Genome Annotation Pipeline

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Description

Annotate one or more genome assemblies using BRAKER2. See the Snakefile for exact commands, the main steps implemented here are:

- Mask repeats in genome using RepeatMasker
- Run braker in mode proteins of any evolutionary distance
- Annotate proteins predicted by braker/augustus with Pfam domains using hmmer (TODO: Should we use InterProScan instead?)

As always: Ensure settings are appropriate to your case, read working program documentation, check reliability of results - This is just a pipeline.

Set up conda env

Assuming conda, bioconda, and optionally mamba have been already installed:

```
conda create --yes -n genomeAnnotationPipeline
conda activate genomeAnnotationPipeline
mamba install -n genomeAnnotationPipeline --yes --file requirements.txt
```

Additional software

Download the appropriate distribution of GeneMark-ES/ET/EP program. The downloaded file should be named something like gmes_linux_64.tar.gz).

GeneMark is free to use but its license prevents from distributing it so it has to be downloaded manually as a tar.gz bundle. The pipeline will take care of its installation. To check the kernel version of your system and download the appropriate distribution execute uname -r.

Run

Input option

• sample_sheet=

Tab-separated file indicating the genome files to be annotated and other input options. You can annotate different genome files and/or the same genome with different settings. Column are:

Column	Description
genome_id	A unique identifier for the output sub-directory of this annotation (no spaces or metacharacters)
genome_fasta protein_database	Fasta file of the genome to be annotated Fasta file of the proteins to use for training or a taxonomy identifier (e.g. Apicomplexa).
repeatmasker_species	NCBI taxonomy for option -species for repeat masker. E.g. <i>plasmodium</i> . Use NA if the genome is already masked

If protein_database is a taxonomy, the pipeline downloads OrthoDB and extracts the proteins belonging to this taxonomy. See OrthoDB for available taxonomies.

- genemark_tar_gz= Full path to genemark tar.gz download as explained above
- -d/--directory Output directory
- -n Dry-run mode omit to actually execute the workflow
- -j Number of jobs to run in parallel

Output

Most relevant files probably are:

- '{genome_id}/hmmer/augustus.hints.gff3': Predicted genes and gene features from braker/augustus annotated with Pfam domains
- {genome_id}/braker/augustus.hints.aa: Fasta file of aminoacid sequences of the mRNAs in augustus.hints.gff3
- {genome_id}/braker/augustus.hints.codingseq: Fasta file of nucleotide sequences of the mRNAs in augustus.hints.gff3

Misc (Ignore me)

Example of mapping proteins to genome using spaln:

```
spaln -W -KP ToxoDB-56_TgondiiRH.fasta
spaln -0:0 -Q7 -dToxoDB-56_TgondiiRH ToxoDB-56_TgondiiRH88_AnnotatedProteins.fasta \
> ToxoDB-56_TgondiiRH.splan.gff
```

For testing annotation of P. berghei ANKA against Apicomplexa without Pb proteins:

```
pigz -cd orthodb/odb10v1_all_fasta.tab.gz \
| ./scripts/getOrthodbProteinsForTaxonomy.py -f - \
    -l2s odb10v1_level2species.tab.gz \
    -l odb10v1_levels.tab.gz \
    -s odb10v1_species.tab.gz \
    -i 'Apicomplexa' -x 'Plasmodium berghei ANKA' > tmp/apicomplexa_wo_pbanka.fasta
```

Download T. gondii assemblies from Xia et al., 2021, NCBI accession PRJNA638608: