## R. Notebook

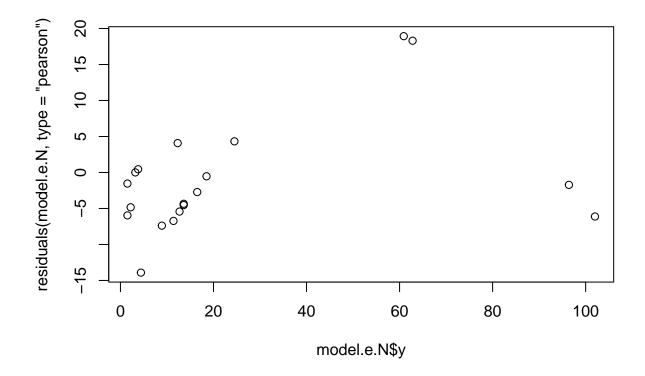
```
data1<-read.table("/Users/phamkhoa/Documents/university/3/stats_model_1/w2/canoladiesel.txt", sep="\t",
attach(data1)
# Problem 1
#a
model.inv1.ex1<-glm(Yield~Time, family = gaussian(link = "inverse"), data = data1)</pre>
summary(model.inv1.ex1)
##
## Call:
## glm(formula = Yield ~ Time, family = gaussian(link = "inverse"),
       data = data1)
##
## Deviance Residuals:
       Min 1Q Median
                                           Max
## -46.845 -7.784 -3.919 3.123
                                        50.755
## Coefficients:
                Estimate Std. Error t value Pr(>|t|)
## (Intercept) 0.145935
                           0.093478
                                     1.561
                                               0.137
## Time
               -0.002809
                          0.002085 -1.347
                                               0.196
##
\#\# (Dispersion parameter for gaussian family taken to be 667.9073)
##
       Null deviance: 17848 on 18 degrees of freedom
## Residual deviance: 11355 on 17 degrees of freedom
## AIC: 181.39
## Number of Fisher Scoring iterations: 7
newdata <-data.frame(Time = 40)</pre>
predict(model.inv1.ex1, newdata = newdata, type = "response")
##
          1
## 29.79662
#b
model.inv1gamma.ex1<-glm(Yield~Time, family = Gamma(link = "inverse"), data = data1)</pre>
model.identity1gamma.ex1<-glm(Yield~Time, family = Gamma(link = "identity"), data = data1)</pre>
model.log1gamma.ex1<-glm(Yield~Time, family = Gamma(link = "log"), data = data1)</pre>
AIC(model.identity1gamma.ex1)
```

```
## [1] 152.502
AIC(model.inv1gamma.ex1)
## [1] 153.5631
AIC(model.log1gamma.ex1)
## [1] 151.7177
# choose (i) log link model
#c
model.log123gamma.ex1<-glm(Yield ~ Time + Temp + Methanol, family = Gamma(link = "log"), data=data1)
summary(model.log123gamma.ex1)
##
## Call:
## glm(formula = Yield ~ Time + Temp + Methanol, family = Gamma(link = "log"),
       data = data1)
##
## Deviance Residuals:
       Min
                 10
                     Median
##
                                   30
                                           Max
## -0.7117 -0.3192 -0.0254 0.2524
                                        0.4851
##
## Coefficients:
                 Estimate Std. Error t value Pr(>|t|)
## (Intercept) -17.402855 2.009810 -8.659 3.21e-07 ***
                            0.007972 8.385 4.80e-07 ***
## Time
                 0.066847
## Temp
                 0.065005
                           0.007776 8.360 4.99e-07 ***
## Methanol
                1.008914
                           0.239162 4.219 0.000745 ***
## Signif. codes: 0 '*** 0.001 '** 0.01 '* 0.05 '.' 0.1 ' 1
## (Dispersion parameter for Gamma family taken to be 0.170404)
       Null deviance: 28.3065 on 18 degrees of freedom
##
## Residual deviance: 2.7836 on 15 degrees of freedom
## AIC: 121.56
## Number of Fisher Scoring iterations: 8
newdata <-data.frame(Time = 40, Temp = 260, Methanol = 1.75)</pre>
pred<-predict(model.log123gamma.ex1, type="response", level=0.95, newdata=newdata, se.fit = TRUE)</pre>
eta<-predict(model.log123gamma.ex1, type="link", newdata=newdata, se.fit = TRUE)
link.lowerbound <- eta $fit-qnorm(0.975) *eta $se.fit
link.upperbound <- eta $fit + qnorm(0.975) * eta $se.fit
upper <- exp(link.upperbound) ### !!!!
lower<-exp(link.lowerbound) ### !!!!</pre>
```

