PBMC Deconvolution

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There is a good reference for PBMCs from a 2015 paper by Newman et al. They call the reference LM22. You can access this data as part of the dtangle.data package available through our webpage: http://dtangle.github.io

First we load the packages

```
library('dtangle')
library('dtangle.data')
library('limma')
```

then we can load the Newman PBMC data set

```
dset = newman_pbmc
```

then load gene expressions (data) and mixture proportions (mix)

```
data = dset$data$log
data[1:5,1:5]
```

```
## A1CF A2M A4GALT A4GNT AAAS
## GSM1587800 4.445675 4.333151 4.297844 4.293793 4.336305
## GSM1587801 4.361775 4.306877 4.263006 4.467572 4.380213
## GSM1587802 4.543298 4.328266 4.302650 4.809243 4.626695
## GSM1587803 4.568712 4.307912 4.302547 4.925474 4.329159
## GSM1587804 4.492878 4.274979 4.375282 4.738910 4.306863
```

```
mix = dset$annotation$mixture
mix[1:5,1:5]
```

```
##
               B cells memory B cells naïve Dendritic cells activated
## GSM1587800
                       0.0296
                                      0.1236
## GSM1587801
                       0.0183
                                      0.0333
                                                                       0
## GSM1587802
                       0.0407
                                      0.1509
                                                                       0
## GSM1587803
                       0.0295
                                      0.1677
                                                                       0
## GSM1587804
                       0.0469
                                      0.0840
                                                                       0
##
              Dendritic cells resting Eosinophils
## GSM1587800
                                      0
                                      0
                                                   0
## GSM1587801
## GSM1587802
                                      0
                                                   0
## GSM1587803
                                      0
                                                   0
## GSM1587804
```

the first 20 rows of data are gene exprs from heterogeneous mixtures to be deconvolved. The remaining rows are references for each of several PBMC cell types (B, NK, T, etc). Because there are so many cell types we collapse some of the sub-types into a fewer number of general leukocyte types:

```
general_types = factor(sapply(strsplit(colnames(mix)," "),"[",1))
mix = sapply(levels(general_types),function(g)rowSums(mix[,general_types==g,drop=FALSE]))
```

We can extract out which rows are pure reference samples:

```
pure_samples = lapply(1:ncol(mix),function(i)which(mix[,i]==1))
names(pure_samples) = colnames(mix)
lapply(pure_samples,head,n=2)
## Bcell.naive.1..HG.U133A...IRIS_GSE22886.GSM565308.
                                                     21
## Bcell.naive.2..HG.U133A...IRIS_GSE22886.GSM565309.
##
                                                     22
##
## $Dendritic
## DendriticCell.Control.1..HG.U133A...IRIS_GSE22886.GSM565366.
## DendriticCell.Control.2..HG.U133A...IRIS_GSE22886.GSM565367.
##
                                                              109
##
##
  $Eosinophils
  {\tt A\_MF\_2hrEosinophils\_U133A..Chtanova\_immune.A\_MF\_2hrEosinophils\_U133A.}
##
##
                                                                        124
         A_MF_ControlEosinophil..Chtanova_immune.A_MF_ControlEosinophil.
##
##
                                                                        125
##
##
  $Macrophages
  Monocyte.Day7.1..HG.U133A...IRIS_GSE22886.GSM565354.
## Monocyte.Day7.2..HG.U133A...IRIS GSE22886.GSM565355.
##
##
##
  $Mast
         A_LW_mastcellctrl_U133A..Chtanova_immune.A_LW_mastcellctrl_U133A.
##
##
  A_MF_ControlMASTCELL_U133A..Chtanova_immune.A_MF_ControlMASTCELL_U133A.
##
##
                                                                          121
##
## $Monocytes
## Monocyte.Day0.1..HG.U133A...IRIS_GSE22886.GSM565330.
##
                                                       78
  Monocyte.Day0.2..HG.U133A...IRIS_GSE22886.GSM565331.
##
##
##
## $Neutrophils
     A_LW_neutrophil_U133A..Chtanova_immune.A_LW_neutrophil_U133A.
##
##
## A MF neutrophils U133A..Chtanova immune.A MF neutrophils U133A.
##
                                                                 127
##
## $NK
  NKcell.control.1..HG.U133A...IRIS_GSE22886.GSM565293.
                                                        63
## NKcell.control.2..HG.U133A...IRIS_GSE22886.GSM565294.
##
                                                        64
##
## $Plasma
## PlasmaCell.FromPBMC.1..HG.U133A...IRIS_GSE22886.GSM565323.
```

```
##
                                                              36
## PlasmaCell.FromPBMC.2..HG.U133A...IRIS_GSE22886.GSM565324.
##
                                                              37
##
## $T
  CD8Tcell.NO.1..HG.U133A...IRIS_GSE22886.GSM565269.
##
##
## CD8Tcell.NO.2..HG.U133A...IRIS_GSE22886.GSM565270.
##
and use those to deconvolve the other samples
dt = dtangle(Y=data,pure_samples=pure_samples,n_choose = 100,data_type='microarray-gene')
matplot(mix[-unlist(pure_samples),],dt$estimates[-unlist(pure_samples),],xlab="truth",ylab="estimate",
        vlim=c(0,1), xlim=c(0,1)); abline(coef=c(0,1), col='orange')
     0.8
     9
estimate
     o.
     0.4
                                                       00 00 00 00
                             8 680
     0.2
     0
            0.0
                          0.2
                                        0.4
                                                       0.6
                                                                     0.8
                                                                                   1.0
```

Since the cell types in the mixtures are known in this case we can subset the cell types to only look for to those cell types that we know exist in the data. First we determine what cell types are present and subset the data appropriately,

truth

and then run dtangle on the subsetted data

