

Estimating Cell-type Proportions Using Gene Expressions

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December 12, 2017

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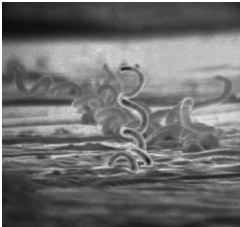
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Understanding The Immune Response to Lyme



Adult deer tick.
Scott Bauer [1].



A typical spirochete.
CDC/Dr. David Cox [2].

Lyme disease: bacterial infection spread by ticks.

1. treatable with antibiotics
2. patients report fatigue, arthritis, muscle soreness and memory problems
3. can lead to worse conditions like Lyme encephalopathy, insomnia, or depression

Bouquet et al: try to understand the immune progression of Lyme.

Study WBCs to Understand Immune Response to Lyme

Bouquet et al: collect gene expression measurements or “profiles” (GEPs) of white blood cells (WBCs) of

1. 28 Lyme patients
2. and 13 healthy controls.

The analysis compares GEPs across groups.

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Understanding these sub-types would be helpful:

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Problem: estimate the cell-type proportions of the samples using the gene expression data.

Gene Expression Data

“Gene expression measurements” = What genes the cells are using

Measure expression using mRNA:

Gene Expressed → mRNA transcript created

Gene 1 → mRNA 1 ①

Gene 2 → mRNA 2 ②

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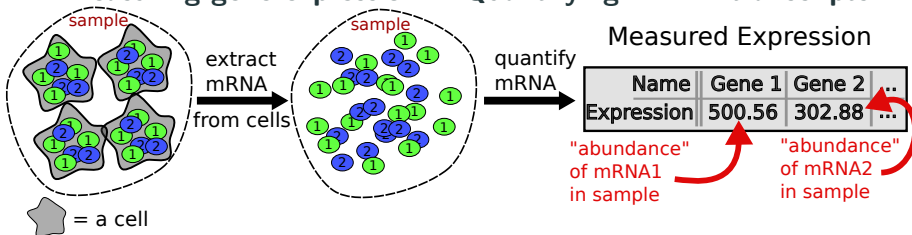
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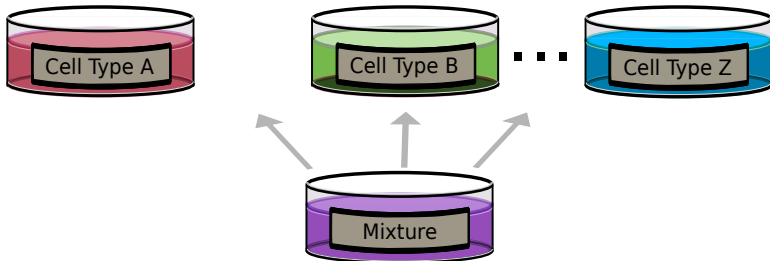
Measuring gene expression = Quantifying mRNA transcripts



Estimating Cell-type Proportions

Given: Gene Expression Profiles (GEPs) of:

1. sample that is mixture of cell types A,B,C,...Z
2. reference samples of types A,B,C,...,Z



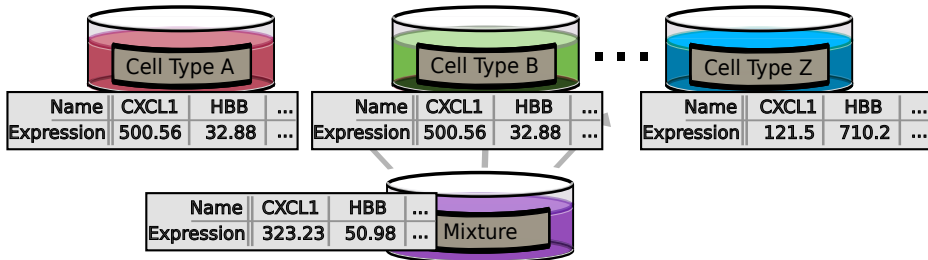
Goal: estimate cell-type proportions

	Type A	Type B	...	Type Y	Type Z
Mixture	5%	20%	...	30%	0%

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Previous Work

A Linear Model is Commonly Used

General model: $M \approx PR$, predict P with known M and R

$$\underbrace{\begin{array}{c} \text{M} \\ \text{gene}_1 \text{ gene}_2 \cdots \text{gene}_G \\ \begin{bmatrix} \text{sample}_1 & 6.6 & 8.9 & \cdots & 3.5 \\ \text{sample}_2 & 3.2 & 5.4 & \cdots & 4.8 \\ \text{sample}_3 & 7.3 & 7.7 & & \\ \vdots & \vdots & & \ddots & \\ \text{samples}_s & 4.1 & & & \end{bmatrix} \end{array}}_{\text{mixture expressions}} \approx \underbrace{\begin{array}{c} \text{P} \\ \text{type}_1 \text{ type}_2 \cdots \text{type}_K \\ \begin{bmatrix} \text{sample}_1 & .5 & .2 & \cdots & .1 \\ \text{sample}_2 & 0 & .01 & \cdots & .95 \\ \text{sample}_3 & .35 & .45 & & 0 \\ \vdots & \vdots & & \ddots & \\ \text{samples}_s & .1 & & & \end{bmatrix} \end{array}}_{\text{mixing proportions}} \underbrace{\begin{array}{c} \text{R} \\ \text{gene}_1 \text{ gene}_2 \cdots \text{gene}_G \\ \begin{bmatrix} \text{type}_1 & 9.3 & 4.1 & \cdots & 3.6 \\ \text{type}_2 & 3.7 & 5.4 & \cdots & 9.3 \\ \text{type}_3 & 2.9 & 3.6 & & \\ \vdots & \vdots & & \ddots & \\ \text{type}_K & 8.6 & & & \end{bmatrix} \end{array}}_{\text{reference expressions}}$$

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Solutions:

1. **Regression:** regress M on R .
(Abbas *et al.*; Gong *et al.*; Lu *et al.*; Wang *et al.*; Qiao *et al.*; Altboum *et al.*; Newman *et al.*)
2. **Bayesian:** Similar to LDA. Estimate as MAP.
(Quon and Morris; Qiao *et al.*; Quon *et al.*)

Marker Genes are Genes Expressed in Only One Cell Type

A **marker gene** is one which is predominantly expressed in one cell type and not the others.

