

## REVIEW ARTICLE

## OPEN



# Adolescent nicotine exposure and persistent neurocircuitry changes: unveiling lifelong psychiatric risks

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Nicotine exposure during adolescence has emerged as a significant risk factor for later psychiatric disease. Notably, adolescence is a critical period for the maturation of acetylcholine and dopamine systems, neuromodulators which tightly regulate cognitive, motivational and emotional behaviors known to contribute to psychiatric vulnerability. This review explores whether long-lasting modifications in these neuromodulatory systems following adolescent nicotine exposure underlie the increased vulnerability to mental health disorders. We discuss evidence that nicotine in adolescence leads to enduring molecular, cellular alterations by perturbing the normal trajectory of cholinergic and dopamine systems, and link these changes with potential adverse behavioral outcomes in adulthood. We propose that persistent alterations in acetylcholine and dopamine signaling caused by adolescent nicotine exposure may contribute to the heightened risk for psychiatric disorders including substance abuse, anxiodepressive disorders, and schizophrenia for which deficits in a large spectrum of motivational domains are highly prevalent. The interaction between nicotine and these developing neurotransmitter systems during adolescence raises important questions about the mechanisms driving these changes. Finally, we discuss limitations in the current research and subsequently identify open questions in the field which will help drive forward research on the psychiatric consequences of adolescent nicotine use. Understanding these maladaptations could pave the way for targeted therapeutic strategies to mitigate the adverse effects of adolescent nicotine exposure on brain development and subsequent psychiatric outcomes.

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## INTRODUCTION

Smoking is a major contributor to disease burden worldwide, driven by addiction to nicotine, the primary psychoactive and addictive component of tobacco [1]. Nicotine addiction is a chronic relapsing disorder, and the prognosis may be particularly bleak for the up to 90% of adult smokers who began in adolescence [2], as early onset nicotine use is associated with longer and heavier smoking careers [3], and smokers with an adolescent onset are less likely to quit smoking than those who began as adults [4].

In addition, nicotine use is associated with multiple psychiatric comorbidities [5]. While deciphering the nature and direction of the relationship between nicotine use and comorbid psychiatric disorders can be challenging, studies increasingly point adolescent onset nicotine use as a strong predictor for later onset of psychiatric disease. Nicotine use in adolescence has been shown to predict the later appearance of depression symptoms [6–10]. Cigarette smoking in adolescence has also been associated with the onset of various anxiety disorders, which include generalized anxiety disorder, panic disorder, and post-traumatic stress disorder, in early adulthood, even after controlling for the presence of anxiety and depressive disorders in adolescence [11], and is also associated with a shorter time to onset of anxiety disorders [12]. Nicotine is also posited to act as a “gateway drug”,

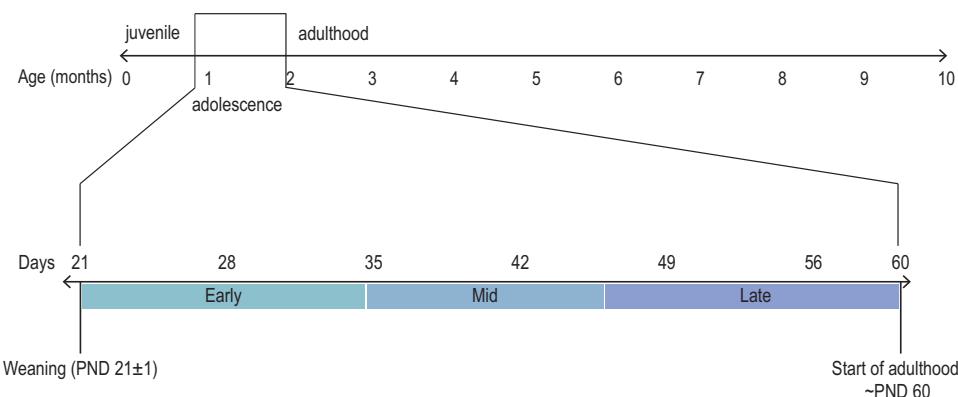
where its use in adolescence is linked with later abuse of different drug categories, such as alcohol, opiate drugs, or cocaine [3, 13–15]. These mental health disorders are a leading contributor to global disease burden, substantially reducing quality of life [16, 17].

Nicotine in adolescence may produce enduring psychiatric vulnerability by interfering with ongoing neurodevelopmental processes. Indeed, adolescence is a period of dramatic brain maturation when the continued formation and pruning of synaptic connections refines precise neuronal networks. Neuroimaging approaches have begun to shed light on complex maturational processes in both healthy subjects and pathological cases. Indeed, evidence from cross sectional and longitudinal structural MRI studies of human brain development show that gray matter thickness in cortical regions decreases across adolescence before stabilizing at adult levels, while white matter volumes increase [18–21]. Postmortem studies suggest that these macroscale changes likely reflect cellular, molecular, and connectivity development [22–25]. Functional imaging studies further indicate that significant changes occur in the function of subcortical structures and in their relationship with cortical regions, notably in nuclei associated with reward and motivation [26–29]. These neurobiological changes underlie the profound maturation in motivational and cognitive domains across

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**Fig. 1 Situating adolescence in the rodent lifespan.** Adolescence can be considered to last from weaning (generally PND 21) until rodents achieve complete sexual, behavioral, and neurobiological maturity around PND 60. It can be further divided into early (~PND 21–34), middle (~PND 35–45), and late (~PND 45–60) periods where animals exhibit different epochs of sexual development and exhibit certain behavioral and neurobiological characteristics.

adolescence [30–34], and interfering with these processes may bias individuals toward a vulnerable psychiatric state. All of these developmental processes are proposed to be highly sensitive to disruption by environmental stimuli.

In the current review we discuss the progress in our understanding of how nicotine in adolescence increases later addiction and psychiatric vulnerability, with a focus on recent advances in understanding how nicotine in adolescence enduringly alters the molecular, neurophysiological, and properties of discrete neuromodulatory circuits in animal models. Given the predominant role that dysregulations in motivational and cognitive processes play in psychiatric conditions, we place a specific emphasis on enduring neuroadaptations in cholinergic and dopaminergic circuits following adolescent nicotine exposure.

#### ADOLESCENT DEVELOPMENT OF NEUROMODULATORY SYSTEMS IN TRANSLATIONAL MODELS

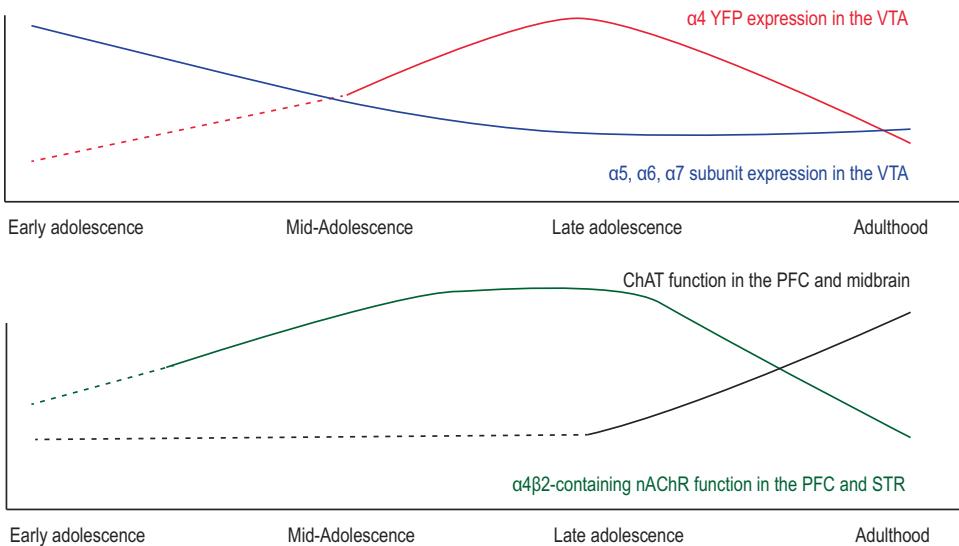
While this period is most often thought of in the context of human teenagers, it is increasingly recognized to be evolutionarily conserved, with animal species also showing distinct markers of an adolescent state [35–40]. Defining the boundaries of adolescence has proved challenging, as there are not clear hallmarks to signal the onset or completion of adolescent development. While puberty and adolescence occur during the same time period, they are not necessarily synonymous: puberty represents the process of attaining sexual maturity, while adolescence is a more diffuse period marked by significant neurobiological, behavioral and social changes. Thus, the age boundaries of the adolescent period are difficult to precisely define. Modern definitions of the human adolescent period range between 10 and 24 years of age [35, 41], an age range where children show dramatic behavioral changes and the human brain undergoes significant restructuring [22, 23, 42, 43]. While there is no clear consensus on the age range of adolescence in animal models, adolescence in rodents is increasingly defined more in line with this larger age window in humans, by counting the period from weaning (~PND 21) where mice and rats begin to interact independently with their environments for the first time, leading to a wealth of experiences with the potential to enduringly impact adult behavioral phenotypes and their underlying neuronal circuits. Rodents show behavioral and neurobiological markers of development until neurobiological maturity is reached around two months of age (~PND 60) [36, 38, 39, 44–47]. The adolescent period can be further sub-divided into early (~PND 21–34), middle (~PND 35–45), and late (~PND 45–60) periods [39, 46], which coincide roughly with pre-, peri-, and post-pubertal periods (Fig. 1). It is

important to define these time windows, as striking, age-dependent differences can be reported in the enduring effects of the same experience on neural circuitry depending on the adolescent period in which this experience occurred [48–51]. Thus, we take great care in this review to be as specific as possible about the age at which nicotine exposure occurred during adolescence.

