



JOHNS HOPKINS
SCHOOL *of* MEDICINE

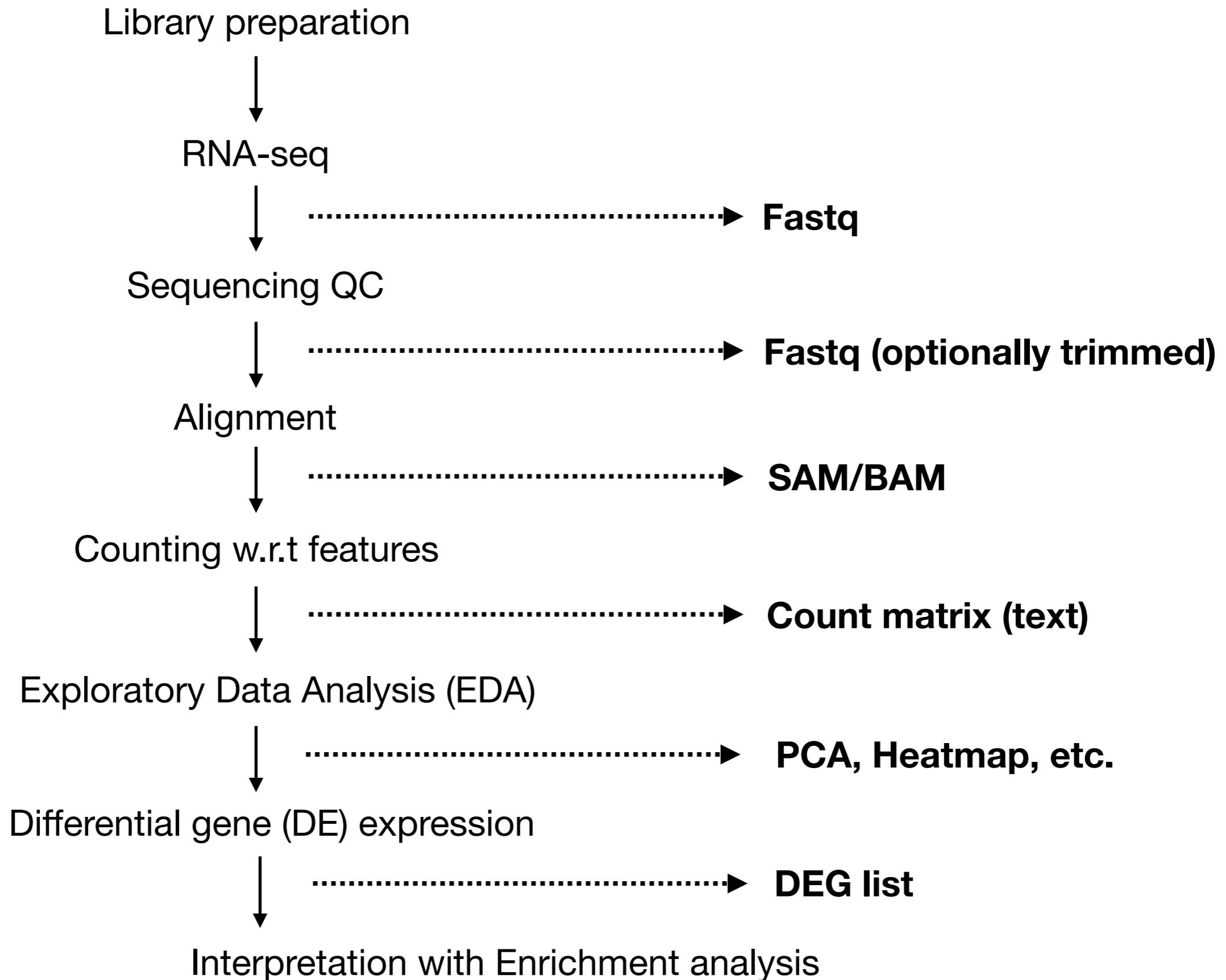
Quantitative Molecular Neurogenomics (ME.440.825)

Module 6 :
Practical RNA-seq workflow &
Enrichment analysis

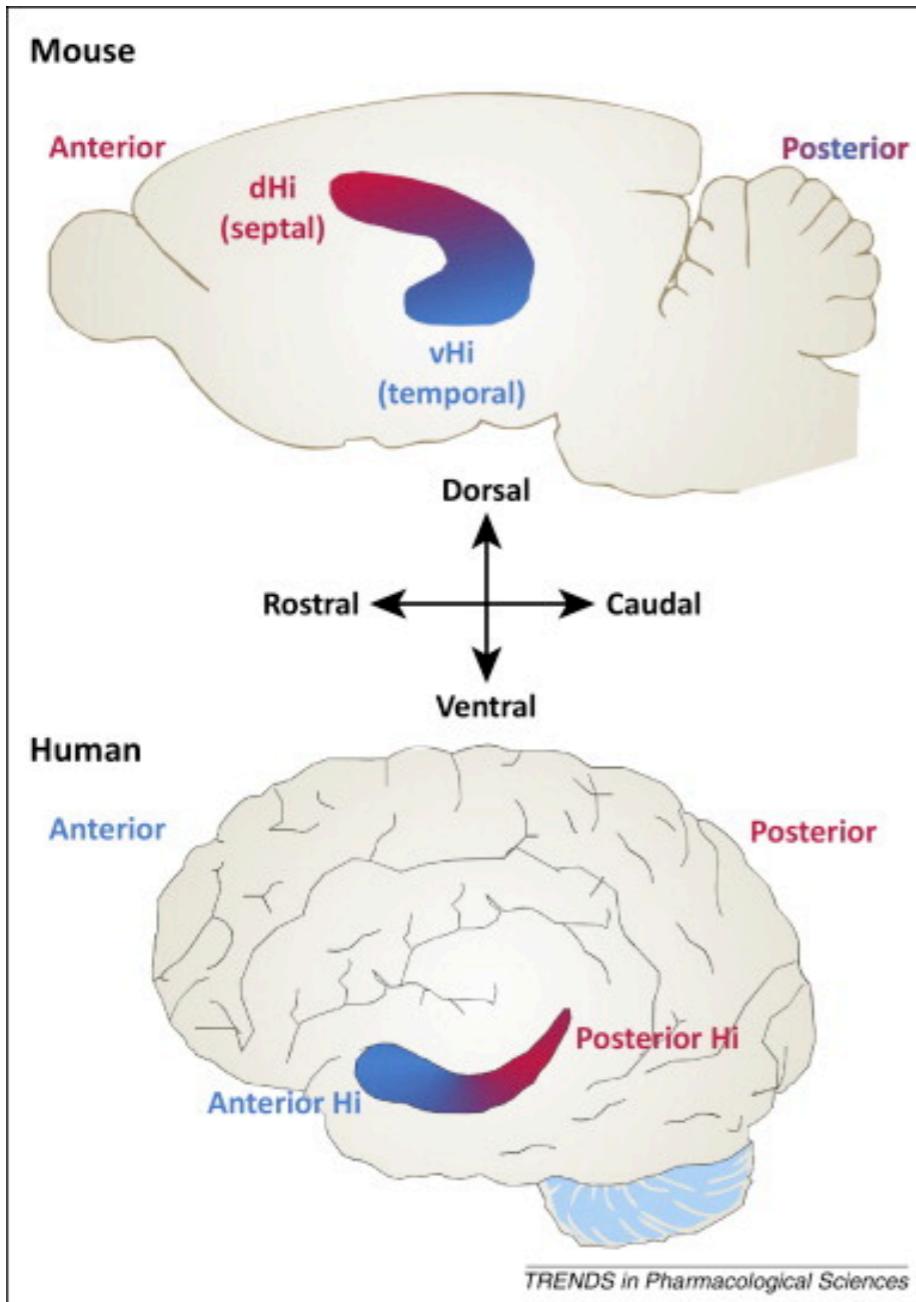
10/10/2022

Taeyoung Hwang, PhD
Lieber Institute for Brain Development,
Department of Neurology and Department of Neuroscience

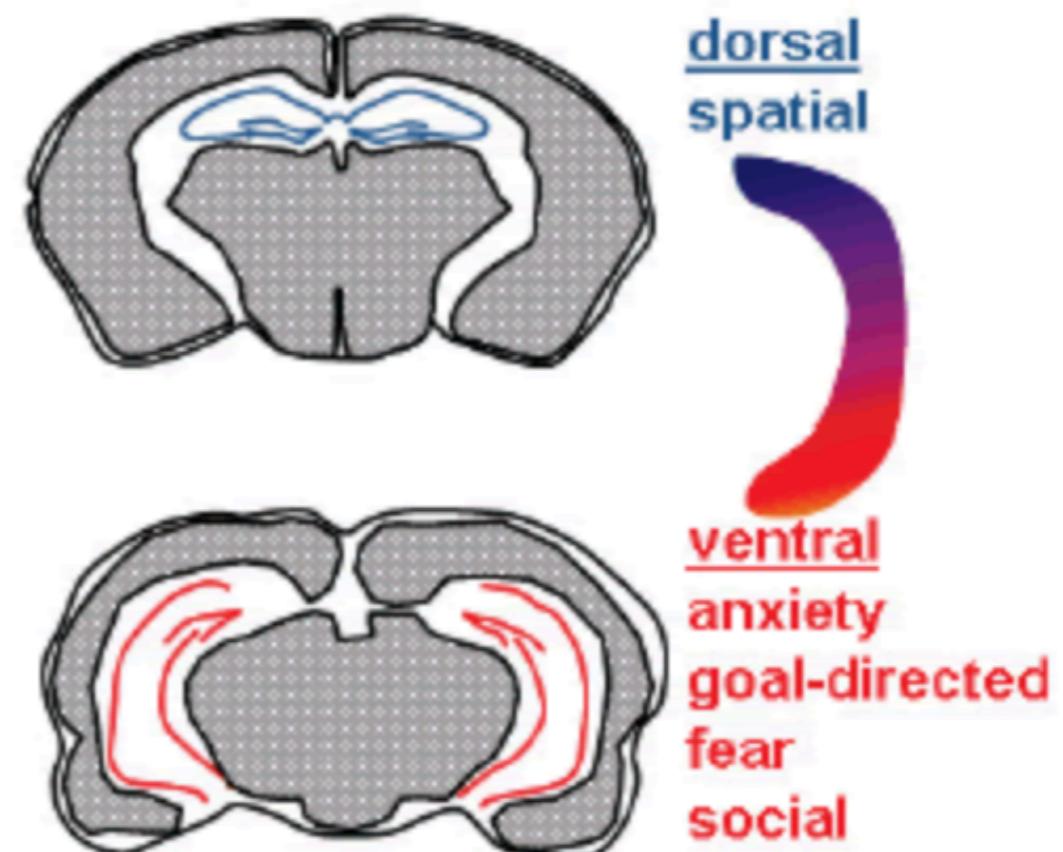
Typical RNA-seq workflow



Sample: mouse hippocampus tissue



The longitudinal axis of the hippocampus



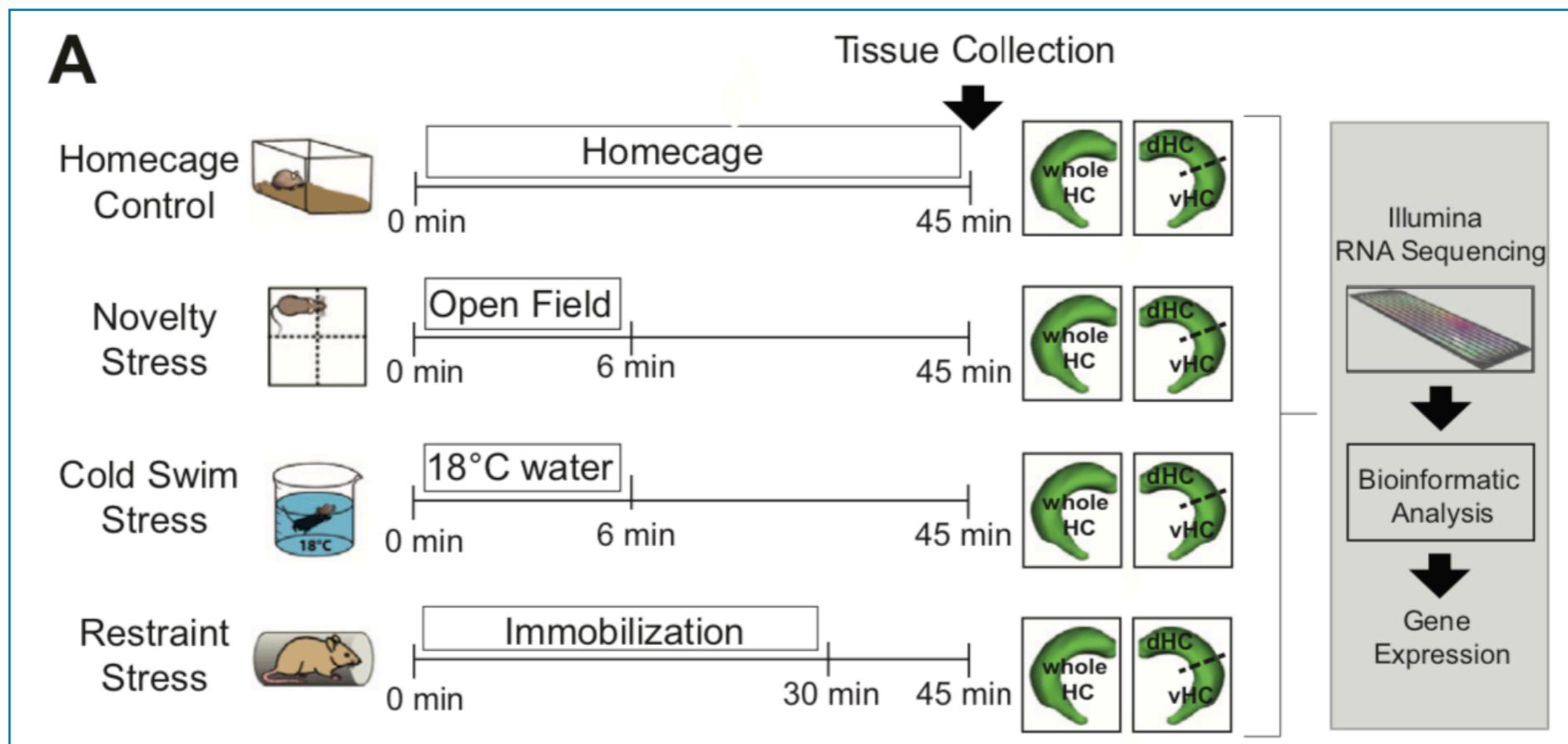
Dataset: RNA-seq of mouse hippocampus

Biological Psychiatry October 1, 2018; 84:531–541

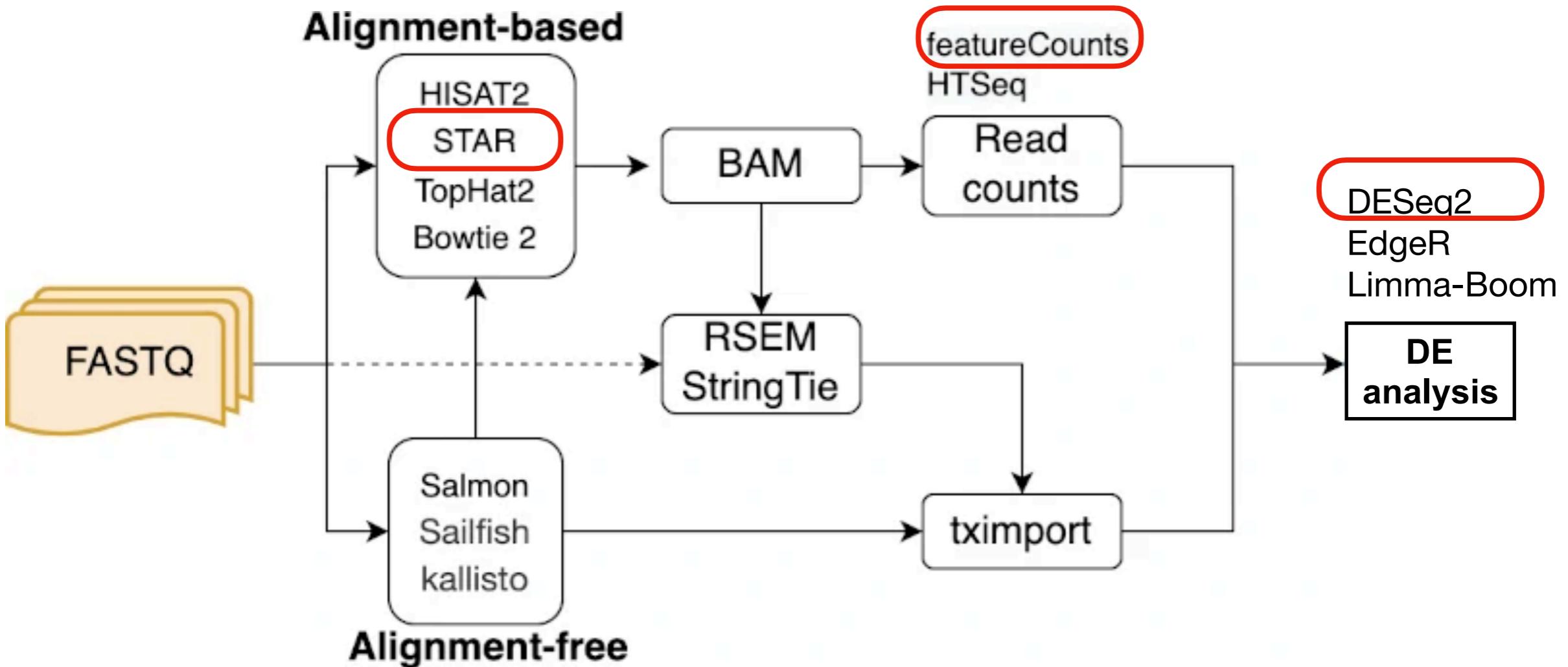
Archival Report

Distinct Proteomic, Transcriptomic, and Epigenetic Stress Responses in Dorsal and Ventral Hippocampus

