

# Design of RNA-Seq and Result Interpretation (II)

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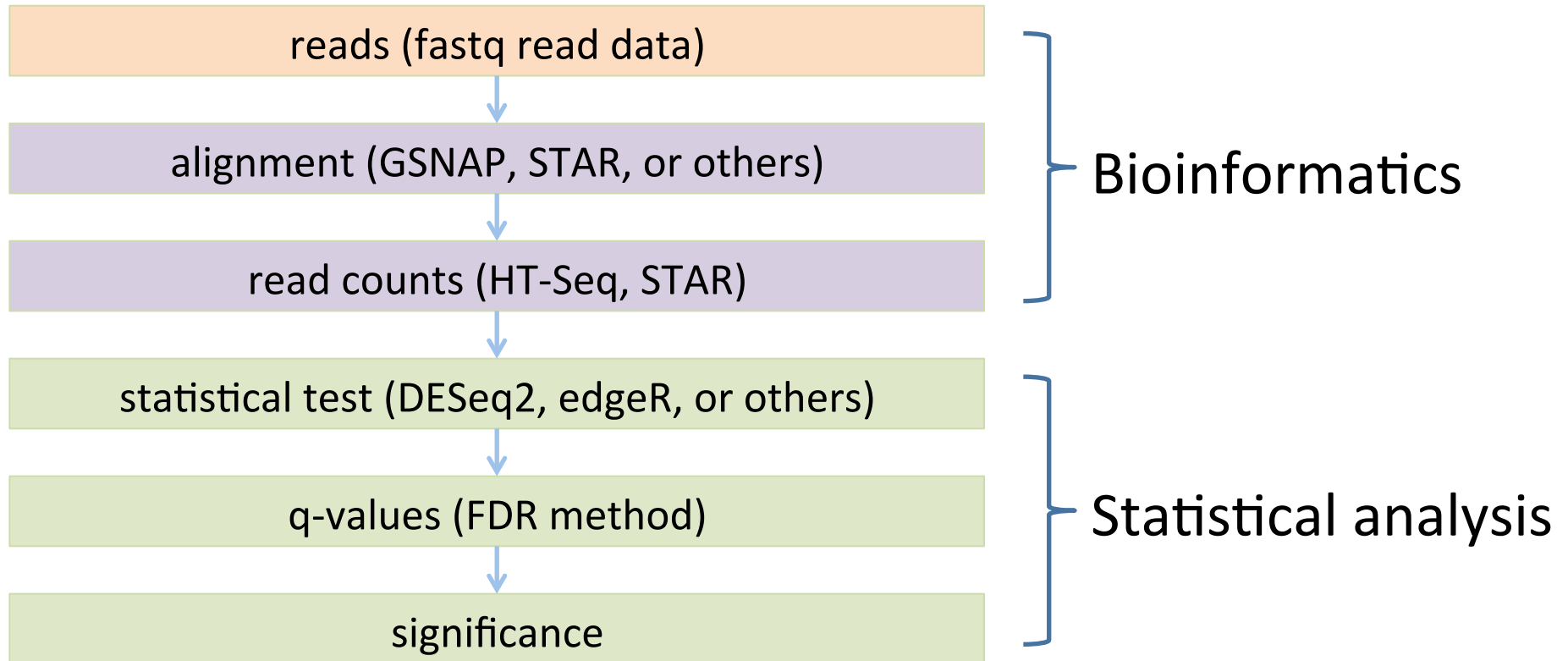
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@K-State IGF RNA-Seq Workshop (PLPTH885)

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# Bioinformatics and Statistics (Illumina data)