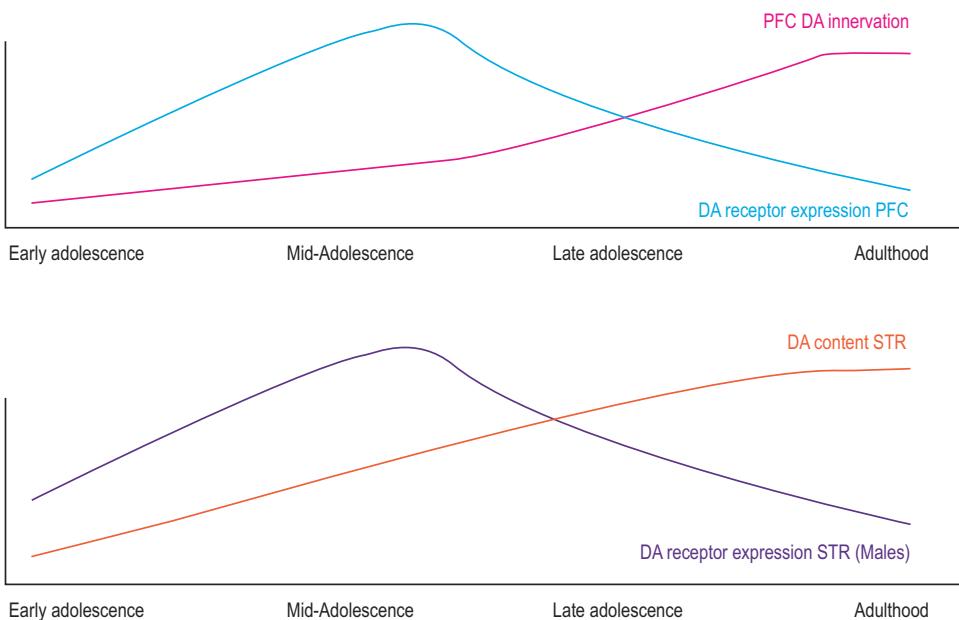
Studies in rodent models have begun to elucidate the complex development of neuromodulatory neurotransmitter circuits in adolescence, including those most likely to be directly affected by adolescent nicotine use. Nicotine usurps the action of endogenous acetylcholine (ACh) by signaling through nicotinic acetylcholine receptors (nAChRs), a diverse family of pentameric ligand-gated cation channels [52]. While nAChRs are present throughout the brain at all developmental stages, maturational patterns in their expression and function have been identified [53, 54]. Notably, expression of the most abundant and high-affinity  $\alpha 4\beta 2$ -containing nAChRs across the brain peaks in adolescence before diminishing to adult levels [55–58], and this peak has been suggested to underlie elevated nicotine self-administration in adolescent animals [59]. This increased nAChR expression was notably observed within the striatum and the midbrain [59]. Studies with greater regional specificity have shown that expression of  $\alpha 4$ -containing nAChRs in the VTA peaks late in adolescence, and is positively correlated with nicotine intake in an oral self-administration task (Fig. 2A) [60]. Rubidium efflux assays, an index of potassium channel activity, indicate higher nAChR receptor functionality in adolescents than in adults in the cortex, striatum, hippocampus, and thalamus (Fig. 2A) [61, 62]. In parallel to the developmental plasticity of cholinergic receptors, activity of choline acetyltransferase (ChAT), a requisite enzyme for ACh synthesis that also reflects the density of cholinergic innervation, increases from adolescence to adulthood in both the cortex and midbrain (Fig. 2A) [63]. At the cellular level, electrophysiological response to nicotine in cholinergic cells from the laterodorsal tegmentum (LDT), a small brain region located in the brainstem that constitutes the primary source of ACh to the ventral tegmental area (VTA) [64, 65], changes across development, with a larger excitatory response in cells from juvenile mice (PND 7–15) than in late juvenile/early adolescent mice (PND 15–34). This change in nicotine responsiveness in LDT neurons is thought to represent a developmental reorganization of the nAChR subunits present on these cells [66, 67].

ACh provides a complex regulation of reward and motivation behaviors through its actions on dopamine (DA) circuitry [68–72]. The release of ACh from LDT projections directly modulates activity of VTA DA neurons, finely tuning their firing rate and burst properties [73, 74]. DA signaling in the striatum is also modulated

## A ACh system changes across adolescence



## B DA system changes across adolescence



**Fig. 2 Adolescent changes in acetylcholine and dopaminergic systems. A** ACh system changes across adolescence. **Top:** regional changes in nAChR subunit expression have been noted across adolescence, with a peak in late adolescence of  $\alpha 4$  receptor expression in the VTA, as assessed with YFP tagging, and with a gradual lessening of  $\alpha 5$ ,  $\alpha 6$ , and  $\alpha 7$  receptors in the VTA between adolescence and adulthood. Dotted lines represent periods where expression has not yet been studied. **Bottom:**  $\alpha 4\beta 2$ -containing nAChR function in the PFC and striatum increases from late adolescence to adulthood. ChAT function changes across adolescence in the PFC and in the midbrain, with a peak in function in mid- to late adolescence before declining to adult levels. **B** Dopamine system changes across adolescence. **Top:** In the PFC, DA innervation increases steadily across adolescence until reaching a plateau around PND 60. DA receptor expression in the PFC peaks in mid-adolescence before declining to adult levels. **Bottom:** DA content in the striatum increases across adolescence, while DA receptor expression peaks in mid-adolescence. This peak appears to be sex specific, as it has been confirmed in male rodents, but was not seen in females.

at the terminal level by ACh release from local striatal cholinergic interneurons [75], but also from an external direct cholinergic input from the LDT [76]. Hence, ACh signaling through presynaptic and somato-dendritic nAChRs tightly regulates DA circuit activity, suggesting that it is another neuromodulatory system profoundly affected by developmental exposure to nicotine. Dopamine

circuits undergo a robust period of maturation in adolescence, including changes to innervation patterns, dopamine synthesis and release, basal dopamine neuron firing rates, and dopamine receptor expression in terminal areas, with particularly pronounced changes in mesocortical and mesolimbic circuits (Fig. 2B) [39, 77–82]. Of note, the segregation of mesocorticolimbic

dopamine pathways continues in adolescence, with a subset of dopamine axons passing through limbic regions before growing to the PFC [83]. The targeting processes of these axons can be disrupted by experience with stimulant drugs of abuse in adolescence, such as amphetamine [49, 51], or by stress [84], resulting in enduring cognitive deficits.

The development of dopaminergic and cholinergic systems may also be inter-related. Adolescent rodents show greater DA release in the NAc in response to acute nicotine when compared to adult counterparts, despite similar baseline measures [85, 86]. Electrophysiological response of VTA DA neurons to acute nicotine also changes across development, with an increased sensitivity in early adolescent animals [87]. Molecular evidence links these changes to maturation in VTA nAChR expression, as mRNA transcripts of  $\alpha 5 \alpha 6$  and  $\alpha 7$  nAChR receptor subunits in the VTA peak early in adolescence (Fig. 2A), in tandem to nicotine-stimulated DA release in the ventral striatum [88]. Together, these results suggest that nicotine in adolescence produces different immediate and long-term outcomes through its actions on these developing neurotransmitter systems.

### **BEHAVIORAL OUTCOMES OF ADOLESCENT NICOTINE EXPOSURE IN ANIMAL MODELS : TRANSLATIONAL EVIDENCE FOR PSYCHIATRIC VULNERABILITY**

These developmental differences in cholinergic and dopaminergic function may underlie the age-specific behavioral responses provoked by an acute exposure to nicotine. In particular, many studies have shown that adolescent rodents show an increased sensitivity to the locomotor activating and rewarding effects of nicotine, blunted withdrawal symptoms, and reduced aversion to high doses of nicotine, compared to adult counterparts. Furthermore, these behavioral outcomes have been linked to differences in response to nicotine across several neurotransmitter systems, as well as recruiting microglial pathways to sculpt circuitry in response to acute exposure [44, 89–91]. These studies on the age-dependent acute effects of nicotine are outside the scope of the current review, but have been well reviewed in the above citations. How nicotine exposure in adolescence affects later sensitivity to nicotine has also been heavily studied, particularly from a behavioral point of view. Nicotine in adolescence has been shown to increase later sensitivity to the rewarding effects of nicotine, while decreasing its aversive effects; these studies have been thoroughly reviewed elsewhere, see: [90, 92]. Nicotine in adolescence has also been proposed to increase sensitivity to other psychostimulant drugs in adulthood [93], leading, notably, to its reputation as a “gateway drug”.

A second focus of behavioral studies with nicotine in adolescence has been its long-term effects of translational behavioral markers for vulnerability to psychiatric disease [94, 95]. Exposure to nicotine in mid-adolescence (PND 35–44) induces anxiety-like behaviors, such as a reduction of time spent in the open arms of the elevated plus maze, in adult rats and promotes a persistent depressive-like state, with an increased immobility in the forced swim test, and reduced sucrose preference [96, 97]. Notably, the consequences of adolescent exposure to this nicotine regimen were sex-specific, with nicotine-induced vulnerability to anxiety- or depression-like behavior specific to male rats [98]. Anxiety- and depression-like behaviors induced by adolescent nicotine exposure can be rescued in adult male rats by re-exposure to treatment with SSRIs [99], or, intriguingly, by re-exposure to nicotine; suggesting that the repression of these negative emotional states may also be a driver for continued nicotine use in adulthood for those that started as adolescents. How exposure to nicotine in adolescence leads to these enduring behavioral changes, which resemble a model of psychiatric vulnerability, remains under investigation – but may result from enduring nicotine-induced changes to developing

cholinergic and dopaminergic circuits, and the interplay between them.

### **ENDURING EFFECTS OF NICOTINE IN ADOLESCENCE ON ADULT CHOLINERGIC SIGNALING**

Acetylcholine acts as a neuromodulator in the brain, where it signals through cationic nAChRs and metabotropic muscarinic receptors, notably regulating cellular excitability, synaptic plasticity, and the coordinated firing of groups of neurons to play a key role in fundamental aspects of brain physiology including emotion, cognition and motivation [100]. Impairments to cholinergic signaling in the adult human brain have been linked to psychiatric disorders in adult patients, including schizophrenia, anxiety, and depression [101–103]. Notably, anomalies in the availability or expression of specific nAChR subunits have been implicated in depression [104] and schizophrenia patients [105, 106]. Whether these alterations to the expression and function of nAChRs are antecedent to the onset of these associated disease states remains an open question. Mouse models have begun to address how manipulating signaling through nAChRs in adult animals may produce symptoms associated with psychiatric diseases other than addiction to nicotine itself [74, 107–110]. However, in addition to its functions in the mature brain, ACh signaling is essential for neuronal development, where it plays a trophic role in establishing and maintaining neuronal connectivity [111, 112]. Thus, developmental exposure to nicotine stands to significantly perturb neuronal maturation by interfering with ACh signaling. In this section, we review evidence from animal models that exposure to nicotine in adolescence produces enduring alterations to nAChR expression and function, altering basal cholinergic signaling, response to later nicotine exposure, and establishing a vulnerable psychiatric phenotype.