Amalia Floriou-Servou, Lukas von Ziegler, Luzia Stalder, Oliver Sturman, Mattia Privitera, Anahita Rassi, Alessio Cremonesi, Beat Thöny, and Johannes Bohacek



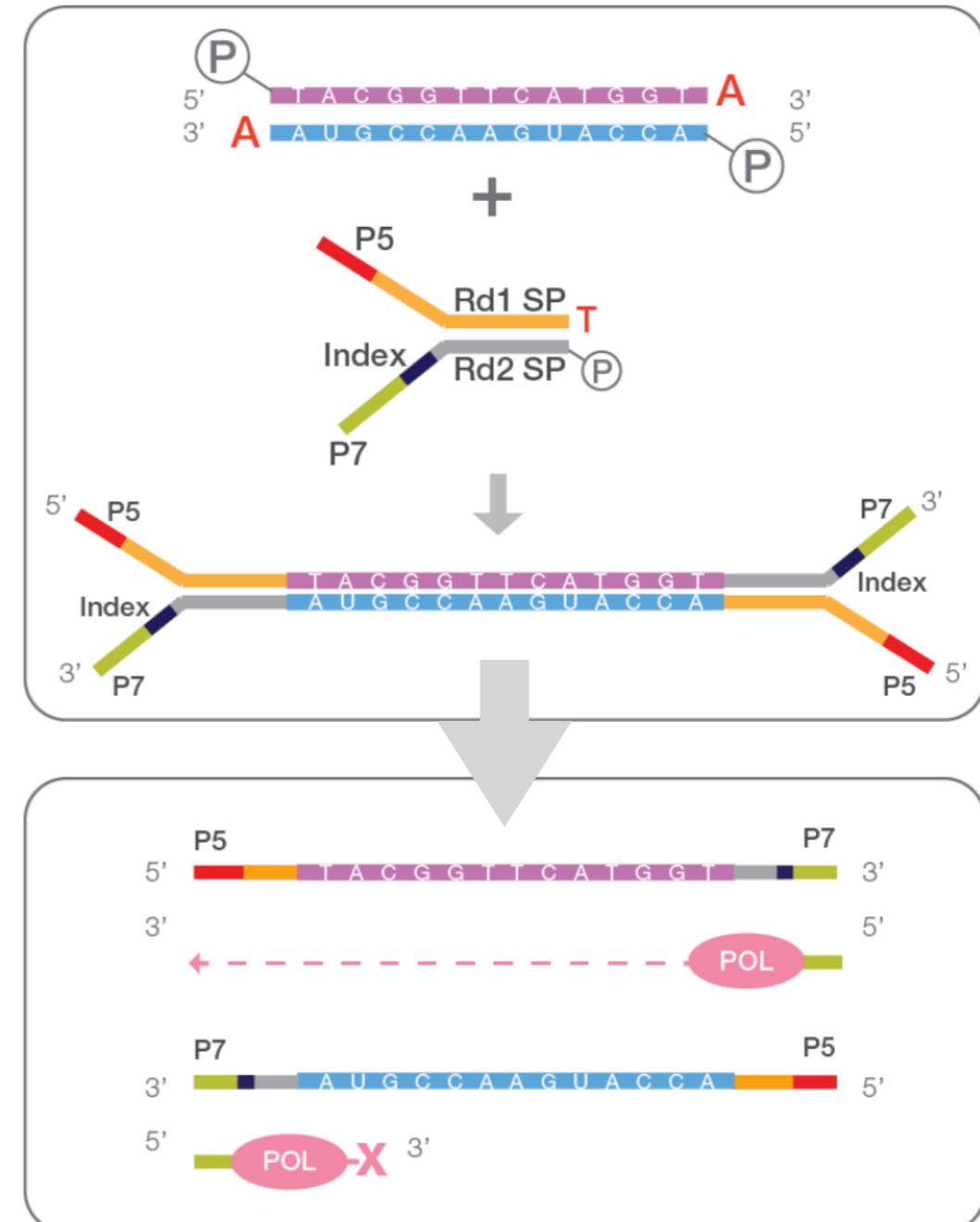
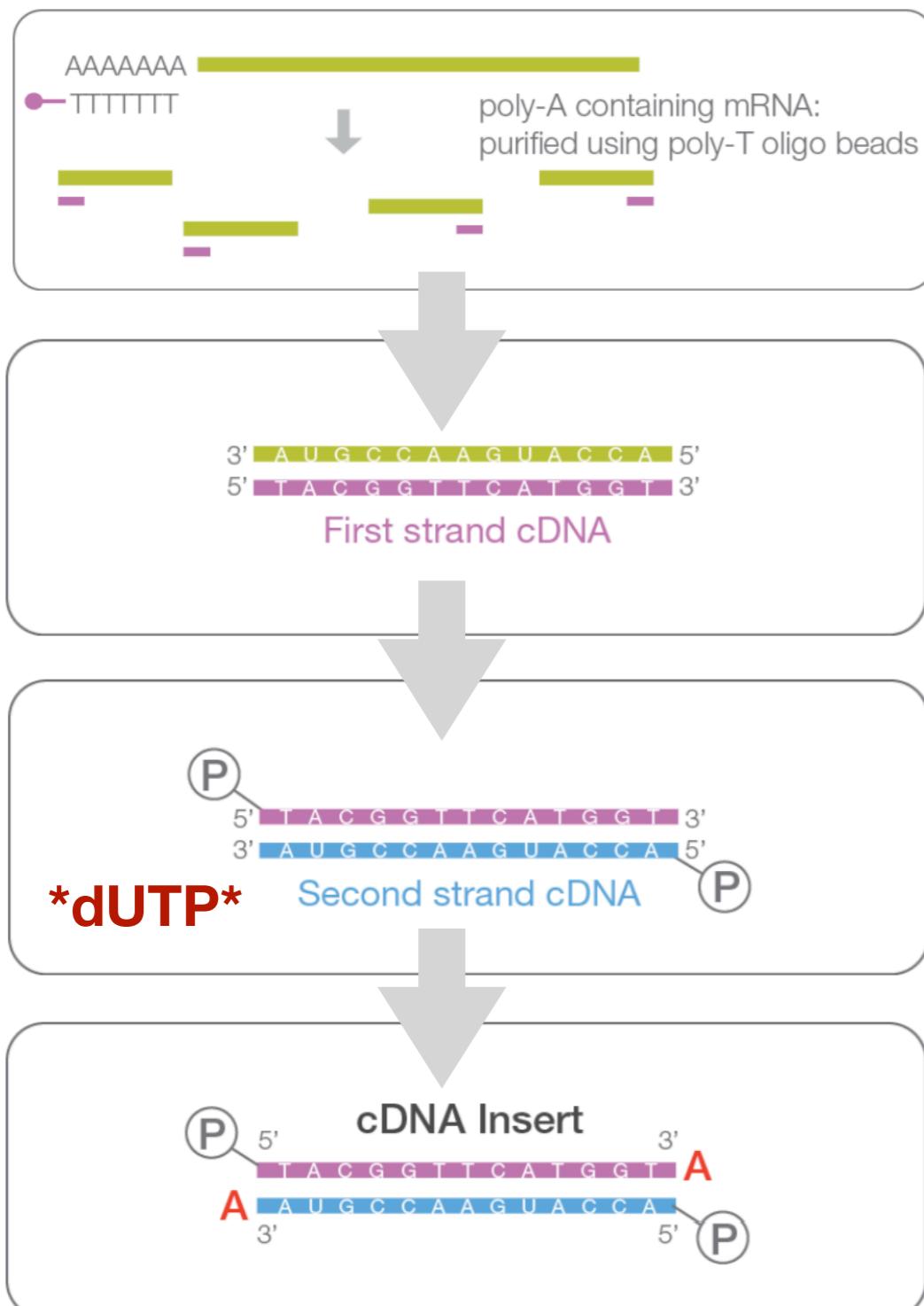
Our RNA-seq pipeline



RNA-seq library preparation

Supplementary Method

“For library preparation, the TruSeq stranded RNA kit (Illumina Inc.) was used according to the manufacturer’s protocol.”



Sequencing QC using Fastqc

```
$ fastqc SRR5715043.fastq.gz
```

 **FastQC Report**

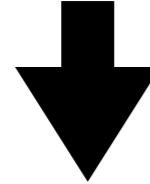
Summary

-  [Basic Statistics](#)
-  [Per base sequence quality](#)
-  [Per sequence quality scores](#)
-  [Per base sequence content](#)
-  [Per sequence GC content](#)
-  [Per base N content](#)
-  [Sequence Length Distribution](#)
-  [Sequence Duplication Levels](#)
-  [Overrepresented sequences](#)
-  [Adapter Content](#)

 **Basic Statistics**

Measure	Value
Filename	SRR5715043.fastq.gz
File type	Conventional base calls
Encoding	Sanger / Illumina 1.9
Total Sequences	36027995
Sequences flagged as poor quality	0
Sequence length	126
%GC	49

 **Per base sequence quality**



****No worries****

Fastqc html report: an example

Summary

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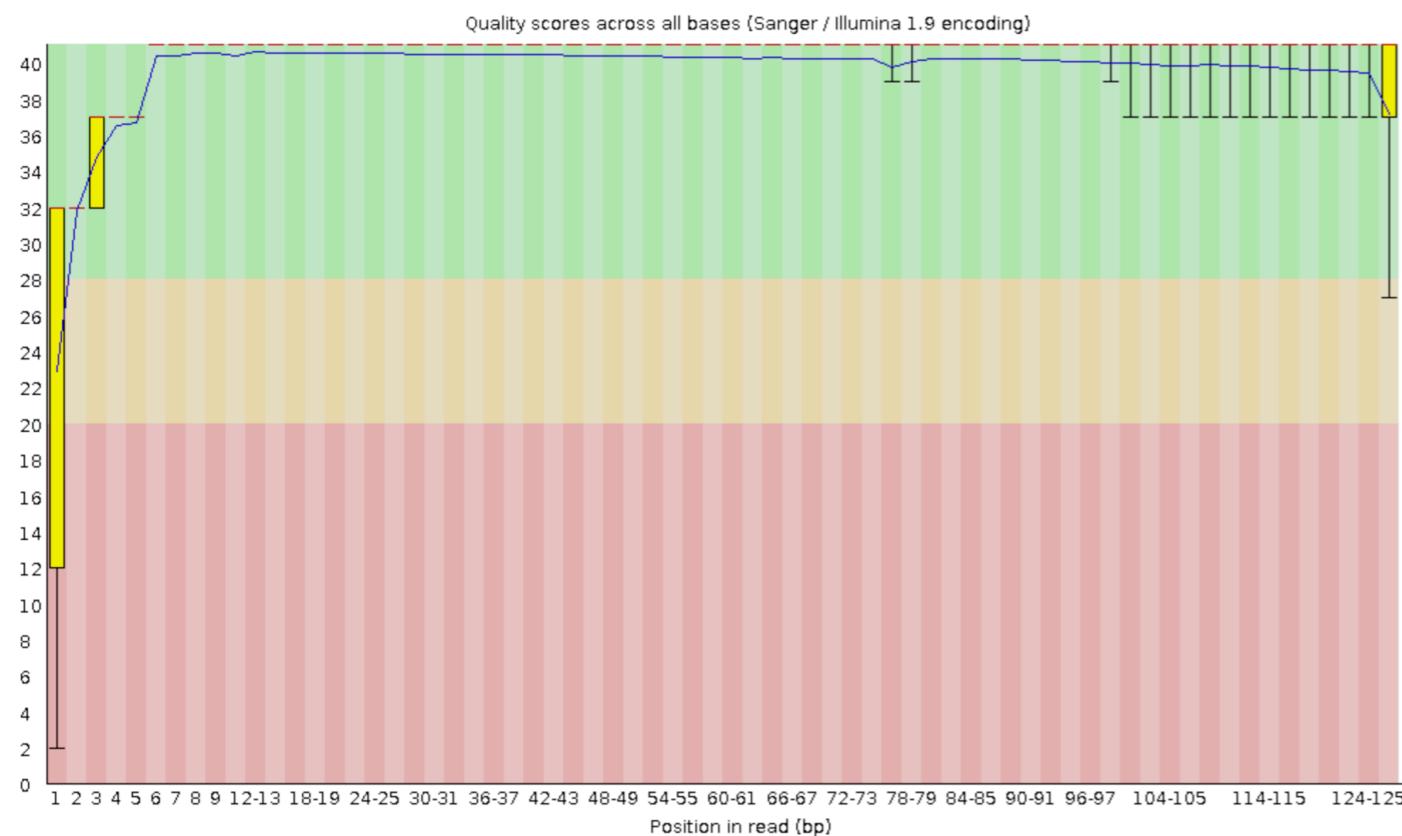
Basic Statistics

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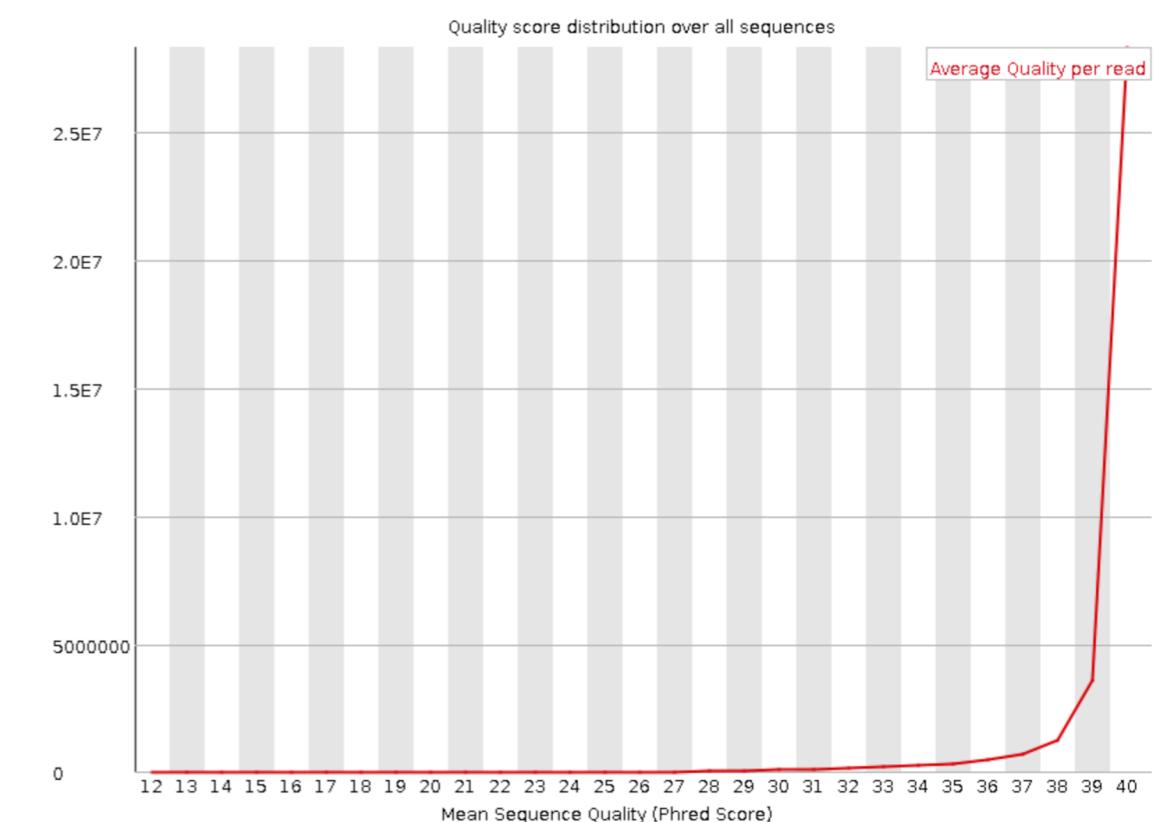
GC % in species

