SDS 383D: Homework 4

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Problem 1. Hierarchical models: Math tests

The data set in "mathtest.csv" shows the scores on a standardized math test from a sample of 10th-grade students at 100 different U.S. urban high schools, all having enrollment of at least 400 10th-grade students. (A lot of educational research involves "survey tests" of this sort, with tests administered to all students being the rare exception.)

Let θ_i be the underlying mean test score for school i, and let y_{ij} be the score for the jth student in school i. Starting with the "mathtest.R" script, you'll notice that the extreme school-level averages \bar{y}_i (both high and low) tend to be at schools where fewer students were sampled.

1. Explain briefly why this would be.

In Figure 1, we notice that extreme average values of the scores are obtained for school with few students sampled. This happens because the distribution of \bar{y}_i for each school has a variance of σ^2/n_i . In fact, $\bar{y}_i \stackrel{\text{iid}}{\sim} N(\mu, \sigma^2/n_i)$. Thus, the smaller n_i , the larger the variability of \bar{y}_i around the grand mean $\frac{1}{I} \sum_{i=1}^{I} \bar{y}_i$.

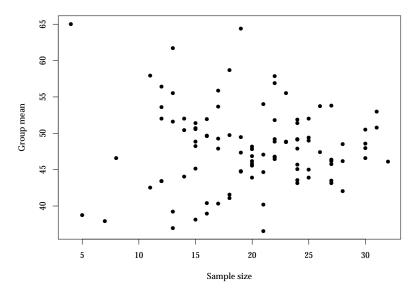


Figure 1: Extreme school-level averages \bar{y}_i *for schools with few students sampled.*

2. Fit a normal hierarchical model to these data via Gibbs sampling:

$$y_{ij} \sim N(\theta_i, \sigma^2)$$

 $\theta_i \sim N(\mu, \tau^2 \sigma^2)$

Decide upon sensible priors for the unknown model parameters (μ, σ^2, τ^2) .

Let us denote with $i=1,\ldots,I$ the schools and with n_i the number of students for school i. The total number of students is $n=\sum_{i=1}^{I}n_i$.

The model is the following:

$$y_{ij}|\theta_i, \sigma^2 \stackrel{\text{iid}}{\sim} N(\theta_i, \sigma^2)$$

along with the priors

$$\theta_{i}|\mu,\sigma^{2},\tau^{2} \sim N(\mu,\tau^{2}\sigma^{2})$$

$$\mu \sim \pi(\mu) = \mathcal{I}_{(-\infty,+\infty)}(\mu)$$

$$\sigma^{2} \sim \pi(\sigma^{2}) = \frac{1}{\sigma^{2}} \cdot \mathcal{I}_{[0,+\infty)}(\sigma^{2})$$

$$\tau^{2} \sim \pi(\tau^{2}) = \mathcal{I}_{[0,+\infty)}(\tau^{2}).$$
(1)

In other words, we used a flat prior for μ and for τ^2 . The flat prior for μ is motivated by the fact that a proper posterior can be achieved. On the other hand, the flat prior for τ^2 has been chosen according to Gelman (2006). This should work well unless the number of groups I is low (below 5, say). In our case, we dispose of 100 different schools and therefore its use is justified.

The likelihood of the model is

$$L(Y|\boldsymbol{\theta}, \sigma^2, \mu, \tau^2) = \prod_{i=1}^{I} \prod_{j=1}^{n_i} \left\{ \left(\frac{1}{2\pi\sigma^2} \right)^{\frac{1}{2}} \exp\left\{ -\frac{1}{2\sigma^2} (y_{ij} - \theta_i)^2 \right\} \right\}$$
$$\propto \left(\frac{1}{\sigma^2} \right)^{\frac{n}{2}} \exp\left\{ -\frac{1}{2\sigma^2} \sum_{i=1}^{I} \sum_{j=1}^{n_i} (y_{ij} - \theta_i)^2 \right\}.$$

Using the prior specification (1) we obtain the following full conditionals:

• the full conditional for each one of the group means θ_i are

$$\begin{split} p(\theta_{i}|y,\sigma^{2},\mu,\tau^{2}) &\propto L(Y|\theta_{i},\sigma^{2})p(\theta_{i}|\mu,\sigma^{2},\tau^{2}) \\ &\propto \exp\left\{-\frac{1}{2\sigma^{2}}\sum_{j=1}^{n_{i}}(y_{ij}-\theta_{i})^{2}\right\} \exp\left\{-\frac{1}{2\tau^{2}\sigma^{2}}(\theta_{i}-\mu)^{2}\right\} \\ &= \exp\left\{-\frac{1}{2\tau^{2}\sigma^{2}}\left[\tau^{2}\sum_{j=1}^{n_{i}}y_{ij}^{2}+n_{i}\tau^{2}\theta_{i}^{2}-2\theta_{i}\tau^{2}\sum_{j=1}^{n_{i}}y_{ij}+\theta_{i}^{2}+\mu^{2}-2\mu\theta_{i}\right]\right\} \\ &\propto \exp\left\{-\frac{n_{i}\tau^{2}+1}{2\tau^{2}\sigma^{2}}\left[\theta_{i}^{2}-2\frac{\mu+\tau^{2}n_{i}\bar{y}_{i}}{n_{i}\tau^{2}+1}\theta_{i}\right]\right\} \\ &= N\left(\frac{\mu+\tau^{2}n_{i}\bar{y}_{i}}{n_{i}\tau^{2}+1};\left(\frac{n_{i}\tau^{2}+1}{\tau^{2}\sigma^{2}}\right)^{-1}\right) \\ &= N\left(\frac{1}{n_{i}\tau^{2}+1}\cdot\mu+\frac{n_{i}\tau^{2}}{n_{i}\tau^{2}+1}\cdot\bar{y}_{i};\left(\frac{n_{i}}{\sigma^{2}}+\frac{1}{\tau^{2}\sigma^{2}}\right)^{-1}\right), \end{split}$$