Exposure to nicotine in adolescence is associated with brain-wide changes to the expression of discrete nAChR subunits, with some transient changes appearing during treatment or withdrawal, but with some persistent changes enduring until adulthood. Transient changes in nAChR expression following adolescent nicotine exposure have been reviewed elsewhere [44, 53, 113, 114]. Persistent changes to cholinergic and nicotinic signaling through nAChRs induced by nicotine in adolescence are of particular interest in the context of long-term psychiatric vulnerability, however there is less data available concerning this topic. Two weeks of exposure to nicotine in mid-adolescence (PND 30–47) increased binding to  $\alpha 4\beta 2$ -containing nAChRs, an effect that persisted into adulthood in the PFC and midbrain [56, 63, 115], while an upregulation of  $\alpha 7$ -containing nAChRs in the striatum, brainstem, and cerebellum was apparent only transiently [116]. A similar mid-adolescent exposure regimen (PND 34–43) lead to an increased expression of  $\alpha 5$ ,  $\alpha 6$ ,  $\beta 2$ , and  $\beta 3$  nAChR subunits in the adult VTA — while the same exposure, given instead during adulthood, only increased the  $\beta 3$  subunit [117]. Brief exposure to nicotine in early adolescence (PND 27–33) increased nAChR function seven weeks later in the striatum, PFC, hippocampus, and thalamus/midbrain, including the VTA [118]. Nicotine treatment in mid-adolescence (PND 35–44) had sex-specific long-term effects on nAChR protein levels in the PFC, with  $\alpha 7$  and  $\beta 2$  protein reduced in adult males (>P75) exposed to nicotine in adolescence and no change the expression of these subunits in female rats exposed at the same age [98].

Choline acetyltransferase (ChAT), an enzyme essential for ACh synthesis, is often used as a measure for cholinergic innervation. Changes in ChAT activity following exposure to nicotine in adolescence appear to be primarily transient, with an upregulation of ChAT activity in the striatum of male rats and in the midbrain of females reported during and shortly following exposure in mid-adolescence (PND 30–47), which were no longer detectable once

the rats reached adulthood [119]. Similarly, oral exposure to nicotine in C57BL/6 mice in mid-adolescence (PND 30–45) increased ChAT activity when measured shortly after treatment at PND 50, an effect which normalized by PND 75 [63]. Persistent changes have been reported in the midbrain, where ChAT activity was reduced during and after nicotine injections during mid-adolescence (PND 30–47), and this reduction persisted until at least PND75 [115, 120].

### PERSISTENT EFFECTS OF ADOLESCENT NICOTINE EXPOSURE ON DA CIRCUITS

DA neurotransmission is essential for diverse behavioral outputs, including motivation, cognition, reward learning, decision making, salience attribution, and voluntary motor control [121–127]. With such varied outputs, it is unsurprising that DA signaling has been heavily implicated in psychiatric disease, and neuroimaging studies in adult patients with schizophrenia, depression, and addiction have noted alterations to dopaminergic receptor expression [128–131]. Functional studies in adult rodent models have linked genetic or experimentally induced alterations in DA neuron activity, DA release, and receptor binding to transdiagnostic changes in motivation and cognitive control [49, 132–135]. As the dopamine system undergoes a period of profound and dynamic development in adolescence [39], it is increasingly proposed to function as a ‘plasticity system’, where experiences can create enduring changes to behavior through their actions on the dopamine system [78, 136]. Nicotine in adolescence is thus poised to alter DA development, as it acts directly on VTA DA neurons through their expression of nAChRs, modulating their firing rate in a population-specific manner in adult rodents [72, 137–139]. Nicotine also has indirect or circuit effects on DA neurotransmission, as nAChRs are also present on DA axon terminals, on neurons present in dopaminoceptive regions, and importantly on VTA GABA neurons, which regulate DA neuron activity [52, 68, 69, 140]. Nicotine may therefore alter interact with the developing DA system at multiple levels to promote later psychiatric vulnerability.

Nicotine in adolescence has been proposed to evoke enduring structural and functional changes in DA neurons and/or in their terminal regions. At the level of the VTA, basal DA neuron firing was increased in adult, male rats following exposure to nicotine in mid-adolescence (PND 35–44) [96, 98], however another group reports that basal DA neuron firing rate was decreased in adult rats following mid-adolescent (PND 38–42) nicotine exposure [141]. These disparate findings may result from differences in rat strains as Jobson, Ng, and colleagues studied Sprague-Dawley rats, while Cadoni and colleagues noted a decrease in firing rate in Lewis rats, with no significant change in Fisher rats. Structural analysis of VTA neuron populations note that nicotine in adolescence increased the proportion of DA neurons that do not express the vesicular glutamate transporter (TH+Vglut-neurons) in the posterior VTA in rats [142]. In this case, rats were exposed to nicotine starting at PND28, and the exposure continued until they were examined as adults, raising the question of when exactly these changes may occur, particularly as no comparison was made with rats whose exposure began as adults. Finally, nicotine-induced changes to local GABA signaling in the VTA may influence DA neuron function, as the concerted action of nicotine on both DA and GABA neurons in the VTA is necessary for nicotine reward [140]. Accordingly, a recent study found that adolescent, but not adult, nicotine exposure alters GABA signaling in the VTA by reorganizing chloride homeostasis, and, in turn, increases inhibitory tone over lateral VTA DA neurons [143].

At the terminal level, regional differences in DA function in the NAc and PFC, the terminal regions of the mesocorticolimbic DA pathway, have been implicated in rodent models of mood disorders and addiction. In the NAc, DA D1 receptors (D1Rs) were

downregulated in the shell subregion of adult male rats that were exposed to nicotine in mid-adolescence (PND 35–44), while, in contrast, D1Rs in the NAc shell were enduringly unregulated in female rats exposed to nicotine in adolescence [97, 98]. Downregulation of D1R expression in the NAc shell of adult male rats following adolescent nicotine exposure was further associated with functional changes to local NAc medium spiny neurons (MSNs). Nicotine in adolescence produced a persistent hyperactive firing state in MSNs, albeit with decreased bursting activity [97]. Nicotine in adolescence has also been shown to change dendrite morphology in DAergic regions including the NAc [144].

DA terminals in the PFC are known to develop throughout adolescence, and their development shapes the maturation of local circuits and calibrates adult cognitive function [83, 145]. Exposure to nicotine in mid-adolescence (PND 34–43) has been shown to increase electrically evoked DA release in the PFC of adult animals [146], an effect that was associated with enduring cognitive deficits including reduced attention and increased impulsivity. This exposure pattern also enduringly altered PFC long term potentiation (LTP) into adulthood, marked by an increased ability of prefrontal synapses to undergo spike-timing-dependent LTP [147]. These electrophysiological and behavioral alterations were associated with transient changes in nAChR expression [57], and enduring changes in mGluR2 signaling [147, 148]. Nicotine exposure in mid-adolescence (PND 35–44) has been shown to have sex-dependent effects on PFC function, as it increased the firing frequency of PFC pyramidal neurons and decreased PFC D1R protein expression in only in male rats [96, 98]. In contrast, this treatment increased D2R expression in the PFC of male rats, while decreasing D2R expression in the PFC of female rats [97, 98]. Finally, exposure to nicotine in mid-adolescence (PND 30–47) increased DA turnover in the and PFC at PND 80 in males and female rats [149], highlighting that female rodents should not be considered invulnerable to the effects of nicotine in adolescence, even if the changes produced by exposure are not the same as those seen in their male counterparts.

Together, these nicotine-induced changes to the developing DA system are thought to potentiate the effects of re-exposure to nicotine in adulthood and lead to the expression of behaviors associated with psychiatric vulnerability, yet few studies have made a causal link between these ideas. Our recent study showed that exposure to nicotine in early adolescence (PND 21–28), but not the same nicotine exposure in adulthood (PND 60+), produced enduring alterations in sensitivity to nicotine, notably defined by an increased sensitivity to its rewarding effects and a decreased sensitivity to its anxiogenic effects. These behavioral outcomes were linked with an electrophysiological signature of hyper-reactivity to nicotine specifically in the VTA-NAc circuit, and not in the VTA-Amygdala circuit, despite the clear role of this pathway in the anxiogenic effects of nicotine [137]. We next provided causal evidence linking nicotine-induced hyperactivity in the VTA-NAc circuit with the reduced anxiogenic effect of nicotine seen in adult mice exposed to nicotine in adolescence, as chemogenetic dampening of this specific pathway unmasked an adult-like behavioral response to nicotine [150]. This is, to our knowledge, the first instance where intervention at the level of circuit changes resulting from adolescent nicotine exposure rescued the expected adult behavior.

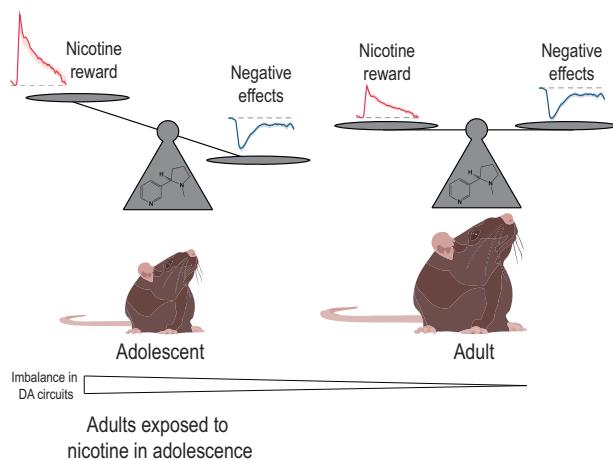
### DOES NICOTINE IN ADOLESCENCE “FREEZE” THE BRAIN IN AN IMMATURE STATE?

How exactly nicotine in adolescence alters DA circuit development is of primary interest. Exposure to stimulant drugs such as amphetamine, for example, has been shown to disrupt adolescent dopamine development, that is it produces outcomes not typically seen in control animals at any age. For example, amphetamine in adolescence misroutes a subset of dopamine axons to the PFC in an

age- and sex-dependent manner, leading to a pathological ectopic innervation associated with cognitive deficits [51]. In contrast, nicotine exposure in adolescence appears to “freeze” dopamine circuitry in an adolescent-like state. This idea was first proposed as mice exposed to nicotine in adolescence showed behavioral responses to methylphenidate as adults that closely resembled the responses of naïve adolescent animals [151]. Recently, our work has given further mechanistic insight into this idea. We showed that exposure to nicotine in early adolescence, but not the same exposure in adulthood, produced a persistent vulnerability profile: adult mice that were exposed to nicotine in their adolescence consumed more nicotine, were less affected by the drug’s negative effects, and experienced an increased sensitivity to its rewarding properties. What was interesting, however, is how closely this vulnerability profile mirrored the response of naïve adolescent mice to nicotine. Notably, adolescent mice were more sensitive to the rewarding effect of nicotine, showing a place preference for nicotine at a dose too low to produce this response in naïve adult mice. Adolescent mice were also impervious to the anxiogenic effect of nicotine, which seems to come online in parallel to the maturation of DA circuitry. Adult mice treated with nicotine in adolescence also showed immature electrophysiological responses to nicotine, with an adolescent-like exaggerated activation of VTA-NAc DA neurons. We thus hypothesized from these findings that exposure to nicotine in adolescence prolongs a naturally-occurring developmental imbalance in dopaminergic signaling between NAc- and amygdala-projecting pathways (Fig. 3). This imbalance, in turn, creates a vulnerable state by dampening negative effects of the drug (e.g. its anxiogenic effects). Finally, we were able to restore the mature behavioral response to nicotine in adolescent-exposed mice by chemogenetically resetting an adult-like balance in dopamine signaling in response to nicotine. We thus came to the initially unexpected conclusion that nicotine does not *disrupt* the adolescent development of dopamine pathways, per se, but rather arrests their development in an immature state [150].