Species	Sequences used (Mb) ¹	CDS count ²	Exon count ³	Intron count	Average Exon length (bp)	Average Exon GC (%)	Average Intron length (bp)	Average Intron GC (%)
<i>A. thaliana</i>	119.2	23488	135697	118838	223.7	44.1	163.7	32.7
<i>B. taurus</i>	2434	16829	162223	151199	162.3	52.0	4516.4	46.9
<i>C. elegans</i>	100.3	27123	171102	149895	208.4	43.0	334.7	29.1
<i>C. familiaris</i>	2445	15960	161238	147118	157.2	50.6	3535.5	46.1
<i>D. melanogaster</i>	118.4	8119	43847	40732	370.0	52.7	1530.7	36.5
<i>D. rerio</i>	1547	20256	173438	156972	156.2	48.7	2276.4	34.5
<i>G. gallus</i>	1032	14367	142329	133690	152.1	48.2	2811.8	42.6
<i>H. sapiens</i>	3077	40430	381122	378089	162.7	50.8	5848.7	45.9
<i>M. musculus</i>	2644	30546	254605	243458	170.5	51.1	4683.6	46.0
<i>O. sativa</i>	372.1	41046	178106	144863	306.0	51.0	396.1	37.7
<i>P. troglodytes</i>	3176	30010	296830	287132	156.2	50.7	6003.7	45.3
<i>R. norvegicus</i>	2719	22243	195940	186864	172.9	51.1	4406.4	46.3
<i>T. nigroviridis</i>	217.3	15455	122253	108901	174.3	54.5	599.5	45.4