# STAR pipeline – from reads to counts

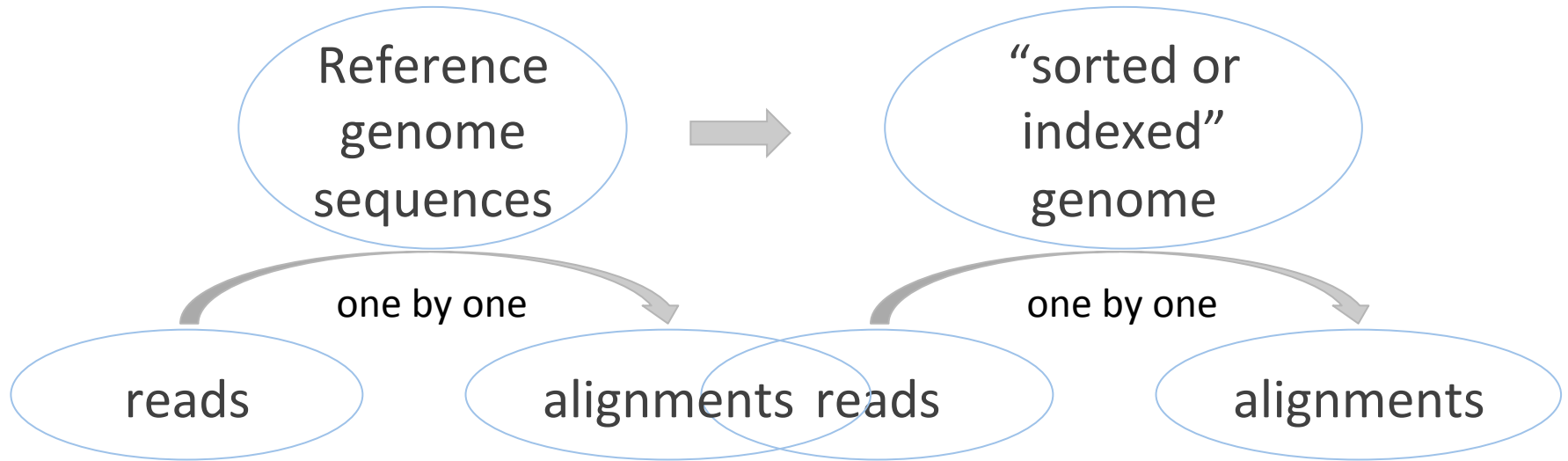
Required files:

1. Reference genome (fasta file)
2. Gene information (gff or gtf gene annotation)
3. Reads (fastq files) – your own data

Many reference genomes and gff/gtf files are available at:  
<http://ensembl.org/info/data/ftp>

Species	DNA (FASTA)	cDNA (FASTA)	CDS (FASTA)	ncRNA (FASTA)	Protein sequence (FASTA)	Annotated sequence (EMBL)	Annotated sequence (GenBank)	Gene sets
<a href="#">Human</a> <i>Homo sapiens</i>	<a href="#">FASTA</a>	<a href="#">FASTA</a>	<a href="#">FASTA</a>	<a href="#">FASTA</a>	<a href="#">FASTA</a>	<a href="#">EMBL</a>	<a href="#">GenBank</a>	<a href="#">GTF</a> <a href="#">GFF3</a>
<a href="#">Mouse</a> <i>Mus musculus</i>	<a href="#">FASTA</a>	<a href="#">FASTA</a>	<a href="#">FASTA</a>	<a href="#">FASTA</a>	<a href="#">FASTA</a>	<a href="#">EMBL</a>	<a href="#">GenBank</a>	<a href="#">GTF</a> <a href="#">GFF3</a>
<a href="#">Zebrafish</a> <i>Danio rerio</i>	<a href="#">FASTA</a>	<a href="#">FASTA</a>	<a href="#">FASTA</a>	<a href="#">FASTA</a>	<a href="#">FASTA</a>	<a href="#">EMBL</a>	<a href="#">GenBank</a>	<a href="#">GTF</a> <a href="#">GFF3</a>

# Reads to counts - reference indexing



```
STAR --runMode genomeGenerate \  
--genomeDir . \  
--genomeFastaFiles reference.fas \  
--sjdbGTFfile genes.gtf \  
--runThreadN 48
```

# Reads to counts – alignment and read counting

```
STAR --genomeDir reference.fas \  
    --readFilesIn read1.fq read2.fq \  
    --alignIntronMax 100000 \  
    --alignMatesGapMax 100000 \  
    --outFileNamePrefix output \  
    --outSAMattrIHstart 0 \  
    --outSAMmultNmax 1 \  
    --outSAMstrandField intronMotif \  
    --outFilterIntronMotifs RemoveNoncanonicalUnannotated \  
    --outSAMtype BAM SortedByCoordinate \  
    --quantMode GeneCounts \  
    --outFilterMismatchNmax 5 \  
    --outFilterMismatchNoverLmax 0.05 \  
    --outFilterMatchNmin 50 \  
    --outSJfilterReads Unique \  
    --outFilterMultimapNmax 1 \  
    --outSAMmapqUnique 60 \  
    --outFilterMultimapScoreRange 2
```