## LIMITATIONS

Research examining the impact of nicotine during adolescence is critical for understanding how early exposure affects brain



**Fig. 3 Nicotine exposure in adolescence leads to immature response to nicotine in adulthood.** Adolescent mice show an increased rewarding effect of nicotine, paired with a blunted anxiogenic effect of the drug in comparison to adults. These differences result from an imbalance in dopamine neuron response to nicotine at the circuit level [150]. Adult mice that were exposed to nicotine in adolescence show similar behavioral and electrophysiological responses to nicotine, suggesting that adolescent exposure arrests the development of dopaminergic circuitry.

development and behavior. When holistically assessing the findings reported to date, however, it is apparent that there are significant limitations which must be taken into account when discussing the existing literature – equally in the case of conflicting findings and in the case of harmonious ones:

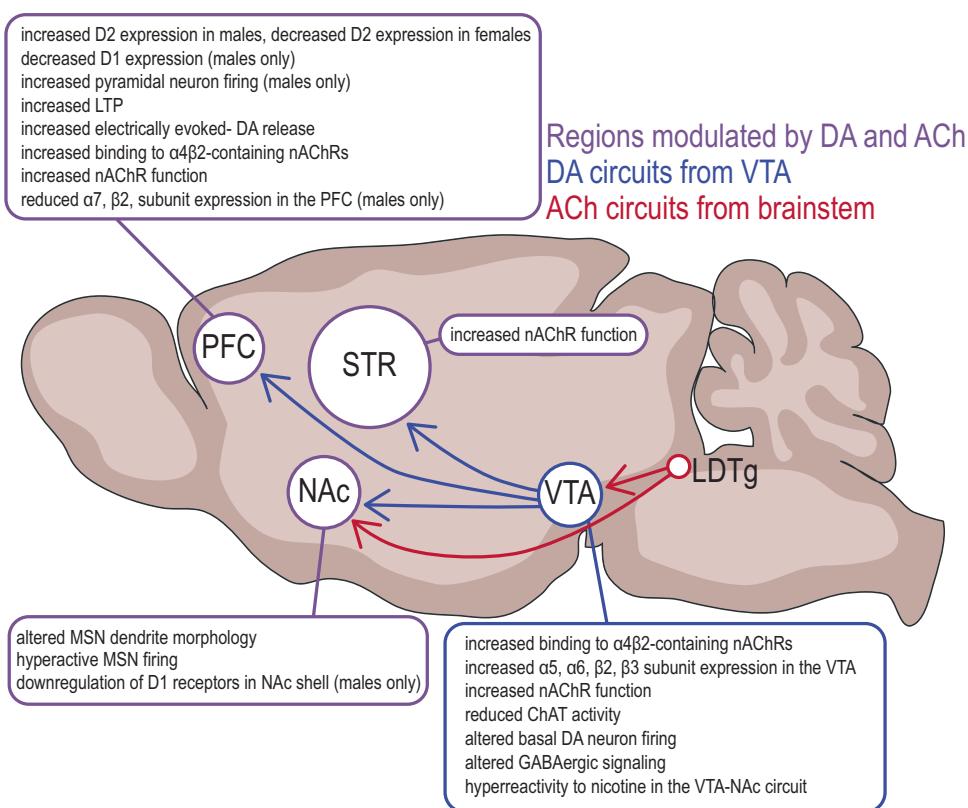
- **Adolescent ages:** As discussed earlier in the review, the definition of adolescence – and particularly of adolescence in animal models – does not necessarily have a strict consensus on its age range. We take care in this review to explicitly state the age ranges of each nicotine treatment, but it is important to keep in mind that the same treatment at different ages can certainly produce disparate outcomes. In the case of amphetamine exposure in adolescence, for example, exposure in early adolescence had opposite sex-dependent outcomes as exposure in mid-adolescence [51]. To the best of our knowledge, no study to date has yet directly compared the consequences of the same nicotine exposure at different adolescent ages. A final, important point on the issue of treatment ages is that not all studies include an adult control group, which is necessary to disentangle the developmental nature of long-term nicotine effects. Without this important control, it is impossible to conclude that the results of an adolescent exposure paradigm are specific to this exposure age.
- **Variability in nicotine exposure:** The routes of administration and doses of nicotine in studies can vary widely (Table 1, [152]). Different administration routes and/or nicotine doses can thus lead to significant variability in reported findings, particularly as different administration routes are known to alter the pharmacokinetic and pharmacodynamic properties of the same drug dose [153]. Indeed, this variability makes it difficult to directly compare results across studies or to replicate the natural patterns of nicotine use in human adolescents.
- **Sex differences:** Sex and gender differences in nicotine use have been reported in human populations [154–156], notably among adolescent users, as well as in rodent studies [157]. While some studies include rodents of both sexes, others may not adequately account for sex differences or may use only one sex, limiting the generalizability of the findings. These differences can be significant in understanding the broader impact on human adolescents.

## OPEN QUESTIONS

This review underscores how even brief nicotine exposure during adolescence can lead to significant and lasting changes in the cholinergic and dopaminergic systems. However, it also highlights how much we have yet to discover about the underlying mechanisms. Further studies are needed, particularly those aimed at determining when in adolescence are the critical moments for nicotine exposure to engender its enduring negative effects. Another major question is indeed how the enduring effects of nicotine in adolescence are perpetuated in the adult brain. One possibility is changes to nAChR receptor trafficking and surface expression patterns in response to nicotine, which may be short- or long-lasting in response to nAChR desensitization or inactivation following exposure [158]. Exposure to nicotine in adolescence may also produce enduring epigenetic changes, like those seen after pre-natal and juvenile exposure to nicotine [159]. Here, we focused on persistent alterations in neuromodulatory circuit function and we end our review on a current, intriguing hypothesis: that nicotine in adolescence may arrest the development of DA systems. Whether this may be true also for ACh circuits or other neurotransmitter systems is currently unknown. Naturally occurring developmental differences between the adolescent and adult brain have been proposed to render

**Table 1.** Treatment details for manuscripts addressing persistent consequences of adolescent nicotine exposure.

Paper	doi	species	strain	sex	adolescent exposure age (PND)	adolescent exposure method	adolescent exposure dose	adolescent exposure dose
Abreu-Villaça et al. [115]	<a href="https://doi.org/10.1038/sj.npp.1300221">https://doi.org/10.1038/sj.npp.1300221</a>	rat	SD	M & F	30–38	minipump OR s.c.	0.6, 2, or 6 mg/kg/d	
Adriani et al. [117]	<a href="https://doi.org/10.1523/jneurosci.23-11-04712.2003">https://doi.org/10.1523/jneurosci.23-11-04712.2003</a>	rat	SD	M	34–43	i.p.	0.4 mg/kg/d	
Cadoni et al. [141]	<a href="https://doi.org/10.1111/adb.12803">https://doi.org/10.1111/adb.12803</a>	rat	Lewis and Fisher	M	38–42	s.c.	0.4 mg/kg (1× day for 5 days)	
Counotte et al. [57]	<a href="https://doi.org/10.1096/fj.11-198994">https://doi.org/10.1096/fj.11-198994</a>	rat	wistar	M	34–43	s.c.	0.4 mg/kg (3× day)	
Counotte et al. [146]	<a href="https://doi.org/10.1038/npp.2008.96">https://doi.org/10.1038/npp.2008.96</a>	rat	wistar	M	34–43	s.c.	0.4 mg/kg (3× day)	
Counotte et al. [148]	<a href="https://doi.org/10.1038/nn2770">https://doi.org/10.1038/nn2770</a>	rat	wistar	M	34–43	s.c.	0.4 mg/kg (3× day)	
Goriounova and Mansvelder, [147]	<a href="https://doi.org/10.1523/jneurosci.5502-11.2012">https://doi.org/10.1523/jneurosci.5502-11.2012</a>	rat	wistar	M	34–43	s.c.	0.4 mg/kg 3x daily × 10 days	
Hudson et al. [97]	<a href="https://doi.org/10.1111/adb.12891">https://doi.org/10.1111/adb.12891</a>	rat	SD	M	35–44	s.c.	0.4 mg/kg 3x daily × 10 days	
Iñiguez et al. [99]	<a href="https://doi.org/10.1038/npp.2008.220">https://doi.org/10.1038/npp.2008.220</a>	rat	SD	M	30–44	s.c.	(0.16, 0.32, and 0.64 mg/kg) twice daily	
Jobson et al. [96]	<a href="https://doi.org/10.1093/cercor/bhy179/5074516">https://doi.org/10.1093/cercor/bhy179/5074516</a>	rat	SD	M	35–44	s.c.	0.4 mg/kg 3x daily × 10 days	
Kota et al. [118]	<a href="https://doi.org/10.1016/j.jbc.p.2009.06.099">https://doi.org/10.1016/j.jbc.p.2009.06.099</a>	mice	ICR	M	27–33	s.c.	0.5 mg/kg per inj 2x day/14 days	
Ng et al. [98]	<a href="https://doi.org/10.1038/s41386-024-01853-y">https://doi.org/10.1038/s41386-024-01853-y</a>	rat	SD	M & F	35–44	s.c.	0.4 mg/kg 3x daily × 10 days	
Nolley and Kelley [151]	<a href="https://doi.org/10.1016/j.jntt.2006.09.026">https://doi.org/10.1016/j.jntt.2006.09.026</a>	mice	C57BL/6J	M	25–57	i.p.	0.3 mg/kg 3.0 mg/kg	
Reynolds et al. [150]	<a href="https://doi.org/10.1101/2023.10.28.564518">https://doi.org/10.1101/2023.10.28.564518</a>	mice	C57BL/6J	M	21–28	oral	100 µg/ml	
Ribeiro-Carvalho et al. [63]	<a href="https://doi.org/10.1016/j.neuroscience.2009.05.032">https://doi.org/10.1016/j.neuroscience.2009.05.032</a>	mice	C57BL/6	M & F	30–45	oral	50 µg/ml in 2% sac	
Slotkin et al. [119]	<a href="https://doi.org/10.1016/j.brainresbull.2007.12.009">https://doi.org/10.1016/j.brainresbull.2007.12.009</a>	rat	SD	M & F	30–47	minipump	6 mg/kg/d	
Slotkin et al. [116]	<a href="https://doi.org/10.1016/j.brainres.2004.10.009">https://doi.org/10.1016/j.brainres.2004.10.009</a>	rat	SD	M & F	30–47	minipump	6 mg/kg/d	
Thomas et al. [143]	<a href="https://doi.org/10.1016/j.ceirep.2018.03.030">https://doi.org/10.1016/j.ceirep.2018.03.030</a>	rat	LE	M	28–42	i.p.	0.4 mg/kg	
Trauth et al. [56]	<a href="https://doi.org/10.1016/s0006-8993(99)01994-0">https://doi.org/10.1016/s0006-8993(99)01994-0</a>	rat	SD	M & F	30–47	minipump	6 mg/kg/d	
Trauth et al. [149]	<a href="https://doi.org/10.1016/s0006-8993(00)03227-3">https://doi.org/10.1016/s0006-8993(00)03227-3</a>	rat	SD	M & F	30–47	minipump	6 mg/kg/d	
Vrettou et al. [142]	<a href="https://doi.org/10.1016/j.jdr.2023.100180">https://doi.org/10.1016/j.jdr.2023.100180</a>	rat	wistar	M	28–63	s.c.	0.35 mg/kg 3x/wk 6 weeks	