Per base sequence quality



Per sequence quality scores



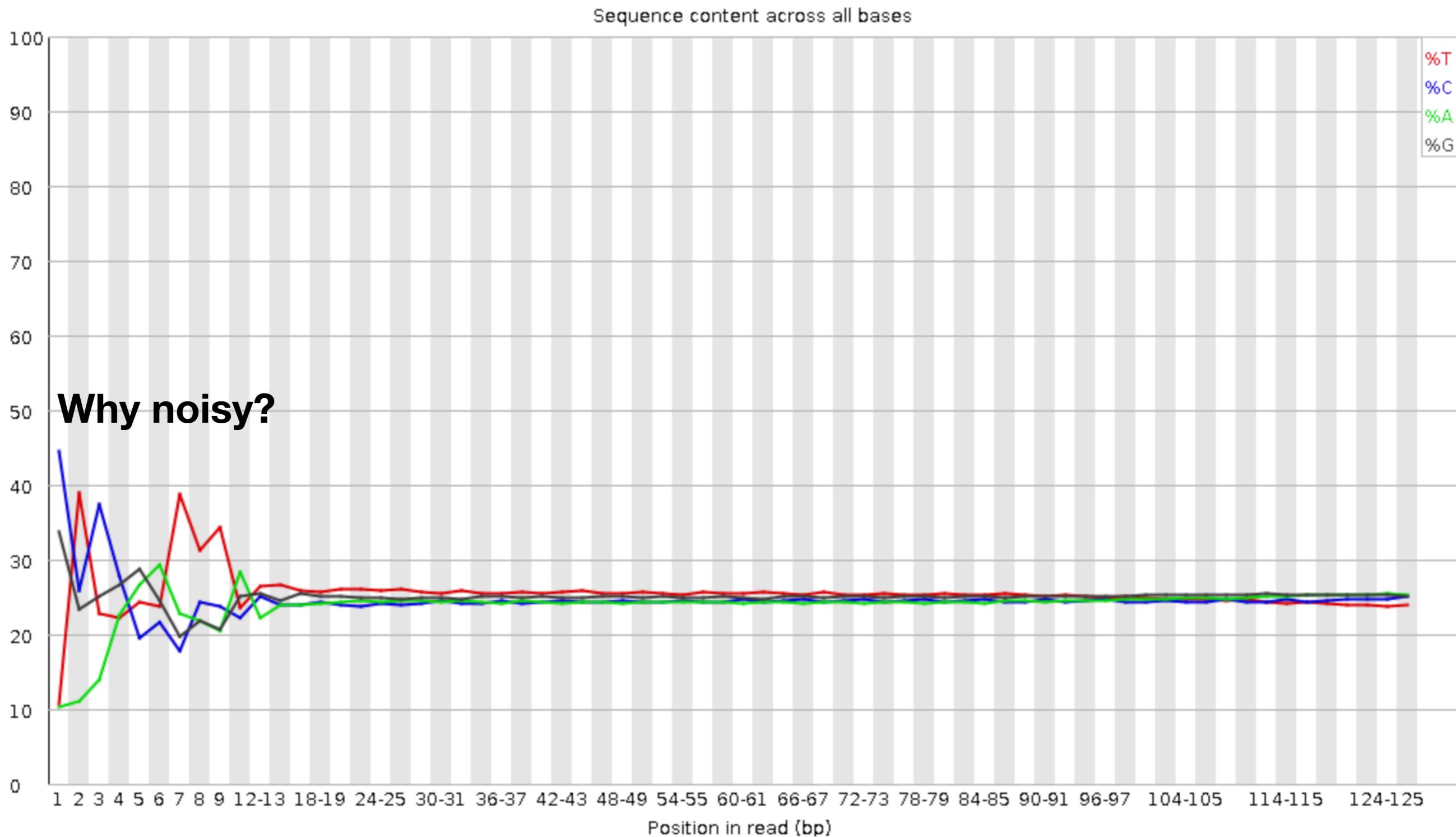
Phred Quality Score

10
20
30
40

Probability of incorrect base call

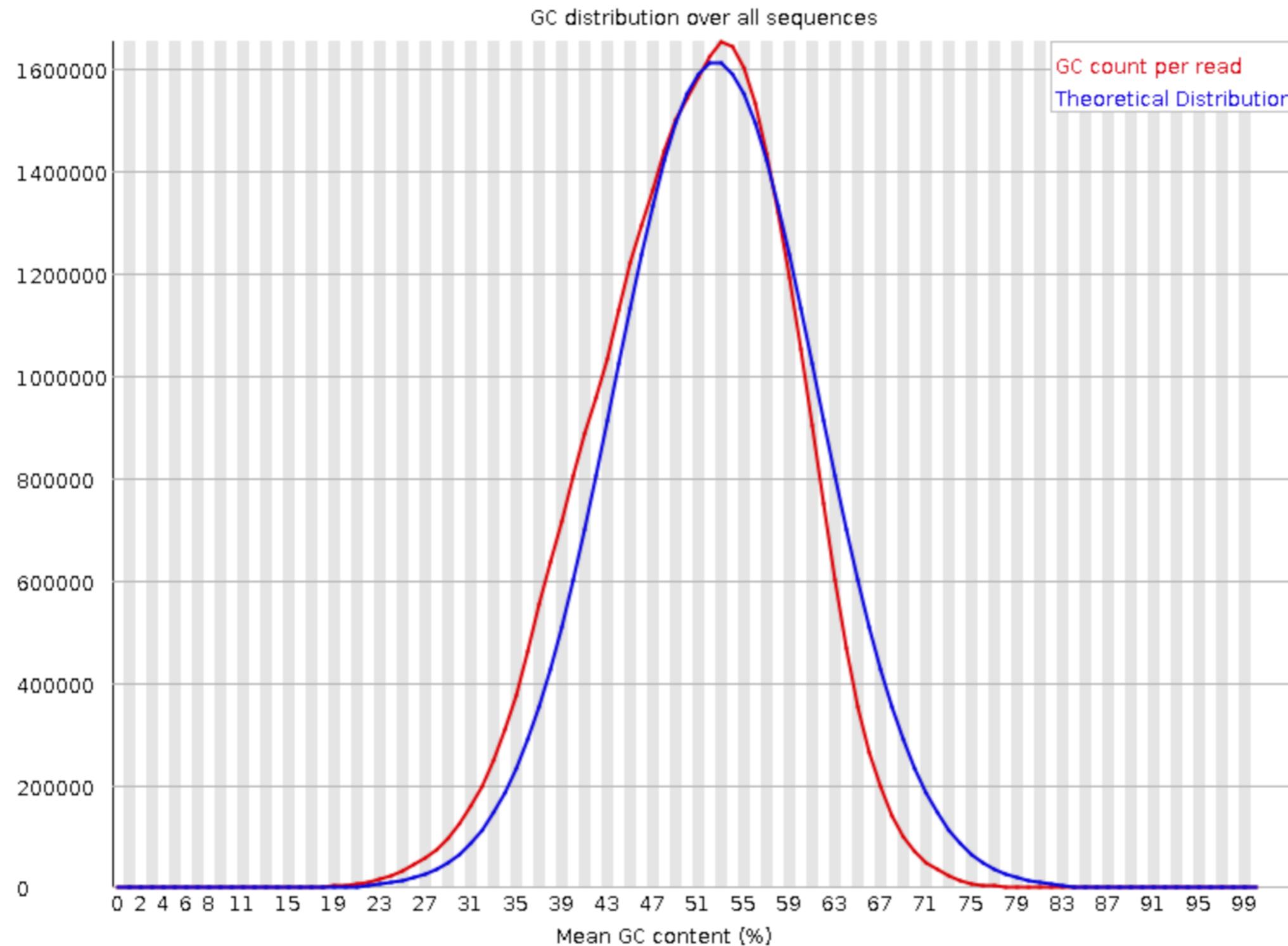
1 in 10
1 in 100
1 in 1000
1 in 10,000

✖ Per base sequence content

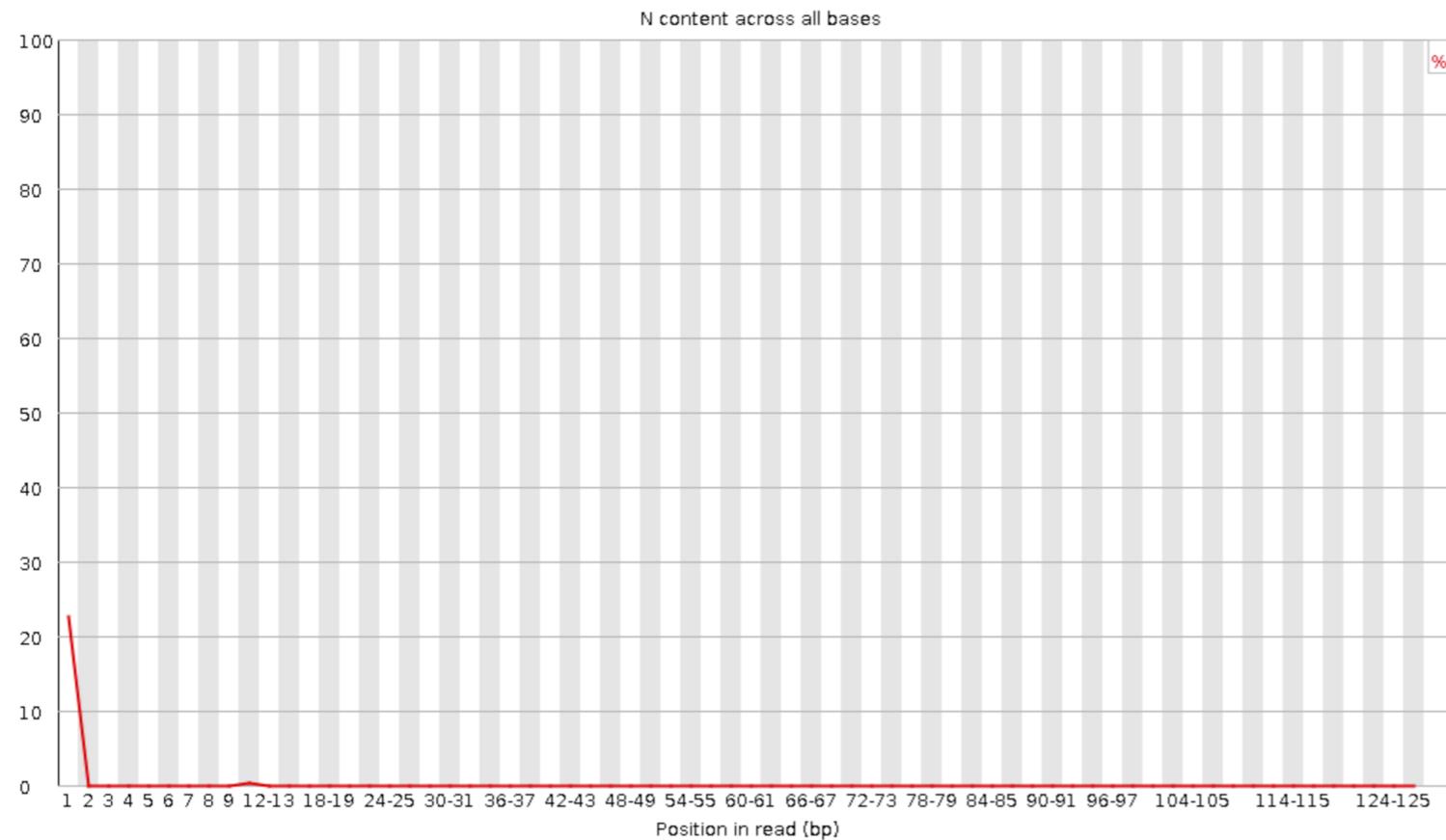




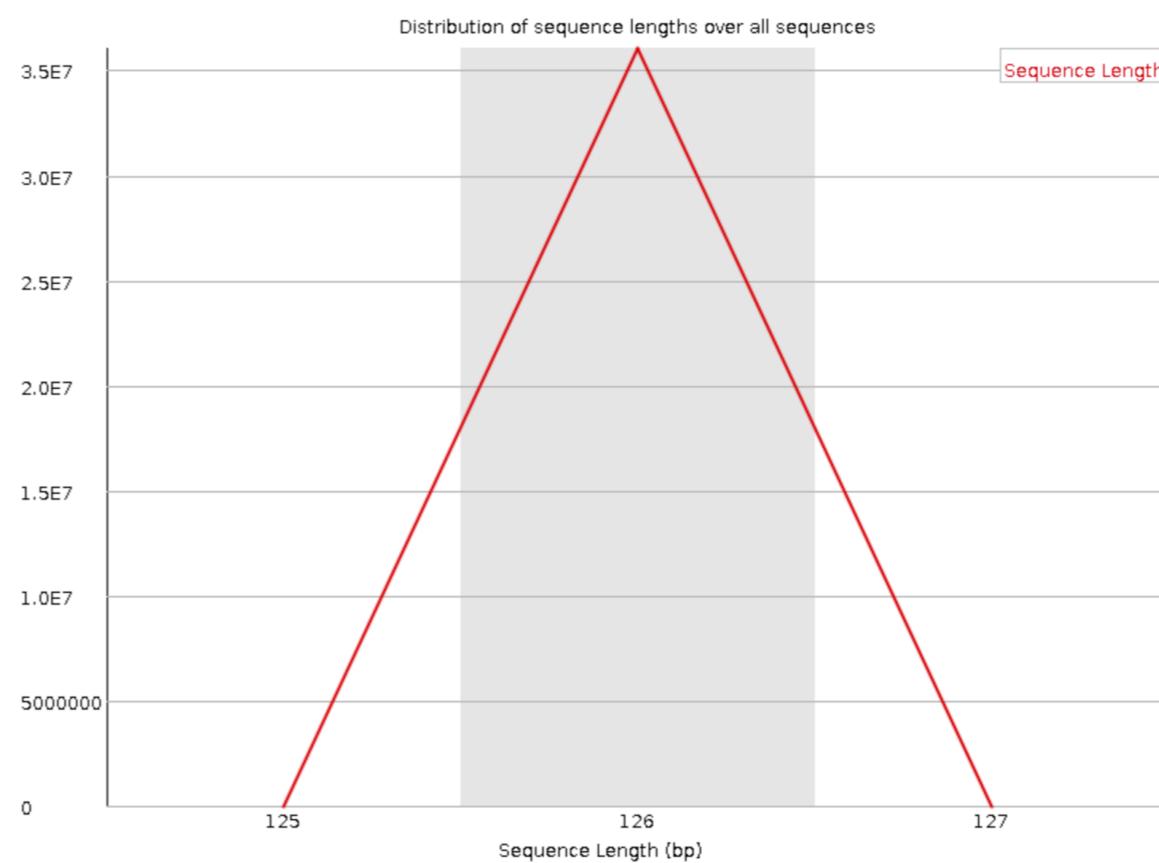
Per sequence GC content



Per base N content

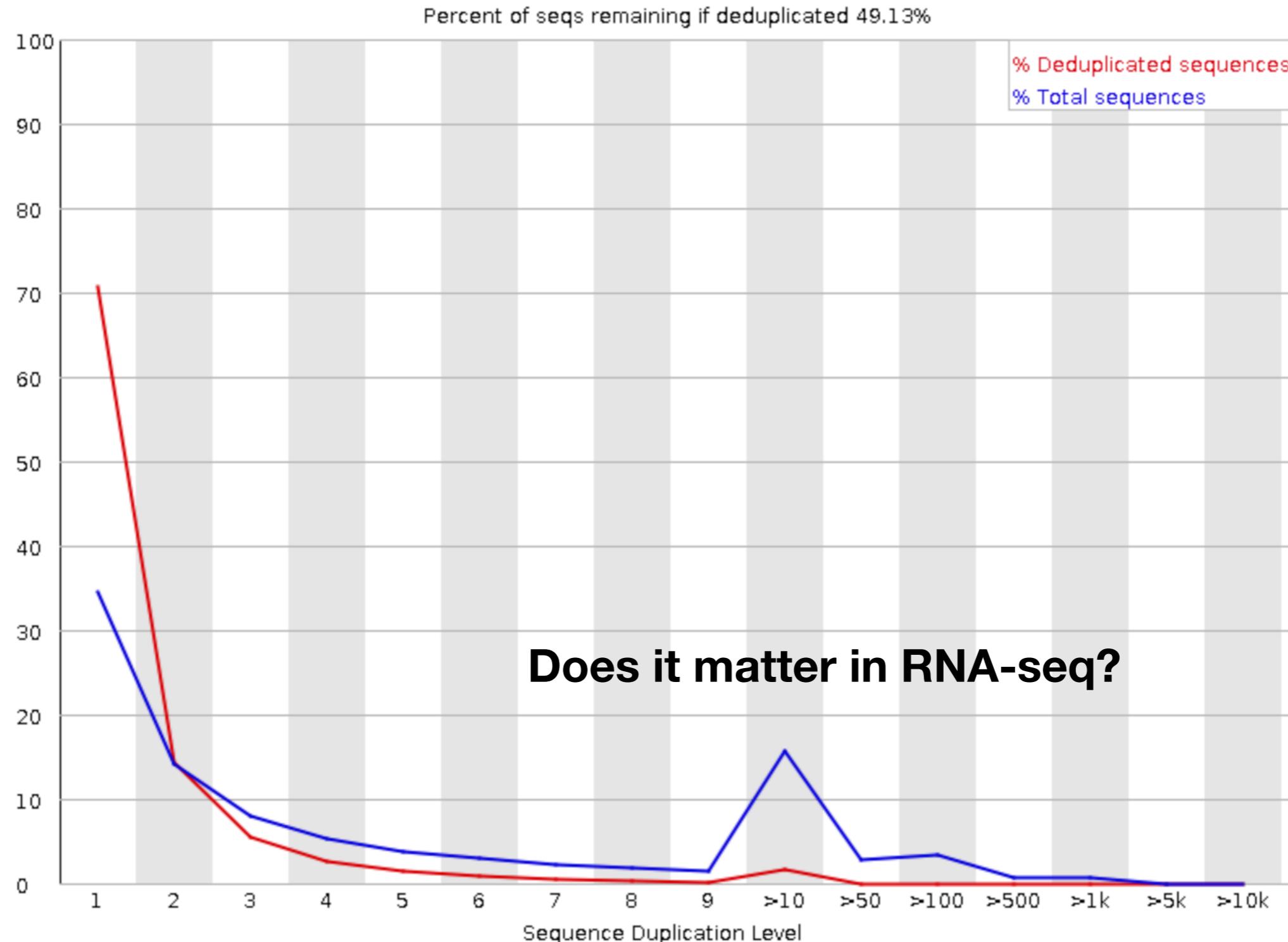


Sequence Length Distribution





Sequence Duplication Levels



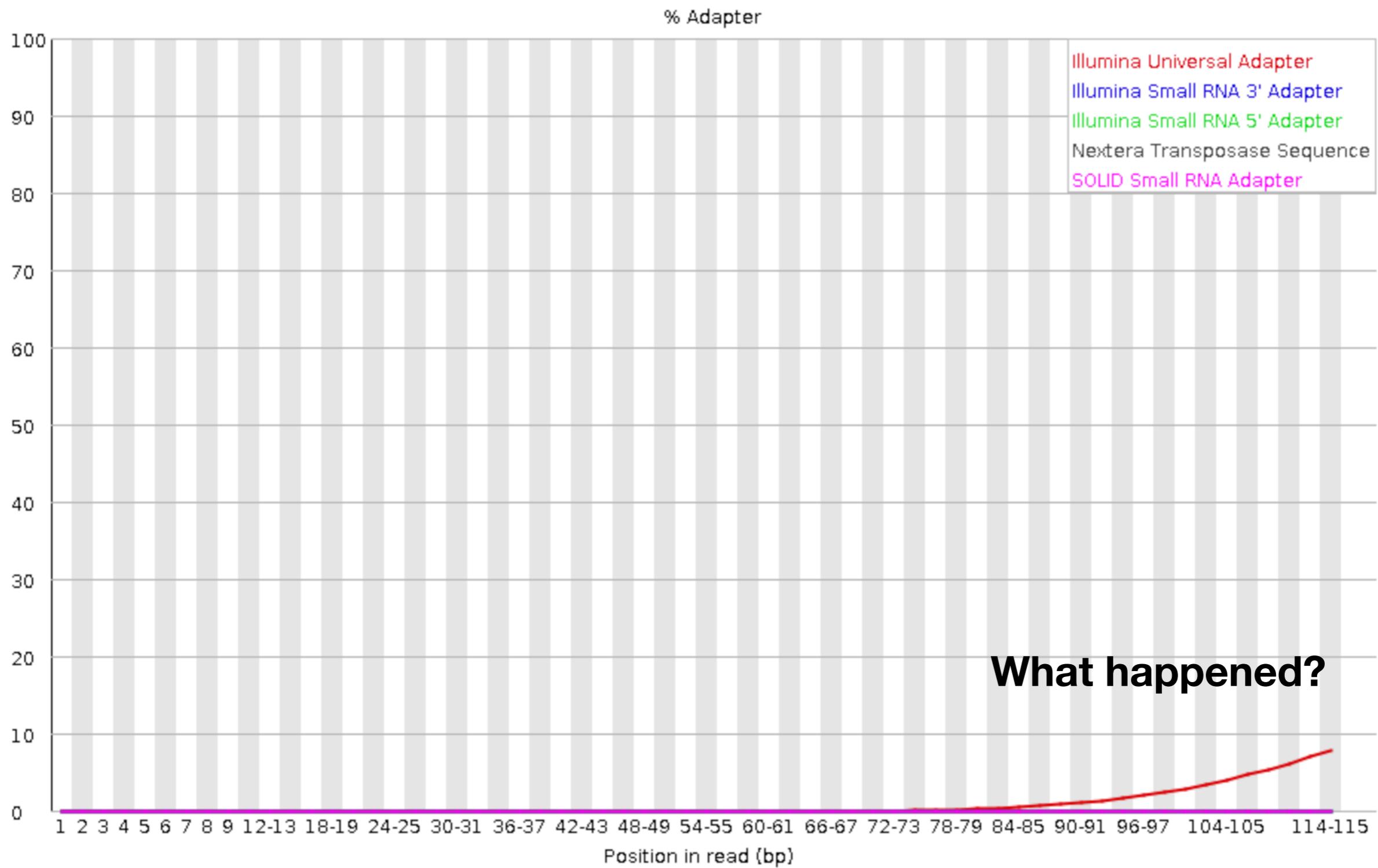


Overrepresented sequences

No overrepresented sequences



Adapter Content



Trimming FastQC using cutadapt

```
$ cutadapt -a AGATCGGAAGAGCACACGTCTGAACTCCAGTCAC -m 20 -o SRR5715043.trimmed.fastq.gz SRR5715043.fastq.gz
```

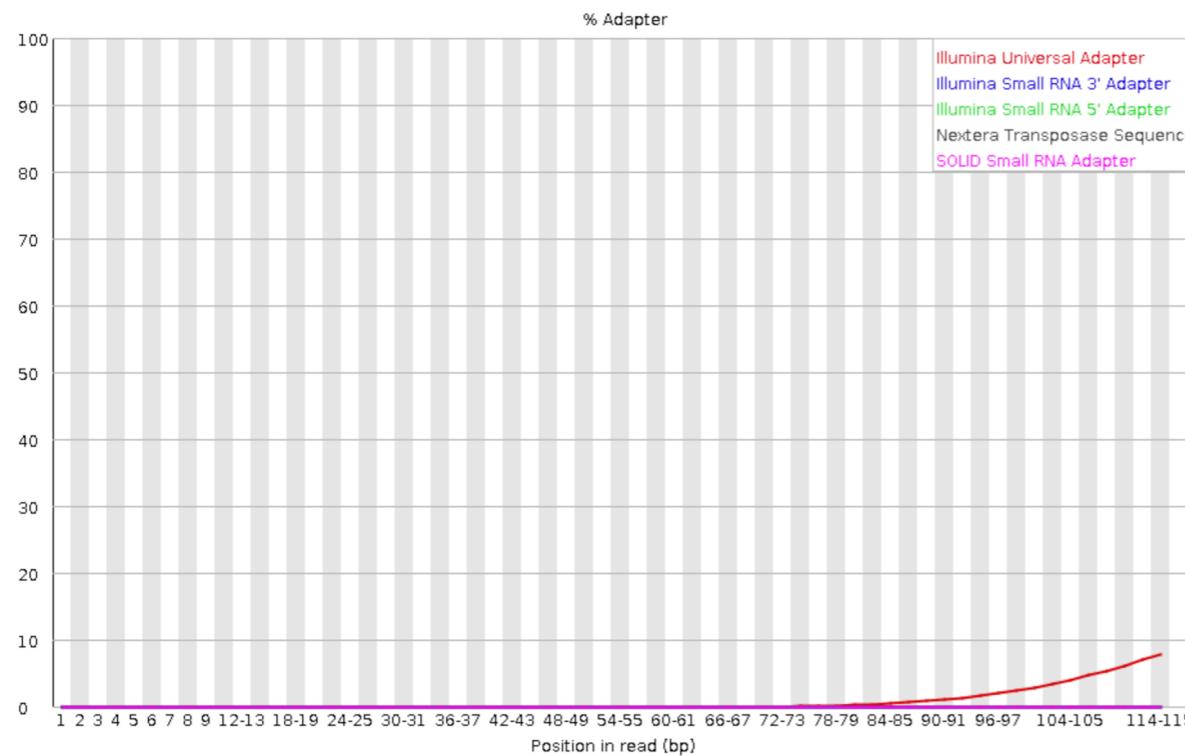
Before

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%GC	49

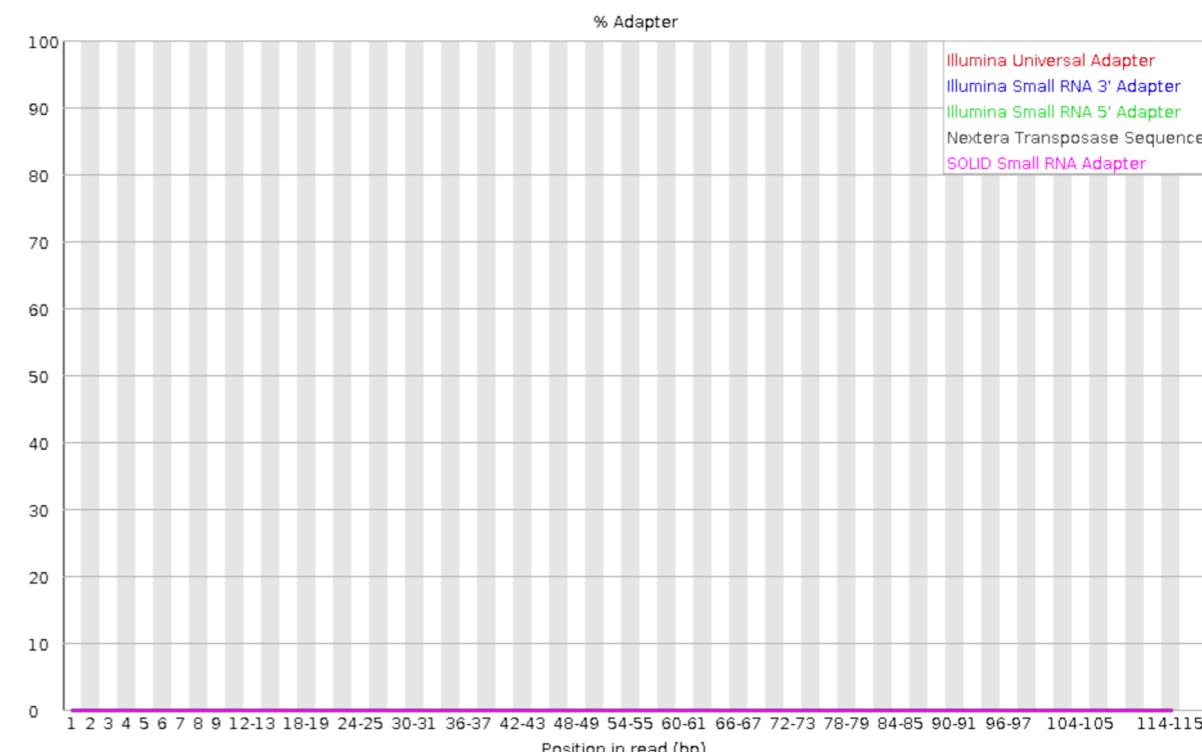
After

Measure	Value
Filename	SRR5715043.trimmed.fastq.gz
File type	Conventional base calls
Encoding	Sanger / Illumina 1.9
Total Sequences	36022775
Sequences flagged as poor quality	0
Sequence length	20-126
%GC	49

Adapter Content



Adapter Content



Alignment using STAR

Spliced Transcripts Alignments to a Reference (STAR)

Indexing (only one time per genome+transcript annotation)

```
$STAR --runMode genomeGenerate \
--runThreadN 6 \
--genomeFastaFiles genome.fa \
--sjdbGTFfile annotation.gtf \
--sjdbOverhang 99 \
--genomeDir STAR_index
```

Mapping (multiple samples)

```
$STAR --genomeDir ./Genome/STAR_Index \
--runThreadN 4 \
--readFilesCommand zcat \
--readFilesIn SRR5715043.trimmed.fastq.gz \
--outSAMtype BAM SortedByCoordinate \
--outFileNamePrefix SRR5715043.
```

A review of module 5

Excerpted from Loyal Goff's slide

Alignment QC - What are we looking for?

- **Number** of reads aligned per sample
- **Percentage** of reads aligned per sample
- **Number/percentage** of reads uniquely mapping
 - % of reads with multiple mapping to reference (ambiguous)
- Paired End read **concordance**
 - # of PE reads uniquely mapping
 - # of PE reads mapping but not together (discordant)
 - # of PE reads where only one mapped [uniquely]
- **Consistent** scores/values across samples
- *Values are often reported after each sample is processed. Useful to try and capture this information in a log file.*