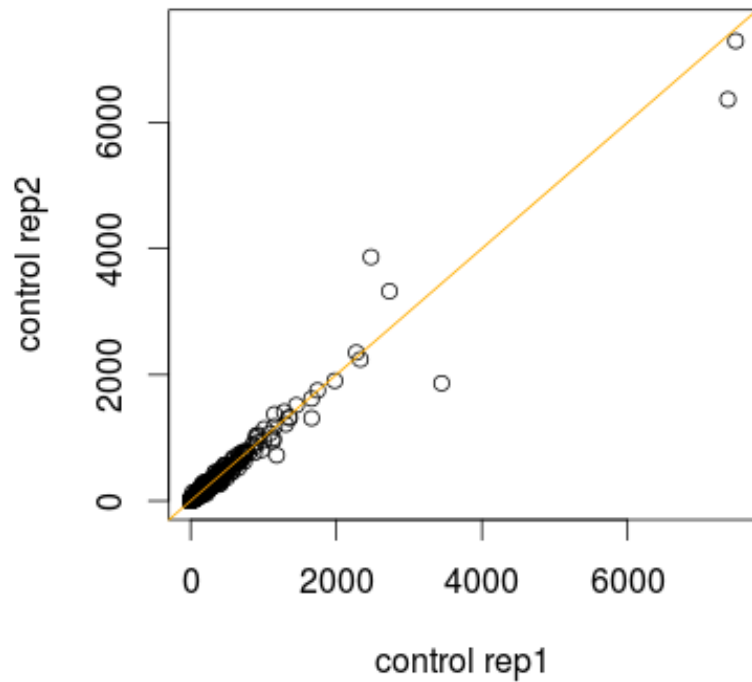
# Count matrix: Read counts (Raw) per gene

Gene	sample 1	sample 2	sample 3
gene 1	6,075	5,934	3,370
gene 2	295	377	169
...	...	...	...

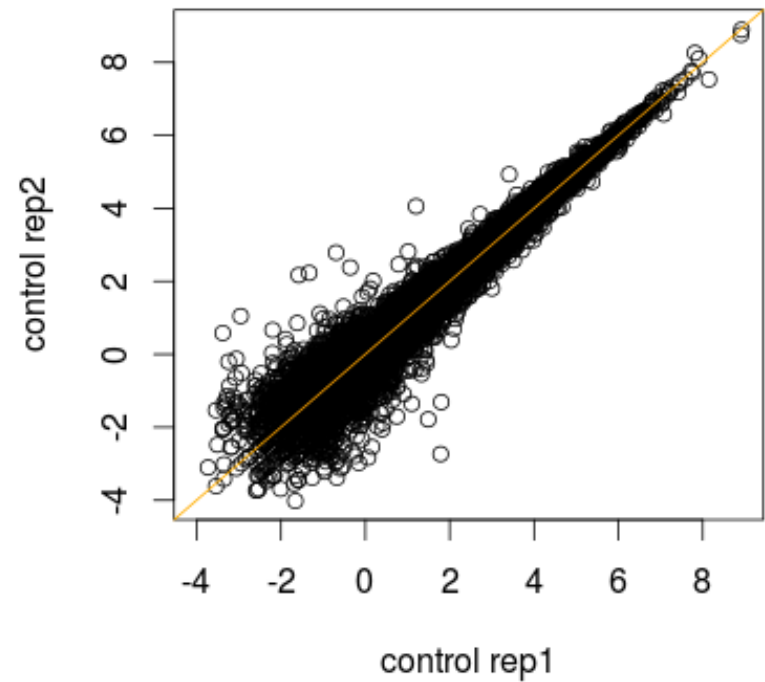
Overall difference of read counts  
among samples