**Fig. 4 Summary of modifications in adult acetylcholine and dopaminergic systems induced by adolescent nicotine exposure.** Alterations in functional outputs in adulthood, such as receptor expression and function, have been noted for both ACh and DA circuits following nicotine in adolescence. Sex-specific changes are noted where that information is available.

adolescents more sensitive to drugs of abuse – in particular, by augmenting rewarding effects of drugs and/or diminishing their negative effects. Adolescents are also thought to be more sensitive to stress. These conjectures again raise the idea that prolonging an adolescent-like state may indeed increase psychiatric vulnerability in adults.

While ACh and DA systems clearly both undergo significant maturation in adolescence, thus making them targets of developmental nicotine exposure, a comprehensive understanding of how the maturation of one neuromodulatory system affects the other during this critical period warrants investigation. Both muscarinic and nicotinic ACh receptors are present on VTA dopamine neurons, as well as on cholinergic and glutamatergic terminals regulating different aspects of dopamine release in output sites, such as the striatum and NAc [160–162]. Conversely, D2 dopamine receptors modulate the activity of striatal cholinergic interneurons [75]. Although these systems mature at different rates, their intertwined regulation suggests that disruptions in one could influence the development and function of the other. There remains a need for longitudinal research that tracks the concurrent development of both systems. Such studies could reveal how early changes in one system might predict alterations in the other and their subsequent behavioral implications, offering valuable insights for psychiatric research.

## CONCLUSION

Nicotine use, and particularly nicotine use during adolescence is associated with an increased risk of addiction and mood disorders in adulthood. While prevention efforts over the past two decades have had some success at curbing adolescent initiation of cigarette smoking, recently vaping has emerged as an alternative nicotine delivery method immensely popular amongst adolescents [163].

While e-cigarettes are viewed as a safer alternative than conventional cigarettes, as they spare exposure to toxic chemical constituents of tobacco smoke, vapers are still chronically exposed to nicotine [164, 165]. Moreover, more adolescents are inclined to transition from vaping to conventional cigarettes, reversing a two-decade-long decline [166–169]. Thus, understanding the neurobiological underpinnings of how nicotine exposure in adolescence increases later psychiatric risk is essential for developing effective prevention and intervention programs to promote healthy transitions from adolescence to adulthood. In this review we cover the state of the literature with regards to studies of the long-term effects of nicotine in adolescence on neuromodulatory systems in animal models (summarized in Fig. 4), identify limitations in our ability to interpret these studies as a whole, and propose open questions that may drive the field forward.

## REFERENCES

- Ezzati M, Lopez AD, Rodgers A, Hoorn SV, Murray CJL, Group CRAC. Selected major risk factors and global and regional burden of disease. *Lancet*. 2002;360:1347–60.
- SAMHSA. Results from the 2011 National Survey on Drug Use and Health: summary of national findings. NSDUH Series H-44, HHS Publication No (SMA) 12-4713. Rockville, MD: Substance Abuse and Mental Health Services Administration; 2012.
- Grant BF. Age at smoking onset and its association with alcohol consumption and DSM-IV alcohol abuse and dependence: results from the national longitudinal alcohol epidemiologic survey. *J Subst Abuse*. 1998;10:59–73.
- Hu T, Gall SL, Widome R, Bazzano LA, Burns TL, Daniels SR, et al. Childhood/Adolescent smoking and adult smoking and cessation: the international childhood cardiovascular cohort (i3C) consortium. *J Am Heart Assoc*. 2020;9:e014381.
- Ziedonis D, Hitsman B, Beckham JC, Zvolensky M, Adler LE, Audrain-McGovern J, et al. Tobacco use and cessation in psychiatric disorders: National Institute of Mental Health report. *Nicotine Tob Res*. 2008;10:1691–715.

