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5. Y. Wang *et al.*, *Phys. Rev. Lett.* **99**, 086402 (2007).
6. M. Feng, J. Lee, J. Zhao, J. T. Yates, H. Petek, *J. Am. Chem. Soc.* **129**, 12394 (2007).
7. T. Hashizume *et al.*, *Phys. Rev. Lett.* **71**, 2959 (1993).
8. K. D. Tsuei, P. D. Johnson, *Solid State Commun.* **101**, 337 (1997).
9. Experiments were carried out in an ultrahigh vacuum (base pressure $<10^{-10}$ mbar) LT-STM. Samples were prepared as described in (6). The templates for the C_{60} monomer, dimer, and wire were occasional 7.6 Å-wide unoxidized rows of single Cu atoms cutting nearly defect-free Cu(110)-(2×1)-O domains. The intermolecular distance within a wire of 10.7 ± 0.2 Å is ~7% larger than the distance in a C_{60} solid (10.05 Å). All measurements were performed at 77.3 K.
10. *dI/dV* images were acquired by positioning the scanning tunneling microscope tip at each point under constant current (engaged-loop conditions) and then measuring the *dI/dV* signal with lock-in detection (tunneling current driven at 267 Hz with 18-mV modulation of the junction bias under open-loop conditions).
11. D. B. Dougherty, P. Maksymovych, J. Lee, J. T. Yates Jr., *Phys. Rev. Lett.* **97**, 236806 (2006).
12. STM topography images were acquired in constant-current mode (0.1-nA current and 0.45-V sample potential referenced to the tip). Tunneling spectroscopy of the unoccupied states was performed by measuring the constant-current *z-V* characteristics as described in (11). Tunneling resonances are identified by the numerical differentiation of the *z-V* data. The similarity between *dz/dV* spectra of C_{60} on the C_{60} /Cu(110) and C_{60} /Cu(110)-(2×1)-O surfaces below 5 V indicates weak perturbation of the electronic structure by the oxide.
13. X. Lu, M. Grobis, K. H. Khoo, S. G. Louie, M. F. Crommie, *Phys. Rev. B* **70**, 115418 (2004).
14. X. Y. Zhu *et al.*, *Phys. Rev. B* **74**, 241401 (2006).
15. DFT calculations predict LUMO+2 and LUMO+3 to be separated by 0.17 eV and to have similar orbital distributions. Experimentally, in the 2.8 to 3.2 V range, *dz/dV* spectra and *dI/dV* imaging do not show evidence for two distinct states. Whereas we attribute our experimental observations to LUMO+2, the observed DOS may also have contributions from LUMO+3.
16. D. B. Dougherty *et al.*, *Phys. Rev. B* **76**, 125428 (2007).
17. J. P. Perdew, K. Burke, M. Ernzerhof, *Phys. Rev. Lett.* **77**, 3865 (1996).
18. D. M. Ceperley, B. J. Alder, *Phys. Rev. Lett.* **45**, 566 (1980).
19. J. L. Martins, N. Troullier, J. H. Weaver, *Chem. Phys. Lett.* **180**, 457 (1991).
20. Using the generalized gradient approximation to DFT with the Perdew-Becke-Ernzerhof functional, we calculated the electronic structure of isolated C_{60} molecules (18). The 30-by-30-by-30 Å unit cell was used to avoid the interaction between C_{60} molecules. The energy cutoff for the plane waves was 400 eV. The results were checked using localized density approximation with the Ceperley and Alder functional, and the difference was negligible (19). SAMO DOS represents the difference between the total DOS and that projected onto the atomic orbitals on C and Li atoms. Calculating the localized and delocalized states on an equal footing is best accomplished with a plane-wave basis set, as was done in (20) for a C_{60} solid. To characterize the diffuse states with an atomic basis requires an appropriately large basis set.
21. H. Shinohara, *Rep. Prog. Phys.* **63**, 843 (2000).
22. J. N. Crain, D. T. Pierce, *Science* **307**, 703 (2005).
23. P. Delaney, J. C. Greer, *Appl. Phys. Lett.* **84**, 431 (2004).
24. P. M. Echenique, J. B. Pendry, *J. Phys. C Solid State Phys.* **11**, 2065 (1978).
25. M. Posternak, A. Baldereschi, A. J. Freeman, E. Wimmer, M. Weinert, *Phys. Rev. Lett.* **50**, 761 (1983).
26. X. Blase, A. Rubio, S. G. Louie, M. L. Cohen, *Phys. Rev. B* **51**, 6868 (1995).
27. E. R. Margine, V. H. Crespi, *Phys. Rev. Lett.* **96**, 196803 (2006).
28. Y. Miyamoto, S. Saito, D. Tománek, *Phys. Rev. B* **65**, 041402 (2001).
29. M. Boyle *et al.*, *Phys. Rev. Lett.* **87**, 273401 (2001).
30. We thank J. Lee and J. T. Yates Jr. for their contributions to templated C_{60} growth, sharing their expertise, and provisioning the LT-STM. We also thank R. W. Field, J. Yang, and D. Pratt for discussions. Grants from the U.S. Department of Energy (DOE DE-FG02-03ER15434), the Petroleum Research Fund (44158-ACS), and Ikerbasque supported this research. Calculations were performed in the Environmental Molecular Sciences Laboratory at the Pacific Northwest National Laboratory, a user facility sponsored by the DOE Office of Biological and Environmental Research.