# Scatter plot

RPKM scatter plot

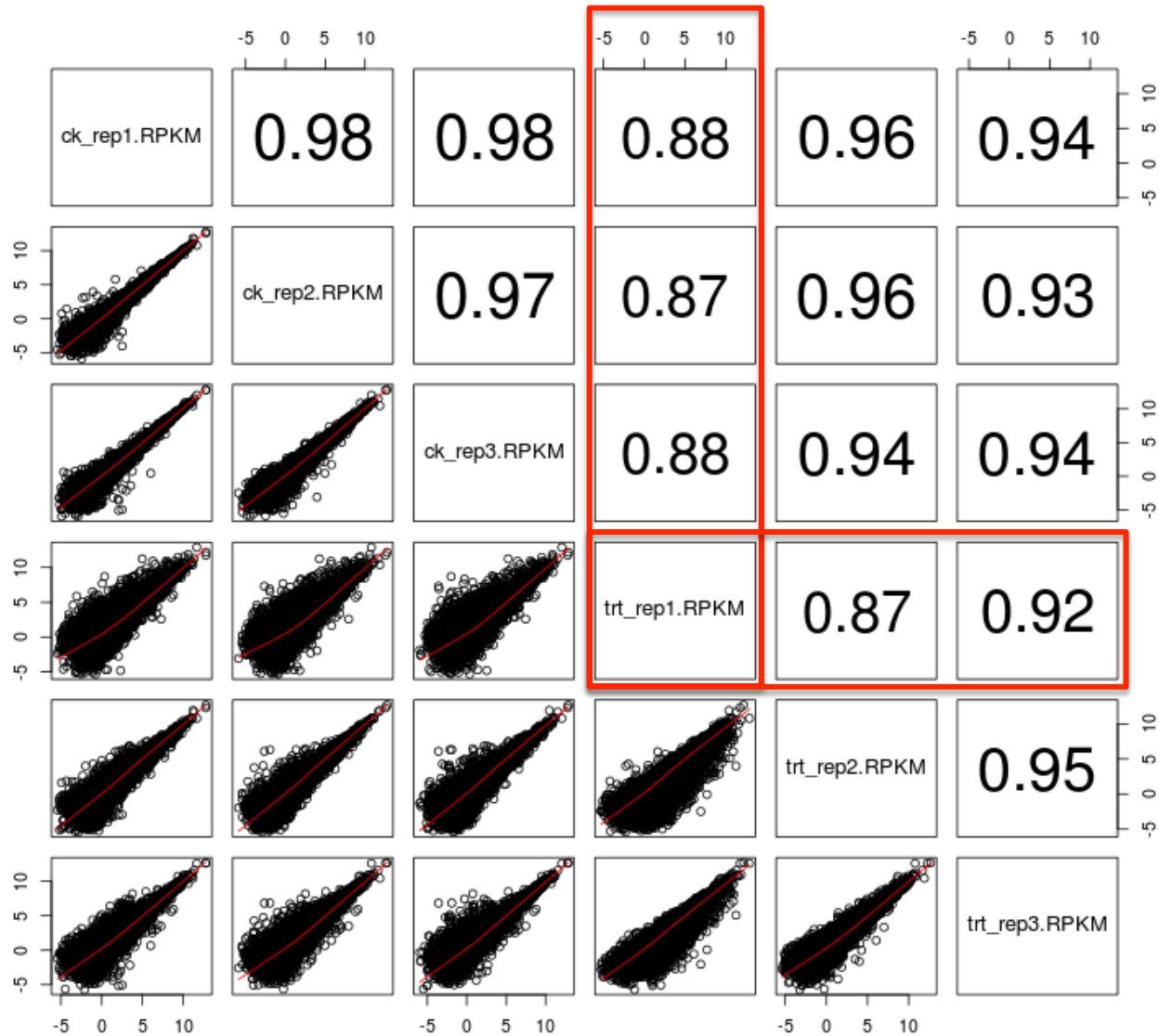


Log RPKM scatter plot





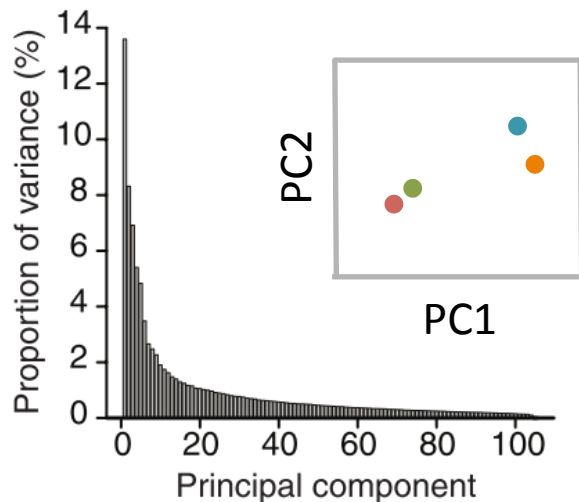
# Pair-wise scatter plot



# Principal Component Analysis (PCA)

PCA is a mathematical algorithm that reduces the dimensionality of the data while retaining most of the variation in the data set.

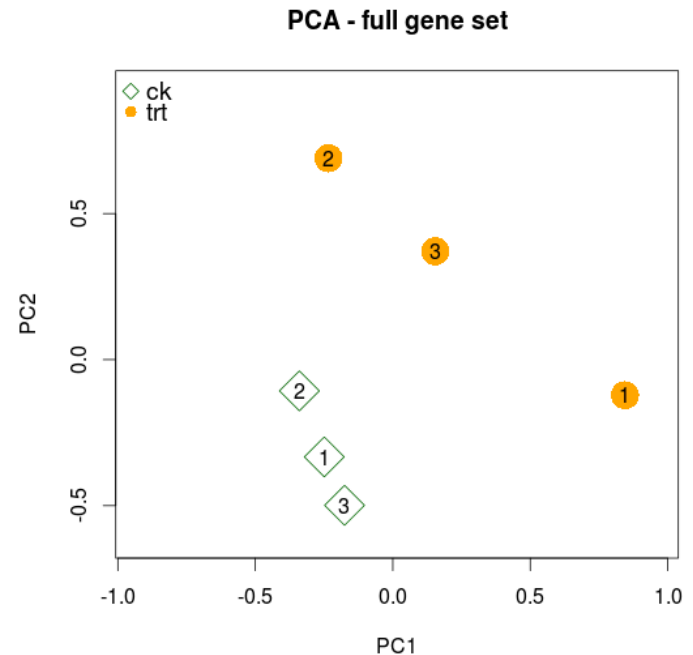
Feature/ variable	John	Mike	Jack	Justin
Weight (lb)	150	243	186	128
Height (cm)	171	190	178	175
...				