6. Choi WS, Patten CA, Gillin JC, Kaplan RM, Pierce JP. Cigarette smoking predicts development of depressive symptoms among U.S. adolescents. *Ann Behav Med*. 1997;19:42–50.
7. Wu LT, Anthony JC. Tobacco smoking and depressed mood in late childhood and early adolescence. *Am J Public Health*. 1999;89:1837–40.
8. Goodman E, Capitman J. Depressive symptoms and cigarette smoking among teens. *Pediatrics*. 2000;106:748–55.
9. Ranjit A, Buchwald J, Latvala A, Heikkilä K, Tuulio-Henriksson A, Rose RJ, et al. Predictive association of smoking with depressive symptoms: a longitudinal study of adolescent twins. *Prev Sci*. 2019;20:1021–30.
10. Baiden P, Szlyk HS, Cavazos-Rehg P, Onyeaka HK, Peoples JE, Kasson E. Use of electronic vaping products and mental health among adolescent high school students in the United States: the moderating effect of sex. *J Psychiatr Res*. 2022;147:24–33.
11. Johnson JG, Cohen P, Pine DS, Klein DF, Kasen S, Brook JS. Association between cigarette smoking and anxiety disorders during adolescence and early adulthood. *JAMA*. 2000;284:2348–51.
12. Jamal M, Does AJ, Penninx BW, Cuijpers P. Age at smoking onset and the onset of depression and anxiety disorders. *Nicotine Tob Res*. 2011;13:809–19.
13. Lai S, Lai H, Page JB, McCoy CB. The association between cigarette smoking and drug abuse in the United States. *J Addict Dis*. 2000;19:11–24.
14. Chen X, Unger JB, Palmer P, Weiner MD, Johnson CA, Wong MM, et al. Prior cigarette smoking initiation predicting current alcohol use evidence for a gateway drug effect among California adolescents from eleven ethnic groups. *Addict Behav*. 2002;27:799–817.
15. Hanna EZ, Grant BF. Parallels to early onset alcohol use in the relationship of early onset smoking with drug use and DSM-IV drug and depressive disorders: findings from the national longitudinal epidemiologic survey. *Alcohol Clin Exp Res*. 1999;23:513–22.
16. Collaborators G 2019 MD. Global, regional, and national burden of 12 mental disorders in 204 countries and territories, 1990–2019: a systematic analysis for the Global Burden of Disease Study 2019. *Lancet Psychiatry*. 2022;9:137–50.
17. Vigo D, Thornicroft G, Atun R. Estimating the true global burden of mental illness. *Lancet Psychiatry*. 2016;3:171–8.
18. Sowell ER, Thompson PM, Holmes CJ, Jernigan TL, Toga AW. In vivo evidence for post-adolescent brain maturation in frontal and striatal regions. *Nat Neurosci*. 1999;2:859–61.
19. Sowell ER, Peterson BS, Thompson PM, Welcome SE, Henkenius AL, Toga AW. Mapping cortical change across the human life span. *Nat Neurosci*. 2003;6:309–15.
20. Giedd JN, Blumenthal J, Jeffries NO, Castellanos FX, Liu H, Zijdenbos A, et al. Brain development during childhood and adolescence: a longitudinal MRI study. *Nat Neurosci*. 1999;2:861–3.
21. Gogtay N, Giedd JN, Lusk L, Hayashi KM, Greenstein D, Vaituzis AC, et al. Dynamic mapping of human cortical development during childhood through early adulthood. *Proc Natl Acad Sci USA*. 2004;101:8174–9.
22. Petanjek Z, Judaš M, Šimic G, Raslin MR, Uylings HBM, Rakic P, et al. Extraordinary neoteny of synaptic spines in the human prefrontal cortex. *Proc Natl Acad Sci USA*. 2011;108:13281–6.
23. Catts VS, Fung SJ, Long LE, Joshi D, Vercammen A, Allen KM, et al. Rethinking schizophrenia in the context of normal neurodevelopment. *Front Cell Neurosci*. 2013;7:60.
24. Fung SJ, Webster MJ, Sivagnanam Sundaram S, Duncan C, Elashoff M, Weickert CS. Expression of interneuron markers in the dorsolateral prefrontal cortex of the developing human and in schizophrenia. *Am J Psychiatry*. 2010;167:1479–88.
25. Lewis DA. Development of the prefrontal cortex during adolescence: insights into vulnerable neural circuits in schizophrenia. *Neuropsychopharmacology*. 1997;16:385–98.
26. Parr AC, Calabro F, Larsen B, Tervo-Clemmens B, Elliot S, Foran W, et al. Dopamine-related striatal neurophysiology is associated with specialization of frontostriatal reward circuitry through adolescence. *Prog Neurobiol*. 2021;201:101997.
27. Galvan A, Hare TA, Parra CE, Penn J, Voss H, Glover G, et al. Earlier development of the accumbens relative to orbitofrontal cortex might underlie risk-taking behavior in adolescents. *J Neurosci*. 2006;26:6885–92.
28. Larsen B, Luna B. In vivo evidence of neurophysiological maturation of the human adolescent striatum. *Dev Cogn Neurosci*. 2014;12C:74–85.
29. Larsen B, Olafsson V, Calabro F, Laymon C, Tervo-Clemmens B, Campbell E, et al. Maturation of the human striatal dopamine system revealed by PET and quantitative MRI. *Nat Commun*. 2020;11:846.
30. Wahlstrom D, White T, Luciana M. Neurobehavioral evidence for changes in dopamine system activity during adolescence. *Neurosci Biobehav Rev*. 2010;34:631–48.
31. Luciana M, Wahlstrom D, Porter JN, Collins PF. Dopaminergic modulation of incentive motivation in adolescence: age-related changes in signaling, individual differences, and implications for the development of self-regulation. *Dev Psychol*. 2012;48:844–61.
32. Tervo-Clemmens B, Calabro FJ, Parr AC, Fedor J, Foran W, Luna B. A canonical trajectory of executive function maturation from adolescence to adulthood. *Nat Commun*. 2023;14:6922.
33. Somerville LH, Hare T, Casey BJ. Frontostriatal maturation predicts cognitive control failure to appetitive cues in adolescents. *J Cogn Neurosci*. 2011;23:2123–34.
34. Liston C, Watts R, Tottenham N, Davidson MC, Niogi S, Ulug AM, et al. Frontostriatal microstructure modulates efficient recruitment of cognitive control. *Cereb Cortex*. 2006;16:553–60.
35. Sawyer SM, Azzopardi PS, Wickremarathne D, Patton GC. The age of adolescence. *Lancet Child Adolesc Health*. 2018;2:223–8.
36. Schneider M. Adolescence as a vulnerable period to alter rodent behavior. *Cell Tissue Res*. 2013;354:99–106.
37. Granata L, Gildawie KR, Ismail N, Brenhouse HC, Kopec AM. Immune signaling as a node of interaction between systems that sex-specifically develop during puberty and adolescence. *Dev Cogn Neurosci*. 2022;57:101143.
38. Spear LP. The adolescent brain and age-related behavioral manifestations. *Neurosci Biobehav Rev*. 2000;24:417–63.
39. Reynolds LM, Flores C. Mesocorticolimbic dopamine pathways across adolescence: diversity in development. *Front Neural Circuit*. 2021;15:735625.
40. Hoops D, Kyne R, Salameh S, MacGowan D, Avramescu RG, Ewing E, et al. The scheduling of adolescence with netrin-1 and UNCSC. *eLife*. 2024;12:RP88261.
41. Hollenstein T, Lougheed JP. Beyond storm and stress. *Am Psychol*. 2013;68:444–54.
42. Miller DJ, Duka T, Stimpson CD, Schapiro SJ, Baze WB, McArthur MJ, et al. Prolonged myelination in human neocortical evolution. *Proc Natl Acad Sci USA*. 2012;109:16480–5.
43. Blakemore S-J. Imaging brain development: the adolescent brain. *Neuroimage*. 2012;61:397–406.
44. Yuan M, Cross SJ, Loughlin SE, Leslie FM. Nicotine and the adolescent brain. *J Physiol*. 2015;593:3397–412.
45. Laviola G, Macrì S, Morley-Fletcher S, Adriani W. Risk-taking behavior in adolescent mice: psychobiological determinants and early epigenetic influence. *Neurosci Biobehav Rev*. 2003;27:19–31.
46. Burke AR, Miczek KA. Stress in adolescence and drugs of abuse in rodent models: role of dopamine, CRF, and HPA axis. *Psychopharmacology*. 2013;231:1557–80.
47. Yetnikoff L, Reichard RA, Schwartz ZM, Parsely KP, Zahm DS. Protracted maturation of forebrain afferent connections of the ventral tegmental area in the rat. *J Comp Neurol*. 2014;522:1031–47.
48. Yetnikoff L, Pokinko M, Arvanitogiannis A, Flores C. Adolescence: a time of transition for the phenotype of *dcc* heterozygous mice. *Psychopharmacology*. 2014;231:1705–14.
49. Reynolds LM, Yetnikoff L, Pokinko M, Wodzinski M, Epelbaum JG, Lambert LC, et al. Early adolescence is a critical period for the maturation of inhibitory behavior. *Cereb Cortex*. 2019;29:3676–86.
50. Yetnikoff L, Almey A, Arvanitogiannis A, Flores C. Abolition of the behavioral phenotype of adult netrin-1 receptor deficient mice by exposure to amphetamine during the juvenile period. *Psychopharmacology*. 2011;217:505–14.
51. Reynolds LM, Hernandez G, MacGowan D, Popescu C, Nouel D, Cuesta S, et al. Amphetamine disrupts dopamine axon growth in adolescence by a sex-specific mechanism in mice. *Nat Commun*. 2023;14:4035.
52. Changeux J-P. Nicotine addiction and nicotinic receptors: lessons from genetically modified mice. *Nat Rev Neurosci*. 2010;11:389–401.
53. Dwyer JB, McQuown SC, Leslie FM. The dynamic effects of nicotine on the developing brain. *Pharmacol Ther*. 2009;122:125–39.
54. Dwyer JB, Broide RS, Leslie FM. Nicotine and brain development. *Birth Defects Res C Embryo Today*. 2008;84:30–44.
55. Doura MB, Gold AB, Keller AB, Perry DC. Adult and periadolescent rats differ in expression of nicotinic cholinergic receptor subtypes and in the response of these subtypes to chronic nicotine exposure. *Brain Res*. 2008;1215:40–52.
56. Trauth JA, Seidler FJ, McCook EC, Slotkin TA. Adolescent nicotine exposure causes persistent upregulation of nicotinic cholinergic receptors in rat brain regions. *Brain Res*. 1999;851:9–19.
57. Counotte DS, Gorounova NA, Moretti M, Smoluch MT, Irth H, Clementi F, et al. Adolescent nicotine exposure transiently increases high-affinity nicotinic receptors and modulates inhibitory synaptic transmission in rat medial prefrontal cortex. *FASEB J*. 2012;26:1810–20.
58. Cano M, Reynaga DD, Belluzzi JD, Loughlin SE, Leslie F. Chronic exposure to cigarette smoke extract upregulates nicotinic receptor binding in adult and adolescent rats. *Neuropharmacology*. 2020;181:108308.
59. Levin ED, Lawrence SS, Petro A, Horton K, Rezvani AH, Seidler FJ, et al. Adolescent vs. adult-onset nicotine self-administration in male rats: duration of