Main Idea: Find marker genes for each cell type. Incorporate them in the model.

1. Can be as simple as fitting using sub-matrices.
2. Many different ways to select markers. Usually chosen by looking at **reference samples**.

Empirically models have better fit if restricted to marker genes.

dtangle

a new cell-type proportion estimator

dtangle in a Simple Setting

1. Two cell types: A and B

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$$\hat{p}_A = \text{logistic}_2 \left(\frac{(M_a - R_{Aa})}{\hat{\gamma}} - \frac{(M_b - R_{Bb})}{\hat{\gamma}} \right)$$

and similarly for p_B where $\text{logistic}_2(x) = 1/(1 + 2^{-x})$, and $\hat{\gamma}$ is a sensitivity parameter.

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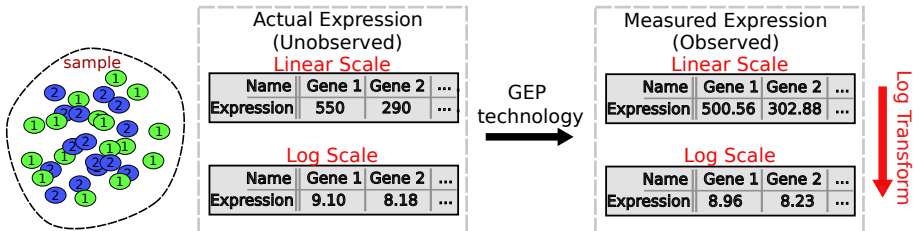
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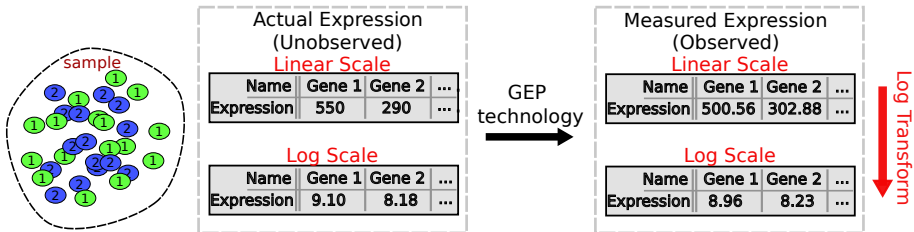
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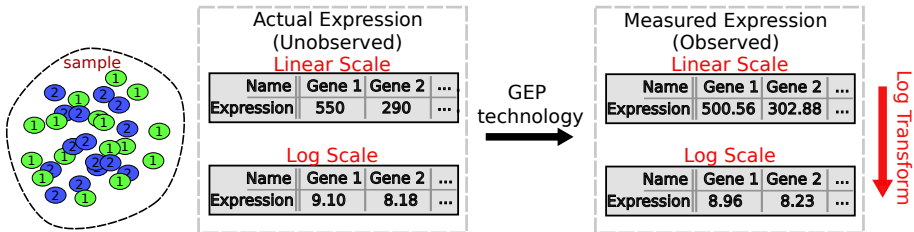


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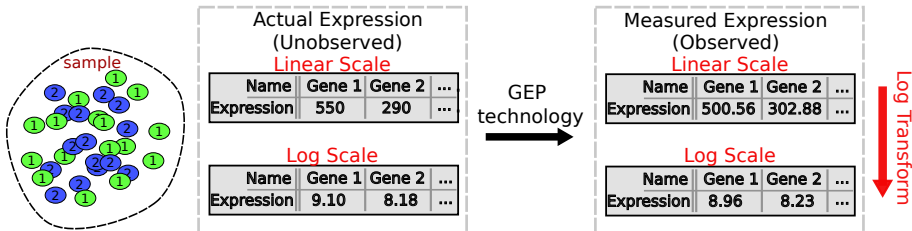
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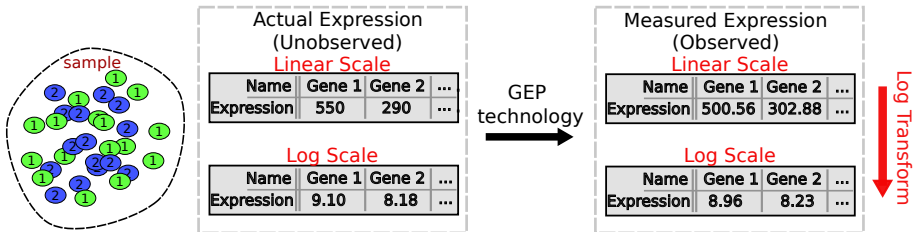
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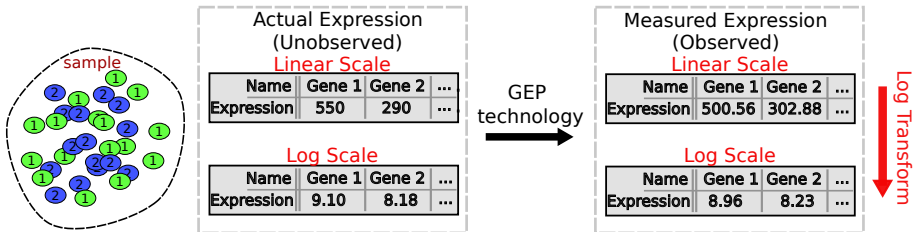
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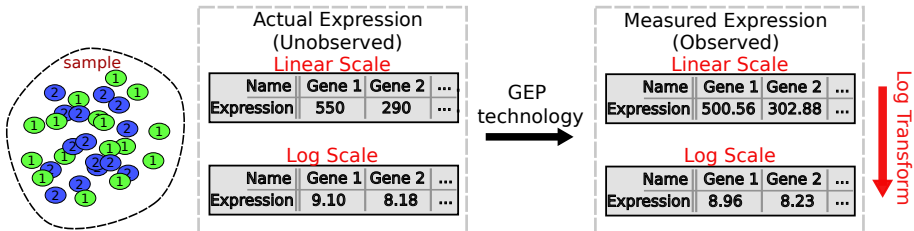
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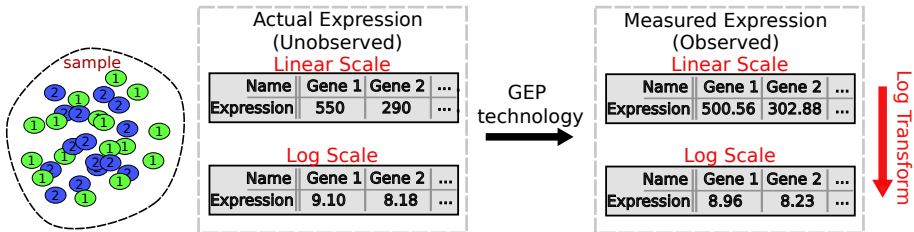
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(Step 1) dtangle Models Actual Expression Mixing