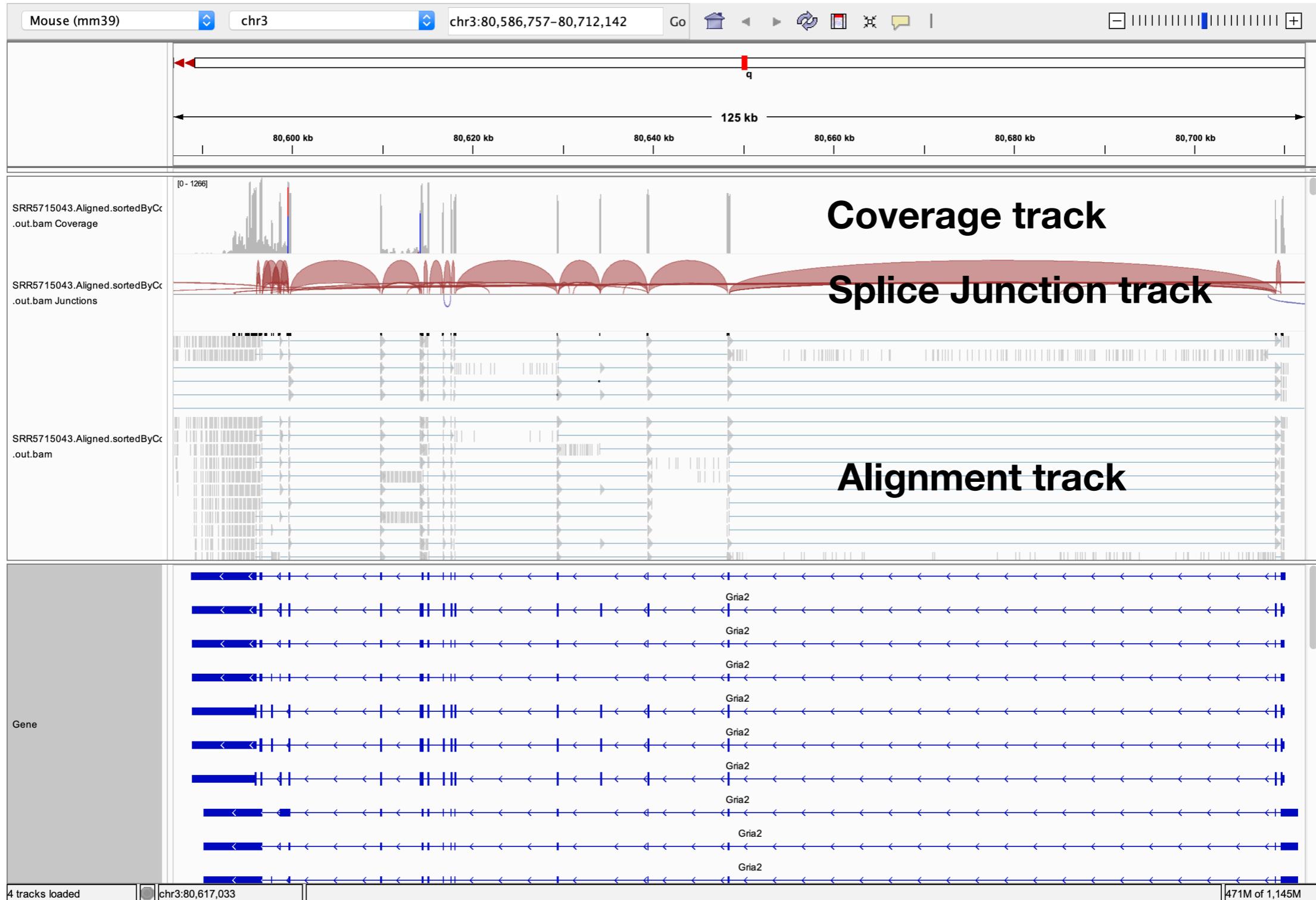
STAR Output example

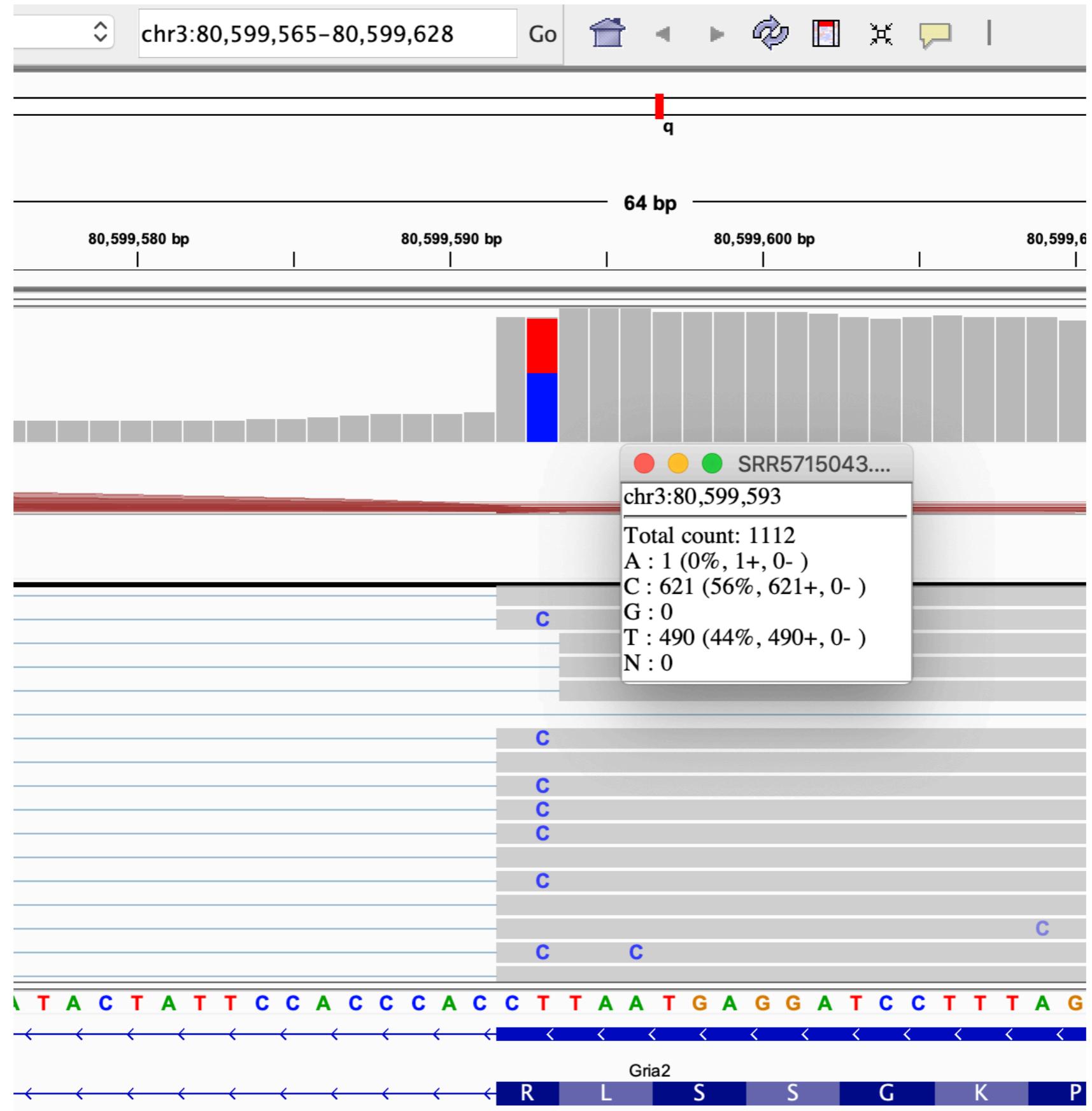
```
$ cat SRR5715043.Log.final.out
```

Started job on	Oct 03 17:10:04
Started mapping on	Oct 03 17:10:55
Finished on	Oct 03 17:16:23
Mapping speed, Million of reads per hour	395.37
Number of input reads	36022775
Average input read length	123
UNIQUE READS:	
Uniquely mapped reads number	33636271
Uniquely mapped reads %	93.38%
Average mapped length	123.01
Number of splices: Total	11521857
Number of splices: Annotated (sjdb)	11418927
Number of splices: GT/AG	11399495
Number of splices: GC/AG	98254
Number of splices: AT/AC	12071
Number of splices: Non-canonical	12037
Mismatch rate per base, %	0.11%
Deletion rate per base	0.01%
Deletion average length	1.96
Insertion rate per base	0.00%
Insertion average length	1.54
MULTI-MAPPING READS:	
Number of reads mapped to multiple loci	1724928
% of reads mapped to multiple loci	4.79%
Number of reads mapped to too many loci	74776
% of reads mapped to too many loci	0.21%
UNMAPPED READS:	
Number of reads unmapped: too many mismatches	0
% of reads unmapped: too many mismatches	0.00%
Number of reads unmapped: too short	536652
% of reads unmapped: too short	1.49%
Number of reads unmapped: other	50148
% of reads unmapped: other	0.14%
CHIMERIC READS:	
Number of chimeric reads	0
% of chimeric reads	0.00%

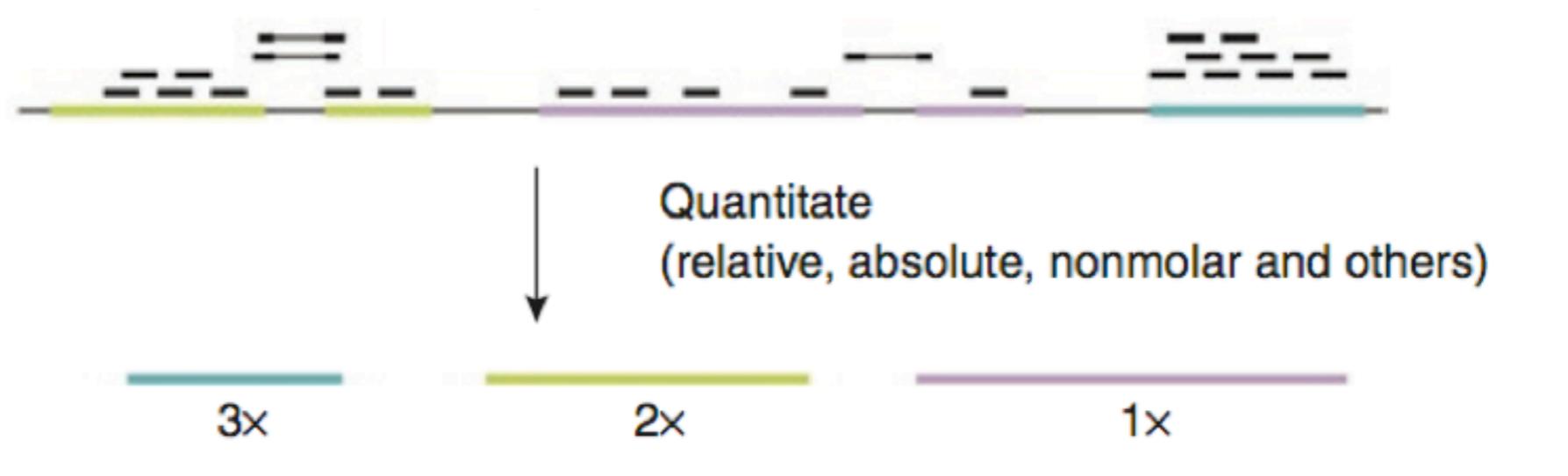
Visualization with IGV

```
$ samtools index  
SRR5715043.Aligned.sortedByCoord.out.bam
```





Counting reads using featureCounts



```
$featureCounts SRR5715043.Aligned.sortedByCoord.out.bam \
-o SRR5715043.fCounts \
-a annotation.gtf \
-s 2 \
-T 4 \
-g gene_id \
-C
```

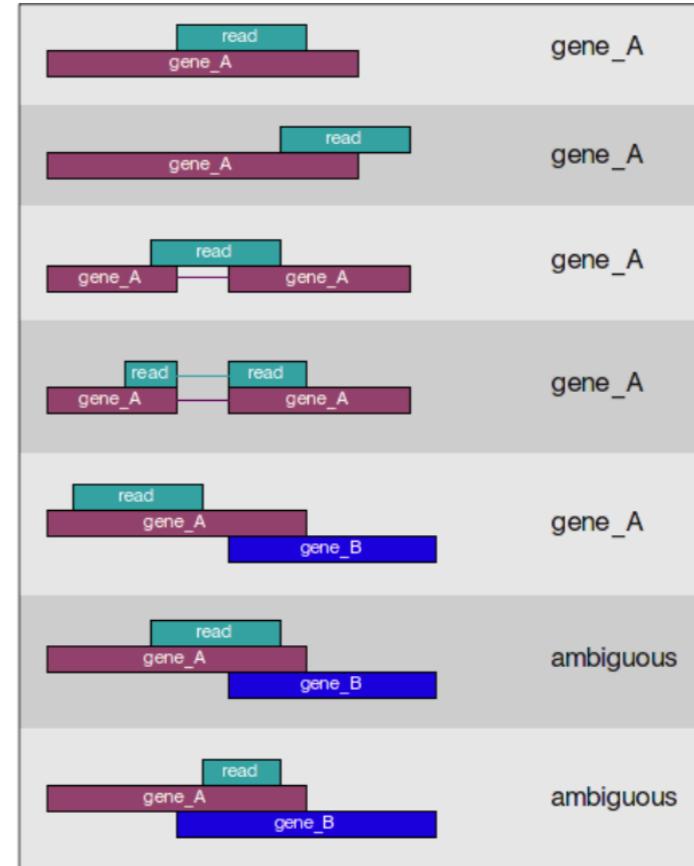
FeatureCounts Output example

```
$ cat SRR5715043.fCounts.summary
```

```
=====
===== 500 READING v2.0.3
=====
//===== featureCounts setting =====\\
Input files : 1 BAM file
              SRR5715043.Aligned.sortedByCoord.out.bam
Output file : SRR5715043.fCounts
Summary : SRR5715043.fCounts.summary
Paired-end : no
Count read pairs : no
Annotation : gencode.vM30.annotation.gtf (GTF)
Dir for temp files : ./3.Count
Threads : 4
      Level : meta-feature level
Multimapping reads : not counted
Multi-overlapping reads : not counted
Min overlapping bases : 1
\\=====
```

```
'Status ./2.Alignment/SRR5715043.Aligned.sortedByCoord.o
Assigned 28832430
Unassigned_Unmapped 0
Unassigned_Read_Type 0
Unassigned_Singleton 0
Unassigned_MappingQuality 0
Unassigned_Chimera 0
Unassigned_FragmentLength 0
Unassigned_Duplicate 0
Unassigned_MultiMapping 4161004
Unassigned_Secondary 0
Unassigned_NonSplit 0
Unassigned_NoFeatures 4381820
Unassigned_Overlapping_Length 0
Unassigned_Ambiguity 422021
```

```
//===== Running =====\\
Load annotation file gencode.vM30.annotation.gtf ...
Features : 868862
Meta-features : 56691
Chromosomes/contigs : 22
Process BAM file SRR5715043.Aligned.sortedByCoord.out.bam...
Strand specific : reversely stranded
Single-end reads are included.
Total alignments : 37797275
Successfully assigned alignments : 28832430 (76.3%)
Running time : 0.16 minutes
Write the final count table.
Write the read assignment summary.
8 Summary of counting results can be found in file "./3.Count/SRR5715043.fCounts.summary"
\\=====
```



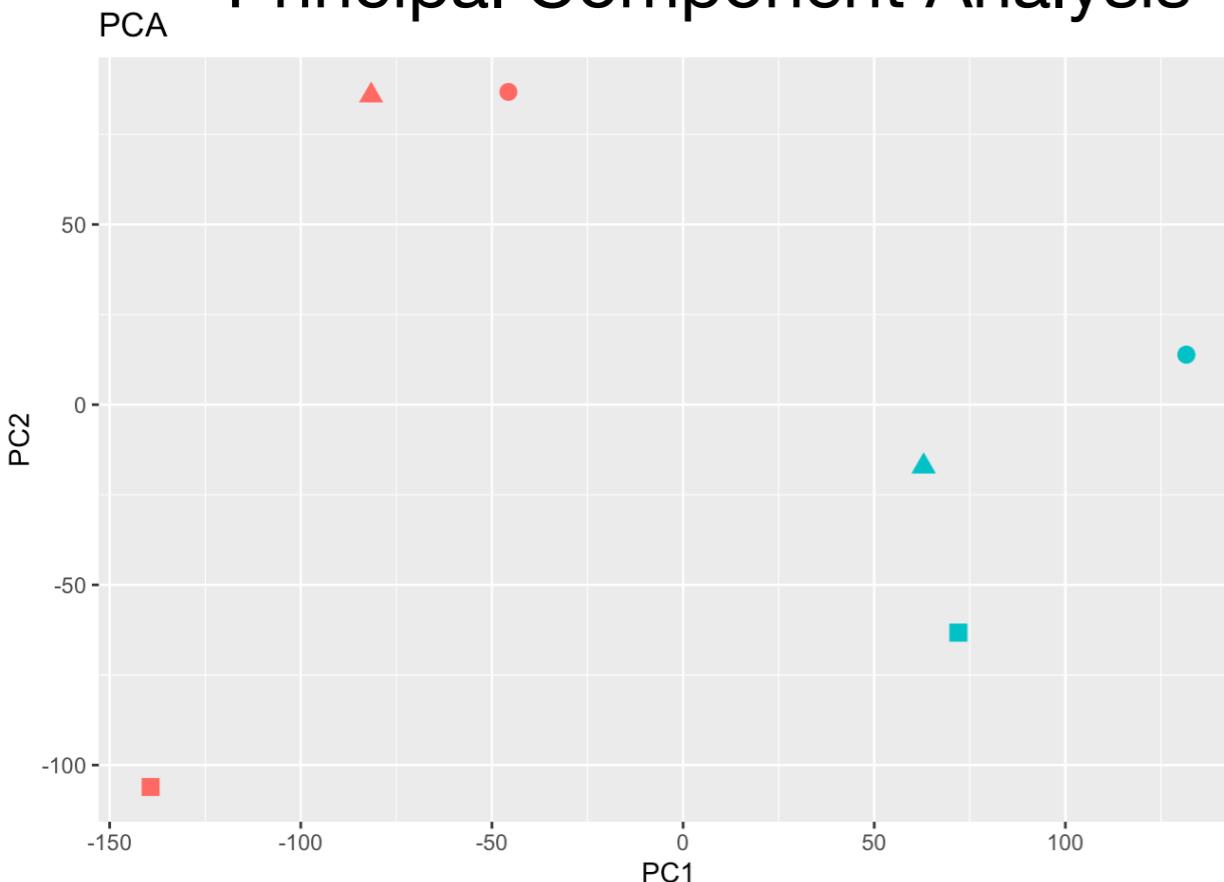
Exploratory Data Analysis (EDA)

Normalize count first !!