- effect and differential nicotinic receptor correlates. *Neurotoxicol Teratol.* 2007;29:458–65.
60. Renda A, Penty N, Komal P, Nashmi R. Vulnerability to nicotine self-administration in adolescent mice correlates with age-specific expression of  $\alpha 4\beta 2$  nicotinic receptors. *Neuropharmacology.* 2016;108:49–59.
  61. Kota D, Martin BR, Robinson SE, Damaj MI. Nicotine dependence and reward differ between adolescent and adult male mice. *J Pharmacol Exp Ther.* 2007;322:399–407.
  62. Britton AF, Vann RE, Robinson SE. Perinatal nicotine exposure eliminates peak in nicotinic acetylcholine receptor response in adolescent rats. *J Pharmacol Exp Ther.* 2007;320:871–6.
  63. Ribeiro-Carvalho A, Lima CS, Medeiros AH, Siqueira NR, Filgueiras CC, Manhães AC, et al. Combined exposure to nicotine and ethanol in adolescent mice: effects on the central cholinergic systems during short and long term withdrawal. *Neuroscience.* 2009;162:1174–86.
  64. Omelchenko N, Sesack SR. Laterodorsal tegmental projections to identified cell populations in the rat ventral tegmental area. *J Comp Neurol.* 2005;483:217–35.
  65. Dautan D, Souza AS, Huerta-Ocampo I, Valencia M, Assous M, Witten IB, et al. Segregated cholinergic transmission modulates dopamine neurons integrated in distinct functional circuits. *Nat Neurosci.* 2016;19:1025–33.
  66. Christensen MH, Kohlmeier KA. Age-related changes in functional postsynaptic nicotinic acetylcholine receptor subunits in neurons of the laterodorsal tegmental nucleus, a nucleus important in drug addiction. *Addict Biol.* 2016;21:267–81.
  67. Christensen MH, Ishibashi M, Nielsen ML, Leonard CS, Kohlmeier KA. Age-related changes in nicotine response of cholinergic and non-cholinergic laterodorsal tegmental neurons: Implications for the heightened adolescent susceptibility to nicotine addiction. *Neuropharmacology.* 2014;85:263–83.
  68. de Kloet SF, Mansvelder HD, De Vries TJ. Cholinergic modulation of dopamine pathways through nicotinic acetylcholine receptors. *Biochem Pharmacol.* 2015;97:425–38.
  69. Mansvelder HD, Rover MD, McGehee DS, Brussaard AB. Cholinergic modulation of dopaminergic reward areas: upstream and downstream targets of nicotine addiction. *Eur J Pharmacol.* 2003;480:117–23.
  70. Naudé J, Dongelmans M, Faure P. Nicotinic alteration of decision-making. *Neuropharmacology.* 2015;96:244–54.
  71. Naudé J, Tolu S, Dongelmans M, Torquet N, Valverde S, Rodriguez G, et al. Nicotinic receptors in the ventral tegmental area promote uncertainty-seeking. *Nat Neurosci.* 2016;19:471–8.
  72. Maskos U, Molles BE, Pons S, Besson M, Guiard BP, Guilloux JP, et al. Nicotine reinforcement and cognition restored by targeted expression of nicotinic receptors. *Nature.* 2005;436:103–7.
  73. Faure P, Tolu S, Valverde S, Naudé J. Role of nicotinic acetylcholine receptors in regulating dopamine neuron activity. *Neuroscience.* 2014;282:86–100.
  74. Fernandez SP, Brousset L, Martí F, Contesse T, Mouska X, Soiza-Reilly M, et al. Mesopontine cholinergic inputs to midbrain dopamine neurons drive stress-induced depressive-like behaviors. *Nat Commun.* 2018;9:4449.
  75. Cragg SJ. Meaningful silences: how dopamine listens to the ACh pause. *Trends Neurosci.* 2006;29:125–31.
  76. Dautan D, Huerta-Ocampo I, Witten IB, Deisseroth K, Bolam JP, Gerdjikov T, et al. A major external source of cholinergic innervation of the striatum and nucleus accumbens originates in the brainstem. *J Neurosci.* 2014;34:4509–18.
  77. Islam KUS, Blaess S. The impact of the mesoprefrontal dopaminergic system on the maturation of interneurons in the murine prefrontal cortex. *Front Neurosci.* 2024;18:1403402.
  78. Reynolds LM, Flores C. Adolescent dopamine development: connecting experience with vulnerability or resilience to psychiatric disease. In: Martin CR, Preedy VR, Rajendram R, editors. *Diagnosis, management and modeling of neurodevelopmental disorders.* Academic Press; 2021. p. 295–304; <https://doi.org/10.1016/B978-0-12-817988-8.00026-9>.
  79. Hoops D, Flores C. Making dopamine connections in adolescence. *Trends Neurosci.* 2017;40:709–19.
  80. O'Donnell P. Adolescent maturation of cortical dopamine. *Neurotox Res.* 2010;18:306–12.
  81. Peters KZ, Naneix F. The role of dopamine and endocannabinoid systems in prefrontal cortex development: adolescence as a critical period. *Front Neural Circuits.* 2022;16:939235.
  82. McCutcheon JE, Conrad KL, Carr SB, Ford KA, McGehee DS, Marinelli M. Dopamine neurons in the ventral tegmental area fire faster in adolescent rats than in adults. *J Neurophysiol.* 2012;108:1620–30.
  83. Reynolds LM, Pokinko M, Torres-Berrio A, Cuesta S, Lambert LC, Pellitero EDC, et al. DCC receptors drive prefrontal cortex maturation by determining dopamine axon targeting in adolescence. *Biol Psychiatry.* 2018;83:181–92.
  84. Vassilev P, Pantoja-Urban AH, Giroux M, Nouel D, Hernandez G, Orsini T, et al. Unique effects of social defeat stress in adolescent male mice on the Netrin-1/DCC pathway, prefrontal cortex dopamine and cognition (Social stress in adolescent vs. adult male mice). *eNeuro.* 2021;8:ENEURO.0045-21.2021.
  85. Corongiu S, Dessi C, Cadoni C. Adolescence versus adulthood: differences in basal mesolimbic and nigrostriatal dopamine transmission and response to drugs of abuse. *Addict Biol.* 2019. <https://doi.org/10.1111/adb.12721>.
  86. Eddins D, Petro A, Levin ED. Impact of acute nicotine exposure on monoaminergic systems in adolescent and adult male and female rats. *Neurotoxicol Teratol.* 2022;93:107122.
  87. Placzek AN, Zhang TA, Dani JA. Age dependent nicotinic influences over dopamine neuron synaptic plasticity. *Biochem Pharmacol.* 2009;78:686–92.
  88. Azam I, Chen Y, Leslie FM. Developmental regulation of nicotinic acetylcholine receptors within midbrain dopamine neurons. *Neuroscience.* 2007;144:1347–60.
  89. Castro EM, Lotfipour S, Leslie FM. Nicotine on the developing brain. *Pharmacol Res.* 2023;190:106716.
  90. Leslie FM. Unique, long-term effects of nicotine on adolescent brain. *Pharmacol Biochem Behav.* 2020;197:173010.
  91. Linker KE, Elabd MG, Tawadrous P, Cano M, Green KN, Wood MA, et al. Microglial activation increases cocaine self-administration following adolescent nicotine exposure. *Nat Commun.* 2020;11:306.
  92. Ren M, Lotfipour S. Nicotine gateway effects on adolescent substance use. *West J Emerg Med.* 2019;20:696–709.
  93. Reed SC, Izenwasser S. Nicotine produces long-term increases in cocaine reinforcement in adolescent but not adult rats. *Brain Res.* 2017;1654:165–70.
  94. Laviolette SR. Exploring the impact of adolescent exposure to cannabinoids and nicotine on psychiatric risk: insights from translational animal models. *Psychol Med.* 2019;1–8. <https://doi.org/10.1017/S0033291719003325>.
  95. Laviolette SR. Molecular and neuronal mechanisms underlying the effects of adolescent nicotine exposure on anxiety and mood disorders. *Neuropharmacology.* 2020;184:108411.
  96. Jobson CLM, Renard J, Szkudlarek H, Rosen LG, Pereira B, Wright DJ, et al. Adolescent nicotine exposure induces dysregulation of mesocorticolimbic activity states and depressive and anxiety-like prefrontal cortical molecular phenotypes persisting into adulthood. *Cereb Cortex.* 2018;29:3140–53.
  97. Hudson R, Green M, Wright DJ, Renard J, Jobson CEL, Jung T, et al. Adolescent nicotine induces depressive and anxiolytic effects through ERK 1-2 and Akt-GSK-3 pathways and neuronal dysregulation in the nucleus accumbens. *Addict Biol.* 2020;26:e12891.
  98. Ng THJ, Sarıkahya MH, Hudson R, Szkudlarek HJ, Pérez-Valenzuela E, Uzuneser TC, et al. Adolescent nicotine exposure induces long-term, sex-specific disturbances in mood and anxiety-related behavioral, neuronal and molecular phenotypes in the mesocorticolimbic system. *Neuropsychopharmacology.* 2024;49:1171–82.
  99. Iñiguez SD, Warren BL, Parise EM, Alcantara LF, Schuh B, Maffeo ML, et al. Nicotine exposure during adolescence induces a depression-like state in adulthood. *Neuropsychopharmacology.* 2009;34:1609–24.
  100. Picciotto MR, Higley MJ, Mineur YS. Acetylcholine as a neuromodulator: cholinergic signaling shapes nervous system function and behavior. *Neuron.* 2012;76:116–29.
  101. Lewis AS, Picciotto MR. High-affinity nicotinic acetylcholine receptor expression and trafficking abnormalities in psychiatric illness. *Psychopharmacology.* 2013;229:477–85.
  102. Higley MJ, Picciotto MR. Neuromodulation by acetylcholine: examples from schizophrenia and depression. *Curr Opin Neurobiol.* 2014;29:88–95.
  103. Terry AV, Jones K, Bertrand D. Nicotinic acetylcholine receptors in neurological and psychiatric diseases. *Pharmacol Res.* 2023;191:106764.
  104. Saricicek A, Esterlis I, Maloney KH, Mineur YS, Ruf BM, Muralidharan A, et al. Persistent  $\beta 2\beta 3$ -nicotinic acetylcholinergic receptor dysfunction in major depressive disorder. *Am J Psychiatry.* 2012;169:851–9.
  105. Martin-Ruiz CM, Haroutunian VH, Long P, Young AH, Davis KL, Perry EK, et al. Dementia rating and nicotinic receptor expression in the prefrontal cortex in schizophrenia. *Biol Psychiatry.* 2003;54:1222–33.
  106. Marutle A, Zhang X, Court J, Piggott M, Johnson M, Perry R, et al. Laminar distribution of nicotinic receptor subtypes in cortical regions in schizophrenia. *J Chem Neuroanat.* 2001;22:115–26.
  107. Koukouli F, Rooy M, Tziotis D, Sailor KA, O'Neill HC, Levenga J, et al. Nicotine reverses hypofrontality in animal models of addiction and schizophrenia. *Nat Med.* 2017;23:347–54.
  108. Morel C, Fernandez SP, Pantouli F, Meye FJ, Martí F, Tolu S, et al. Nicotinic receptors mediate stress-nicotine detrimental interplay via dopamine cells' activity. *Mol Psychiatry.* 2017;36:1418.
  109. Ortiz V, Campos RC, Fofo H, Fernandez SP, Barik J. Nicotinic receptors promote susceptibility to social stress in female mice linked with neuroadaptations within VTA dopamine neurons. *Neuropsychopharmacology.* 2022;47:1587–96.
  110. Mineur YS, Soares AR, Etherington IM, Abdulla ZI, Picciotto MR. Pathophysiology of nAChRs: Limbic circuits and related disorders. *Pharmacol Res.* 2023;191:106745.