Existing approach: model mixing of **measured** expressions:

$$M_g = p_A R_{Ag} + p_B R_{Bg}$$

on either the log or linear scale.

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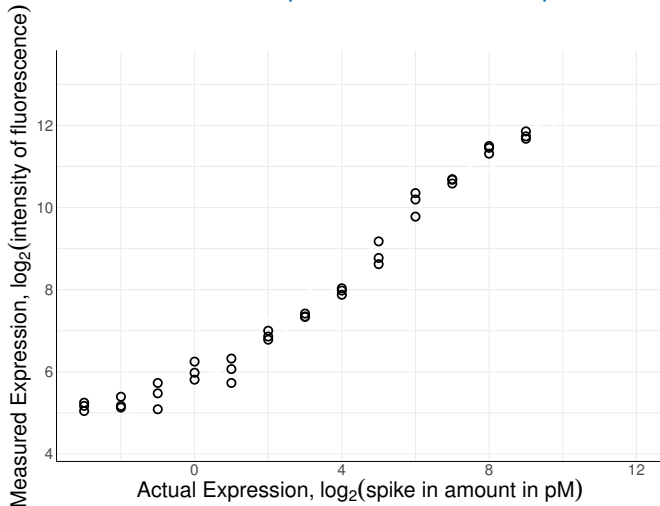
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Compare:

$$M_g = \log_2 (\text{measured expression of gene } g \text{ in mixture sample})$$

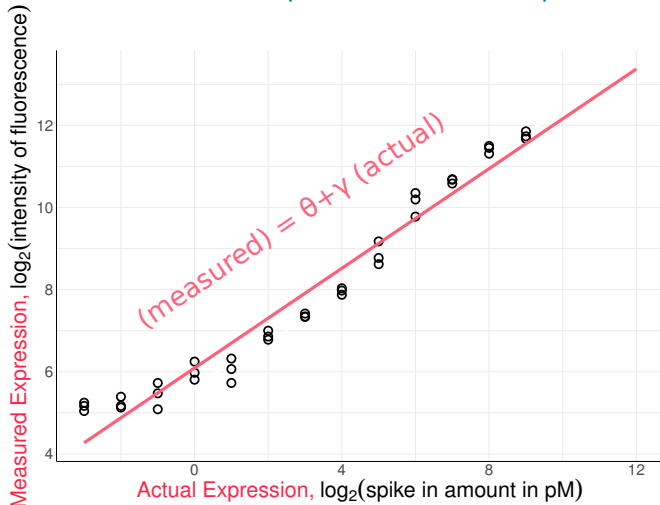
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We model **measured expression** on **actual expression** as linear:



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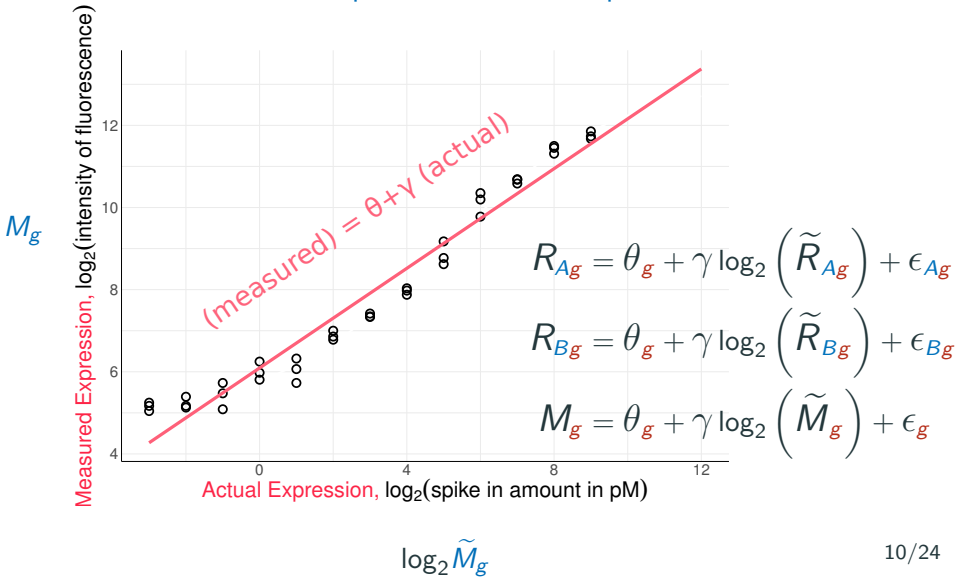
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$$\log_2 \tilde{M}_g$$