Normalization method	Description	Accounted factors	Recommendations for use
CPM (counts per million)	counts scaled by total number of reads	sequencing depth	gene count comparisons between replicates of the same sample group; NOT for within sample comparisons or DE analysis
TPM (transcripts per kilobase million)	counts per length of transcript (kb) per million reads mapped	sequencing depth and gene length	gene count comparisons within a sample or between samples of the same sample group; NOT for DE analysis
RPKM/FPKM (reads/fragments per kilobase of exon per million reads/fragments mapped)	similar to TPM	sequencing depth and gene length	gene count comparisons between genes within a sample; NOT for between sample comparisons or DE analysis
DESeq2's median of ratios [1]	counts divided by sample-specific size factors determined by median ratio of gene counts relative to geometric mean per gene	sequencing depth and RNA composition	gene count comparisons between samples and for DE analysis; NOT for within sample comparisons
EdgeR's trimmed mean of M values (TMM) [2]	uses a weighted trimmed mean of the log expression ratios between samples	sequencing depth and RNA composition	gene count comparisons between samples and for DE analysis; NOT for within sample comparisons

Exploratory Data Analysis (EDA)

Principal Component Analysis



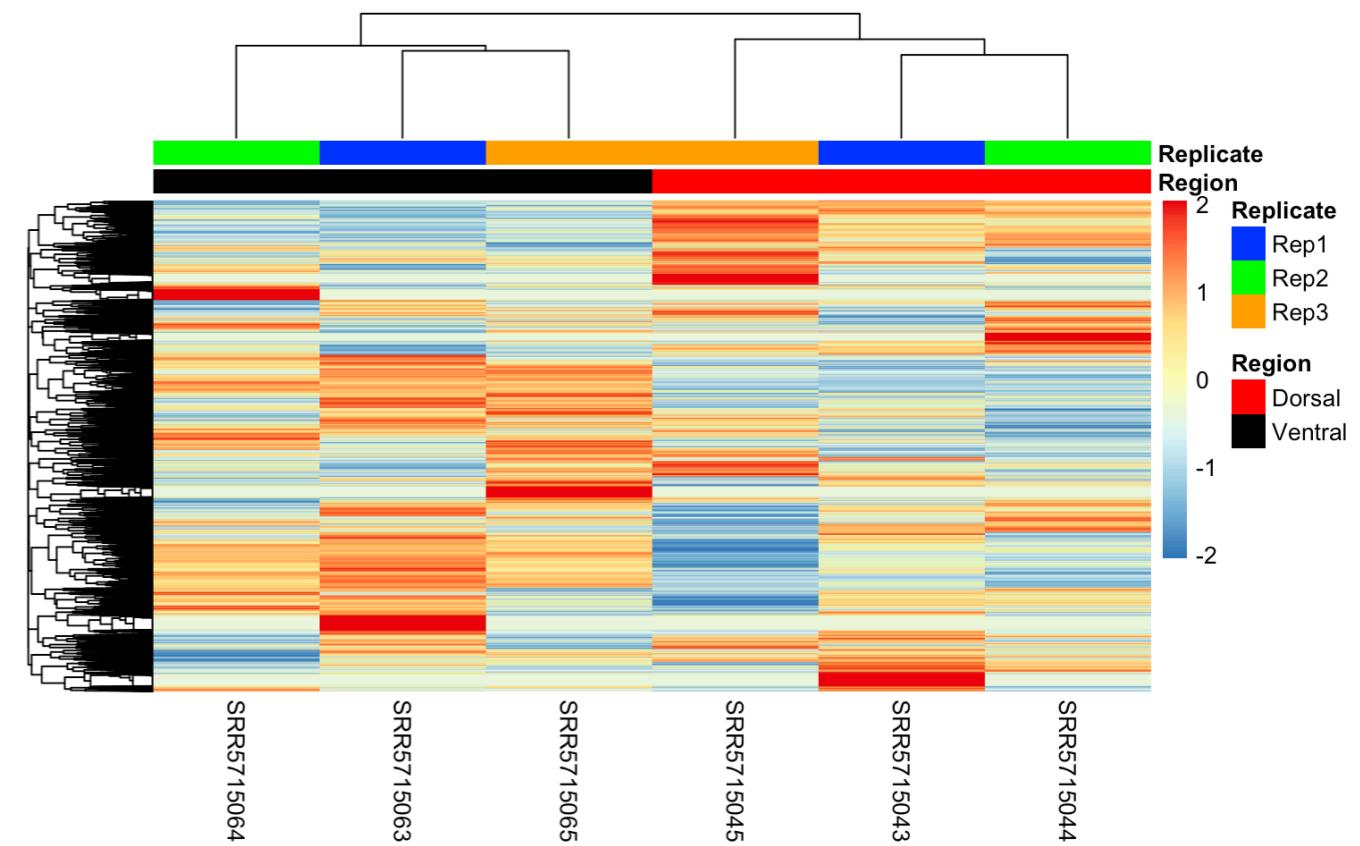
Replicate

- Rep1
- ▲ Rep2
- Rep3

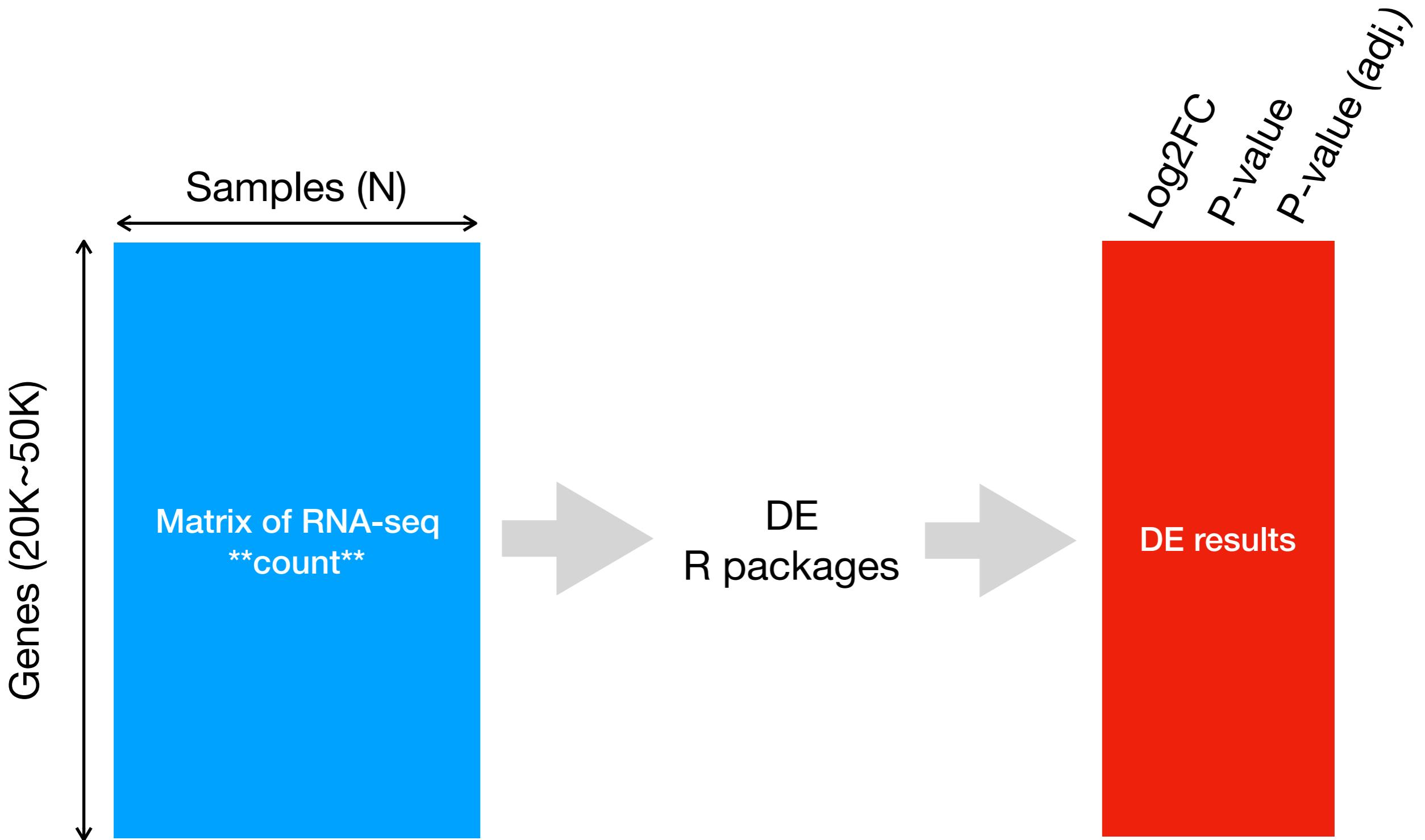
Region

- Dorsal
- Ventral

Heatmap



Differential Expression (DE)

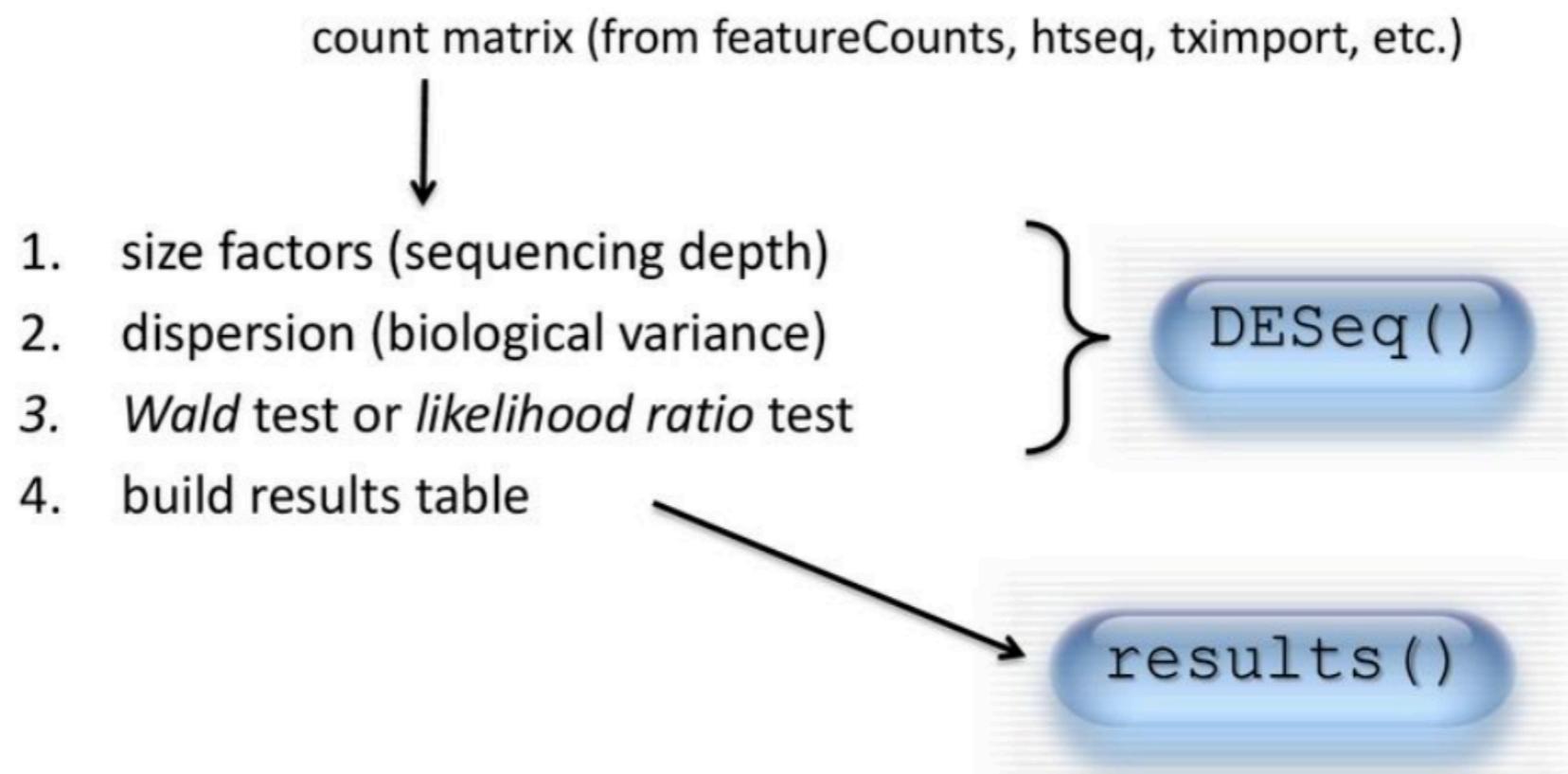


DE R packages

DE tools	Normalization	Count model	Statistical Test (Null: LFC==0)
EdgeR	TMM	Negative Binomial	Exact test or LRT
DESeq2	DESeq's median of ratio	Negative Binomial	Wald test or LRT
Limma-Voom	TMM	Log Normal	Moderated t-test

DESeq2

```
dds <- DESeqDataSetFromMatrix(countM, colData=sampleSheet,  
design= ~ Region)  
dds <- DESeq(dds)  
Res <- results(dds, contrast=c("Region", "Dorsal", "Ventral"))
```



Design matrix

```
dds <- DESeqDataSetFromMatrix(countM, colData=sampleSheet,  
design= ~ Region)
```