where in the last line we recognize the usual formulation as posterior mean as weighted average of prior mean and data sample mean.

• the full conditional for the grand mean μ is

$$\begin{split} p(\mu|y,\pmb{\theta},\sigma^2,\tau^2) &\propto p(\pmb{\theta}|\mu,\tau^2,\sigma^2)p(\mu) \\ &\propto \exp\left\{-\frac{1}{2\sigma^2\tau^2}\sum_{i=1}^I(\theta_i-\mu)^2\right\}p(\mu) \\ &\propto \exp\left\{-\frac{1}{2\sigma^2\tau^2}\left(I\mu^2+\sum_{i=1}^I\theta_i^2-2\mu\sum_{i=1}^I\theta_i\right)\right\} \\ &= N\left(\bar{\pmb{\theta}};\frac{\sigma^2\tau^2}{I}\right). \end{split}$$

• the full conditional for the variance σ^2 is

$$\begin{split} p(\sigma^2|y,\pmb{\theta},\mu,\tau^2) &\propto p(Y|\pmb{\theta},\sigma^2,\mu,\tau^2) p(\pmb{\theta}|\mu,\tau^2,\sigma^2) p(\sigma^2) \\ &\propto \left(\frac{1}{\sigma^2}\right)^{\frac{n+I}{2}+1} \exp\left\{-\frac{1}{2\sigma^2} \sum_{i=1}^{I} \sum_{j=1}^{n_i} (y_{ij}-\theta_i)^2\right\} \exp\left\{-\frac{1}{2\sigma^2\tau^2} \sum_{i=1}^{I} (\theta_i-\mu)^2\right\} \\ &= \left(\frac{1}{\sigma^2}\right)^{\frac{n+I}{2}+1} \exp\left\{-\frac{1}{\sigma^2} \left[\frac{1}{2} \sum_{i=1}^{I} \sum_{j=1}^{n_i} (y_{ij}-\theta_i)^2 + \frac{1}{2\tau^2} \sum_{i=1}^{I} (\theta_i-\mu)^2\right]\right\} \\ &= \text{inv-Gamma}\left(\frac{n+I}{2}; \frac{1}{2} \sum_{i=1}^{I} \sum_{j=1}^{n_i} (y_{ij}-\theta_i)^2 + \frac{1}{2\tau^2} \sum_{i=1}^{I} (\theta_i-\mu)^2\right). \end{split}$$

• the full conditional for the variance τ^2 is

$$\begin{split} p(\tau^2|y,\pmb{\theta},\mu,\sigma^2) &\propto p(Y|\pmb{\theta},\sigma^2,\mu,\tau^2) p(\pmb{\theta}|\sigma^2,\mu,\tau^2) p(\tau^2) \\ &\propto \left(\frac{1}{\tau^2}\right)^{\frac{I}{2}} \exp\left\{-\frac{1}{2\sigma^2\tau^2} \sum_{i=1}^{I} (\theta_i-\mu)^2\right\} \\ &= \text{inv-Gamma}\left(\frac{I}{2}-1; \frac{\sum_{i=1}^{I} (\theta_i-\mu)^2}{2\sigma^2}\right). \end{split}$$

3. Suppose you use the posterior mean $\hat{\theta}_i$ from the above model to estimate each school-level mean θ_i . Define the shrinkage coefficient κ_i as

$$\kappa_i = \frac{\bar{y}_i - \hat{\theta}_i}{\bar{y}_i} \,,$$

which tells you how much the posterior mean shrinks the observed sample mean. Plot this shrinkage coefficient (in absolute value) for each school as a function of that school's sample size, and comment.

In Figure 2, we notice that groups with low sample size get shrunk the most, whereas groups with large sample size hardly get shrunk at all. In fact, the larger the sample size for a group, the more information we have for that group and the less information we need to borrow from the rest of the population.

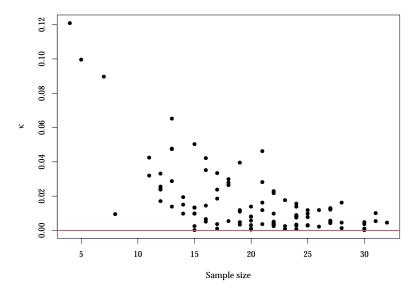


Figure 2: Shrinkage coefficient plotted versus the sample size of the groups.

