

Giordan Kitts | Curriculum Vitae

✉ gkitts@ucsc.edu

Education

University of California, Santa Cruz

Ph.D. - Microbiology and Environmental Toxicology

Santa Cruz, CA

2018 - Present

University of California, Santa Cruz

B.S. - Biochemistry and Molecular Biology, Minor - Bioinformatics

Santa Cruz, CA

2012 - 2017

Research Experience

Yildiz Lab, UCSC

Graduate researcher; Advisor: Dr. Fitnat Yildiz

Santa Cruz, CA

2018 - Present

My research focuses on understanding of human pathogen *Vibrio cholerae*'s infection cycle and how this pathogen senses and adapts to environmental insults. My goal is to understand transcriptional network circuitries enable *V. cholerae* environmental adaptation programs. I study mechanisms of activation of two-component signal transduction systems that regulate virulence and biofilm formation in *V. cholerae*. I also develop tools to develop a predictive model using deep learning methods utilizing transcriptional profiling data sets. My project is interdisciplinary and integration of molecular genetics, functional genomics approaches will enhance understanding of *V. cholerae*'s transcriptional landscape.

Yildiz Lab, UCSC

Junior Research Specialist

Santa Cruz, CA

April 2017 - September 2018

Characterized the conserved regulatory circuit, LapDG, controlling reversible cell attachment in *V. cholerae*. First identified in pseudomonads, this mechanism combines c-di-GMP-mediated transcription of an adhesin with post-translational proteolytic control of adhesin cell surface display by second messenger levels. Continued to develop genetic tools and use in screens to identify novel methods of c-di-GMP regulation and modulation. Identified signals and corresponding signal transduction pathways regulating biofilm formation.

Associated publication: <https://doi.org/10.1128/mBio.02822-19>

Yildiz Lab, UCSC

Undergraduate researcher; Advisor: Dr. Fitnat Yildiz

Santa Cruz, CA

April 2016 - March 2017

Worked on project entitled "Identification of regulators of c-di-GMP responsible for increased biofilm formation in the absence of flagellum", supervised by Dr. David Zamorano-Sanchez. Identified a previously uncharacterized diguanylate cyclase (CdgO) contributing to modulation of c-di-GMP levels seen in absence of flagellum (termed FDBR - Flagellar Dependent Biofilm Response). Project used forward genetics to identify novel regulators of c-di-GMP, a second messenger molecule ubiquitously correlated to biofilm formation in bacteria, by use of a dual-fluorescent c-di-GMP biosensor coupled with transposon mutagenesis.

Associated publication: <https://doi.org/10.1371/journal.pgen.1008703>

UCSC iGEM 2016

Head of metabolic engineering; Advisor: Dr. David Bernick

Santa Cruz, CA

Summer 2016

Partook in the synthetic biology competition International Genetically Engineered Machine (iGEM), as part of the UCSC 2016 team. Team's goal was to create a mutual solution to the crises of excessive agricultural waste and rising obesity rates by engineering a strain of *B. subtilis* to produce the non-caloric alternative sweetener, erythritol, via degradation of an agricultural waste product, almond hulls. Engineered strain was used in a team-created chemostat system leading into a filtration unit, yielding erythritol as end product. As head of metabolic engineering, led team in charge of molecular cloning, necessary genetic manipulation of organism, and optimization of various growth conditions.

Website: <https://2016.igem.org/Team:UCSC>

Wang Lab, CNCS, HPU

Research internship

Kailua, HI

Summer 2015

Studied the neuroplasticity associated protein ARC (activity-regulated cytoskeleton-associated protein). Research focused on identifying active site of this protein, and better understanding possible interactions with the protein Dynamin.

Teaching Experience

Microbiology Teaching Lab

Teaching Assistant

UCSC

Multiple Academic Quarters

Led an undergraduate microbiology teaching lab class. Students learn methods for isolation, cultivation, characterization, and identification of unknown microbes. Taught various standard microbiological techniques: Plate isolation methods, aseptic technique, membrane filtration, gram-staining, 16S rRNA sequencing, PCR, gel electrophoresis, etc.