Nature Biotech, 2008, 26:303-4

	Control			Treatment		
GeneID	Rep1	Rep2	Rep3	Rep1	Rep2	Rep3
1	2679	2360	2573	2563	3398	3012
2	177	161	171	154	137	152
3	381	371	397	541	723	635
...						
30000	990	1073	1236	850	672	859

Normalized and standardized data



# Statistical test for differential expression

$\pi_G$ : Proportion of transcript fragments of *gene G* among all transcripts

Sample one read, the distribution of the read from *gene G* is Bernoulli( $\pi_G$ )

$$\Pr(\text{read from } G) = \pi_G$$

$$\Pr(\text{read not from } G) = 1 - \pi_G$$

Sample N read, the distribution of the number of reads from *gene G* is:

$$\text{Binomial}(N, \pi_G) \approx \text{Poisson}(N\pi_G)$$

In the Poisson distribution, mean = variance =  $N\pi_G$

However, the Poisson distribution can not well explain data variance (overdispersion issue)

That is why a Negative-Binomial (NB)distribution was introduced

Counts of reads from a gene  $\sim \text{NB}(\text{mean} = \mu, \text{variance} = \sigma^2)$

$$\sigma^2 = \mu + \varphi \mu^2$$

$\varphi$  is a dispersion parameter

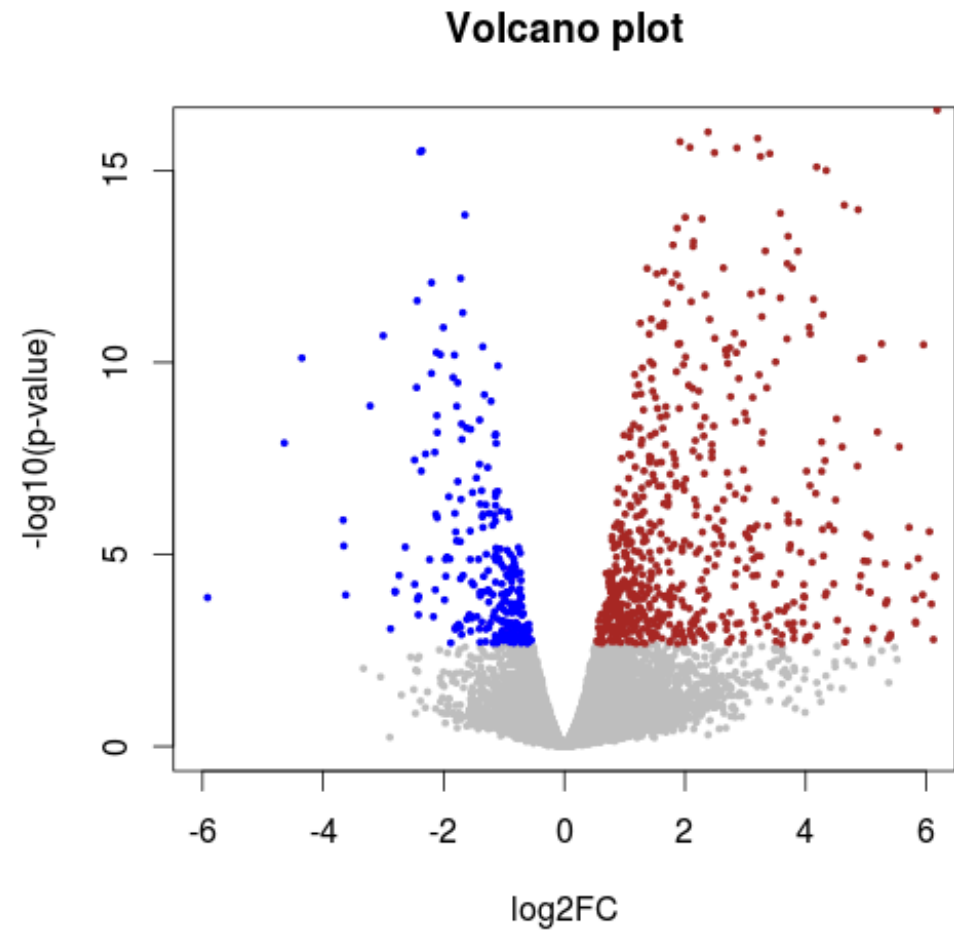
- Generalized Linear Model (GLM) to deal with count data
- **NB-GLM** to incorporate dispersion into the model