Problem 2. Hierarchical models: Price elasticity of demand

The data in "cheese.csv" are about sales volume, price, and advertisting display activity for packages of Borden sliced "cheese." The data are taken from Rossi, Allenby, and McCulloch's textbook on Bayesian Statistics and Marketing. For each of 88 stores (store) in different US cities, we have repeated observations of the weekly sales volume (vol, in terms of packages sold), unit price (price), and whether the product was advertised with an in-store display during that week (disp = 1 for display).

Your goal is to estimate, on a store-by-store basis, the effect of display ads on the demand curve for cheese. A standard form of a demand curve in economics is of the form $Q = \alpha P^{\beta}$, where Q is quantity demanded (i.e. sales volume), P is price, and α and β are parameters to be estimated. You'll notice that this is linear on a log-log scale,

$$\log P = \log \alpha + \beta \log Q$$

which you should feel free to assume here. Economists would refer to β as the price elasticity of demand (PED). Notice that on a log-log scale, the errors enter multiplicatively.

There are several things for you to consider in analyzing this data set.

- 1. The demand curve might shift (different α) and also change shape (different β) depending on whether there is a display ad or not in the store.
- 2. Different stores will have very different typical volumes, and your model should account for this.
- 3. Do different stores have different PEDs? If so, do you really want to estimate a separate, unrelated β for each store?
- 4. If there is an effect on the demand curve due to showing a display ad, does this effect differ store by store, or does it look relatively stable across stores?

5. Once you build the best model you can using the log-log specification, do see you any evidence of major model mis-fit?

April 24, 2017

Propose an appropriate hierarchical model that allows you to address these issues, and use Gibbs sampling to fit your model.

First of all, we analyze this problem with a hierarchical model using the package lme4.

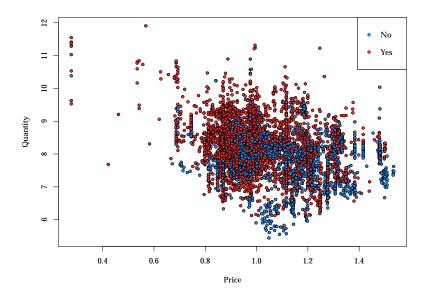


Figure 3: Scatterplot of the data on a log-log scale. The colors correspond to the presence/absence of advertisement.

By a simple descriptive analysis, in Figure 3, we see that there seems to be an effect due to the advertising (red points shifted up). However, we need to be careful because there might be confounding. In particular, red points seem to be shifted on the left, that is, display happens when the cheese is cheaper. Price is a confounder for this model, as it is correlated with both the predictor (display) and the response (quantity).

The necessity of using a hierarchical model is evident in Figure 4. In fact, in some of the groups there are no observations (or only one observation) corresponding to the presence/absence of advertising. Not pooling among the groups can lead to distorted estimates.

Using the command

```
hlm = lmer(logvol \sim (1 + logprice + disp + logprice:disp | store))
```

we fit a hierarchical model to the cheese data, after having log-transformed both price and volume. Remark that the matrix of the covariates is four dimensional: apart from the intercept, we are trying to measure the effect of the log-price, of the advertisement and of their interaction on the log-volume of sales. Thus, the demand curve is allowed to shift and to change shape depending on whether there is a display ad or not. Moreover, the estimates can vary store by store, around a common grand mean of the population. This allows the stores that have different typical volumes to be modelled differently, albeit maintaining a common structure of the population of the stores.

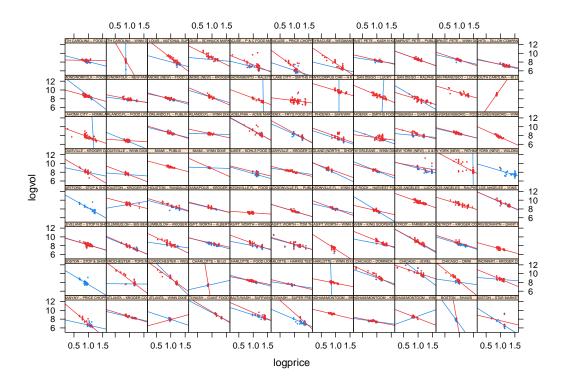


Figure 4: Observation plotted according to the different stores. The colors correspond to the presence/absence (red/blue) of advertisement. The lines represent the OLS estimates with no pooling.

The results are not shown because they are similar to the ones obtained via Gibbs sampling, which is detailed in the following.