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The Chemical Genomic Portrait of Yeast: Uncovering a Phenotype for All Genes

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Genetics aims to understand the relation between genotype and phenotype. However, because complete deletion of most yeast genes (~80%) has no obvious phenotypic consequence in rich medium, it is difficult to study their functions. To uncover phenotypes for this nonessential fraction of the genome, we performed 1144 chemical genomic assays on the yeast whole-genome heterozygous and homozygous deletion collections and quantified the growth fitness of each deletion strain in the presence of chemical or environmental stress conditions. We found that 97% of gene deletions exhibited a measurable growth phenotype, suggesting that nearly all genes are essential for optimal growth in at least one condition.

Small molecules are potent probes to help understand cellular physiology [for review, see (1)]. The emergent field of chemical genomics promises that, by understanding the relations between small molecules and genes on a systems level, we might understand genomic responses to small molecule perturbants. We show that the global response of all protein-coding gene deletions tested with a panel of several hundred perturbations yields insight into gene dispensability, multidrug resistance, and gene functions within the *Saccharomyces cerevisiae* cell.

The diploid yeast deletion collections comprise ~6000 heterozygous gene deletion strains and ~5000 homozygous gene deletion strains (~1000 genes are essential) (2, 3). We tested the growth responses of these cells to over 400 small molecules and diverse environmental stresses.

Surveying a large swath of ecological space allowed us to identify genes required for growth in each tested condition. Essential genes are a potential source of new drug targets (4), whereas nonessential genes have been proposed to contribute to genetic robustness (via compensation by redundant pathways) (5, 6) or to be required for growth in particular conditions (7). Our results provide an experimental framework to test these hypotheses. We also identified previously unknown genes that function in multidrug resistance (MDR), that is, those genes required for growth in the presence of multiple drugs.