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(Step 3) dtangle Precisely Defines Marker Genes

(Defn) Marker gene: actually expressed in only one type.

$$\tilde{R}_{Ab} = 0 \text{ and } \tilde{R}_{Ba} = 0.$$

i.e. the actual expression of *a* in ref *B* is zero
and the actual expression of *b* in ref *A* is zero

Combining dtangle's Models

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$$\frac{M_a - R_{Aa}}{\gamma} = \log_2(p_A) + \epsilon$$

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$$\exp_2 \left(\frac{M_a - R_{Aa}}{\gamma} \right) \approx p_A$$

dtangle: A Re-normalization of the Exponential Terms.

We can show that,

$$p_A \approx \exp_2 \left(\frac{M_a - R_{Aa}}{\gamma} \right) \text{ and } p_B \approx \exp_2 \left(\frac{M_b - R_{Bb}}{\gamma} \right)$$

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$$\begin{aligned} \widehat{p}_A &= \frac{\exp_2 \left(\frac{M_a - R_{Aa}}{\gamma} \right)}{\exp_2 \left(\frac{M_a - R_{Aa}}{\gamma} \right) + \exp_2 \left(\frac{M_b - R_{Bb}}{\gamma} \right)} \\ &= \text{logistic}_2 \left(\frac{(M_a - R_{Aa})}{\gamma} - \frac{(M_b - R_{Bb})}{\gamma} \right) \end{aligned}$$

dtangle: A Re-normalization of the Exponential Terms.

We can show that,

$$p_A \approx \exp_2 \left(\frac{M_a - R_{Aa}}{\gamma} \right) \text{ and } p_B \approx \exp_2 \left(\frac{M_b - R_{Bb}}{\gamma} \right)$$

they are not nice since

1. they are not bounded above by 1
2. they do not sum to 1.

We can fix this by re-normalizing each by their sum:

$$\begin{aligned} \hat{p}_A &= \frac{\exp_2 \left(\frac{M_a - R_{Aa}}{\hat{\gamma}} \right)}{\exp_2 \left(\frac{M_a - R_{Aa}}{\hat{\gamma}} \right) + \exp_2 \left(\frac{M_b - R_{Bb}}{\hat{\gamma}} \right)} \\ &= \text{logistic}_2 \left(\frac{(M_a - R_{Aa})}{\hat{\gamma}} - \frac{(M_b - R_{Bb})}{\hat{\gamma}} \right) \end{aligned}$$

dtangle is Generalizable

The general setting: (1) K cell types, (2) ν_k reference samples of type k , (3) set of marker genes G_k for each cell type. Want to estimate mixing proportions p_1, \dots, p_K . For the simple case we had

$$\widehat{p}_A = \text{logistic}_2 \left(\frac{(M_a - R_{Aa})}{\widehat{\gamma}} - \frac{(M_b - R_{Bb})}{\widehat{\gamma}} \right)$$

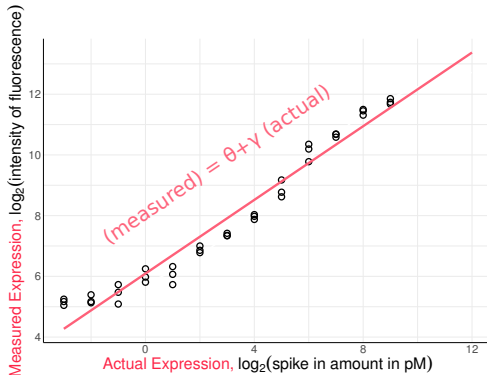
there is a direct generalization

$$\widehat{p}_k = L_k \left(\frac{(\overline{M_{G_k}} - \overline{R_{G_k}})}{\widehat{\gamma}} - \frac{(\overline{M_{G_1}} - \overline{R_{G_1}})}{\widehat{\gamma}}, \dots, \frac{(\overline{M_{G_k}} - \overline{R_{G_k}})}{\widehat{\gamma}} - \frac{(\overline{M_{G_K}} - \overline{R_{G_K}})}{\widehat{\gamma}} \right)$$

1. $L_k(x) = 1/(1 + \sum_{t \neq k} 2^{-x_t})$, a generalized logistic function
2. $\overline{M_{G_k}} = \frac{1}{|G_k|} \sum_{g \in G_k} M_g$, average marker genes in the mixture sample
3. $\overline{R_{G_k}} = \frac{1}{|G_k| \nu_k} \sum_{g \in G_k} \sum_{r=1}^{\nu_k} Z_{kr} g$, average marker genes in references