We could also fit a fully Bayesian hierarchical model. In fact, let us assume

$$Y_{i}|\beta, \gamma_{i}, \lambda \sim N_{n_{i}} \left(X_{i}\beta + Z_{i}\gamma_{i}, \lambda^{-1}\mathcal{I}_{n_{i}}\right)$$

$$\gamma_{i}|D \stackrel{\text{iid}}{\sim} N_{q} \left(\mathbf{0}, D\right)$$

$$D \sim \mathcal{IW} \left(\nu, \Psi\right)$$

$$\beta \sim N_{p}(\mathbf{0}, \lambda_{0}^{-1}\mathcal{I}_{p})$$

$$\lambda \sim \frac{1}{\lambda}\mathcal{I}_{[0, +\infty)}(\lambda).$$

In this case, it is easy to find the full conditional distributions. In fact,

• the full conditional for β is

$$p(\boldsymbol{\beta}|Y, \boldsymbol{\gamma}_1, \dots, \boldsymbol{\gamma}_I, \lambda) \propto L(Y|\boldsymbol{\beta}, \boldsymbol{\gamma}_1, \dots, \boldsymbol{\gamma}_I, \lambda)p(\boldsymbol{\beta})$$

$$\propto \prod_{i=1}^{I} \left\{ \exp\left\{ -\frac{\lambda}{2} (\boldsymbol{y}_i - X_i \boldsymbol{\beta} - Z_i \boldsymbol{\gamma}_i)^T (\boldsymbol{y}_i - X_i \boldsymbol{\beta} - Z_i \boldsymbol{\gamma}_i) \right\} \right\} \exp\left\{ -\frac{\lambda_0}{2} \boldsymbol{\beta}^T \boldsymbol{\beta} \right\}.$$

By defining the shifted quantity $w_i = y_i - Z_i \gamma_i$ we fall in the normal linear model case, that

is,

$$p(\boldsymbol{\beta}|Y, \boldsymbol{\gamma}_1, \dots, \boldsymbol{\gamma}_I, \lambda) \propto \exp\left\{-\frac{\lambda}{2} \sum_{i=1}^{I} (\boldsymbol{w}_i - X_i \boldsymbol{\beta})^T (\boldsymbol{w}_i - X_i \boldsymbol{\beta})\right\} \exp\left\{-\frac{\lambda_0}{2} \boldsymbol{\beta}^T \boldsymbol{\beta}\right\}$$
$$\sim N\left(\left(\lambda_0 \mathcal{I} + \lambda \sum_{i=1}^{I} X_i^T X_i\right)^{-1} \sum_{i=1}^{I} X_i^T \boldsymbol{w}_i, \left(\lambda_0 \mathcal{I} + \lambda \sum_{i=1}^{I} X_i^T X_i\right)^{-1}\right).$$

• the full conditionals for the random effects $\gamma_1, \dots, \gamma_I$ are, $\forall i \in \{1, \dots, I\}$,

$$p(\boldsymbol{\gamma}_{i}|\boldsymbol{y}_{i},\boldsymbol{\beta},\lambda) \propto p(\boldsymbol{Y}_{i}|\boldsymbol{\gamma}_{i},\boldsymbol{\beta},\lambda)p(\boldsymbol{\gamma}_{i}|D)$$

$$\propto \exp\left\{-\frac{\lambda}{2}(\boldsymbol{y}_{i}-X_{i}\boldsymbol{\beta}-Z_{i}\boldsymbol{\gamma}_{i})^{T}(\boldsymbol{y}_{i}-X_{i}\boldsymbol{\beta}-Z_{i}\boldsymbol{\gamma}_{i})\right\} \exp\left\{-\frac{1}{2}\boldsymbol{\gamma}_{i}^{T}D^{-1}\boldsymbol{\gamma}_{i}\right\}$$

$$= \exp\left\{-\frac{\lambda}{2}(\boldsymbol{m}_{i}-Z_{i}\boldsymbol{\gamma}_{i})^{T}(\boldsymbol{m}_{i}-Z_{i}\boldsymbol{\gamma}_{i})\right\} \exp\left\{-\frac{1}{2}\boldsymbol{\gamma}_{i}^{T}D^{-1}\boldsymbol{\gamma}_{i}\right\}$$

$$\sim N\left((\lambda Z_{i}^{T}Z_{i}+D^{-1})^{-1}\lambda Z_{i}^{T}\boldsymbol{m}_{i},(\lambda Z_{i}^{T}Z_{i}+D^{-1})^{-1}\right)$$

where $\forall i \in \{1, \dots, I\}, \boldsymbol{m}_i = \boldsymbol{y}_i - X_i \boldsymbol{\beta}$.

• the full conditional for the precision parameter λ is

$$p(\lambda|Y, \boldsymbol{\beta}, \boldsymbol{\gamma}_1, \dots, \boldsymbol{\gamma}_I) \propto p(Y|\boldsymbol{\gamma}_1, \dots, \boldsymbol{\gamma}_I, \boldsymbol{\beta}, \lambda)p(\lambda)$$

$$\sim \operatorname{Gamma}\left(\frac{n}{2}, \frac{1}{2} \sum_{i=1}^{I} ||\boldsymbol{y}_i - X_i \boldsymbol{\beta} - Z_i \boldsymbol{\gamma}_i||_2^2\right).$$

• the full conditional for the covariance matrix of the random effects is

$$p(D|Y, \gamma_1, \dots, \gamma_I) \propto p(\gamma_1, \dots, \gamma_I|D)p(D)$$
$$\sim \mathcal{IW}\left(\nu + I, \Psi + \sum_{i=1}^{I} \gamma_i \gamma_i^T\right).$$

With the model specification just described, we get results that are similar to the ones of the lme4 package. In particular, in Figure 6 one can see that the partial pooling allows the estimation of the lines also for stores that have no observation with or without advertisement. In Figure 5, instead, the regression lines that are estimated separately for each of the the same stores are shown. In the top left panel, we see a store with a balanced sample size with and without advertising. In this case, the shrinkage effect is less evident. The other three panels show pathological cases in which only a few observations (or none) are present for one of the two groups (usually the non-display group).