We screened small molecules from diverse sources and libraries, including drugs approved by the World Health Organization and the U.S. Food and Drug Administration, well-characterized chemical probes, and compounds with uncertain

biological activity (tables S1 and S2). The structural diversity of these compounds is comparable to that of approved drugs (fig. S1). We also assayed the effects of various environmental treatments and stresses (for example, depletion of amino acids or vitamins). We performed 726 treatment experiments in each of the heterozygous deletion strains and 418 separate experiments in each of the homozygous strains, for a total of more than 6 million single-gene measurements. These sets include some repeated experiments in which drug dose or exposure time was varied. Collapsing such repeats yielded a total of 354 unique conditions for the heterozygous collection and 178 for the homozygous collection (124 of which were tested against both collections). A gene deletion strain was defined as sensitive to a treatment if it showed a growth defect in the treatment relative to its growth in control (no drug) conditions. We defined significant sensi-

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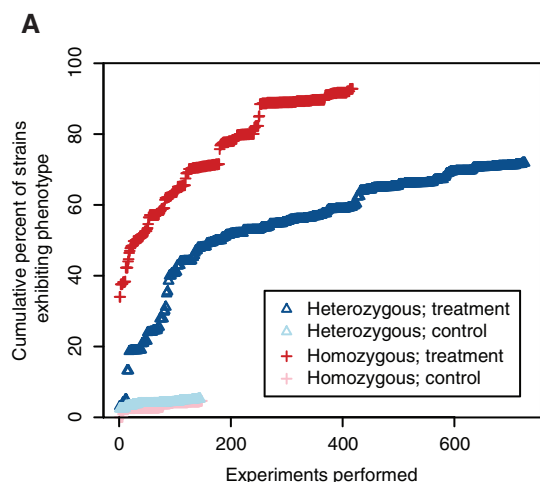
tivity and corrected for multiple comparisons by controlling the false discovery rate (FDR) to ~ 0.1 for genes exhibiting any phenotype (8) (figs. S2 and S3).

Previous studies revealed that 34% of homozygous deletion strains display a distinct phenotype (19% lethality and 15% fitness defect) when grown in rich medium (2, 3, 9). Three percent of heterozygous strains display a fitness defect (9). One interpretation of these observations is that the majority of the yeast genome is dispensable for growth. However, it is unlikely that yeast encounters such ideal conditions outside of the

laboratory. In our experiments, nearly all of the deletion strains manifested a phenotype in one or more conditions (Fig. 1).

In the experiments using homozygous deletion strains, only 416 strains (containing deletions of 7% of the genome) did not manifest a phenotype that was different from the no-drug control phenotype (hereafter “exhibited no phenotype”). Because more than half of these 7% of the genome are either of unknown function or designated as dubious (10), it is likely the majority do not encode proteins. Of the heterozygous deletion strain experiments, 2049 (34%

of the genome) exhibited no phenotype. Nearly a third of these are either of unknown function or designated as dubious (10) and are also unlikely to be protein-coding. Only 205 strains (3%) failed to exhibit a phenotype under either homozygous or heterozygous conditions. Thus, considering all ~ 6000 yeast genes, 97% manifested a significant fitness defect when deleted. In a control analysis of rich-medium experiments, only about 10% of strains manifested a false positive fitness defect [FDR of ~ 0.1 (Fig. 1A and figs. S2 and S3)]. We further tested the small set of 205 deletion strains that showed no



B Consequences of deletions:

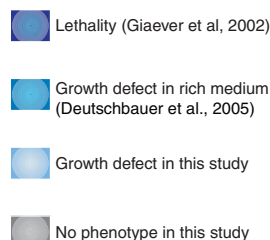


Fig. 1. Fraction of genome required for optimal growth under experimental conditions. **(A)** Percent of gene deletion strains that exhibit significant sensitivity in at least one treatment as a function of number of experiments performed, ordered by date. We used a significance threshold (z score $P < 1 \times 10^{-5}$

and $P < 1 \times 10^{-6}$ for homozygous and heterozygous experiments, respectively) that limited the FDR of genes exhibiting any phenotype to ~ 0.1 (figs. S2 and S3). Treatment experiments measure the growth of the deletion strains in a drug or altered environmental conditions; control experiments measure growth of the same deletion strains in no-drug rich medium (8).