111. Ananth MR, Rajebhosale P, Kim R, Talmage DA, Role LW. Basal forebrain cholinergic signalling: development, connectivity and roles in cognition. *Nat Rev Neurosci.* 2023;24:233–51.
112. Role LW, Berg DK. Nicotinic receptors in the development and modulation of CNS synapses. *Neuron.* 1996;16:1077–85.
113. Ren M, Lotfipour S, Leslie F. Unique effects of nicotine across the lifespan. *Pharmacol Biochem Behav.* 2022;214:173343.
114. Goriounova NA, Mansvelder HD. Nicotine exposure during adolescence alters the rules for prefrontal cortical synaptic plasticity during adulthood. *Front Synaptic Neurosci.* 2012;4:3.
115. Abreu-Villaça Y, Seidler FJ, Qiao D, Tate CA, Cousins MM, Thillai I, et al. Short-term adolescent nicotine exposure has immediate and persistent effects on cholinergic systems: critical periods, patterns of exposure, dose thresholds. *Neuropharmacology.* 2003;28:1935–49.
116. Slotkin TA, Cousins MM, Seidler FJ. Administration of nicotine to adolescent rats evokes regionally selective upregulation of CNS α7 nicotinic acetylcholine receptors. *Brain Res.* 2004;1030:159–63.
117. Adriani W, Spijkerman S, Deroche-Gamonet V, Laviola G, Moal ML, Smit AB, et al. Evidence for enhanced neurobehavioral vulnerability to nicotine during peradolescence in rats. *J Neurosci.* 2003;23:4712–6.
118. Kota D, Robinson SE, Damaj MI. Enhanced nicotine reward in adulthood after exposure to nicotine during early adolescence in mice. *Biochem Pharmacol.* 2009;78:873–9.
119. Slotkin TA, Ryde IT, MacKillop EA, Bodwell BE, Seidler FJ. Adolescent nicotine administration changes the responses to nicotine given subsequently in adulthood: Adenylyl cyclase cell signaling in brain regions during nicotine administration and withdrawal, and lasting effects. *Brain Res Bull.* 2008;76:522–30.
120. Trauth JA, McCook EC, Seidler FJ, Slotkin TA. Modeling adolescent nicotine exposure: effects on cholinergic systems in rat brain regions. *Brain Res.* 2000;873:18–25.
121. Wise RA. Dopamine, learning and motivation. *Nat Rev Neurosci.* 2004;5:483–94.
122. Schultz W. Behavioral dopamine signals. *Trends Neurosci.* 2007;30:203–10.
123. Bromberg-Martin ES, Matsumoto M, Hikosaka O. Dopamine in motivational control: rewarding, aversive, and alerting. *Neuron.* 2010;68:815–34.
124. Orsini CA, Moorman DE, Young JW, Setlow B, Floresco SB. Neural mechanisms regulating different forms of risk-related decision-making: insights from animal models. *Neurosci Biobehav Rev.* 2015;58:147–67.
125. Coddington LT, Dudman JT. Learning from action: reconsidering movement signaling in midbrain dopamine neuron activity. *Neuron.* 2019;104:63–77.
126. Cools R, D'Esposito M. Inverted-U-shaped dopamine actions on human working memory and cognitive control. *Biol Psychiatry.* 2011;69:e113–25.
127. Berke JD. What does dopamine mean? *Nat Neurosci.* 2018;21:1–793.
128. Volkow ND, Wise RA, Baler R. The dopamine motive system: implications for drug and food addiction. *Nat Rev Neurosci.* 2017;18:741–52.
129. Toda M, Abi-Dargham A. Dopamine hypothesis of schizophrenia: making sense of it all. *Curr Psychiatry Rep.* 2007;9:329–36.
130. Dunlop BW, Nemeroff CB. The role of dopamine in the pathophysiology of depression. *Arch Gen Psychiatry.* 2007;64:327–37.
131. Schulz J, Zimmermann J, Sorg C, Menegaux A, Brandl F. Magnetic resonance imaging of the dopamine system in schizophrenia – a scoping review. *Front Psychiatry.* 2022;13:925476.
132. Gurvich C, Rossell SL. Dopamine and cognitive control: sex-by-genotype interactions influence the capacity to switch attention. *Behav Brain Res.* 2015;281:96–101.
133. Papaleo F, Erickson L, Liu G, Chen J, Weinberger DR. Effects of sex and COMT genotype on environmentally modulated cognitive control in mice. *Proc Natl Acad Sci USA.* 2012;109:20160–5.
134. Haddon JE, Killcross S. Rat prefrontal dopamine and cognitive control: impaired and enhanced conflict performance. *Behav Neurosci.* 2011;125:344–9.
135. Baarendse PJJ, Counotte DS, O'Donnell P, Vanderschuren LJMJ. Early social experience is critical for the development of cognitive control and dopamine modulation of prefrontal cortex function. *Neuropharmacology.* 2013;38:1485–94.
136. Barth B, Portella AK, Dubé L, Meaney MJ, Silveira PP. The interplay between dopamine and environment as the biological basis for the early origins of mental health. In: Early Life Origins of Ageing and Longevity. Springer; 2019. p. 121–40. [https://doi.org/10.1007/978-3-030-24958-8\\_7](https://doi.org/10.1007/978-3-030-24958-8_7).
137. Nguyen C, Mondoloni S, Borgne TL, Centeno I, Come M, Jehl J, et al. Nicotine inhibits the VTA-to-amygdala dopamine pathway to promote anxiety. *Neuron.* 2021. <https://doi.org/10.1016/j.neuron.2021.06.013>.
138. Eddine R, Valverde S, Tolu S, Dautan D, Hay A, Morel C, et al. A concurrent excitation and inhibition of dopaminergic subpopulations in response to nicotine. *Sci Rep.* 2015;5:8184.
139. Exley R, Maubourguet N, David V, Eddine R, Evrard A, Pons S, et al. Distinct contributions of nicotinic acetylcholine receptor subunit α4 and subunit α6 to the reinforcing effects of nicotine. *Proc Natl Acad Sci USA.* 2011;108:7577–82.
140. Tolu S, Eddine R, Marti F, David V, Graupner M, Pons S, et al. Co-activation of VTA DA and GABA neurons mediates nicotine reinforcement. *Mol Psychiatry.* 2013;18:382–93.
141. Cadoni C, Felice MD, Corongiu S, Dessì C, Espa E, Melis M, et al. Role of genetic background in the effects of adolescent nicotine exposure on mesolimbic dopamine transmission. *Addict Biol.* 2019;25:e12803.
142. Vrettou M, Thalhammer SB, Svensson A-L, Dumas S, Nilsson KW, Wallén-Mackenzie Å, et al. Vesicular glutamate transporter 2 expression in the ventral tegmental area of outbred male rats following exposure to nicotine and alcohol. *Drug Alcohol Depend Rep.* 2023;8:100180.
143. Thomas AM, Ostroff A, Kimmy BA, Taormina MB, Holden WM, Kim K, et al. Adolescent nicotine exposure alters GABA<sub>A</sub> receptor signaling in the ventral tegmental area and increases adult ethanol self-administration. *Cell Rep.* 2018;23:68–77.
144. Smith RF, McDonald CG, Bergstrom HC, Ehlinger DG, Brielmaier JM. Adolescent nicotine induces persisting changes in development of neural connectivity. *Neurosci Biobehav Rev.* 2015;55:432–43.
145. Manitt C, Eng C, Pokinko M, Ryan RT, Torres-Berrío A, Lopez JP, et al. dcc orchestrates the development of the prefrontal cortex during adolescence and is altered in psychiatric patients. *Transl Psychiatry.* 2013;3:e338.
146. Counotte DS, Spijkerman S, Van de Burgwal LH, Hogenboom F, Schoffelmeer AN, De Vries TJ, et al. Long-lasting cognitive deficits resulting from adolescent nicotine exposure in rats. *Neuropharmacology.* 2009;54:299–306.
147. Goriounova NA, Mansvelder HD. Nicotine exposure during adolescence leads to short- and long-term changes in spike timing-dependent plasticity in rat prefrontal cortex. *J Neurosci.* 2012;32:10484–93.
148. Counotte DS, Goriounova NA, Li KW, Loos M, van der Schors RC, Schetters D, et al. Lasting synaptic changes underlie attention deficits caused by nicotine exposure during adolescence. *Nat Neurosci.* 2011;14:417–9.
149. Trauth JA, Seidler FJ, Ali SF, Slotkin TA. Adolescent nicotine exposure produces immediate and long-term changes in CNS noradrenergic and dopaminergic function. *Brain Res.* 2001;892:269–80.
150. Reynolds LM, Gulmez A, Fayad SL, Campos RC, Rigoni D, Nguyen C, et al. Transient nicotine exposure in early adolescent male mice freezes their dopamine circuits in an immature state. *Nat Commun.* 2024;15:9017.
151. Nolley EP, Kelley BM. Adolescent reward system perseveration due to nicotine: studies with methylphenidate. *Neurotoxicol Teratol.* 2007;29:47–56.
152. Matta SG, Balfour DJ, Benowitz NL, Boyd RT, Buccafusco JJ, Caggia AR, et al. Guidelines on nicotine dose selection for in vivo research. *Psychopharmacology.* 2007;190:269–319.
153. Allain F, Minogianis E-A, Roberts DCS, Samaha A-N. How fast and how often: the pharmacokinetics of drug use are decisive in addiction. *Neurosci Biobehav Rev.* 2015;56:166–79.
154. Alam F, Silveyra P. Sex differences in E-cigarette use and related health effects. *Int J Environ Res Public Health.* 2023;20:7079.
155. Yimsaard P, McNeill A, Yong H-H, Cummings KM, Chung-Hall J, Hawkins SS, et al. Gender differences in reasons for using electronic cigarettes and product characteristics: findings from the 2018 ITC four country smoking and vaping survey. *Nicotine Tob Res.* 2020;23:678–86.
156. Wong DN, Fan W. Ethnic and sex differences in E-cigarette use and relation to alcohol use in California adolescents: the California Health Interview Survey. *Public Health.* 2018;157:147–52.
157. Moen JK, Lee AM. Sex differences in the nicotinic acetylcholine receptor system of rodents: impacts on nicotine and alcohol reward behaviors. *Front Neurosci.* 2021;15:745783.
158. Picciotto MR. Nicotine as a modulator of behavior: beyond the inverted U. *Trends Pharmacol Sci.* 2003;24:493–9.
159. Jung Y, Hsieh LS, Lee AM, Zhou Z, Coman D, Heath CJ, et al. An epigenetic mechanism mediates developmental nicotine effects on neuronal structure and behavior. *Nat Neurosci.* 2016;19:905–14.
160. Miller AD, Blaha CD. Midbrain muscarinic receptor mechanisms underlying regulation of mesoaccumbens and nigrostriatal dopaminergic transmission in the rat. *Eur J Neurosci.* 2005;21:1837–46.
161. Forster GL, Blaha CD. Laterodorsal tegmental stimulation elicits dopamine efflux in the rat nucleus accumbens by activation of acetylcholine and glutamate receptors in the ventral tegmental area. *Eur J Neurosci.* 2000;12:3596–604.
162. Yeomans J, Forster G, Blaha C. M5 muscarinic receptors are needed for slow activation of dopamine neurons and for rewarding brain stimulation. *Life Sci.* 2001;68:2449–56.
163. Glantz S, Jeffers A, Winickoff JP. Nicotine addiction and intensity of e-cigarette use by adolescents in the US, 2014 to 2021. *JAMA Netw Open.* 2022;5:e2240671.

164. Volesky KD, Maki A, Scherf C, Watson LM, Cassol E, Villeneuve PJ. Characteristics of e-cigarette users and their perceptions of the benefits, harms and risks of e-cigarette use: survey results from a convenience sample in Ottawa, Canada. *Heal Promot Chronic Dis Prev Can.* 2016;36:130–8.
165. Coleman BN, Johnson SE, Tessman GK, Tworek C, Alexander J, Dickinson DM, et al. "It's not smoke. It's not tar. It's not 4000 chemicals. Case closed": exploring attitudes, beliefs, and perceived social norms of e-cigarette use among adult users. *Drug Alcohol Depend.* 2016;159:80–85.
166. Leventhal AM, Stone MD, Andrabí N, Barrington-Trimis J, Strong DR, Sussman S, et al. Association of e-cigarette vaping and progression to heavier patterns of cigarette smoking. *JAMA.* 2016;316:1918.
167. Kinnunen JM, Ollila H, Minkkinen J, Lindfors PL, Timberlake DS, Rimpelä AH. Nicotine matters in predicting subsequent smoking after e-cigarette experimentation: a longitudinal study among finnish adolescents. *Drug Alcohol Depend.* 2019;201:182–7.
168. Martinelli T, Candel MJM, de Vries H, Talhout R, Knapen V, van Schayck CP, et al. Exploring the gateway hypothesis of e-cigarettes and tobacco: a prospective replication study among adolescents in the Netherlands and Flanders. *Tob Control.* 2023;32:170–8.
169. Goldenson NI, Leventhal AM, Stone MD, McConnell RS, Barrington-Trimis JL. Associations of electronic cigarette nicotine concentration with subsequent cigarette smoking and vaping levels in adolescents. *JAMA Pediatr.* 2017;171:1192.

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## AUTHOR CONTRIBUTIONS

LMR, PF and JB wrote the manuscript. All authors approved its content.

## COMPETING INTERESTS

The authors declare no competing interests.

## ADDITIONAL INFORMATION

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