- Simple comparison

design = ~ region

- Controlling covariates

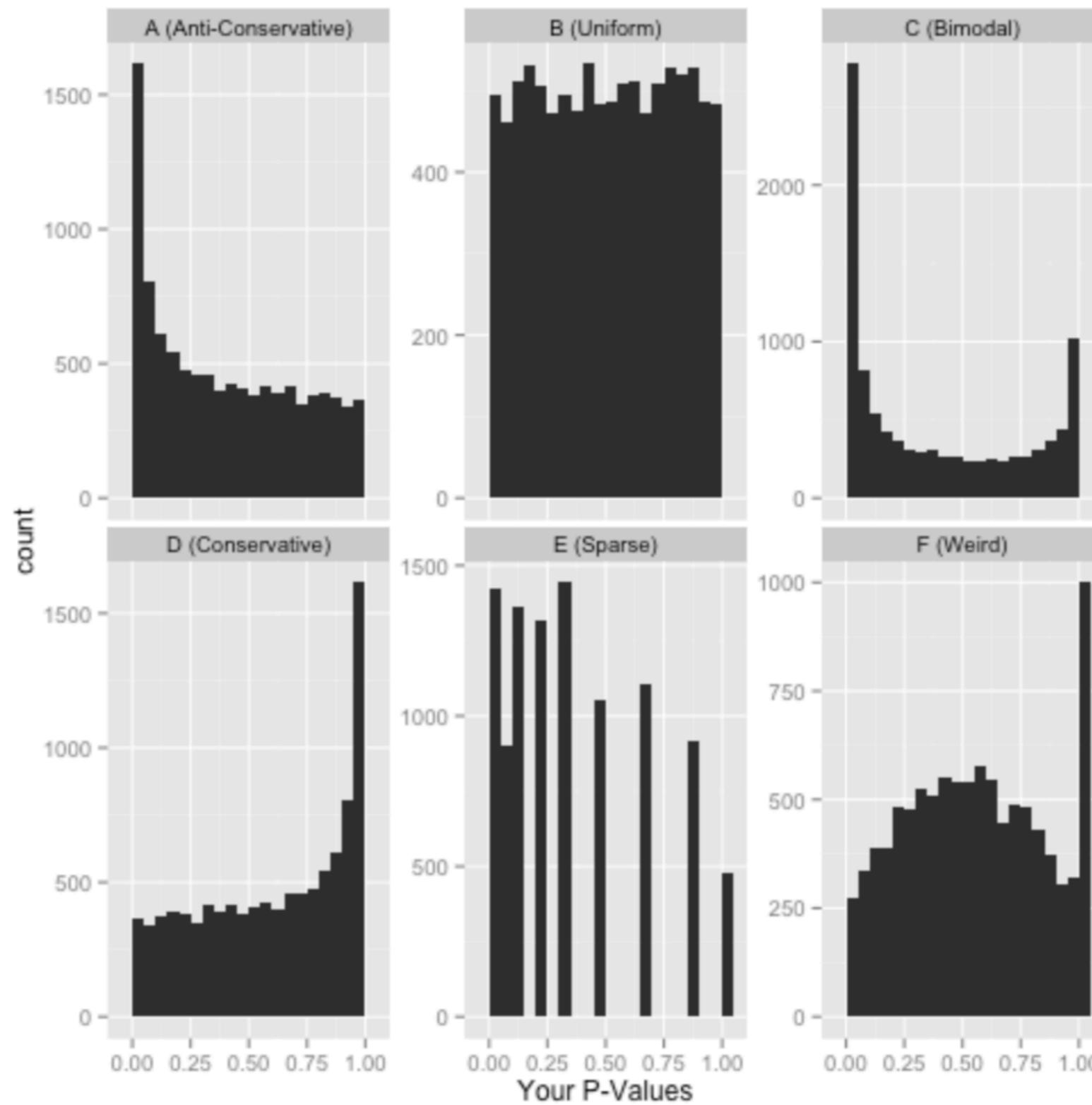
design = ~ replicate + region

design = ~ replicate + sex + region

- Investigating interaction effects

design = ~ replicate + sex + region + sex : region

DE QC: p-value distribution

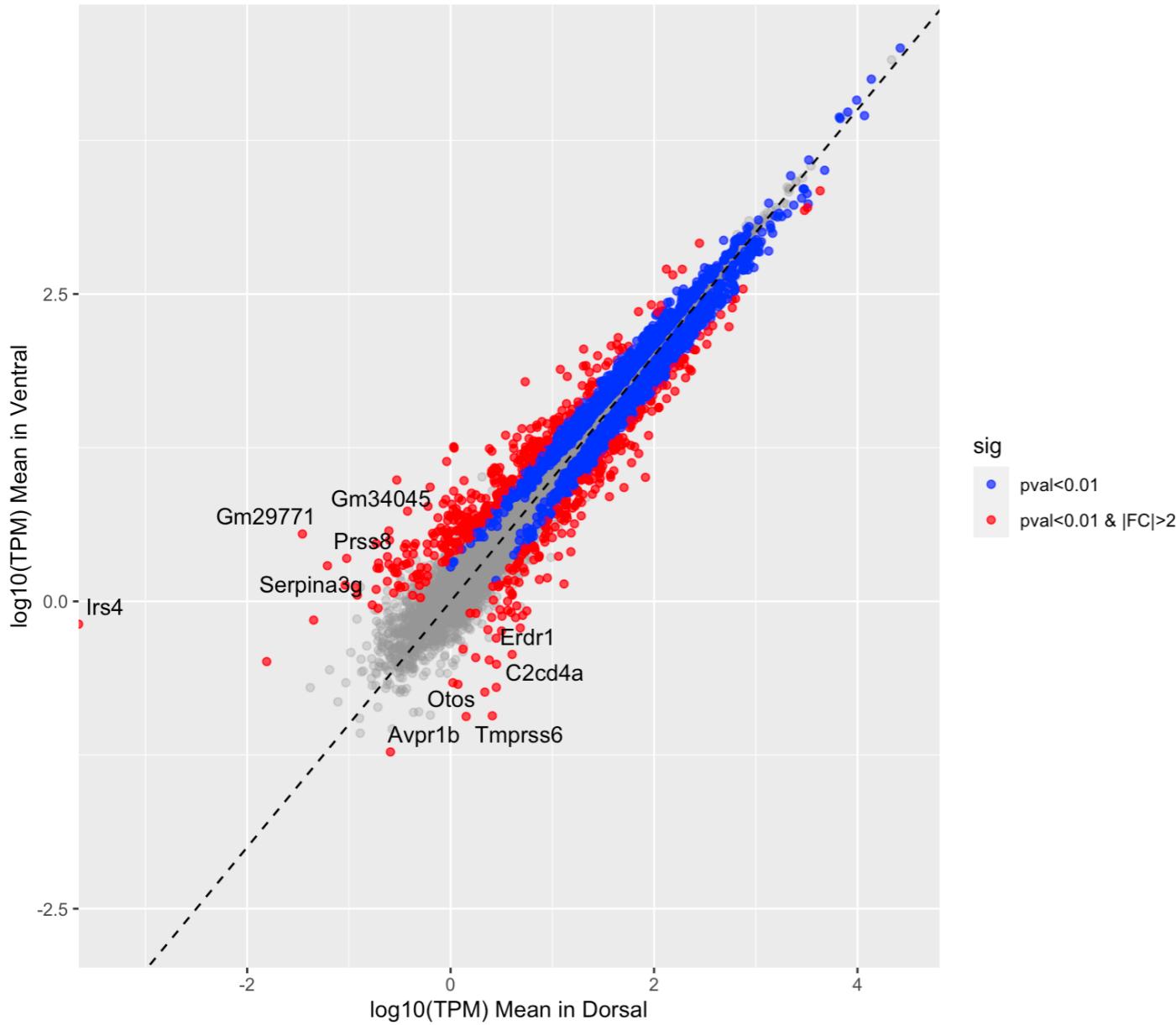


DE Results

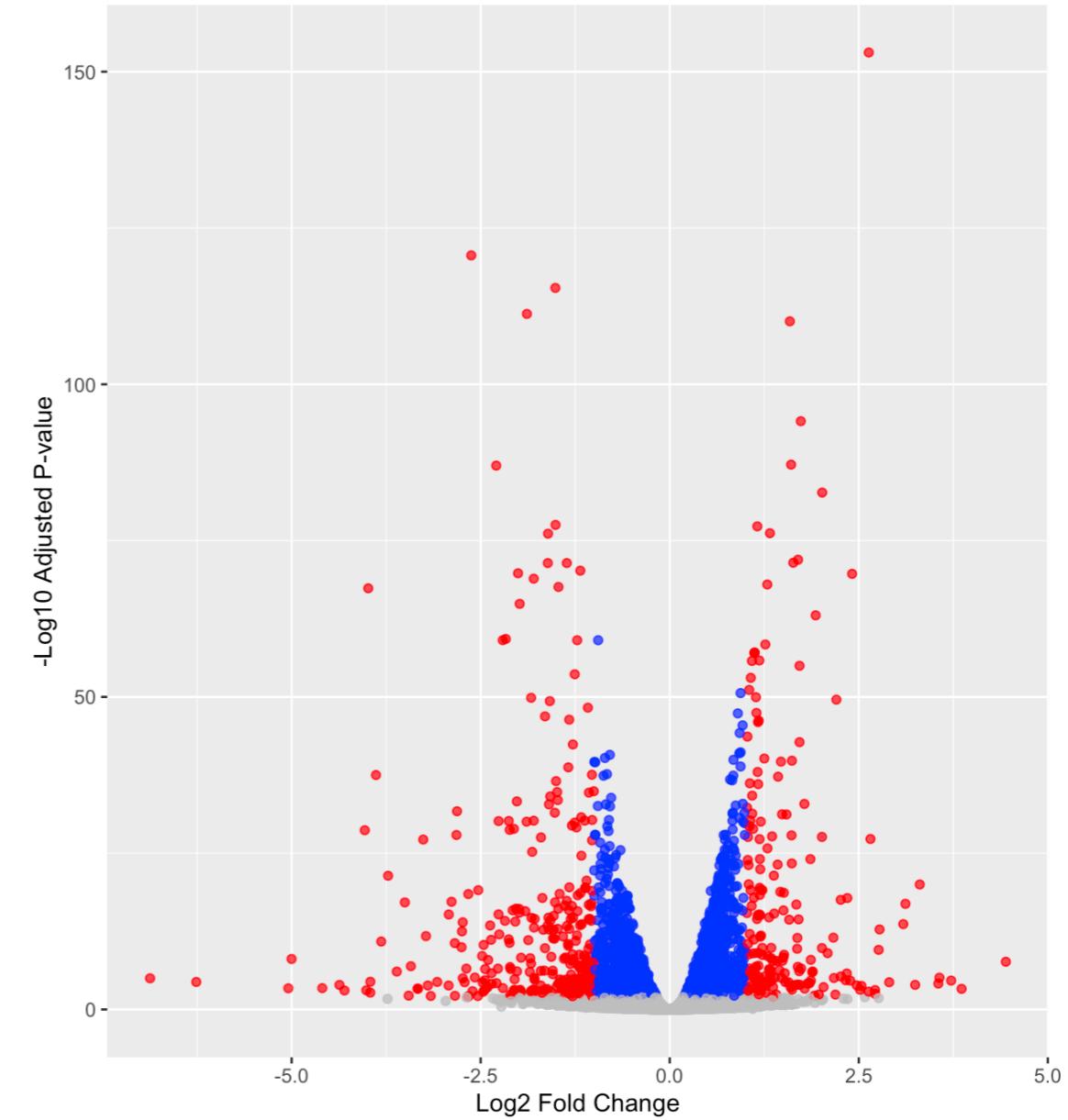
```
> res
log2 fold change (MLE): Region Dorsal vs Ventral
Wald test p-value: Region Dorsal vs Ventral
DataFrame with 56691 rows and 6 columns
  baseMean log2FoldChange lfcSE      stat      pvalue      padj
  <numeric>    <numeric> <numeric>    <numeric>    <numeric>    <numeric>
ENSMUSG00000102693.2      0        NA       NA       NA       NA       NA
ENSMUSG0000064842.3      0        NA       NA       NA       NA       NA
ENSMUSG0000051951.6  934.374636735581 -0.3552436281259 0.103883171365474 -3.41964558317256 0.000627027707598258 0.0042158466553528
ENSMUSG00000102851.2      0        NA       NA       NA       NA       NA
ENSMUSG00000103377.2  0.965337795076986 -0.10445910151227 2.64821564471098 -0.0394450888925515 0.968535532124425       NA
...
...          ...
ENSMUSG0000064368.1  554.832874720361 -0.268513399642424 0.121424238061778 -2.21136573659874 0.0270105205370121 0.0973450770345984
ENSMUSG0000064369.1  4.70528683513818 -0.229770388610795 1.14361158725448 -0.200916457275861 0.840763899060845       NA
ENSMUSG0000064370.1  153824.906100841 -0.188788470434184 0.0613087510925515 -3.07930706579212 0.00207482709210156 0.0118964348163462
ENSMUSG0000064371.1  14.6385304120105 0.333703279931054 0.656922522781579 0.507979660246796 0.611467607501022 0.808762996851356
ENSMUSG0000064372.1  29.7519207584107 -0.239430038273013 0.465265290440962 -0.51460971448373 0.606825775339794 0.806035794669969
```

DE Results Plot

- Scatter plot

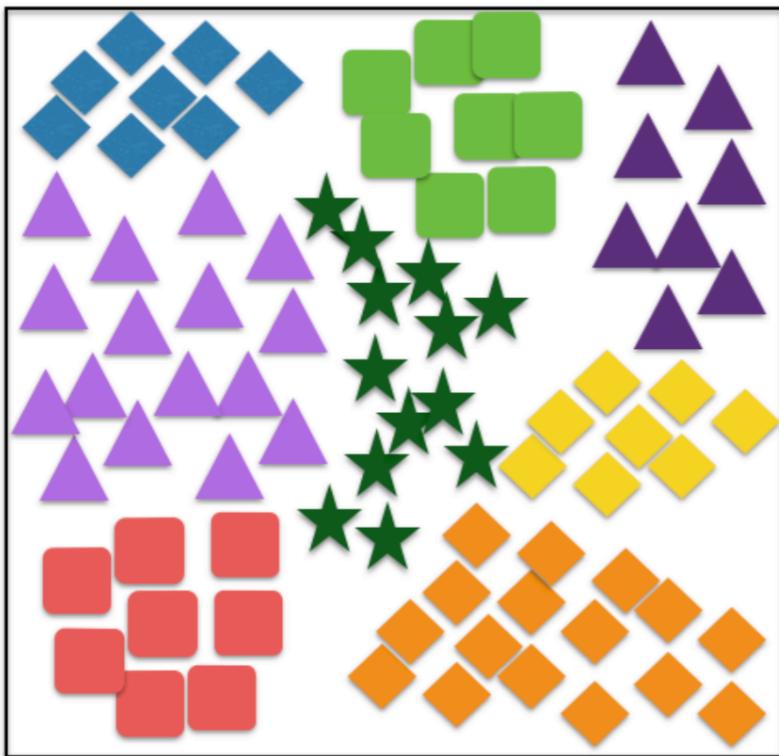


- Volcano plot

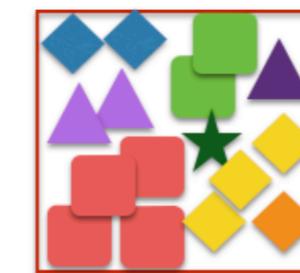


Enrichment analysis

All known genes in a species
(categorized into groups)



DE analysis



DEGs

Genes categories	Organism-specific Background	DE results	Over-represented?
Functional category 1	35/13000	25/1000	Likely
Functional category 2	56/13000	4/1000	Unlikely
Functional category 3	90/13000	8/1000	Unlikely
Functional category 4	15/13000	10/1000	Likely
...			

Hypergeometric test

Genes categories	Organism-specific Background	DE results	Over-represented?
Functional category 1	35/13000	25/1000	Likely
Functional category 2	56/13000	4/1000	Unlikely
Functional category 3	90/13000	8/1000	Unlikely
Functional category 4	15/13000	10/1000	Likely
...			

Null distribution: hypergeometric distribution

$$\Pr(X = k) = \frac{\binom{K}{k} \binom{N-K}{n-k}}{\binom{N}{n}}$$

For category 1, N=13000, n=25, K=35

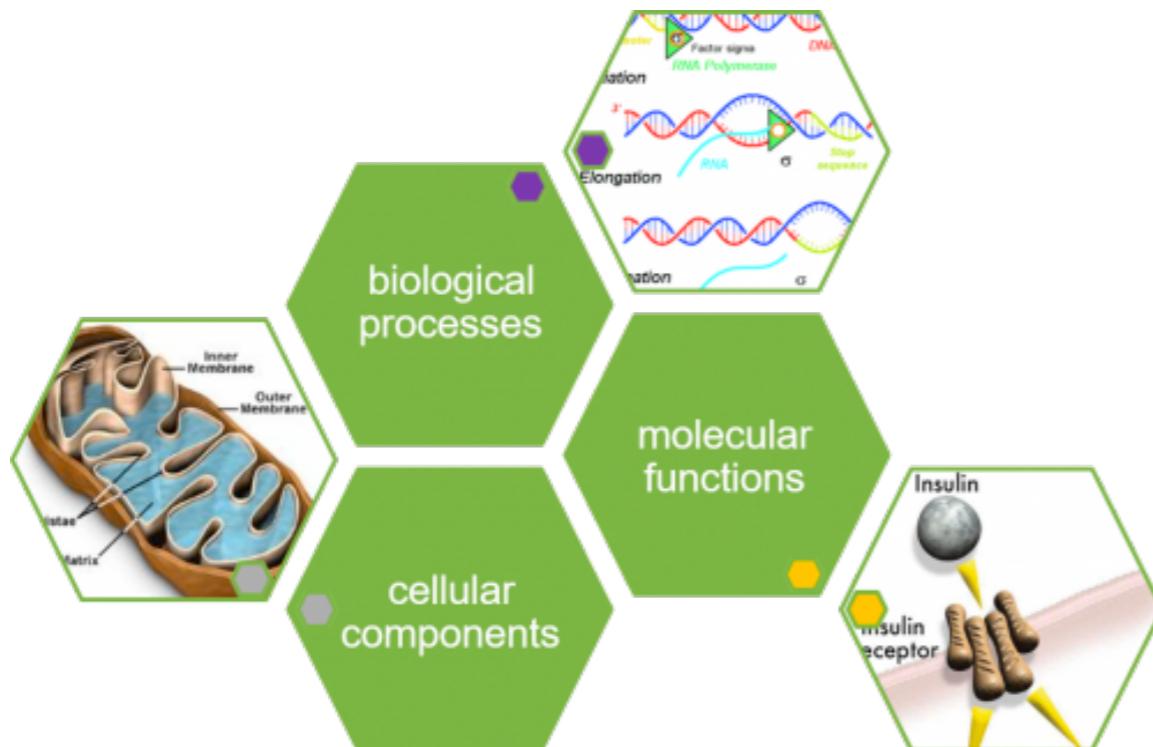
For category 2, N=13000, n=25, K=56

Gene Ontology database

GO:1903204

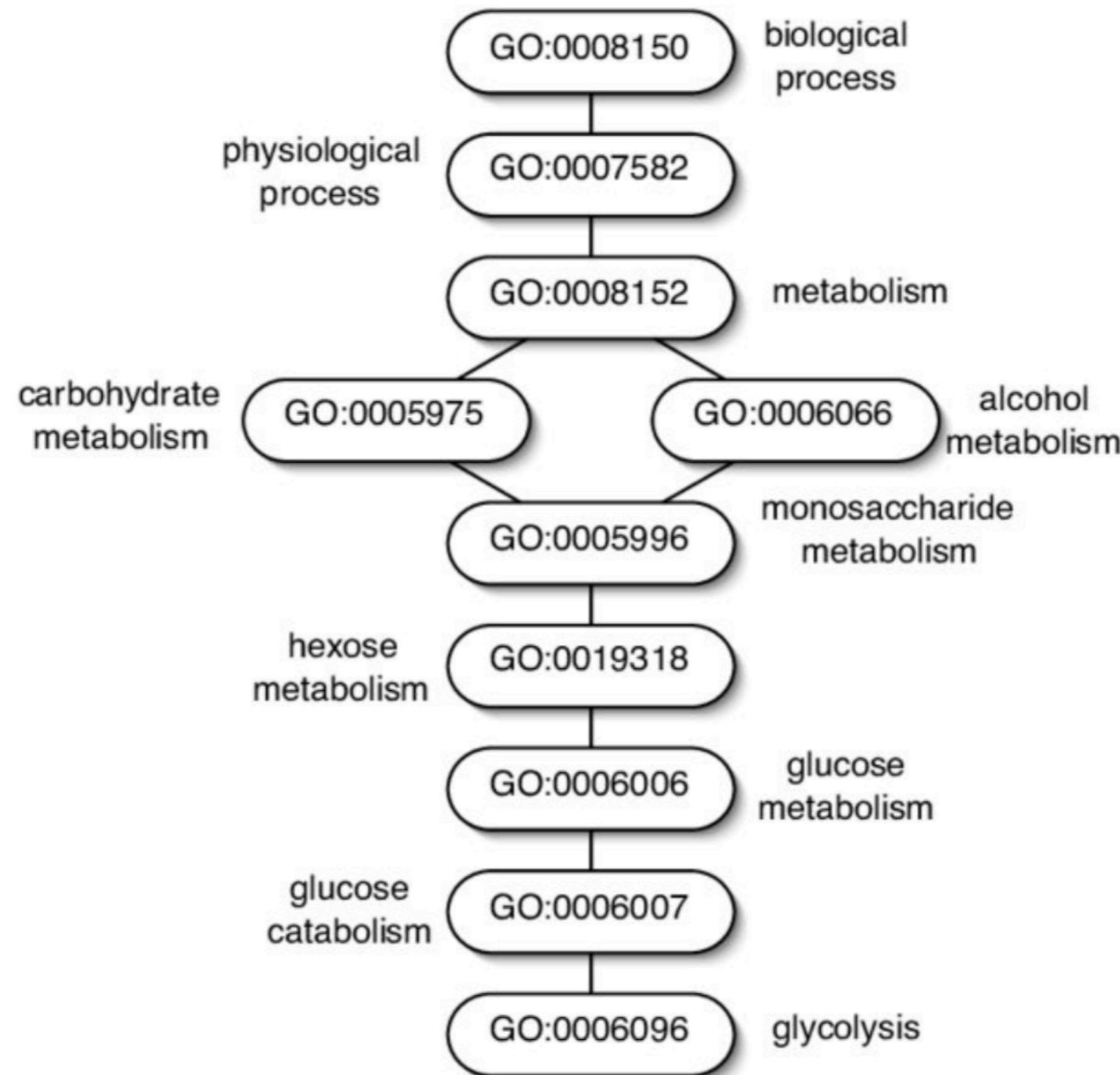
negative regulation of oxidative stress-induced neuron death

(#18) FBXO7, NONO, PARK7, RACK1, WNT1, PINK1, REST, IL10, NCOA7, MEAK7, HIF1A, SLC7A11, FZD1, GFER, CTNNB1, NR4A3, OXR1, ATF4.