The percent of strains exhibiting a phenotype begins at the percentage previously observed in rich medium (3% for heterozygotes and 34% for homozygotes). **(B)** Percentage of yeast genes with a phenotype under particular conditions: 19% are essential genes, 15% exhibited a growth defect as homozygous deletions in rich medium, and 63% exhibited a phenotype as either homozygous or heterozygous deletions under particular conditions in this study.

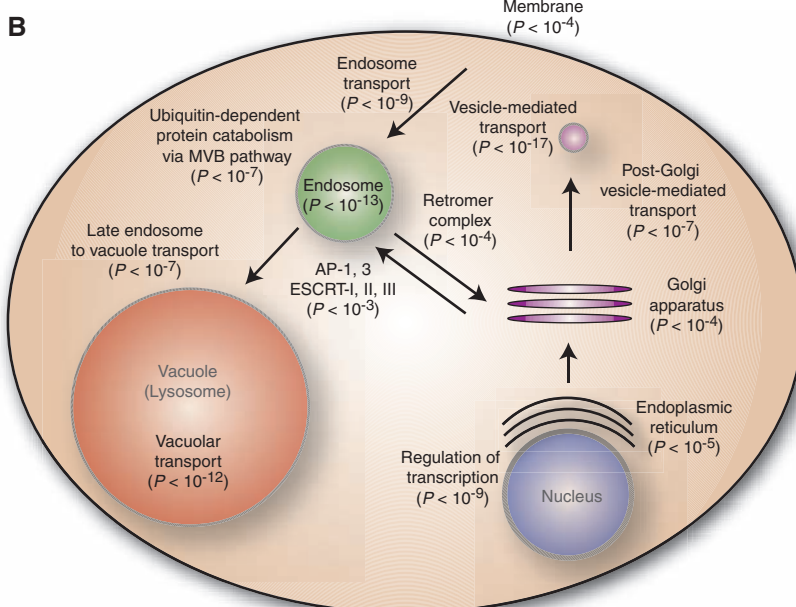
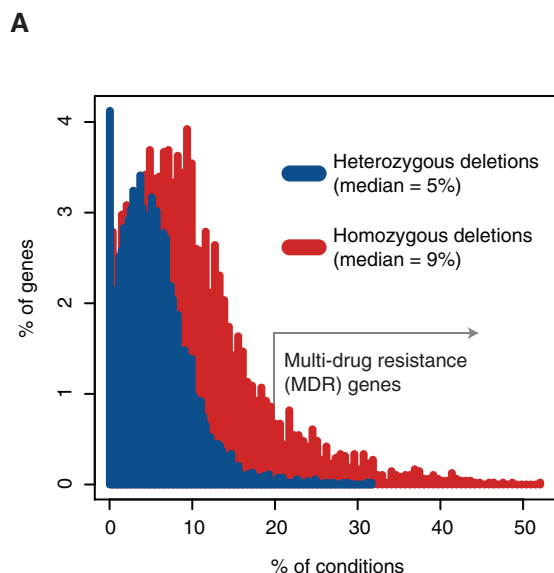


Fig. 2. Genes required for optimal growth in multiple conditions. **(A)** The percent of deletion strains inhibited by the given percent of unique conditions at $P < 0.01$ (z score). Most strains are perturbed by multiple distinct conditions. Genes whose deletion strains showed sensitivity to at least 20% of the unique small molecules are defined as MDR genes. **(B)**

Enriched functions of the homozygous MDR genes. Shown are the important GO biological processes, molecular functions, and cellular components, the hypergeometric enrichment P value, and the locations of the processes in the cell. AP, adaptor protein; ESCRT, endosomal sorting complex required for transport.

growth defect: 62 deletions do not appear to encode proteins (10), and 155 are expressed at low levels or not at all (11). This gene set was not enriched for duplicated genes (12), as might have been predicted by the redundancy hypothesis (5–7), nor were duplicated gene deletion strains sensitive in a greater or fewer number of conditions than nonduplicated strains on average (fig. S5).