Marker Genes and γ

1. Estimate γ from benchmark data sets:

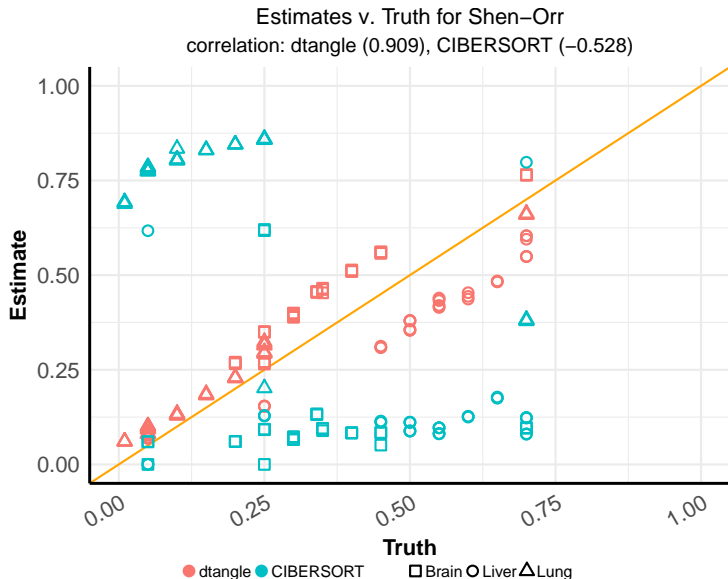


2. We find marker genes through differential expression analysis on the references.

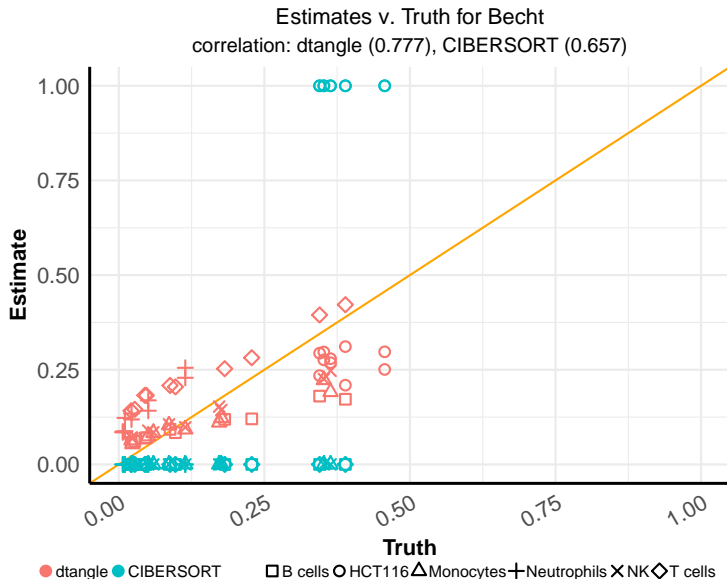
dtangle is robust to changes in γ and marker genes.

Benchmarking dtangle

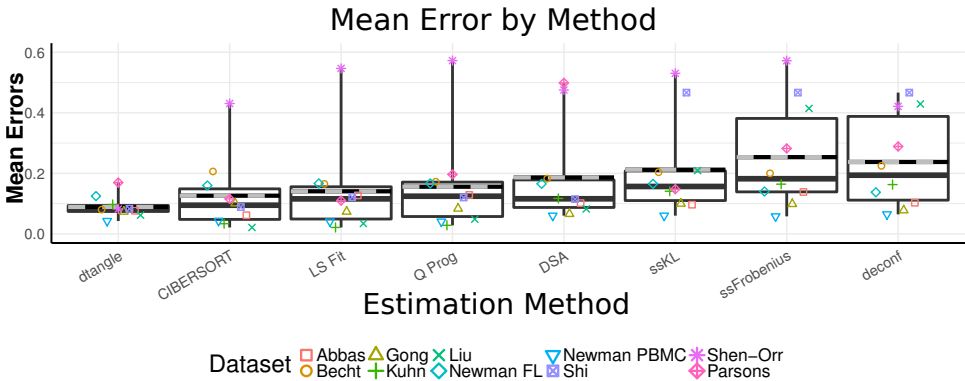
dtangle Works Well (Shen-Orr et al.)



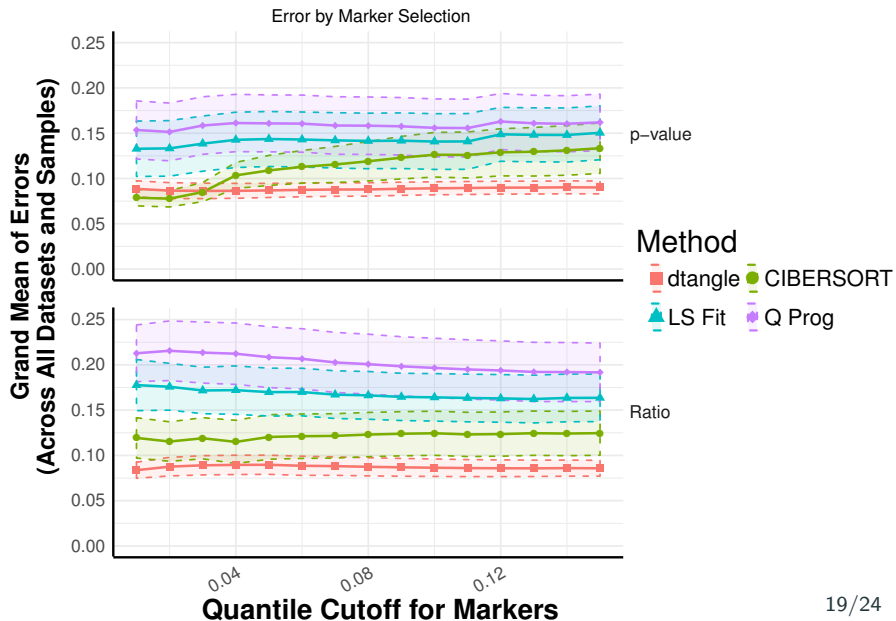
dtangle Works With Complicated Data (Becht et al.)



