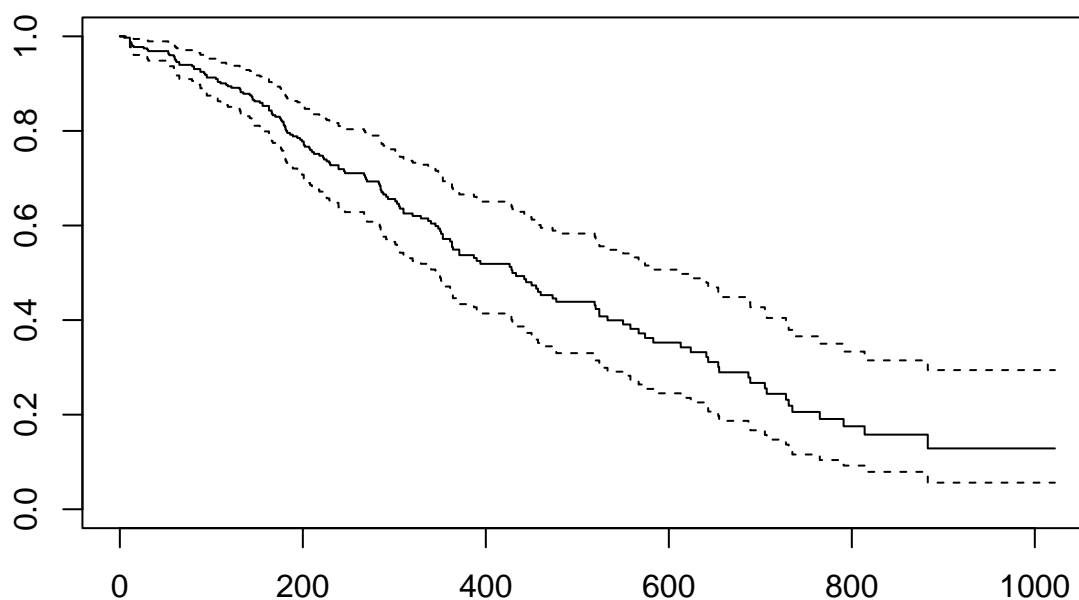
```
summary(coxph(Surv(time, status) ~ sex, data = lung))
```

```
## Call:
## coxph(formula = Surv(time, status) ~ sex, data = lung)
##
##    n= 228, number of events= 165
##
##           coef exp(coef) se(coef)      z Pr(>|z|)
## sex -0.5310    0.5880   0.1672 -3.176  0.00149 **
## ---
## Signif. codes:  0 '***' 0.001 '**' 0.01 '*' 0.05 '.' 0.1 ' ' 1
##
##      exp(coef) exp(-coef) lower .95 upper .95
## sex      0.588      1.701    0.4237    0.816
##
## Concordance= 0.579 (se = 0.021 )
## Likelihood ratio test= 10.63 on 1 df,  p=0.001
## Wald test               = 10.09 on 1 df,  p=0.001
## Score (logrank) test = 10.33 on 1 df,  p=0.001
```

6. (2) Combine categories 2 and 3 in variable ph.ecog and report survival curves by this new variable

```
lung$ph.ecog.females <- factor(ifelse(lung$ph.ecog >2, 2, lung$ph.ecog))
surv <- survfit(coxph(Surv(time, status) ~ ph.ecog.females, data = lung))

plot(surv)
```



7. (3) Calculate the hazard ratios comparing each level to patients in category 1.

```
lung$ph.ecog.females <- relevel(lung$ph.ecog.females,ref="1")
hr <- coxph(Surv(time, status) ~ ph.ecog.females,data = lung)
exp(cbind(hr$coef,confint(hr)))
```

```
##                2.5 %   97.5 %
## ph.ecog.females0 0.6919883 0.4688171 1.021396
## ph.ecog.females2 1.7554989 1.2184847 2.529188
```

8. (3) Calculate the hazard ratio for a 1 unit change in this variable (e.g. going from 1 to 2 or 2 to 3).

```
hra <- coxph(Surv(time,status) ~ ph.ecog, data=lung)
exp(cbind(hra$coef,confint(hra)))
```

```
##                2.5 %   97.5 %
## ph.ecog 1.609532 1.288833 2.01003
```

9. (10) Examine the association of the variables age, ph.karno, pat.karno, meal.cal and wt.loss with survival using univariable Cox models and comment on the significance of the association with each.

```
summary(oslog1 <- survreg(Surv(time, status) ~ age, dist="lognormal", data = lung))
```

```
##  
## Call:  
## survreg(formula = Surv(time, status) ~ age, data = lung, dist = "lognormal")  
##           Value Std. Error      z      p  
## (Intercept)  7.33027    0.53979 13.58 <2e-16  
## age         -0.02666    0.00849 -3.14 0.0017  
## Log(scale)   0.07155    0.05621  1.27 0.2030  
##  
## Scale= 1.07  
##  
## Log Normal distribution  
## Loglik(model)= -1164.4   Loglik(intercept only)= -1169.3  
##  Chisq= 9.83 on 1 degrees of freedom, p= 0.0017  
## Number of Newton-Raphson Iterations: 3  
## n= 228
```