Certain deletion strains were sensitive to multiple drug treatments (Fig. 2A) (13–15), and therefore the corresponding deleted genes appear to be required for resistance to diverse perturbations. We therefore refer to them as multidrug resistance (MDR) genes, with “multiple” defined as greater than 20% of all unique treatments. We predicted that MDR genes in our data would include well-characterized pleiotropic efflux pumps such as *PDR5* (human homolog is *MDR1*) and their regulators such as the transcription factor *PDR1* (16). These transporters do score as MDR in our assay, but several hundred additional genes behaved similarly: In total, 51 genes were defined as MDR as heterozygous deletion strains, and 510 genes were defined as MDR as homozygous deletion strains (Fig. 2A). These genes are highly enriched for certain Gene Ontology functions (17), particularly endosome transport, vacuolar degradation, and transcription; together these cellular processes compose a coherent system (Fig. 2B). This coordinated system of endocytosis and vacuolar or lysosomal degradation, conserved from yeast to humans, is a mechanism whereby the cell regulates transmembrane proteins, including the transporters *PDR5*, *FUR4*, and *TAT1* and signaling receptors (18). Subsets of these highly conserved pathways have been previously associated with MDR in yeast (13–15, 19). These results are consistent with recent findings in mammalian cells that MDR is correlated with changes in intracellular trafficking (20, 21), although the exact contribution of these pathways to drug resistance is not known (22–25). The most frequently observed MDR gene was *IRS4*, and although it has not previously been associated with any drug response, we found that this gene conferred resistance to 55% of the unique compounds. *IRS4* regulates phosphoinositides (PIs), well-conserved second messengers that regulate vesicular trafficking, membrane transporters, and membrane lipid composition (18). These functions largely encompassed the remaining MDR genes’ functions (Fig. 2B), suggesting that *IRS4* signaling may coordinate MDR-specific pathways. To address this, we examined the MDR genes that conferred resistance to the same compounds as *IRS4*; these genes were enriched for vesicle transport function (hypergeometric $P < 1 \times 10^{-20}$), whereas those genes not related to *IRS4* were not enriched. Additional interesting MDR genes included nine genes involved in aromatic amino acid biosynthesis (*TRP1*, *TRP2*, *TRP3*, *TRP4*, *TRP5*, *ARO1*, *ARO2*, *ARO7*, and *TKL1*) and *GCN4*, a master transcriptional regulator of amino acid biosynthesis (15). The sensi-

tivity profiles of these strains were nearly identical to one another, and the strains were sensitive to nearly 30% of all tested compounds. Lastly, although transcription factors (TFs) are underrepresented in the set of essential genes in rich media (~3% of

TFs are essential) (26), we found that 16 TFs (~10%) conferred MDR. Our compendium of conditions under which TFs are required for growth suggests experimental conditions to better characterize these proteins.