Problem 3. Hierarchical models: a hierarchical probit model via data augmentation

Read the following paper:

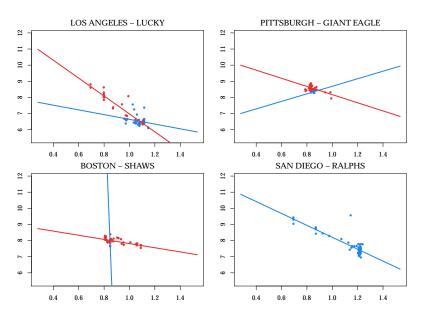


Figure 5: Regression lines fitted for four different stores separately.

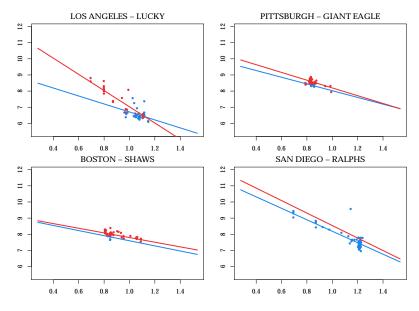


Figure 6: Regression lines fitted for four different stores in the bayesian model (that accounts for partial pooling).

"Bayesian Analysis of Binary and Polychotomous Response Data." James H. Albert and Siddhartha Chib. Journal of the American Statistical Association, Vol. 88, No. 422 (Jun., 1993), pp. 669-679

The paper describes a Bayesian treatment of probit regression (similar to logistic regression) using the trick of data augmentation—that is, introducing "latent variables" that turn a hard problem into a much easier one. Briefly summarize your understanding of the key trick proposed by this paper. Then see you if you can apply the trick in the following context, which is more complex than ordinary probit regression.

In "polls.csv" you will find the results of several political polls from the 1988 U.S. presidential election.

The outcome of interest is whether someone plans to vote for George Bush (senior, not junior). There are several potentially relevant demographic predictors here, including the respondent's state of residence. The goal is to understand how these relate to the probability that someone will support Bush in the election. You can imagine this information would help a great deal in poll re-weighting and aggregation (ala Nate Silver).

Use Gibbs sampling, together with the Albert and Chib trick, to fit a hierarchical probit model of the following form:

$$Pr(y_{ij} = 1) = \Phi(z_{ij})$$

$$z_{ij} = \mu_i + x_{ij}^T \beta_i.$$

Here y_{ij} is the response (Bush=1, other=0) for respondent j in state i; $\Phi(\cdot)$ is the probit link function, i.e. the CDF of the standard normal distribution; μ_i is a state-level intercept term; x_{ij} is a vector of respondent-level demographic predictors; and β_i is a vector of regression coefficients for state i.

Notes:

- 1. There are severe imbalances among the states in terms of numbers of survey respondents. Following the last problem, the key is to impose a hierarchical prior on the state-level parameters.
- 2. The data-augmentation trick from the Albert and Chib paper above is explained in many standard references on Bayesian analysis. If you want to get a quick introduction to the idea, you can consult one of these. A good presentation is in Section 8.1.1 of "Bayesian analysis for the social sciences" by Simon Jackman, available as an ebook through lib.utexas.edu.
- 3. You are welcome to use the logit model instead of the probit model. If you do this, you'll need to read the following paper, rather than Albert and Chib: Polson, N.G., Scott, J.G. and Windle, J. (2013). Bayesian inference for logistic models using Polya-Gamma latent variables. J. Amer. Statist. Assoc. 108 1339–1349.

The augmenting strategy for the probit Gibbs sampler is widely known in literature. The model is fairly easy. In fact, the original model is

$$Y_{ij}|oldsymbol{eta}_i \stackrel{ ext{iid}}{\sim} \operatorname{Be}(p_{ij}) \ p_{ij} = \Phi(oldsymbol{x}_{ij}^Toldsymbol{eta}_i)$$

where $i=1,\ldots,I$ is the index corresponding to the state (in this example) and $j=1,\ldots,n_i$ an index corresponding to the poll respondent. It is straightforward to prove that this model is equivalent to the augmented model

$$Z_{ij}|\boldsymbol{\gamma}_i \stackrel{\text{ind}}{\sim} N(\boldsymbol{\gamma}_{ij}^T \boldsymbol{\beta}_i, 1)$$

$$Y_{ij} = 1 \text{ if } Z_{ij} \ge 0, \quad Y_{ij} = 0 \text{ if } Z_{ij} < 0.$$

In fact,

$$P(Y_{ij} = 1) = p_{ij} = P(Z_{ij} \ge 0) = 1 - \Phi(-\boldsymbol{x}_{ij}^T \gamma_i) = \Phi(\boldsymbol{x}_{ij}^T \gamma_i)$$

which is exactly the model we started from.