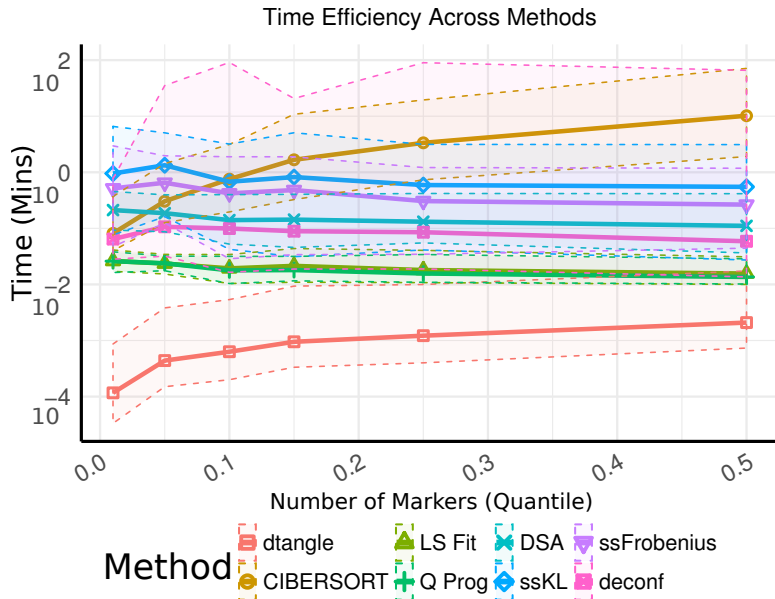
dtangle is Consistently Good



dtangle is Robust



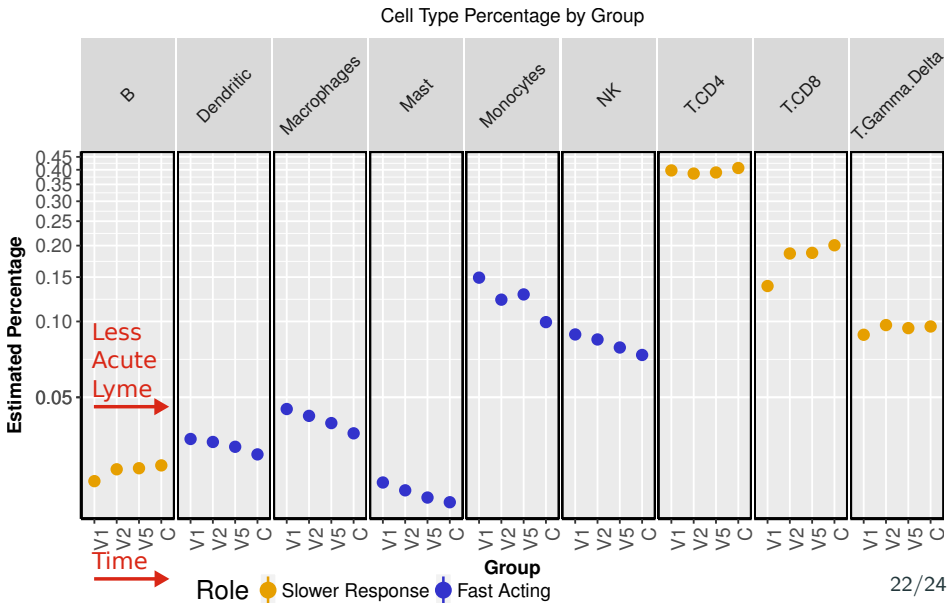
dtangle is Fast



Gene expression measurements of white blood cells from Bouquet et al.

1. Gene expression measurements of 28 patients at three points:
 - (V1) at diagnosis
 - (V2) after antibiotic treatment
 - (V5) 6 months post treatment
2. Gene expressions of 13 healthy controls (C)

dtangle on the Lyme Data



Future research directions:

1. estimating proportion of unknown cell-types
2. removing unwanted latent factors as part of estimation
3. extension to high-throughput methylation data
4. variance estimate and goodness-of-fit

dtangle is Available!

An R package is available
on github

[dtangle.github.io](https://github.com/dtangle/dtangle)

or on CRAN

cran.r-project.org/package=dtangle

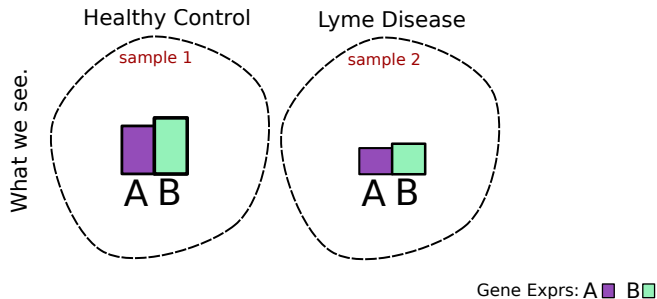
Hopefully rolling out to stemformatics soon!

www.stemformatics.org

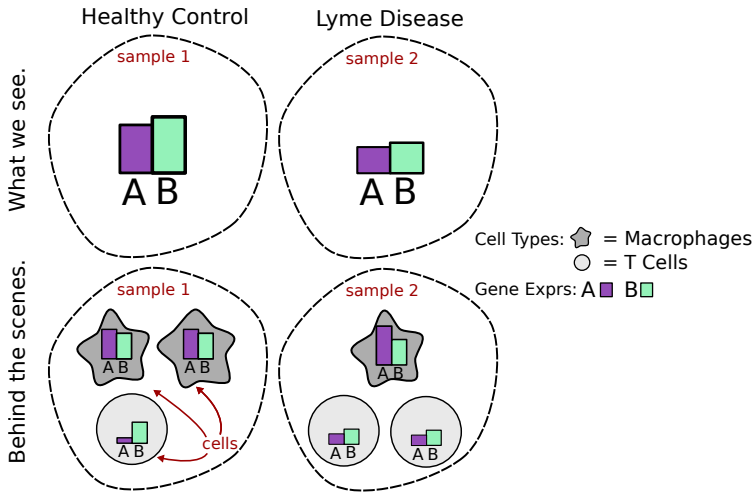
Thanks!