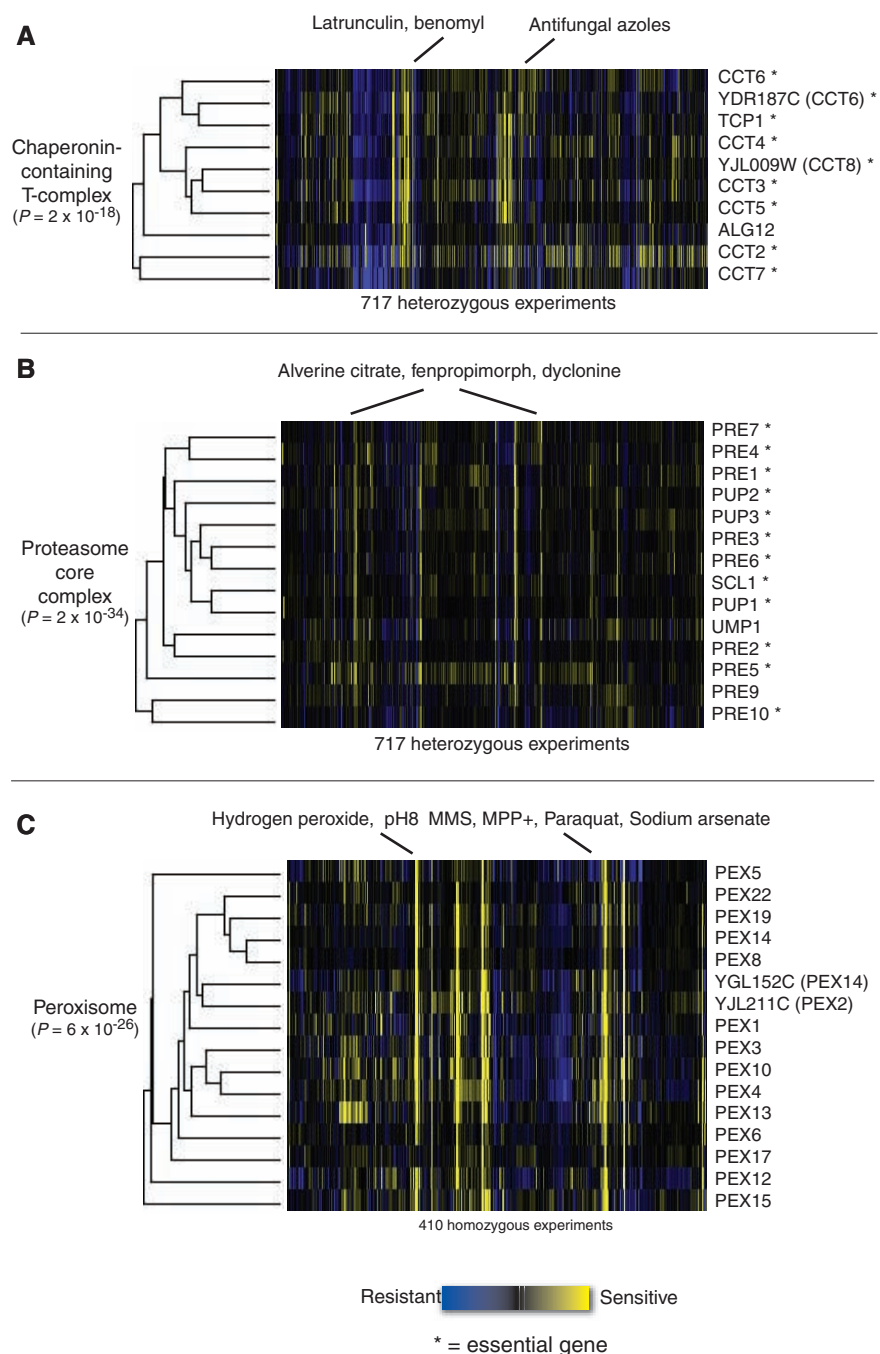


Fig. 3. Gene clusters with similar co-fitness profiles and similar biological functions, extracted from two-way hierarchical clustering on the complete data set (using all genes and all experiments). Each cell in the matrix is a sensitivity score of the deletion strain in the experiment: yellow indicates that the strain was sensitive; blue indicates resistance. Essential genes are marked with asterisks, and open reading frames (ORFs) that overlap a dubious ORF are in parentheses. (A) CCT complex genes, mediators of cytoskeletal assembly, cluster as heterozygous deletion strains primarily because of sensitivity to the cytoskeletal disrupting agents latrunculin and benomyl. (B) Genes of the proteasome core complex cluster in the heterozygous deletion assays because of sensitivity to three structurally similar compounds that target the lipid synthesis pathway. (C) Peroxisomal genes clustered as homozygous deletions because of sensitivity to hydrogen peroxide, high pH, and oxidative stress-inducing conditions.