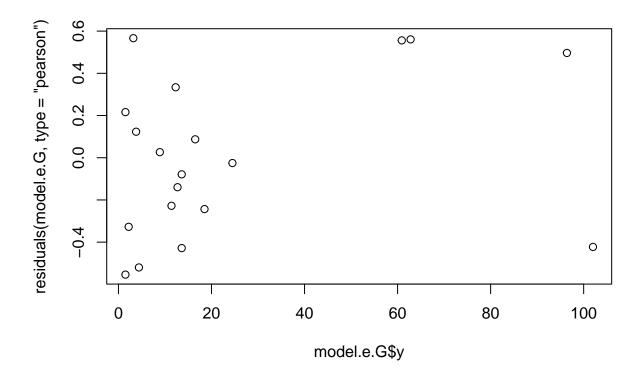
upper

```
##
## 68.33616
lower
## 38.51991
# 95% CI = (38.51991, 68.33616)
gammaH0 <- glm(Yield ~ Time + Temp, family = Gamma(link = "log"), data=data1)</pre>
test<-anova(gammaH0, model.log123gamma.ex1, test = "F")</pre>
test$`Pr(>F)`
## [1]
                NA 0.001575748
#e
model.e.N <- glm(Yield ~ Time + Temp + Methanol, family = gaussian(link = "log"), data=data1)
model.e.G <- glm(Yield ~ Time + Temp + Methanol, family = Gamma(link = "log"), data=data1)</pre>
model.e.IG <- glm(Yield ~ Time + Temp + Methanol, family = inverse.gaussian(link = "log"), data=data1)</pre>
AIC(model.e.N)
## [1] 142.6993
AIC(model.e.G)
## [1] 121.5555
AIC(model.e.IG)
## [1] 128.6878
## MSE
mean(residuals(model.e.N, type="pearson")^2)
## [1] 63.19944
mean(residuals(model.e.G, type="pearson")^2)
## [1] 0.1345295
mean(residuals(model.e.IG, type="pearson")^2)
## [1] 0.018071
```

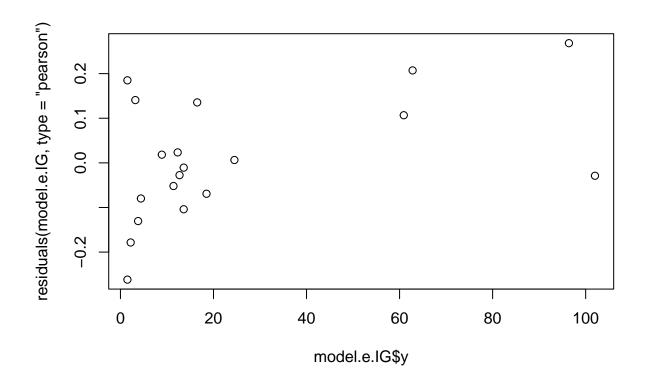
```
shapiro.test(residuals(model.e.N, type="pearson"))
##
##
   Shapiro-Wilk normality test
## data: residuals(model.e.N, type = "pearson")
## W = 0.83734, p-value = 0.004203
shapiro.test(residuals(model.e.G, type="pearson"))
##
   Shapiro-Wilk normality test
##
##
## data: residuals(model.e.G, type = "pearson")
## W = 0.93471, p-value = 0.2114
shapiro.test(residuals(model.e.IG, type="pearson"))
##
## Shapiro-Wilk normality test
##
## data: residuals(model.e.IG, type = "pearson")
## W = 0.98053, p-value = 0.9483
#plot(fitted(model.e.N, type="response"), residuals(model.e.N, type="pearson")^2)
#plot(fitted(model.e.G, type="response"), residuals(model.e.G, type="pearson")^2)
#plot(fitted(model.e.IG, type="response"), residuals(model.e.IG, type="pearson")^2)
plot(model.e.N$y, residuals(model.e.N, type="pearson"))
```



plot(model.e.G\$y, residuals(model.e.G, type="pearson"))



plot(model.e.IG\$y, residuals(model.e.IG, type="pearson"))



# Based on my analysis, I choose the inverse gaussian distrbution #as it yield the highest p for normality test for Pearson's residual. #IG distribution also have the lowest MSE

```
#Problem 2
```

data2<-read.table("/Users/phamkhoa/Documents/university/3/stats\_model\_1/w2/ratstime.txt", sep="\t", dec attach(data2)

```
#a
```

N.p2<-glm(time~factor(poison) + factor(treat), family = gaussian("identity"), data = data2)
G.p2<-glm(time~factor(poison) + factor(treat), family = Gamma("identity"), data = data2)
IG.p2<-glm(time~factor(poison) + factor(treat), family = inverse.gaussian("identity"), data = data2)
AIC(N.p2)</pre>