# Visualization of DE results

# Volcano plot



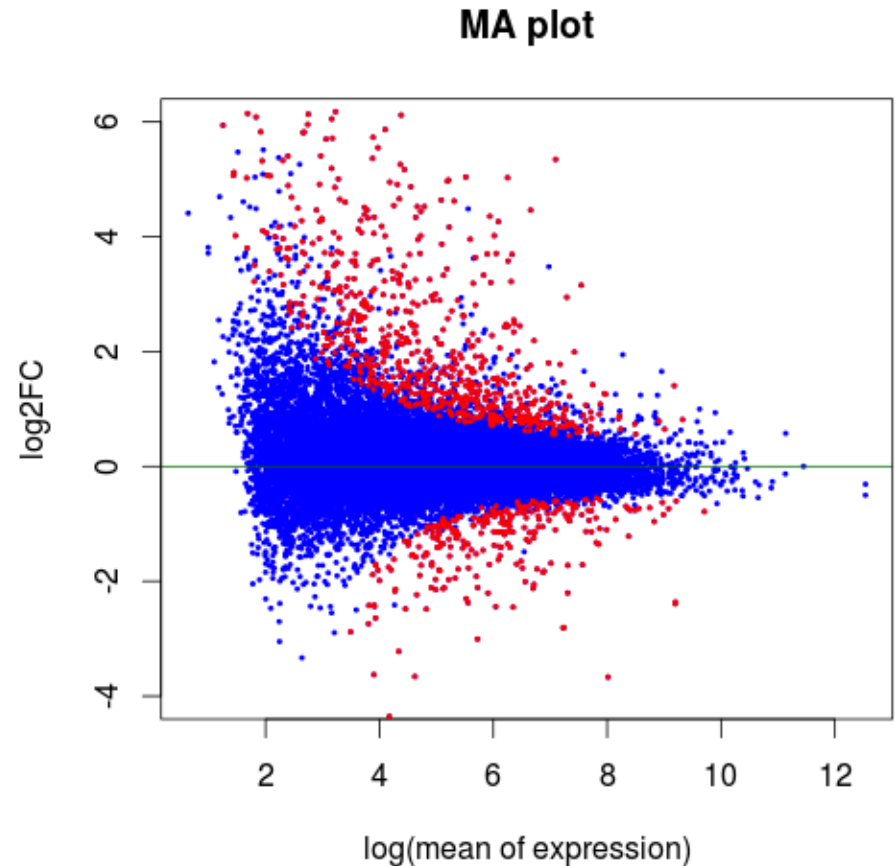
DE Result			
GeneID	Log2FC	p-value	$-\log_{10}(\text{pvalue})$
1	-0.40	0.037	1.43
2	0.03	0.916	0.04
3	-0.89	2.42E-05	4.62
4	0.30	0.130	0.89
5	-0.36	0.140	0.85
6	-0.07	0.811	0.09
...			



# MA plot

M (log ratios) and A (mean average)

GeneID	Mean RPKM	log mean	log2FC
1	0.51	-0.29	-0.40
2	1.25	0.10	0.03
3	3.52	0.55	-0.89
4	0.19	-0.72	0.30
5	2.34	0.37	-0.36
6	6.14	0.79	-0.07
...			