We defined the co-fitness between any two genes as the correlation of their fitness profiles across all experiments (19). We clustered genes by this metric and found that those that are co-fit often clustered by known function, having shared Gene Ontology biological processes and molecular functions (17). Three examples of function-enriched clusters were extracted from genome-wide clustering, and they serve to validate co-fitness as an informative metric that provides biological insight into the function of these genes. One heterozygous cluster (Fig. 3A) comprised all eight chaperonin-containing T (CCT) complex genes: *TCPI*, *CCT2*, *CCT3*, *CCT4*, *CCT5*, *CCT6* (and its overlapping gene *YDR187C*), *CCT7*, and *YJL009W*, which overlaps *CCT8*. The CCT cytoskeletal-folding complex is essential and conserved between yeast and human. These strains were especially sensitive to the cytoskeletal poisons latrunculin and benomyl, illustrating cases where drugs and gene deletions act synergistically in an essential pathway to produce a phenotype. Another heterozygous deletion strain cluster included 13 of the 14 proteasome core complex genes and *UMP1*, a chaperone required for maturation of that complex (Fig. 3B). These strains were sensitive to alverine citrate, fenpropimorph, dyclonine, and dihydromotuporamine C, all of which share a nitrogen-containing structural motif and may target the lipid synthesis pathway (4). This observation suggests a possible relation between lipid synthesis and proteasome-mediated degradation. The cluster of homozygous strains shown in Fig. 3C contains 15 genes encoding components of the peroxisome, an organelle conserved from yeast to human that is responsible for breakdown of peroxides and other metabolites (27). Not surprisingly, the deletion strains were sensitive to hydrogen peroxide, presumably because the defective peroxisome could not properly metabolize it. The peroxisome requires a low pH, and the deletion strains were also sensitive to high pH. There is also growing evidence for peroxisomal involvement in managing oxidative stress (27); interestingly, the deletion strains were sensitive in the presence of chemicals that induce oxidative stress, such as MPP+ (1-methyl-4-phenylpyridinium), paraquat, and sodium arsenate.

Our finding that nearly all genes in yeast are required for wild-type growth in at least one experimental condition addresses the debate concerning the purpose of nonessential genes. The definition of essentiality is controversial, but under laboratory conditions over 80% of the genome is dispensable for life. We have found conditions that render the remaining genes essential for optimal growth in some condition and show that genetic redundancy is therefore limited in providing tolerance of the tested conditions (5–7). The chemical-genetic interactions defined here are complementary to genetic interactions (28) and should improve the resolution of genetic and chemical-genetic interaction maps, with applications such as predicting drug activities and synergies (29, 30).