## [1] -33.21821

AIC(G.p2)

## [1] -62.23033

AIC(IG.p2)

## [1] -71.183

```
## MSE
mean(residuals(N.p2, type="response")^2)
## [1] 0.02189297
mean(residuals(G.p2, type="response")^2)
## [1] 0.02379278
mean(residuals(IG.p2, type="response")^2)
## [1] 0.02535005
shapiro.test(residuals(N.p2, type="pearson"))
##
    Shapiro-Wilk normality test
##
##
## data: residuals(N.p2, type = "pearson")
## W = 0.92242, p-value = 0.003622
shapiro.test(residuals(G.p2, type="pearson"))
##
##
    Shapiro-Wilk normality test
## data: residuals(G.p2, type = "pearson")
## W = 0.90006, p-value = 0.0006314
shapiro.test(residuals(IG.p2, type="pearson"))
##
    Shapiro-Wilk normality test
##
## data: residuals(IG.p2, type = "pearson")
## W = 0.90504, p-value = 0.0009185
#I choose the model N.p2 (normal distribution identity link)
#as it have the smallest MSE and highest p for shapiro test
p2b.identity<-glm(time~factor(poison) + factor(treat), family = Gamma("identity"), data = data2)
p2b.log<-glm(time~factor(poison) + factor(treat), family = Gamma("log"), data = data2)</pre>
p2b.inverse<-glm(time~factor(poison) + factor(treat), family = Gamma("inverse"), data = data2)
mean(residuals(p2b.identity, type="response")^2)
```

## [1] 0.02379278

```
mean(residuals(p2b.log, type="response")^2)
## [1] 0.01939997
mean(residuals(p2b.inverse, type="response")^2)
## [1] 0.01900666
# From the MSE value, choose inverse link with gramma distribution as it has the smallest MSE
ig.HO<-glm(time~factor(poison) + factor(treat), family = inverse.gaussian("log"), data = data2)
ig.H1<-glm(time~factor(poison)*factor(treat), family = inverse.gaussian("log"), data = data2)
anova(ig.H0, ig.H1, test = "F")
## Analysis of Deviance Table
## Model 1: time ~ factor(poison) + factor(treat)
## Model 2: time ~ factor(poison) * factor(treat)
## Resid. Df Resid. Dev Df Deviance
                                            F Pr(>F)
## 1
            42
                   4.6835
## 2
                   3.6418 6 1.0417 1.8462 0.1176
            36
\#Pr(>F) = 0.1176
newdata<-data.frame(poison = "II", treat = "B")</pre>
pred<-predict(p2b.log, newdata=newdata, type="response")</pre>
pred
##
## 0.7304192
xf < -cbind(c(1,1,0,1,0,0))
Var.Yf<-summary(p2b.log)$dispersion</pre>
D.f<-pred
Var.ef<-Var.Yf+(D.f^2)*t(xf)%*%vcov(p2b.log)%*%xf</pre>
lower.yf<-pred-qnorm(0.9)*sqrt(Var.ef)</pre>
upper.yf<-pred+qnorm(0.9)*sqrt(Var.ef)
lower.yf
             [,1]
## [1,] 0.4027708
```

```
#[1,] lower 0.4027708
upper.yf
             [,1]
## [1,] 1.058068
#[1,] upper 1.058068
newdata<-data.frame(poison = c("I","II"), treat = c("D","B"))</pre>
pred<-predict(p2b.log, newdata=newdata, type="response")</pre>
pred
##
## 0.7028960 0.7304192
x1 < -cbind(c(1,0,0,0,0,1))
x2 < -cbind(c(1,1,0,1,0,0))
Xf \leftarrow t(cbind(x1,x2))
k < -cbind(c(-1,1))
phi<-summary(p2b.log)$dispersion</pre>
Var.Y1f<-phi*(pred[1]^2)</pre>
Var.Y2f<-phi*(pred[2]^2)</pre>
D.f<-diag(pred)</pre>
Var.ef<-Var.Y1f+Var.Y2f+t(k)%*%D.f%*%Xf%*%vcov(p2b.log)%*%t(Xf)%*%D.f%*%k
lower.diff<-(pred[2]-pred[1])-qnorm(0.9)*sqrt(Var.ef)</pre>
upper.diff<-(pred[2]-pred[1])+qnorm(0.9)*sqrt(Var.ef)
lower.diff
##
               [.1]
## [1,] -0.3167109
upper.diff
              [,1]
## [1,] 0.3717572
#(-0.3167109, 0.3717572)
```