Extras

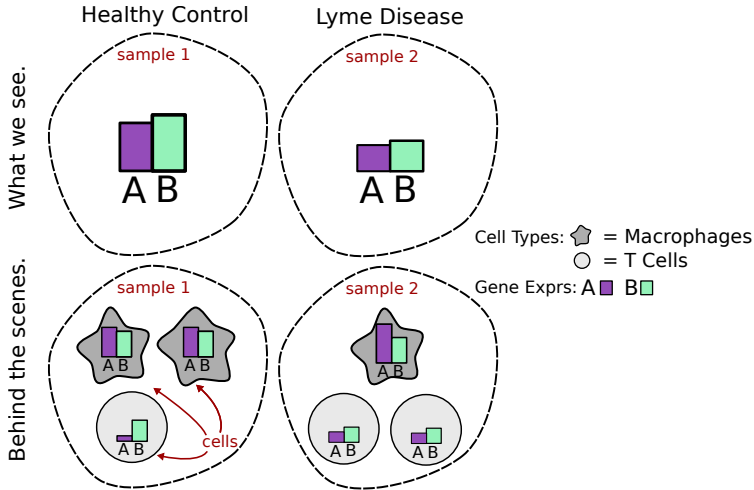
Cell-types Can Confound Differential Expression Analysis



Cell-types Can Confound Differential Expression Analysis



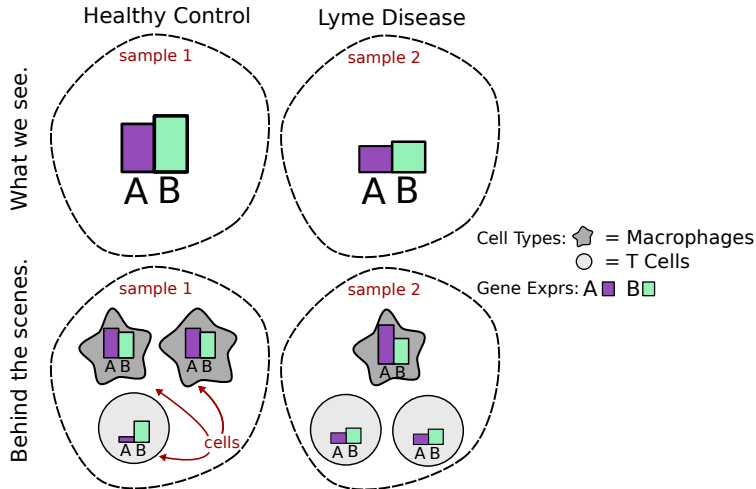
Cell-types Can Confound Differential Expression Analysis



Differences we see come from

1. differences across samples of GEPs for each cell type

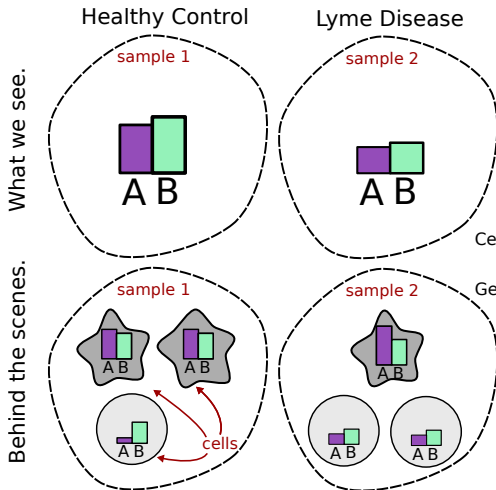
Cell-types Can Confound Differential Expression Analysis



Differences we see come from

1. differences across samples of GEPs for each cell type
2. differences across samples of cell-type composition

Cell-types Can Confound Differential Expression Analysis



Solution: Estimate the cell-type proportions. De-confound analysis with estimates.

Cell Types: ★ = Macrophages
○ = T Cells
Gene Exprs: A ■ B ■

Differences we see come from

1. differences across samples of GEPs for each cell type
2. differences across samples of cell-type composition

Accounting for Cell Types Drastically Changes Results

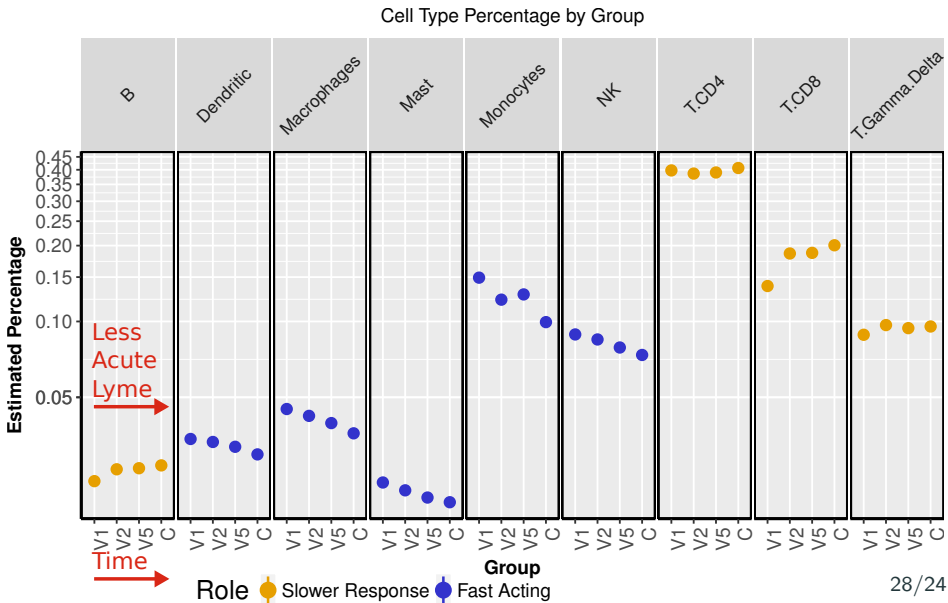
We compare the control group to Lyme patients:

1. **un-adjusted:** there are 399 differentially expressed genes
2. **cell-type adjusted:** there are 158 differentially expressed genes after adding in covariates to account for cell types

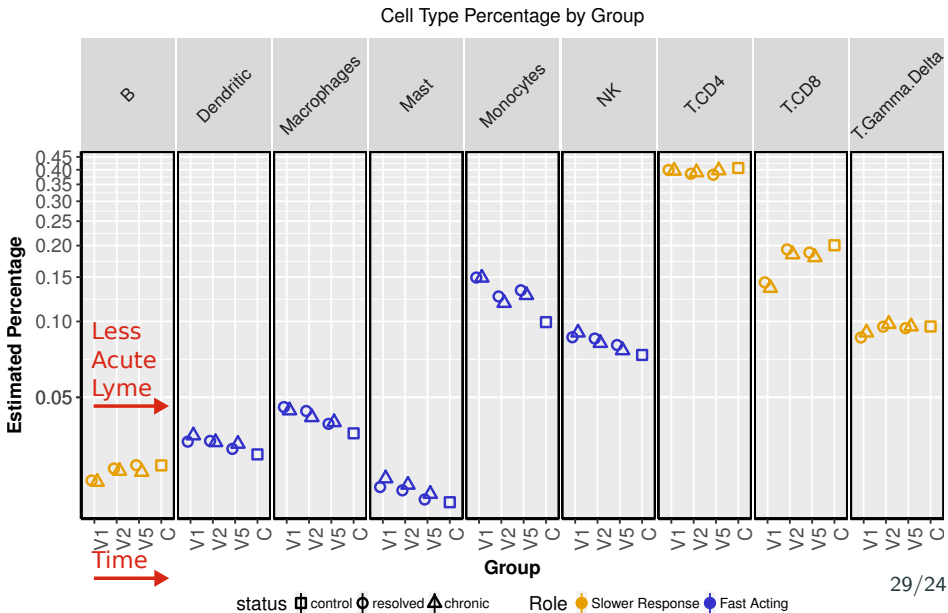
Number of diff. expressed genes changes by a factor of 2.5!

Some of the un-adjusted genes probably due to cell type.

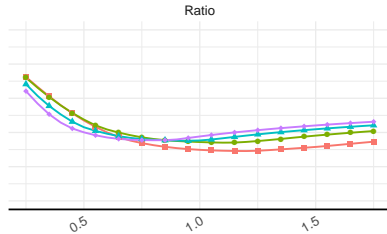
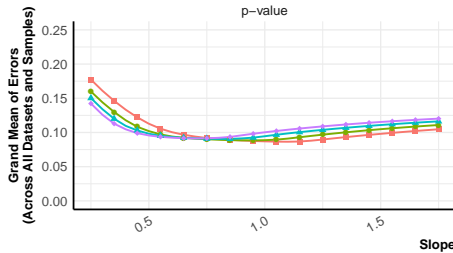
dtangle on the Lyme data



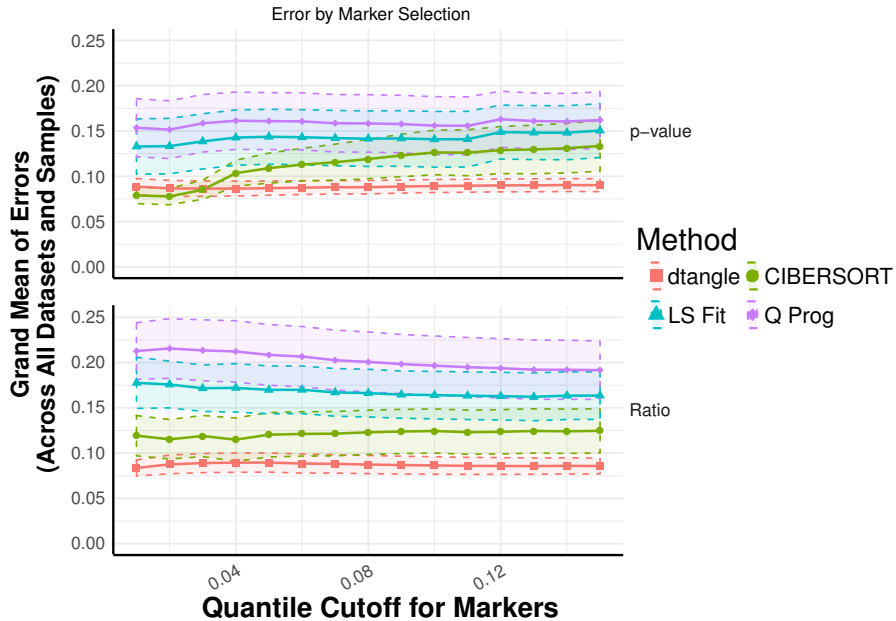
dtangle on the Lyme data



Error by Slope



Quantile 0.01 0.05 0.1 0.15



- 1 Adult deer tick, "*Ixodes scapularis*" .;Source:
<http://www.ars.usda.gov/is/graphics/photos/mar98/k8002-3.htm>;Image Number: K8002-3 ;Credits: Photo by Scott Bauer. PD-USGov-USDA-ARS
- 2 Electron micrograph of "*Treponema pallidum*". From
<http://phil.cdc.gov/phil/home.asp> ID 1977.

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