References and Notes

- S. L. Schreiber, *Nat. Chem. Biol.* **1**, 64 (2005).
- E. A. Winzeler et al., *Science* **285**, 901 (1999).
- G. Giaever et al., *Nature* **418**, 387 (2002).
- G. Giaever et al., *Proc. Natl. Acad. Sci. U.S.A.* **101**, 793 (2004).
- Z. Gu et al., *Nature* **421**, 63 (2003).
- D. Deutscher, I. Meilijson, M. Kupiec, E. Rupp, *Nat. Genet.* **38**, 993 (2006).
- B. Papp, C. Pal, L. D. Hurst, *Nature* **429**, 661 (2004).
- Materials and methods are available as supporting material on Science Online and at <http://chemogenomics.stanford.edu/supplements/global>.
- A. M. Deuschbauer et al., *Genetics* **169**, 1915 (2005).
- www.yeastgenome.org/.
- L. David et al., *Proc. Natl. Acad. Sci. U.S.A.* **103**, 5320 (2006).
- M. Kellis, B. W. Birren, E. S. Lander, *Nature* **428**, 617 (2004).
- A. B. Parsons et al., *Nat. Biotechnol.* **22**, 62 (2004).
- A. B. Parsons et al., *Cell* **126**, 611 (2006).
- R. C. Fry, T. J. Begley, L. D. Samson, *Annu. Rev. Microbiol.* **59**, 357 (2005).
- H. Jungwirth, K. Kuchler, *FEBS Lett.* **580**, 1131 (2006).
- E. I. Boyle et al., *Bioinformatics* **20**, 3710 (2004).
- D. J. Katzmman, G. Odorizzi, S. D. Emr, *Nat. Rev. Mol. Cell Biol.* **3**, 893 (2002).
- A. M. Dudley, D. M. Janse, A. Tanay, R. Shamir, G. McDonald Church, *Mol. Systems Biol.* **1** (2005); www.nature.com/msb/journal/v1/n1/full/msb4100004.html.
- X. J. Liang, S. Mukherjee, D. W. Shen, F. R. Maxfield, M. M. Gottesman, *Cancer Res.* **66**, 2346 (2006).
- A. Rajagopal, S. M. Simon, *Mol. Biol. Cell* **14**, 3389 (2003).
- R. D. Bagshaw, D. J. Mahuran, J. W. Callahan, *Mol. Cell. Proteomics* **4**, 133 (2005).
- Y. Gong, M. Duvvuri, J. P. Krise, *J. Biol. Chem.* **278**, 50234 (2003).
- R. Egner, K. Kuchler, *FEBS Lett.* **378**, 177 (1996).
- J. Ferte, *Eur. J. Biochem.* **267**, 277 (2000).
- G. Chua, M. D. Robinson, Q. Morris, T. R. Hughes, *Curr. Opin. Microbiol.* **7**, 638 (2004).
- M. Schrader, H. D. Fahimi, *Histochem. Cell Biol.* **122**, 383 (2004).
- A. H. Y. Tong et al., *Science* **303**, 808 (2004).
- J. R. Sharom, D. S. Bellows, M. Tyers, *Curr. Opin. Chem. Biol.* **8**, 81 (2004).
- P. E. Blower et al., *Pharmacogenomics J.* **2**, 259 (2002).
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Supporting Online Material

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Materials and Methods
SOM Text
Figs. S1 to S11
Tables S1 to S3
References

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Wnt5a Control of Cell Polarity and Directional Movement by Polarized Redistribution of Adhesion Receptors

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Mechanisms by which Wnt pathways integrate the organization of receptors, organelles, and cytoskeletal proteins to confer cell polarity and directional cell movement are incompletely understood. We show that acute responses to Wnt5a involve recruitment of actin, myosin IIB, Frizzled 3, and melanoma cell adhesion molecule into an intracellular structure in a melanoma cell line. In the presence of a chemokine gradient, this Wnt-mediated receptor–actin–myosin polarity (W-RAMP) structure accumulates asymmetrically at the cell periphery, where it triggers membrane contractility and nuclear movement in the direction of membrane retraction. The process requires endosome trafficking, is associated with multivesicular bodies, and is regulated by Wnt5a through the small guanosine triphosphatases Rab4 and RhoB. Thus, cell-autonomous mechanisms allow Wnt5a to control cell orientation, polarity, and directional movement in response to positional cues from chemokine gradients.

Wnt signaling controls cell polarity and directional cell movement in developmental systems, as well as cell invasion in certain cancers. Features shared between noncanonical Wnt pathways include recruitment of Frizzled (Fz) receptors to the posterior end of cells, and asymmetric distribution of atypical cell adhesion molecules, often associated with Fz

(1, 2). Thus, receptors redistribute, in response to Wnt, to define an axis of asymmetry. In developmental systems, these processes can be regulated by interactions with adjacent cells, which confer orientation with respect to surrounding tissues (1, 3). For example, during endoderm specification in *Caenorhabditis elegans*, the division plane in the four-cell blastomere is determined by a positional Wnt signal from a nearby P₂ cell (4). In contrast, Wnt pathway mutations in *C. elegans* that disrupt neuronal cell migration and polarity can be rescued by Wnt overexpression without requiring a localized source of ligand (5, 6). This suggests that directional presentation of Wnt to cells is not always needed for

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