Microbiology Lecture

Teaching Assistant

UCSC

Multiple Academic Quarters

Teaching assistant for Intro To Microbiology 5-unit lecture course. Lead discussion sections with applied case studies or lecture material review, graded, and helped proctor exams.

Grants and Awards

ARCS Scholar

ARCS Foundation

2020

Publications

Giordan Kitts, Krista M. Giglio, David Zamorano-Sánchez, Jin Hwan Park, Loni Townsley, Richard B. Cooley, Benjamin R. Wucher, Karl E. Klose, Carey D. Nadell, Fitnat H. Yildiz, and Holger Sondermann. A Conserved Regulatory Circuit Controls Large Adhesins in *Vibrio cholerae*. *mBio*, 10(6), 12 2019.

Patrick J. Woida, Giordan Kitts, Stephanie Shee, Adam Godzik, and Karla J. F. Satchell. Actin Cross-Linking Effector Domain of the *Vibrio vulnificus* F-Type MARTX Toxin Dominates Disease Progression During Intestinal Infection. *Infection and Immunity*, 90(4), 4 2022.

Daniel C. Wu, David Zamorano-Sánchez, Fernando A. Pagliai, Jin Hwan Park, Kyle A. Floyd, Calvin K. Lee, Giordan Kitts, Christopher B. Rose, Eric M. Bilotta, Gerard C. L. Wong, and Fitnat H. Yildiz. Reciprocal c-di-GMP signaling: Incomplete flagellum biogenesis triggers c-di-GMP signaling pathways that promote biofilm formation. *PLOS Genetics*, 16(3):e1008703, 3 2020.

Posters and Presentations

Graduate Recruitment Symposium

Seymour Center, Santa Cruz, CA

Poster

January, 2020

Title: A conserved regulatory circuit controls large cell surface adhesins in *Vibrio cholerae*.

ASM Microbe 2019 - American Society for Microbiology

Moscone Convention Center, San Francisco, CA

Poster

June, 2019

Title: A conserved regulatory circuit controls large cell surface adhesins in *Vibrio cholerae*.

BAMPS 2017 - Bay Area Microbial Pathogenesis Symposium

Genentech Hall - UCSF, San Francisco, CA

Poster

March, 2017

Title: Designing genetic tools to study c-di-GMP in *Vibrio cholerae*.

Skills

- **Code and Software:** R, Python, Bash, Markdown, LaTeX, Microsoft Office, FIJI, COMSTAT, Imaris, Adobe Illustrator, Adobe Photoshop, Inkscape, CytoScape, Notion, Pandoc
- **Bioinformatic tools and databases:** Salmon, DESeq2, Jalview, Bowtie2, SAMtools, BEDtools, UCSC Genome Browser, BLAT, BLAST, NCBI, MicrobesOnline, KEGG, BioCyc, PATRIC, Benchling, Geneious, CLC Genomics Workbench
- **Molecular biology:** Multiple cloning methods, sequence design, PCR, qPCR, RNA-sequencing, various media preparations, FPLC
- **Microscopy:** Confocal Laser Scanning Microscopy (CLSM), Fluorescence microscopy, Phase-contrast, Brightfield
- **Microbiology** Bacterial genetics and genome manipulation, Forward genetic screens, growth curves, metabolic screens, transcriptional reporter assays

References

- **Dr. Fitnat Yildiz** **fyildiz@ucsc.edu**
Principal investigator, Yildiz Lab, UCSC 408-396-0133
- **Dr. Karen Ottemann** **ottemann@ucsc.edu**
Principal investigator, Ottemann Lab, UCSC
- **Dr. Karl Klose** **Karl.Klose@utsa.edu**
Principal investigator, Klose Lab, UT San Antonio