Therefore it is evident that the strategy to follow is the implementation of a linear model (or hierarchical linear model) for the latent Z_i 's, adding on top a further level of hierarchy for the Y_i 's. The full conditionals turn to be straightforward.

Let us write the model with the prior specification in the case of the hierarchical linear model. Let X_i and W_i be the covariates of the fixed and random effects, respectively, for the i^{th} state. Then,

$$egin{align} Y_{ij}|Z_{ij} &= 1 \cdot \mathbb{1}(Z_{ij} \geq 0) + 0 \cdot \mathbb{1}(Z_{ij} < 0) \ Z_i|oldsymbol{eta}, oldsymbol{\gamma}_i \sim N_{n_i}(X_ioldsymbol{eta} + W_ioldsymbol{\gamma}_i, \mathcal{I}_{n_i}) \ oldsymbol{eta} \propto 1 \ oldsymbol{\gamma}_i|\Sigma \stackrel{ ext{iid}}{\sim} N_q(oldsymbol{0}, \Sigma) \ \Sigma \sim \mathcal{IW}(
u, \Psi). \end{split}$$

We obtain:

• the full conditional for γ_i , $\forall i = 1, ..., I$ are

$$p(\boldsymbol{\gamma}_{i}|\boldsymbol{z}_{i},\boldsymbol{\beta},\boldsymbol{\Sigma}) \propto p(\boldsymbol{Z}_{i}|\boldsymbol{\beta},\boldsymbol{\gamma}_{i}) \cdot p(\boldsymbol{\gamma}_{i}|\boldsymbol{\Sigma})$$

$$\propto \exp\left\{-\frac{1}{2}(\boldsymbol{z}_{i} - X_{i}\boldsymbol{\beta} - W_{i}\boldsymbol{\gamma}_{i})^{T}(\boldsymbol{z}_{i} - X_{i}\boldsymbol{\beta} - W_{i}\boldsymbol{\gamma}_{i})\right\} \exp\left\{-\frac{1}{2}\boldsymbol{\gamma}_{i}^{T}\boldsymbol{\Sigma}^{-1}\boldsymbol{\gamma}_{i}\right\}$$

$$\sim N_{q}\left((W_{i}^{T}W_{i} + \boldsymbol{\Sigma}^{-1})^{-1}W_{i}^{T}\boldsymbol{m}_{i}, (W_{i}^{T}W_{i} + \boldsymbol{\Sigma}^{-1})^{-1}\right),$$

where $\boldsymbol{m}_i = \boldsymbol{z}_i - X_i \boldsymbol{\beta}$.

• the full conditional for β is

$$p(\boldsymbol{\beta}|\boldsymbol{\gamma}_{1},...,\boldsymbol{\gamma}_{I},Z) \propto \prod_{i=1}^{I} p(\boldsymbol{Z}_{i}|\boldsymbol{\beta},\boldsymbol{\gamma}_{i}) \cdot p(\boldsymbol{\beta})$$

$$= \exp \left\{ -\frac{1}{2} \sum_{i=1}^{I} (\boldsymbol{w}_{i} - X_{i}\boldsymbol{\beta})^{T} (\boldsymbol{w}_{i} - X_{i}\boldsymbol{\beta}) \right\}$$

$$\sim N_{p} \left(\left(\sum_{i=1}^{I} X_{i}^{T} X_{i} \right)^{-1} \sum_{i=1}^{I} X_{i}^{T} \boldsymbol{b}_{i}, \left(\sum_{i=1}^{I} X_{i}^{T} X_{i} \right)^{-1} \right),$$

where $\boldsymbol{b}_i = \boldsymbol{z}_i - W_i \boldsymbol{\gamma}_i$.

• the full conditional for the covariance matrix Σ is

$$p(\Sigma|\boldsymbol{\gamma}_1,\ldots,\boldsymbol{\gamma}_I) \propto \prod_{i=1}^{I} p(\boldsymbol{\gamma}_i|\Sigma) \cdot p(\Sigma)$$

 $\sim \mathcal{IW}(\nu+I,\Psi+\sum_{i=1}^{I} \boldsymbol{\gamma}_i \boldsymbol{\gamma}_i^T)$

• the full conditional for the latent variables

$$Z_{ij}|y_{ij}, \boldsymbol{\beta}, \boldsymbol{\gamma}_i \sim N(\boldsymbol{x}_{ij}^T \boldsymbol{\beta} + \boldsymbol{w}_{ij}^T \boldsymbol{\gamma}_i, 1) \cdot [\mathbb{1}(Z_{ij} \geq 0) \cdot \mathbb{1}(y_{ij} = 1) + \mathbb{1}(Z_{ij} < 0) \cdot \mathbb{1}(y_{ij} = 0)],$$

which is easy to sample from (truncated normal distribution) via inverse cdf method.