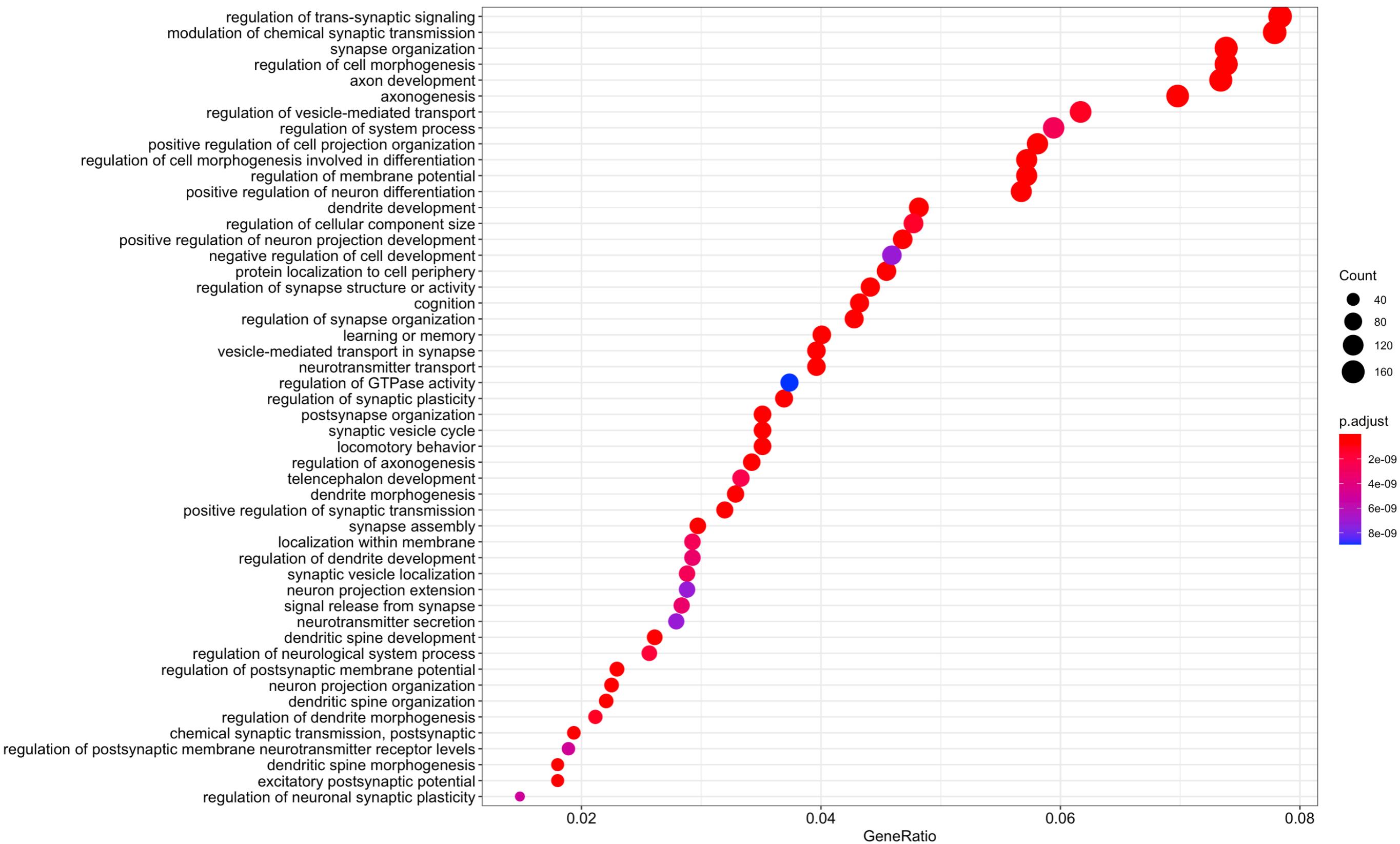


“cytochrome c” can be described by the **molecular function oxidoreductase activity**, the **biological process oxidative phosphorylation**, and the **cellular component mitochondrial matrix**.

GO: Hierarchical structure



GO terms enriched in DE genes



KEGG pathway database

<https://www.genome.jp/kegg/pathway.html>

KEGG PATHWAY is a collection of manually drawn [pathway maps](#) representing our knowledge of the molecular interaction, reaction and relation networks for:

1. Metabolism

[Global/overview](#) [Carbohydrate](#) [Energy](#) [Lipid](#) [Nucleotide](#) [Amino acid](#) [Other amino](#) [Glycan](#)

[Cofactor/vitamin](#) [Terpenoid/PK](#) [Other secondary metabolite](#) [Xenobiotics](#) [Chemical structure](#)

2. Genetic Information Processing

3. Environmental Information Processing

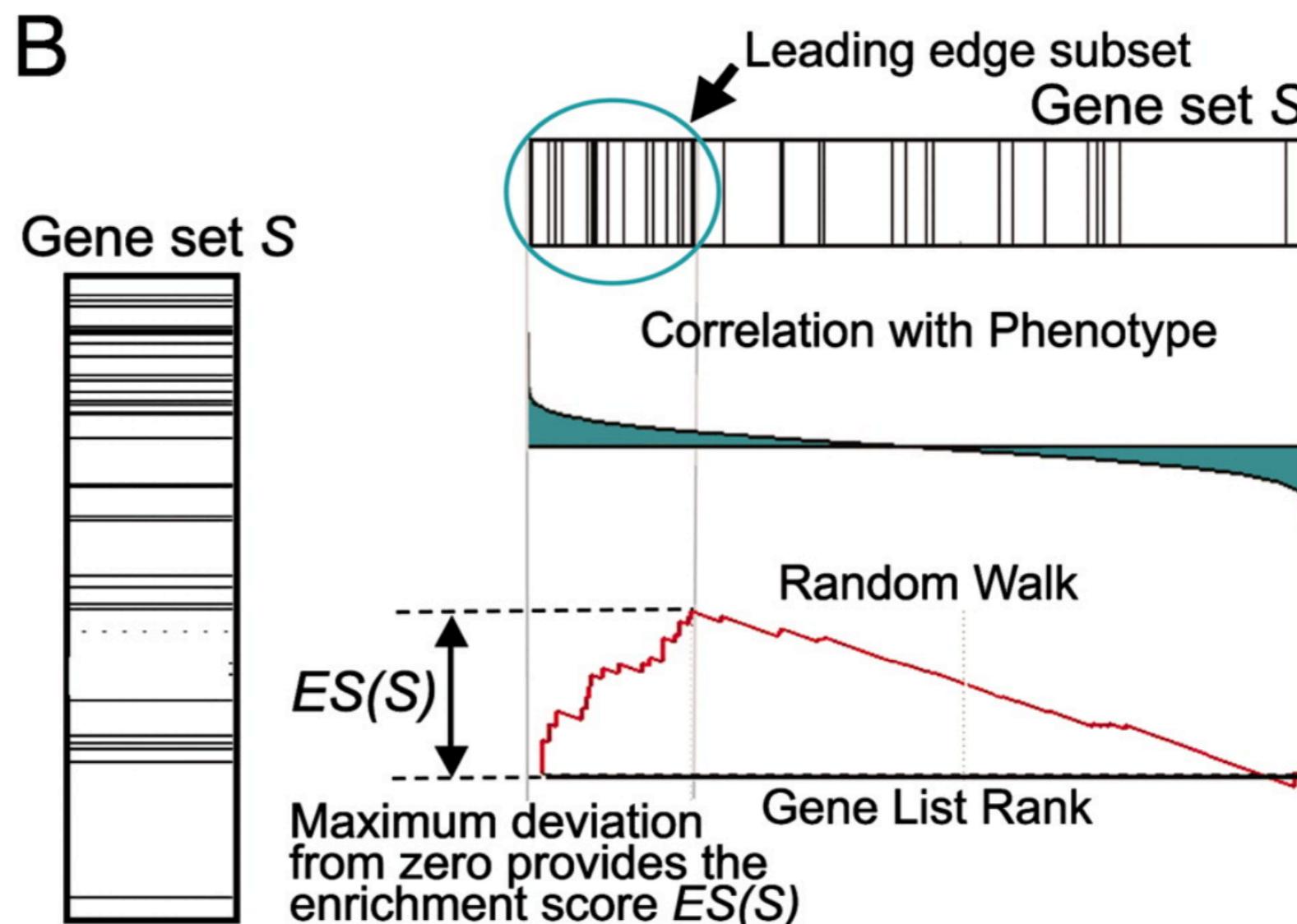
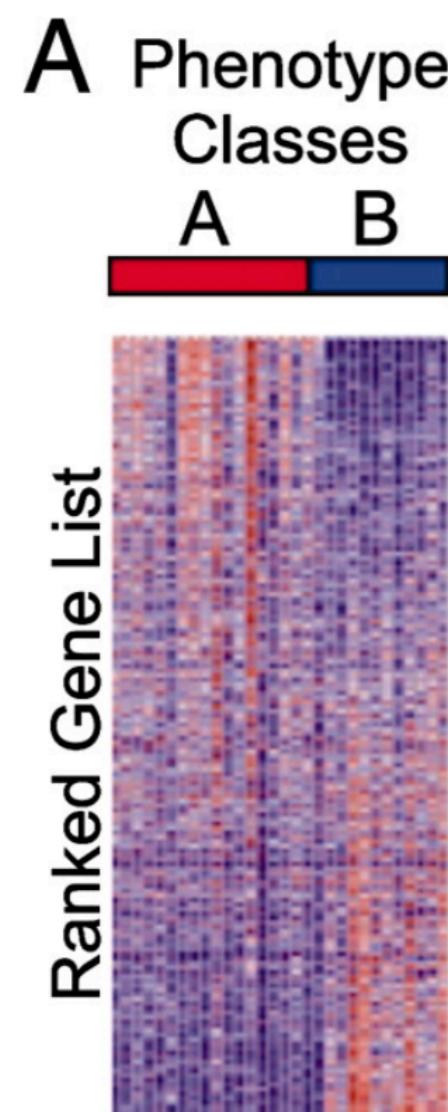
4. Cellular Processes

5. Organismal Systems

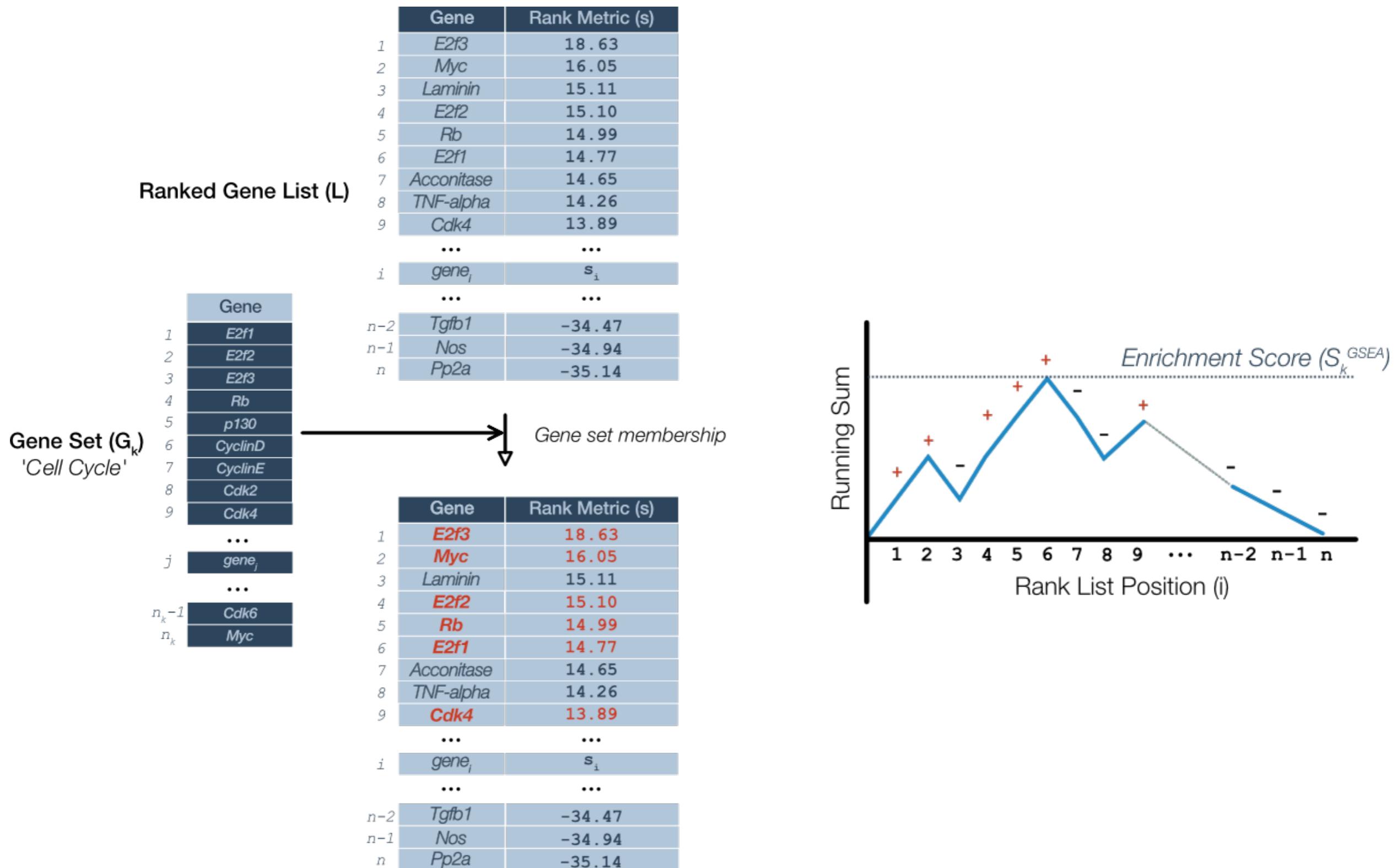
6. Human Diseases

7. Drug Development

Gene Set Enrichment Analysis (GSEA)

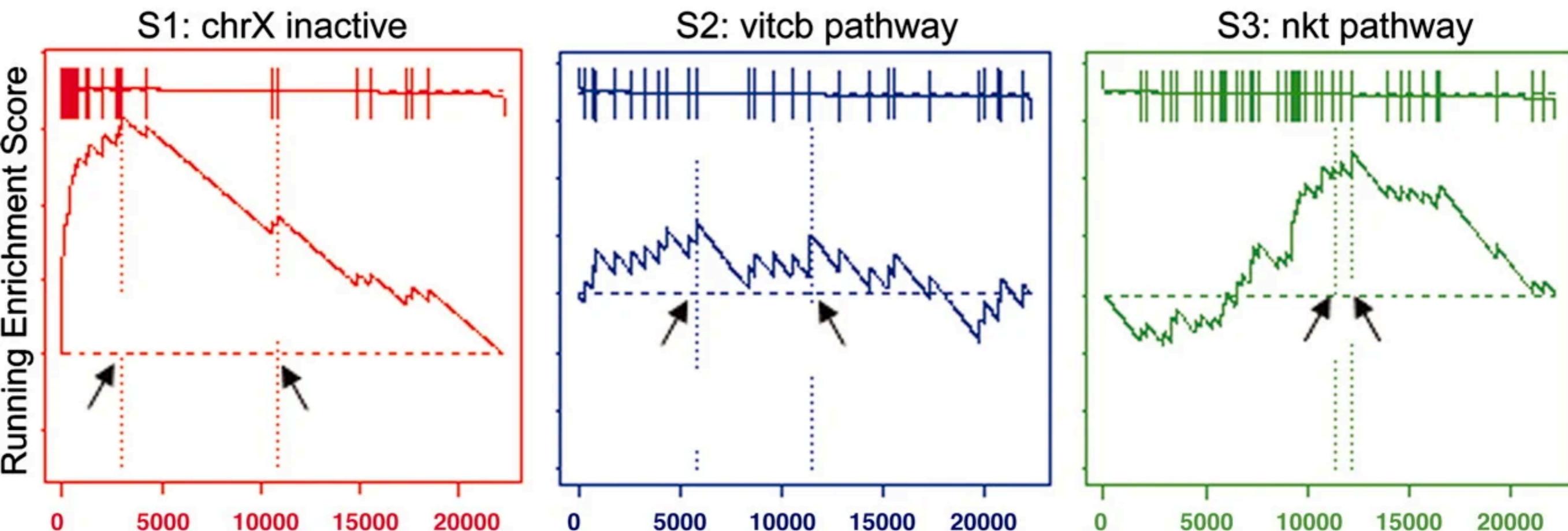


GSEA enrichment score



GSEA enrichment examples

the list of genes in the male/female lymphoblastoid cell line example ranked by their correlation with gender



Readings

- **Teaching materials at the Harvard Chan Bioinformatics Core (recommended)**
https://github.com/hbctraining/DGE_workshop_salmon_online/blob/master/schedule/links-to-lessons.md
- **UC-Davis Workshop material**
https://ucdavis-bioinformatics-training.github.io/2019_March_UCSF_mRNAseq_Workshop/data_reduction/alignment.html
- **Galaxy training**
<https://training.galaxyproject.org/archive/2019-02-07/topics/transcriptomics/tutorials/ref-based/tutorial.html>