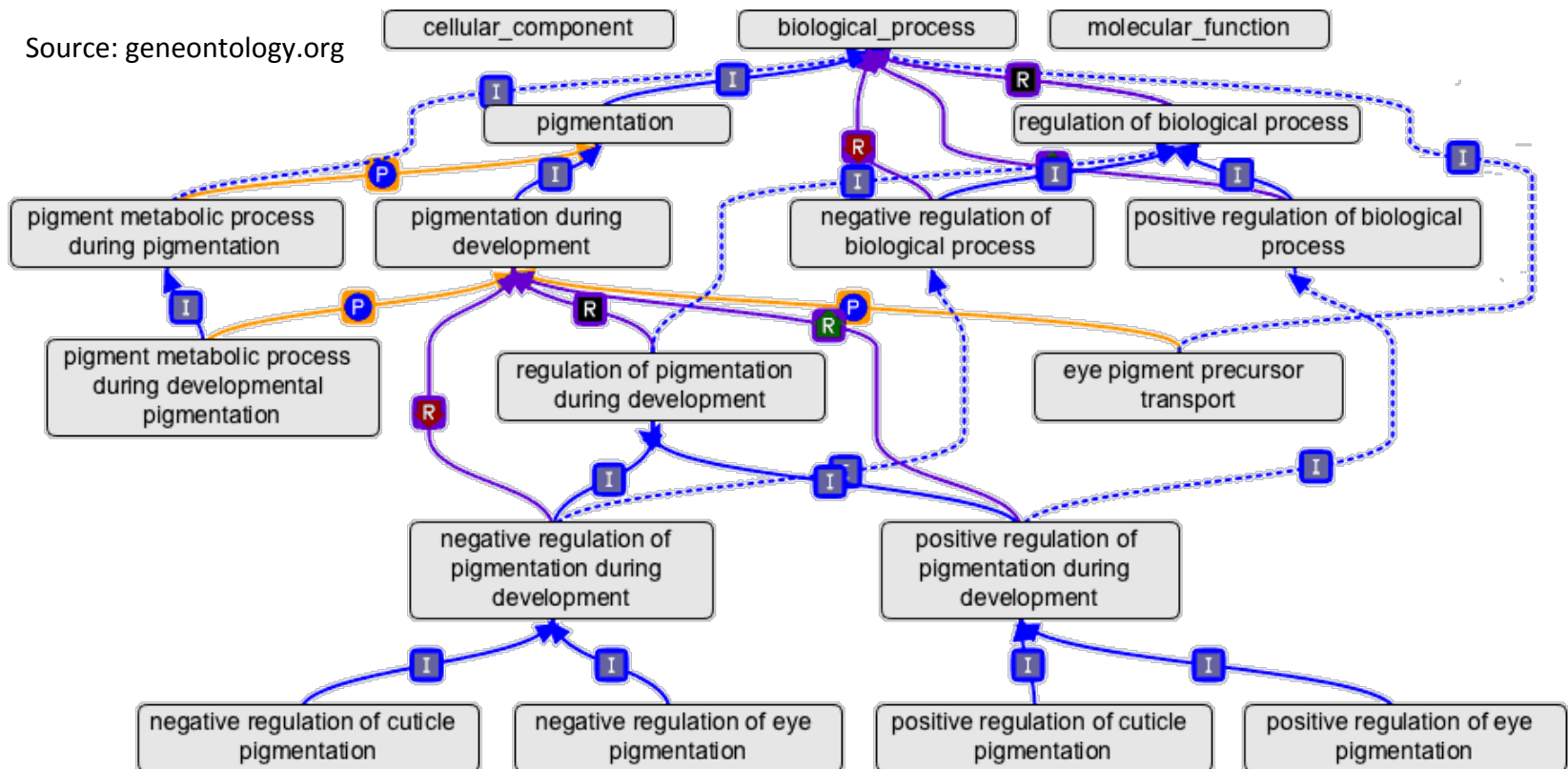


Functional interpretation

# Gene ontology (GO)

An ontology is a representation of a body of knowledge, within a given domain. Ontologies usually consist of a set of classes or terms with relations that operate between them.

Source: [geneontology.org](http://geneontology.org)



Three domains, three roots

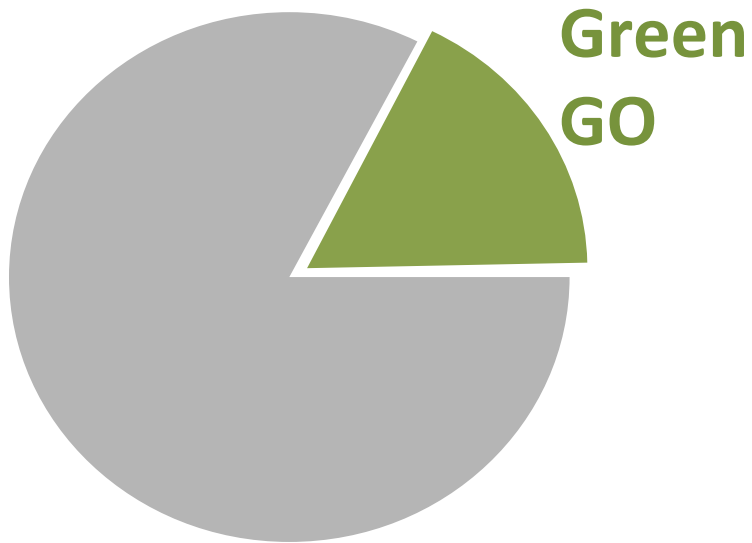
Node: GO term (e.g., cell growth, GO:0016049, biological process)

Edge: term-term connection

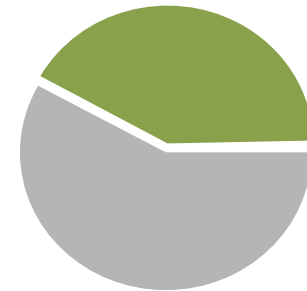
Each GO term can be traced back to a root



# Category enrichment



All genes



significant  
gene set

Is **Green GO** enriched in the  
significant gene set ?