Remark that in this model missing data are easy to handle, since covariates are available. We can think of missing data as people whose personal information is given but who refused to answer to the question whether they would vote Bush or not. It is sufficient to insert those responses Y^* in the parameter space and to impute them at each iteration. This is a standard procedure in Bayesian models.

A Gibbs sampler has been implemented, with hyperparameters $\Psi = \mathcal{I}_p, \nu = p+1$.

Problem 4. Hierarchical models: gene expression over time

In droslong.csv, you will find a small subset of a time-course DNA microarray experiment. The gene-expression profiles of 2000 different genes in the fruit fly (Drosophila) genome are tracked over time during embryogenesis; you are getting data on 14 of these genes, organized in three groups (think of these as marking which cellular pathway that gene influences). For each gene at each time point, there are 3 "technical replicates"—that is, three copies of the same biological material from the same fly, run through the same process to measure gene expression.

The question of interest is: how does each gene's expression profile change over time, as the process of embryogenesis unfolds? Propose a hierarchical model for this data that properly reflects its structure. Fit this model using Gibbs sampling.

A nice graphics package is the "lattice" library. Install and load this; then try commands such as

```
xyplot(log2exp~time | gene, data=droslong)
xyplot(log2exp~time | group, data=droslong)
```

to begin exploring the structure of this data. If you know ggplot2, you can easily accomplish similar things.

An exploratory analysis of the data suggests the existence of a natural hierarchy for this problem. In particular, the gene-expression profiles seem to be different among the three different groups. Within the groups, instead, gene expression curves seem to share a common population structure, that diversifies in each gene. Moreover, each gene has three different replicates (that are ideally identical but are distorted by measurement error)

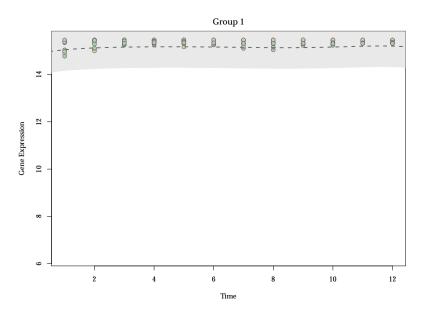


Figure 7: Reconstructed mean function for the expression of all the genes in the first group.

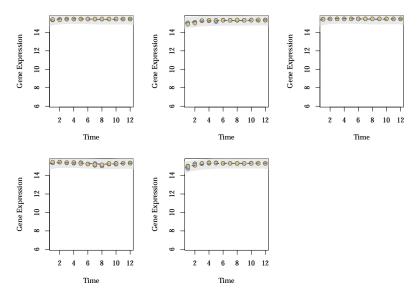


Figure 8: Reconstructed mean functions for the expression of each gene in the first group.

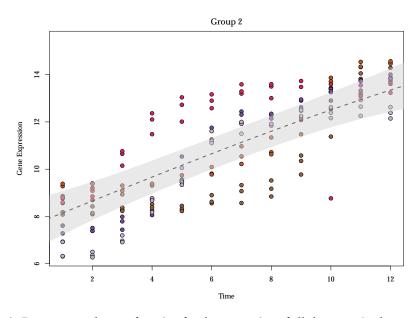


Figure 9: Reconstructed mean function for the expression of all the genes in the second group.

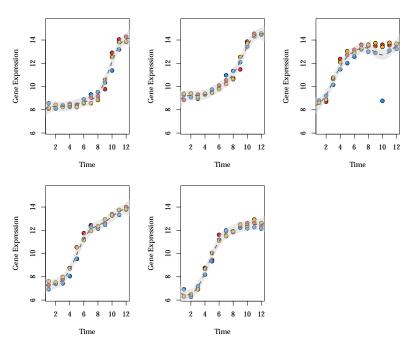


Figure 10: Reconstructed mean functions for the expression of each gene in the second group.

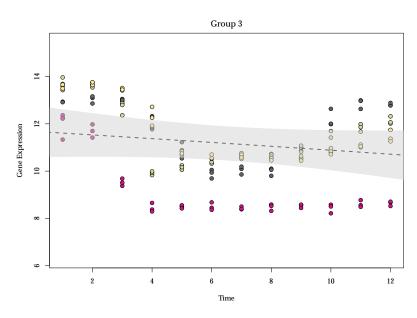


Figure 11: Reconstructed mean function for the expression of all the genes in the third group.

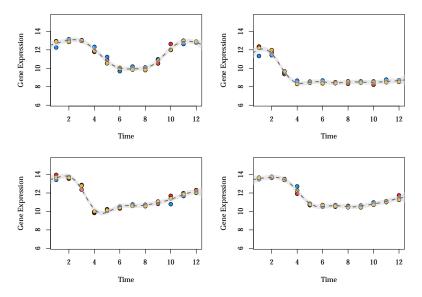


Figure 12: Reconstructed mean functions for the expression of each gene in the third group.

Appendix A

R code

```
# ==== Math Tests ====
 3
    math <- read.csv(file = 'SDS383D-master/data/mathtest.csv')</pre>
 7
    y <- math$mathscore
   ybar <- aggregate(y, list(math$school), mean)$x</pre>
   ni <- as.numeric(table(math$school))</pre>
10
   n <- sum(ni)
11
    I <- length(unique(math$school))</pre>
12
13
    # Let us see the distribution of the scores for each school
   par (mar=c(2,2,1,1))
   boxplot(y ~ math$school, col = 'gray', pch = 16, cex = 0.8, lwd = 1.2)
16
    abline(h = mean(y), col = 'indianred3', lwd = 2)
17
18
   # Let us plot the average scores for each school vs the sample size of that school
19
   par(mar=c(4,4,2,2), family = 'Palatino', cex = 1.1)
20 | plot(ni, ybar, pch = 16, xlab = 'Sample size', ylab = 'Group mean')
21 # We notice that extreme average values of the scores are obtained for school with few
22 | # students sampled. This happens because the distribution of ybar for each school has a
23
   # variance of sigma^2/ni: the smaller ni, the larger the variability of ybar around the
24
    # grand mean.
25
26
27
   # Run the Gibbs Sampler
28 Niter <- 11000
29 burnin <- 1000
   thin <- 2
31
32
   # Initialize the chain
33 | thetas.chain <- array(NA, dim = c(Niter, I))
34 mu.chain <- array(NA, dim = Niter)
35 | sigma2.chain <- array(NA, dim = Niter)
36 tau2.chain <- array(NA, dim = Niter)
    thetas.chain[1,] <- rep(0, I)
   mu.chain[1] <- 0
39 | sigma2.chain[1] <- 1
40 tau2.chain[1] <- 1
```

```
41
42
    for (i in 2:Niter) {
43
      # Update thetas
44
     var.post <- tau2.chain[i-1] * sigma2.chain[i-1] / (ni * tau2.chain[i-1] + 1)</pre>
45
      mean.post <- (mu.chain[i-1] + tau2.chain[i-1] * ni * ybar) / (ni * tau2.chain[i-1] + 1)
46
      thetas.chain[i,] <- rnorm(I, mean.post, sqrt(var.post))</pre>
47
48
      # Update mu
49
      theta.bar <- mean(thetas.chain[i,])</pre>
50
      \verb|mu.chain[i]| <- \verb|rnorm(1, theta.bar, sqrt(sigma2.chain[i-1] * tau2.chain[i-1] / I))|
51
52
      # Update sigma2
53
      S.theta <- sum((thetas.chain[i,] - mu.chain[i])^2)</pre>
54
      S.y <- sum((y - rep(thetas.chain[i,], times = ni))^2)</pre>
      rate.new <- (1/2) * (S.y + S.theta / tau2.chain[i-1])
56
      sigma2.chain[i] \leftarrow 1/rgamma(1, (n + I)/2, rate.new)
57
58
      # Update tau2
59
     rate.new <- S.theta / (2 * sigma2.chain[i])</pre>
60
     tau2.chain[i] <- 1/rgamma(1, I/2 - 1, rate.new)
61
62
    # Thin the chains
63
    thetas.chain <- thetas.chain[seq(burnin + 1, Niter, by = thin),]</pre>
    mu.chain <- mu.chain[seq(burnin + 1, Niter, by = thin)]</pre>
65
    sigma2.chain <- sigma2.chain[seq(burnin + 1, Niter, by = thin)]</pre>
66 tau2.chain <- tau2.chain[seq(burnin + 1, Niter, by = thin)]
67
68
69
    # Let us see how the Bayesian estimates differ from the sample means
70 par(mar=c(4,4,2,2), family = 'Palatino', cex = 1.1)
71 | plot(ybar, colMeans(thetas.chain), xlab=bquote(bar(y)), ylab=bquote(hat(theta)), pch = 16)
72 | abline(0, 1, col = 'indianred3', lwd = 2)
73
    # The slope of this line is smaller than 1, that is, high values of ybar_i correspond to
74
    # slightly less high values of the Bayesian estimates of theta_i; low values
75
    # of ybar_i correspond to slightly less low values of the Bayesian estimates of
76
    # theta_i. This is the shrinkage effect towards the grand mean (partial pooling).
77
78 par(mar=c(4,4,2,2), family = 'Palatino', cex = 1.1)
79
    kappa <- (ybar - colMeans(thetas.chain)) / ybar</pre>
80 plot(ni, abs(kappa), ylab=bquote(kappa), xlab="Sample size", pch = 16)
81
    abline(h=0, lwd = 2, col = 'indianred3')
    \sharp Groups with low sample size get shrunk the most, whereas groups with large
83
    # sample size hardly get shrunk at all. The larger the sample size for a group, the more
84 | # information we have for that group and the less information we need to borrow from the
85 # rest of the population.
```

Listing A.1: Math tests analysis.