🐧 Day 1, PM practical - More advanced shell

To begin:

After logging into Hydra, navigate to the directory you created this morning at `/pool/cluster0/workshop/<username>`

Hint: Day 1 AM Practical Step 3

1. Using Core utilities

A. grep: globally search a regular expression and print

- `man grep` Everything you need to know.

- `grep 'EAS' sequences.fa`

- `grep 'Contig' sequences.fa`

- `grep 'contig' sequences.fa`

- grep -i : grep,case insensitive `grep -i 'contig' sequences.fa`

- grep -r: recursive grep `grep -r 'Contig' ./`

- You may also combine options, e.g.: `grep -ir 'conTIG' ./`

- grep -A: show lines after the grep hit `grep -i -A 1 'contig' sequences.fa` Useful if the sequences are in one line.

    - Also try:

        - `grep -B 10 '<searchterm>' <filename>` (10 lines Before)
        - `grep -C 5 '<searchterm>' <filename>` (5 lines of Context)
        - Combinations are possible: `grep -A 1 -B 2 '<searchterm>' <filename>` (1 line After, 2 lines Before the hit)

B. Output redirection: > (write to file)

- `grep 'Contig' sequences.fa > selection.fa` Creates a file,if it does not exists. Writes the output into the file. *Caution: existing content will be overwritten. Do not write to the same file that you read from.*

- `grep 'Contig' sequences.fa >> selection.fa` Creates a file, if it does not exists. Appends the output to the end of thef ile. Existing content is not overwritten. Rather, the file grows. (You should still not write to the same file that you read from.)

C. sed: stream editor

- Add taxa name to end of all header lines:

    `sed 's/>.*/& Escherichia coli/' sequences.fa > outfile_sed.fa`

    D. awk: 'awk' stands for the names of its authors "Aho, Weinberger, and Kernighan"

- Clean up a fasta file so only first column of the header is outputted:

    `awk '{print $1}' outfile_sed.fa > output_awk.fa`

2. Working with FASTA files

- Mnemonic: wc → word count. `-l` (for counting lines) is the most commonly used option. More information: `man wc` .

- **Exercise: sequence count**

  - Find a way to output the number of Sequences in a FASTA file. You may use 'temporary' files. Tip: individual sequences are identified by their headers. Header lines in a FASTA file begin with '>'.

    - **Possible solutions: sequence count** `man grep` `grep -c '>' sequences.fa`

      `grep '>' sequences.fa > tempfile.txt` `wc -l tempfile.txt`

- `ls -1 | wc -l` 'Pipes' the output of `ls -1` into the input of `wc -l` . Prints out the number of items in this directory. No temporary file necessary.

- `grep '>' sequences.fa | less` 'Pipes' the output of grep into the input of less. Allows to comfortably read the list of headers in a FASTA file. No temporary file necessary.

  - Displayed using less: Search using `/` Quit with `q`

- **Exercise: filter;** Output the first five headers in a FASTA file

  - Possible solution: `grep '>' sequences.fa | head -n 5`

- **Exercise: one specific line;** Output line number 19 of a given file.

  - Possible solution: `head -n 19 sequences.fa | tail -n 1`

- **Exercise: collecting sequences;** Save the first three and the last three headers from a FASTA file in a new file. This cannot be solved (easily) with only one command.

  - Possible solution: `grep '>' sequences.fa | head -n 3 > newfile.fa` and
    `grep '>' sequences.fa | tail -n 3 >> newfile.fa`

3.Managing Files

`tar -cvf new_tar_archive.tar <list>`

- Mnemonic: `tar` → tape archive
- `c` : create new archive
- `v` : verbose output
- `f` : file to use; can take files and directories as arguments

- **Exercise: create a tar archive;**

  - `tar -cvf sequences.tar sequences.fa`

Show the content of a tar archive: `tar -tvf new_tar_archive.tar`

Unpack a tar archive: `tar -xvf tar_archive.tar`

- **Exercise: unpack a tar archive;**

  - `tar -xvf sequences.tar`

- **Exercise: compress fasta file;**

  - `ls -lh sequences.fa`
  - `gzip sequences.fa`
  - `ls -lh sequences.fa.gz`

Compression reduces file size. Useful for copying over network, Mail attachments, storing on removable media, . . .

- **Uncompress:** `gzip -d sequences.fa.gz` Mnemonic: `d` → decompress

4. Transfer a file from /pool/cluster0/workshop/ to your local computer

`scp [options] <username1>@source_host:/directory1/filename1 <username2>@distination_host:/directory2/filename2`

- Name of the account on the host computer (username1)
- [options]

  - `-p` for preserve time info (can be very helpful to keep track of what is what)
  - `-r` copy directory

- Hostname of the computer on which the source file resides (source_host = hydra-login01)

- Name of the directory containing the source file (directory1)

- Filename of the source file (filename1)

- The location to which the source file will be copied is specified by username2@destination_host:directory2/filename2, which includes the:

  - Name of the account on the destination computer (username2)
  - Hostname of the computer to which the source file will be copied (destination_host)
  - Name of the directory to which the source file will be copied (directory2)
  - Filename of the copy (filename2)

- For more information: `man scp`

- **Exercise: compress and transfer to your local computer**

  - In hydra-3: `gzip sequences.fa`
  - Open a new terminal session/window
  - Confirm that you are on your local computer: `ls`
  - In local terminal:
    `scp -p <username>@hydra-login01:/pool/cluster0/workshop/<username>/Day1/data/sequences.fa.gz ./`
  - Confirm that the file has been transfered: `ls`

  Note: Windows users do not have a built-in scp program. Try Cyberduck, Filezilla...

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