# GO enrichment test – Fisher's Exact test

Gene	GO accession
GRMZM2G001475	<b>GO:0006519</b>
GRMZM2G001475	GO:0016831
GRMZM2G001500	GO:0005524
GRMZM2G001500	GO:0006457
GRMZM2G001500	GO:0051082
GRMZM2G001508	GO:0003993
GRMZM2G001514	GO:0003677
GRMZM2G001514	GO:0004879
GRMZM2G001514	GO:0005634
GRMZM2G001514	GO:0006355
...	...



GRMZM2G001475	1
GRMZM2G002652	2
GRMZM2G006480	3
...	...
GRMZM5G868038	40

Gene	Significant?
GRMZM2G001475	no
GRMZM2G002652	no
<b>GRMZM2G006480</b>	<b>yes</b>
...	...
GRMZM5G868038	no



Question: Are the genes of this GO term enriched in the significant gene set?

Assumption: all genes are independent and equally likely to be selected as DEs.

2x2 Table for GO:0006519

	<b>GO:0006519</b>	Others
Significant	5	210
Not significant	35	39416

Fisher's Exact Test:  
p-value = 2.518e-06

**Name**  
**Ontology**  
**Definition**

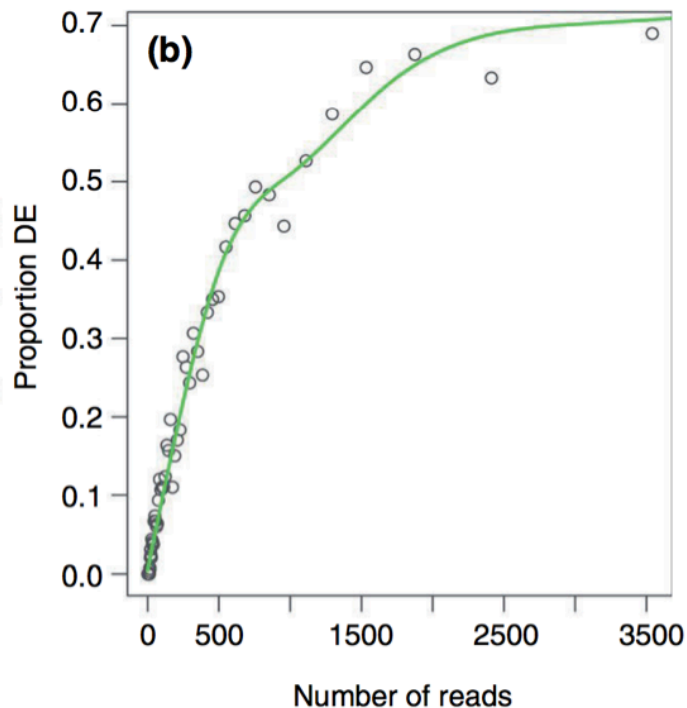
cellular amino acid metabolic process

Biological Process

The chemical reactions and pathways involving amino acids, carboxylic acids containing one or more amino groups, as carried out by individual cells.

# GOSeq

Not all genes are equally likely to be selected as DEs.



1. The likelihood of DE as a function of number of reads is quantified through fitting a monotonic function to “proportion of DE” versus “number of reads”.
2. The function is incorporated into the enrichment statistical test

Gene	Significant?	Read counts	Proportion
GRMZM2G001475	no	224	0.16
GRMZM2G002652	no	51	0.05
<b>GRMZM2G006480</b>	<b>yes</b>	<b>536</b>	<b>0.38</b>
...	...	...	...
GRMZM5G868038	no	0	0

3. Weighted sampling to perform enrichment test

<b>GO:0006519</b>	# DE
Obs (from the DE analysis)	5
1 <sup>st</sup> weighted sampling	1
2 <sup>nd</sup> weighted sampling	0
3 <sup>rd</sup> weighted sampling	2
...	...

→ p-value

# Summary

- R is an excellent tool for DE analysis and data visualization.
- Many bioinformatics pipelines and statistical methods have been developed. Methods and parameters need to be carefully selected.
- A proper GO enrichment test needs to be used.
- Dobin A, Davis CA, Schlesinger F, Drenkow J, Zaleski C, Jha S, Batut P, Chaisson M, Gingeras TR. 2013. STAR: ultrafast universal RNA-seq aligner. *Bioinformatics* 29:15-21.
- Love MI, Huber W, Anders S. 2014. Moderated estimation of fold change and dispersion for RNA-seq data with DESeq2. *Genome Biol* 15:550.
- Young MD, Wakefield MJ, Smyth GK, Oshlack A. 2010. Gene ontology analysis for RNA-seq: accounting for selection bias. *Genome Biol* 11:R14.